Biosynthesis of porphyrins and related macrocycles. Part $47.^{1,2}$ Synthesis and chemistry of 2H-pyrroles (pyrrolenines) related to the proposed spiro-intermediate for porphyrin biosynthesis



Craig J. Hawker, W. Marshall Stark, Alan C. Spivey, Paul R. Raithby, Finian J. Leeper and Alan R. Battersby*

University Chemical Laboratory, Lensfield Road, Cambridge, UK CB2 1EW

It is proposed that the biosynthesis of uroporphyrinogen III 3, the parent precursor of the natural porphyrins, chlorins and corrins, involves a pyrrolenine 2 as a key intermediate, yet methods for the synthesis of such systems are not available. Novel routes for the synthesis of pyrrolenines by desulfurisation of unsaturated thiolactams have now been devised and the chemistry of such compounds has been explored. Enzymic experiments are carried out using a model pyrrolenine indicating that deletion of one of the pyrrole rings of the putative intermediate 2 leads to loss of tight binding.

Uroporphyrinogen III synthase (E.C. 4.2.1.75), usually called cosynthetase, catalyses the conversion of hydroxymethylbilane 1 into uroporphyrinogen III 3, shortened to uro'gen III. This macrocycle is the parent from which haem, chlorophyll and vitamin B₁₂ are all biosynthesised and much interest has centred on the mechanism of its formation. Intriguingly, the cyclisation of hydroxymethylbilane 1 to produce uro'gen III 3 is accompanied by a rearrangement which leads to inversion of ring D in the product 3. Extensive earlier studies have been made both of the biosynthesis of hydroxymethylbilane 1 and of its ringclosure by cosynthetase; this work has been reviewed.³ Of central importance for the present paper was the discovery 4,5 that the inversion of ring D occurs by an intramolecular mechanism that only involves ring D of the bilane 1. An attractive sequence for this inversion involves the spiro-pyrrolenine 2 as a key intermediate, Scheme 1. Such pyrrolylmethylpyrrolenines are known⁶ to undergo the indicated fragmentation and this step

Scheme 1 Mechanism proposed for the production of uro'gen III by cosynthetase; some ¹³C-labelling experiments which established the intramolecular rearrangement ⁴ are illustrated by the black spots and squares

 $A = CH_2CO_2H$, $P = CH_2CH_2CO_2H$

could be followed by formation of a new carbon–carbon bond as illustrated. This proposed mechanism is based on a suggestion by Mathewson and Corwin,⁷ as shown in Scheme 1 in a simplified form.

The intermediacy of the spiro-pyrrolenine 2 for the biosynthesis of uro'gen III 3 was given strong support by synthesis of the racemic spirolactam 4 and the finding that it

 $A^R = CH_2CO_2R, \ P^R = CH_2CH_2CO_2R$

inhibits strongly the action of cosynthetase in cyclising hydroxymethylbilane 1 to uro'gen III 3. This support was further strengthened by preparing both enantiomers of the spirolactam 4 and demonstrating that one enantiomer competitively inhibits cosynthetase over twenty times more strongly than the other. Finally, the strongly inhibiting enantiomer of 4 has been shown 10 to have the R-configuration and therefore, if the spiro-intermediate 2 is in fact formed in the enzymic process, the evidence points to its absolute configuration also being R, as illustrated in Scheme 1.

A synthesis of the spiro-pyrrolenine 2 itself, even as a racemate, would allow a decisive test of the mechanism in Scheme 1, since cosynthetase should convert one enantiomer of the synthetic product into uro'gen III 3 without formation of any significant amount of uro'gen I 6. In contrast, non-enzymic rearrangement of 2 would be expected to yield a mixture of the type III 3 and type I 6 isomers (see the second in this series of four papers 11). An earlier paper 6 described our first experiments aimed at developing chemistry necessary for the synthesis of 2. The central problem was to devise routes to what were expected to be rather labile pyrrolylmethylpyrrolenines and bis(pyrrolylmethyl)pyrrolenines exemplified by the ring A-ring D-ring C system of 2. In that earlier paper 6 the pyrrolenine ring of a model pyrrolenine was built up by conjugate addition of a nitro alkane to an acetylenic ester, followed

by reduction of the ester and nitro groups and cyclisation. This route does not permit substituents at C-4 of the pyrrolenine. Although a methyl group at C-4 was introduced in a roundabout fashion by alkylation of an enolate, such chemistry would clearly be incompatible with the ester side-chains required for the synthesis of 2. Furthermore, we were keen to exploit the coupling of iodopyrroles with acetoxymethylpyrroles to give 2-(pyrrolylmethyl)-5-halopyrrolenines (e.g. 11 + 12→13, Scheme 3), which has been so successful in the formation of mono- and bis-(pyrrolylmethyl)lactams, culminating in the synthesis of spirolactam 4.8 The present paper describes how these 2,2,3,4-tetra-substituted pyrrolenines can be constructed and also some studies of their chemistry.

Results and discussion

Exploration of radical and other methods to reduce 5-substituted pyrrolenines

Our first approach to 5-unsubstituted mono- and bis-(pyrrolylmethyl)pyrrolenines 10, Scheme 2, was by reduction of

Scheme 2

5-substituted pyrrolenines 8 (X = I or PhSe), derived from lactams 7. It was hoped that generation of radicals 9 with quenching by hydrogen abstraction might afford the desired pyrrolenines.

When the reaction of acetoxymethylpyrrole 11 with iodopyrrole 12 is catalysed by stannic chloride,8 the product is mainly the chloropyrrolenine 13 with lesser amounts of the iodide 14, Scheme 3. The iodo analogue 14 was the preferred

Scheme 3 Reagents: i, SnCl₄; ii, BF₃; iii, AgOAc, H₃O⁺ or SmI₂, THF, H₂O; iv, COCl₂ or triphosgene; v, PhSeH (or PhSH); vi, Ph₃SnH, AIBN (or hv)

material for radical generation yet under none of the wide variety of conditions tested was it possible to displace chloride from 13 by iodide. The rather unstable iodopyrrolenine 14

was obtained, albeit in 19% yield, by changing the Lewis acid catalyst for the reaction of 11 with 12 to boron trifluoride. Sufficient iodo derivative 14 was obtained to allow it to be photolysed in the presence of triphenyltin hydride but the major product, 59%, was the pyrrole 27¹² together with a small amount of the α -free pyrrole 13 25. These arise either by the cationic fragmentation pathway (e.g. see Scheme 8; also fully exemplified in the following paper 11), with reduction of the intermediate azafulvene by triphenyltin hydride, or by an analogous process involving radicals. Essentially the same result was obtained when the iodo derivative 14 was heated with triphenyltin hydride and a catalytic amount of 2,2'-azobisisobutyronitrile (AIBN).

The phenylseleno group has been widely used for the generation of radicals 14 and so attention now focused on the selenopyrrolenine 16, Scheme 3. Despite the lack of reactivity of chloropyrrolenine 13 towards iodide ion, we expected that displacement of chloride by selenophenol or its anion might be successful. It was also important to develop a way to prepare chloropyrrolenines e.g. 13, from lactams, e.g. 21, since the final application of any reductive method would be to the spirolactam ester 5.

Lactam 21 had been synthesised earlier⁸ from pyrroles 11 and 12, as in Scheme 3. An improved procedure described for a close relative 10 has now doubled the yield of 21 from the earlier ca. 30% to 62%. The best conversion of lactam 21 into the chloropyrrolenine 13 was by treatment with phosgene 15 but importantly with 4-dimethylaminopyridine (DMAP) in place of the originally used pyridine; use of triphosgene rather than phosgene gave identical results and was experimentally preferable. The reaction was not straightforward in that three compounds were formed in differing amounts depending on the conditions. All three materials were unstable but could be separated under argon for spectroscopic characterisation. The rapidly formed kinetic product appeared to be the enolised N-chloroformyl lactam 24, the second product was thought to be the O-chloroformyl derivative 15, and the chloropyrrolenine 13 only predominated after heating at reflux in CH₂Cl₂. The presence of an N-chloroformyl species was supported by the results of attempted reductions of the crude reaction mixture with ZnBH₄; reaction for 3 h followed by aqueous work-up gave the N-methyl lactam 22, 44%, whereas reaction for 12 h followed by treatment with acetic acid gave the N-methyl dihydropyrrole 23, 25%.

Treatment of chloropyrrolenine 13 (prepared from lactam 21 using phosgene) with selenophenol (prepared by reduction of diphenyl diselenide)¹⁶ or with thiophenol gave the phenylselenopyrrolenine 16, 92%, and the sulfur analogue 17, 81%, respectively. Interestingly, these reactions occurred more rapidly and were higher yielding if no base (triethylamine) was added, presumably because the initial addition step is catalysed by protonation of the pyrrolenine nitrogen by the two phenols under the non-basic conditions.

Irradiation of the phenylseleno compound 16 in benzene with triphenyltin hydride or heating it at reflux in benzene with AIBN and either triphenyltin hydride or tributyltin hydride gave the same two products, pyrroles 26 and 27, in good yield. Again these had been formed by fragmentation followed by reduction.

Strictly analogous steps to those just described were also carried out in the dipyrrolic series on lactam 288 (Scheme 4). The chloropyrrolenine 30 was derived from it using phosgene and reacted with selenophenol to give the phenyl selenide 31 in low yield (34%) because the now familiar fragmentation also occurred. One of the products of this competing process was the phenylselenomethylpyrrole ¹⁷ 33 but the analogue 34 that would arise by the alternative fragmentation was not formed, no doubt due to the significant electron-withdrawing effect of the tribromoethyl group.

The same chemistry was also performed on lactam 29 which

Scheme 4 Reagents: i, COCl₂; ii, PhSeH

only differs from isomer 35 synthesised earlier 8 (Scheme 5) in the arrangement of the side-chains on one pyrrole ring. This change made the synthesis, described in the following paper, 11 easier and more reproducible than the earlier one. The phenyl-selenopyrrolenine 32 was prepared from lactam 29 in the usual way but without isolation of the chloro intermediate. Similar studies using tin hydrides to those described above for the monopyrrolic series were also carried out briefly with the dipyrrolic system 32 but here too, appreciable fragmentation occurred under all conditions tested, so this work is not further described.

The conclusion from these experiments was that either the desired radical is formed but the molecule then undergoes rapid homolytic fragmentation or that heterolytic fragmentation dominates the chemistry under the conditions used.

Reduction of the monopyrrolic chloropyrrolenine 13 under many conditions with a wide variety of hydride and other reducing agents was also attempted. These experiments usually led either to recovery of starting material or only to reduction of the side-chain ester(s). A few reactions did occur, however. Thus treatment of the halopyrrolenine mixture 13/14 with samarium(II) iodide and water gave the lactam 21, Scheme 3, whereas treatment with lithium triethylborohydride resulted in the formation of a boronic acid complex of 21.

We then aimed to replace the chlorine atom of 13 by another group for further reductive experiments. It proved impossible to displace the chlorine using methoxide or cyanide ions but the methoxy derivative 18 was available by methylation of lactam 21 with either trimethyloxonium tetrafluoroborate (Meerwein's reagent), in 83% yield, or methyl iodide and silver carbonate, 75% yield. Similarly, p-nitrobenzyl bromide or p-bromobenzyl bromide together with silver carbonate and 18-crown-6 afforded the imino ethers 19 and 20, respectively, in yields of 71 and 62%. Earlier work 6 had shown that a close analogue of 18 lacking the acetate and propionate side chains could be smoothly reduced with DIBAL but the use of this reducing agent is precluded in the case of 18 by the presence of the ester groups. Many other reducing agents were tested but again they either left 18 unchanged or just reduced one or more of the side-chain esters. The only exception was the high-yielding conversion of 18 into the original lactam 21 by samarium(II) iodide, with or without tributyltin hydride.

Exploration of the thionation route

The obvious and apparently simple way to prepare the spiropyrrolenine 2 would be to reduce the spiro-lactam ester 5, followed by a final hydrolysis step. Clearly the reduction of the lactam would have to be effected in the presence of the eight more reactive ester groups and therefore some means of activation of the lactam would be required. We thought that conversion of a lactam into the corresponding thiolactam might allow selective reduction using Raney nickel ¹⁸ or some similar reagent. Alternatively alkylation of the sulfur atom might be used to increase the reactivity of the thiolactam towards different reducing agents. The pyrrole rings of spiro-lactam 5, however, lack any electron-withdrawing ester groups to deactivate them and it was necessary to test out the required chemistry on simpler model pyrrolylmethyllactams having similarly electronrich pyrrole rings.

We therefore explored thionation of bis(pyrrolylmethyl)-lactams having non-deactivated α -free pyrrole group(s). Hydrogenolysis of the previously synthesised ⁸ lactam dibenzyl ester 35 gave the diacid 37 which was decarboxylated using trifluoroacetic acid (TFA), Scheme 5. The product 38 was then treated

Scheme 5 Reagents: i, 43; ii, Pd/C, H₂; iii, TFA; iv, 44; v, Zn, AcOH

with Lawesson's reagent ¹⁹ **43** in tetrahydrofuran, conditions that had been found to allow relatively mild conversion of **35** into the thiolactam **36**. However, the bis- α -free system **38** did not react and even more forcing conditions left it largely unaffected; the thiolactam **39** was not produced. This may be due to association of the two electron-rich pyrrole rings with the lactam residue by π - π interaction and hydrogen bonding, a phenomenon previously observed.⁶

One final hope for this approach was to use Davy's reagent 44 for thionation 20 since it is milder than Lawesson's reagent 43. Indeed, this reagent converted the closely related bis(pyrrolylmethyl)lactam 28 into thiolactam 40 at room temperature in 94% yield. Because of the foregoing results with 38, just one α -free pyrrolylmethyl group was generated by removal of the tribromoethyl group from 28 using zinc and acetic acid and then decarboxylation of the resultant acid using TFA. The product 41 was treated directly with 44 under the same conditions as before; 42 was not formed and there was essentially complete decomposition of the material. Thus, on the one hand, there was insufficient reactivity for the system 38 with two α -free pyrroles whereas, on the other hand, there was destructive lability for the analogue 41 with one α -free pyrrole.

Synthesis of pyrrolenines

Because of the failure to thionate pyrrolylmethyl lactams having a non-deactivated pyrrole ring, we changed to studying thionation of systems having deactivated pyrroles, with the thought that it might be possible to remove or use the deactivating groups at a later stage and further steps would then allow formation of the macrocycle. Our initial studies were performed on the more readily available monopyrrolic lactams before we progressed to dipyrrolic systems.

Lactam 21 was smoothly converted into the thiolactam 46 using Lawesson's reagent (Scheme 6), which clearly demonstrates the difference that the deactivating ester groups can make. An alternative procedure for the preparation of this thiolactam was found to be treatment of halopyrrolenine mixture 13/14 with H₂S. Desulfurisation of thiolactam 46 with Raney

Scheme 6 Reagents: i, 43; ii, H₂S; iii, TMOF, TFA; iv, Raney nickel; v, nickel boride

nickel, or more reproducibly with nickel boride,²¹ then yielded the desired pyrrolenine **49**, 45%. Alternatively, methylation of the thiolactam with trimethyl orthoformate (TMOF) and trifluoroacetic acid (TFA) afforded the thioimino ether **48**, 95%, which was desulfurised using Raney nickel yielding the pyrrolenine **49**, 41%. When nickel boride replaced Raney nickel in the foregoing experiment, the yield of **49** was only 10–16% and a second product proved to be the amine **47**. The latter became the sole product when either **46** or **48** was treated with nickel boride for 15–24 h, yields 51 and 37%, respectively.

The pyrrolenine **49** was reasonably stable if handled carefully but it decomposed completely after storage for one month at 0 °C under argon. It was thought that an adduct of the pyrrolenine with a nucleophile might produce a more stable compound, amenable to further synthetic steps. However, neither thiols, selenophenol nor cyanide could be added to the imine residue of pyrrolenine **49** by any of the many methods tested, including *e.g.* trimethylsilyl cyanide–zinc iodide.²² Clearly the 5-position of these 2,2,3,4-substituted pyrrolenines is strongly hindered.

In the dipyrrolic series, the synthesis of pyrrolenines was just as successful as in the monopyrrolic series. Reduction of thiolactam **36** with nickel boride gave pyrrolenine **51** (46%). Similar reduction of the isomeric thiolactam **50** to give the corresponding pyrrolenine **52** (52%) is reported in the following paper.¹¹

Chemistry of thiolactams

Having established an effective route to pyrrolenines such as 49 and 51 from the corresponding thiolactams, we wished to explore some of the chemistry that would be necessary if a thiolactam similar to 51 were to be converted into the thiolactam corresponding to spirolactam 5. For this to be possible it would be necessary to make a bis(pyrrolylmethyl)thiolactam with a pyrrolic α -formyl group to allow attachment of the third pyrrole ring. The chemistry was first explored in the mono-(pyrrolylmethyl) series.

The initial approach was to make the thiolactam first and then cleave the pyrrolic α -ester group, decarboxylate and formylate. This demanded a different ester, as it would probably not be possible to hydrogenolyse a benzyl group in a sulfur-containing molecule. Because tribromoethyl ester 53 was available, lactam tribromoethyl ester 54, with a reversed substitution pattern on the pyrrolic ring, was synthesised as before, Scheme 7. It was converted into the thiolactam 55 and the ester was cleaved with zinc and acetic acid to yield the thiolactam

Scheme 7 Reagents: i, SnCl₄ then AgOAc, H₃O⁺; ii, 43; iii, Zn, AcOH; iv, TFA, TMOF; v, TFA; vi, DMF, POCl₃; vii, TsOH; viii, KI₃, (NH₂)₂C=S then KOH, MeI

acid **56**. Unexpectedly, this underwent fragmentation under standard decarboxylation–formylation conditions, TMOF and TFA, to afford the monopyrrole **59** in 75% yield and the same product was also formed under Vilsmeier conditions (Me₂-NCHO, POCl₃). Thiolactam **56** was also broken down by TFA alone, the major product being pyrrole **25**. The two pyrroles **25** and **59** were identified by comparison with authentic samples prepared by published methods.^{13,23} A rationalisation of the outcome of these reactions is shown in Scheme 8; acid-

Scheme 8 Proposed mechanism for the formation of pyrroles 25 and 59 from thiolactam 56

catalysed decarboxylation of **56** is probably the first step, generating an undeactivated pyrrole, which upon protonation of the sulfur atom would fragment as shown. A similar fragmentation by a related mechanism occurred when the thioimino ether **58**, prepared from thiolactam **55** by methylation followed by reductive ester cleavage, was treated with toluene-*p*-sulfonic acid. The sole product, apart from polymeric material, was the methylthiopyrrole **62**, 49%, identified by comparison with authentic material synthesised ²⁴ from the acid **61**, ²⁵ Scheme 7.

Surprisingly, treatment of acid **63**, prepared by hydro-

Surprisingly, treatment of acid 63, prepared by hydrogenolysis of imino ether 18, with either TFA or toluene-p-sulfonic acid gave the α -free pyrrole 64 in up to 87% yield whereas only fragmentation was observed with the sulfur analogue 58. This result is somewhat puzzling but does fit in

with the greater stability of pyrrolylmethyl lactams compared with the corresponding thiolactams.

Concurrently with the above studies in the monopyrrolic series, analogous experiments were also tried in the dipyrrolic series. Thus treatment of lactam 65, synthesised earlier, with Lawesson's reagent afforded the thiolactam 66, from which the tribromoethyl group was reductively removed, Scheme 9. The

Scheme 9 Reagents: i, 43; ii, Zn, AcOH; iii, TsOH; iv, PtO₂, H₂

resultant acid 67, on treatment with acid, underwent fragmentation as in the simpler mono-pyrrolyl case, to yield the dipyrromethane 68. The mechanism of its formation is clearly analogous to that shown in Scheme 8; 68 was identified by comparison with a standard sample prepared from iodide 69 11 by hydrogenolysis.

It was clear from the foregoing experiments that, due to the lability under acidic conditions of pyrrolylmethyl thiolactams having an undeactivated pyrrole ring, it would not be possible to set up the thiolactam system early in the synthesis and carry it forward through the remaining steps necessary ⁸ to generate the spirothiolactam ester **70**, Scheme 10.

Scheme 10 Proposed mechanism for the formation of uroporphyrin octamethyl esters upon thionation of spirolactam ester 5

The alternative to early formation of the thiolactam system was to attempt the transformation of the spirolactam ester 5 into the thiolactam 70 at the end of the synthesis; all the experiments were carried out with racemic material. Naturally this conversion was tried as soon as the thiolactam route to pyrrolenines was developed, indeed before some of the studies described above, but without success. All the experience gained from the subsequent work led us to return to this desired con-

version. For example, if the explanation offered for the lack of reactivity of the bis- α -free pyrrole system 38 is correct (i.e. steric hindrance from the pyrrole rings), then the spirolactam 5 should not be similarly affected because the pyrrole rings are tied back away from the lactam. The foregoing results with the mono-α-free pyrrole model 41 and the acid 56 forewarned of possible problems but even the smallest chance for the preparation of the spirothiolactam 70 could not be ignored. Accordingly, the spirolactam ester 5 was heated with Lawesson's reagent or with phosphorus pentasulfide. However the only products, in each case after aerial oxidation, were uroporphyrin octamethyl esters, 48% yield, shown by HPLC and ¹H NMR spectroscopy to contain the statistical mixture of the four possible isomers, *i.e.* type III **72**, 50%, type I, 12.5%, type II, 12.5% and type IV, 25%. Presumably these arise, Scheme 10, by fragmentation analogous to that shown in Scheme 8 followed by recyclisation and loss of sulfur (also similar to that in Scheme 8) to give uro'gen esters of both type I and type III (only the type III one 71 is shown). Then scrambling of the pyrrole rings at the uro'gen stage under the acidic conditions leads to the statistical mixture of isomers.²⁶ Davy's reagent 44 at room temperature only caused slow decomposition of 5, and raising the temperature simply increased the rate of decomposition. The p-tolyl analogue 27 45 left the lactam 5 unchanged up to 50 °C and decomposition set in at 60 °C. At roughly the half-way point of the last experiment, it was shown by ¹H NMR spectroscopy that the undecomposed substance was unchanged starting material.

Thus we were reluctantly forced to conclude that the route to pyrrolylmethylpyrrolenines from lactams by thionation followed by reduction, so successful when the pyrrole rings are deactivated, cannot be used for synthesis of the spiropyrrolenine 2 itself.

Synthesis of α-formyl- and α-cyano-pyrrolic pyrrolenines

Finally, in view of the fact that the spiro-pyrrolenine 2 itself was not yet synthetically accessible, we wanted to make alternative pyrrolenines in which the necessary electron-withdrawing groups on the pyrrolic ring(s) are smaller than the benzyloxy-carbonyl residue of 49 and 51. This was because, in addition to studying the chemistry of pyrrolenines, a further aim was to explore how they interacted with the enzyme, cosynthetase. Formyl and cyano groups were chosen as being suitable for the envisaged enzymic experiments.

As in previous cases, we initially investigated the required chemistry in the monopyrrolic series. In view of the fragmentation that occurred upon attempted decarboxylation–formylation of thiolactam acid **56**, we investigated the alternative sequence of reactions, *i.e.* formylation first then thionation. Hydrogenolysis of lactam benzyl ester **21** gave the acid **73** which was treated with TMOF and TFA to afford the aldehyde **74** (Scheme 11). However, thionation using Lawesson's reagent converted this into a mixture of the lactam thioaldehyde **75** and the corresponding thiolactam thioaldehyde **79**. It was clear that the monothio product was thioaldehyde **75** and not the desired thiolactam **80** from the ¹H NMR chemical shift of the aldehyde proton, which had moved to δ 10.57 from 9.51 in the starting aldehyde **74**.

Since thioketones can be hydrolysed to give ketones,²⁸ we hoped to hydrolyse the bis-thio system **79** to the thiolactam aldehyde **80**. Trial experiments were carried out by converting the aldehyde **59** into the thioaldehyde **60** using Lawesson's reagent, Scheme 7. This thioaldehyde **60** crystallised as red rhombohedra and its structure, shown in Fig. 1, was determined by X-ray analysis.† It is monomeric in contrast to the case

[†] The experimental details and coordinates for the crystal structure determinations of **60** and **77** have been deposited with the Cambridge Crystallographic Data Centre (CCDC). Deposition numbers: **60**, 100546; **77**, 100545.

$$\begin{array}{c} C(13) \\ C(13) \\ C(12) \\ C(12) \\ C(10) \\ C(2) \\ C(3) \\ C(11) \\ C(4) \\ C(5) \\ C(6) \\ S(1) \\ \end{array}$$

Fig. 1 X-Ray crystal structure of thioaldehyde 60

Scheme 11 Reagents: i, Pd/C, H₂; ii, TFA, TMOF; iii, NH₂OH; iv, Ac₂O; v, 43; vi, DMF, POCl₃; vii, morpholine, H₂O; viii, Raney nickel; ix, nickel boride; x, KOH

 $A^{R} = CH_{2}CO_{2}R, P^{R} = CH_{2}CH_{2}CO_{2}R$

studied by Baker²⁹ which existed as the trimer 84. Presumably the greater electron release from the pyrrole ring to the thioaldehyde residue in the case of 60 accounts for its greater stability.

Thioaldehyde 60 could be hydrolysed in aqueous morpholine to yield the starting aldehyde 59. These conditions were then applied to the thiolactam thioaldehyde 79 and gave the thiolactam aldehyde 80. Desulfurisation by Raney nickel then gave the desired formylpyrrolenine 82.

The second target in the monopyrrolic series was the nitrile 83, Scheme 11. Treatment of aldehyde 74 with hydroxylamine yielded a mixture of syn- and anti-oximes 76 and subsequent reaction with hot acetic anhydride then gave a mixture of

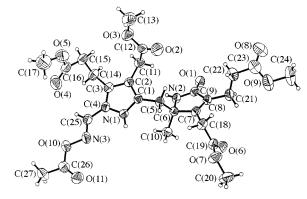


Fig. 2 X-Ray crystal structure of anti-O-acetyloxime 77

products, some of them inseparable. However, mass spectrometry showed that the mixture contained the nitrile 78 and the acetylated oxime 77. The latter was obtained pure by fractional crystallisation and an X-ray structure determination was carried out.† Fig. 2 shows that this product is the anti-isomer, which is understandable since the syn-isomer presumably undergoes rapid elimination of acetic acid to give the nitrile. This structure fully confirms all the features of this molecule and indeed of the entire monopyrrolic series, previously dependent on spectroscopic evidence. An alternative method for dehydration of oximes 30 (Me2NCHO and POCl3) smoothly converted the mixture of isomeric oximes 76 into the nitrile 78 in good yield. Preparation of the corresponding thiolactam 81, as earlier, and desulfurisation with nickel boride then readily afforded the cyanopyrrolenine 83.

Turning to the dipyrrolic series, hydrogenolysis of the benzyl groups of lactam 29 gave the diacid 85, which with TMOF and TFA was converted into the required dialdehyde 86. However, it was not possible to prepare the corresponding thiolactam dithioaldehyde cleanly using Lawesson's reagent and attempted hydrolysis (as in the monopyrrolic series) of the product mixture aiming to produce some of the thiolactam dialdehyde was unsuccessful. Attention therefore turned to the dinitrile 87 which was synthesised smoothly from dialdehyde 86 by forming the mixture of oximes followed by dehydration as earlier. Further, the lactam dinitrile 87 was converted into the thiolactam 88 by Lawesson's reagent ready for desulfurisation with nickel boride. Unlike the monopyrrolic case 81→83, desulfurisation generated a mixture and caused serious loss of material. In order to obtain pure pyrrolenine 89 for the enzymic experiments, purification by HPLC was needed and we had to accept a yield of 14%.

Enzymic studies

The plan was to hydrolyse the six ester groups of the dicyanopyrrolenine 89 under conditions that left the cyano and pyrrolenine systems unaffected. These conditions were developed using the simpler monopyrrolic analogue 83, Scheme 11, which on treatment with aqueous methanolic potassium hydroxide gave a homogeneous solution. Part of this solution was repeatedly evaporated and redissolved in deuterium oxide; the final solution gave a 400 MHz ¹H NMR spectrum fully consistent with the tetrapotassium salt derived from 83. The remainder of the hydrolysate was re-esterified using diazomethane to give almost pure starting material 83, together with trace amounts of higher $R_{\rm f}$ fragmentation products. Thus, the hydrolytic conditions met the requirements and were used to prepare the hexapotassium salt of the acid 90 from the ester 89.

Hart studied the effect of the hexaacid 90 in buffered solution on the rate of conversion of synthetic hydroxymethylbilane 12 1 into uro'gen III 3 by purified cosynthetase31 using kinetic experiments exactly as had been used 8 to demonstrate strong inhibition of the enzyme by the spirolactam 4. The outcome was that, even at high concentrations (500 μmol dm⁻³), the acid

90 had no effect on the enzymic rate and is thus not an inhibitor of cosynthetase. This interlocks with earlier results which showed that molecules lacking some part or all of the lactam ring present in the inhibitory spirolactam 4 did not inhibit the enzyme. The present results suggest either that the entire macrocyclic system is also necessary for strong binding to occur or that 90 cannot readily adopt the same conformation as that of rings A, D and C of 4.

Conclusions

The probable involvement of a 2,2-disubstituted pyrrolenine (2*H*-pyrrole) in the biosynthesis of uro'gen III 3, the parent of all the natural porphyrins, chlorins and corrins, has focused attention on the chemistry of such systems. However, methods for synthesis of close analogues of the proposed natural system were not available. The novel methods described in this paper have provided a variety of the required pyrrolenines carrying one or two pyrrolylmethyl groups at the disubstituted 2-position of the 3,4-substituted pyrrolenine ring. This has allowed extensive study of the chemistry of these systems and also of how one example interacted with the enzyme cosynthetase. It was found that whereas the spirolactam 4 strongly inhibited cosynthetase, the open-chain pyrrolenine 90, lacking ring B of 4, did not.

Experimental

General directions

General directions are as in ref. 32. Coupling constants, *J*, are quoted in Hz. For ¹³C NMR spectra in which the numbers of hydrogens attached to carbons were determined, this was achieved by heteronuclear *J*-resolved 2D spectra, off-resonance decoupling or DEPT experiments.

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10H)-one 21

A mixture of a solution of 3-(2-methoxycarbonylethyl)-4 $methoxy carbonyl methyl-5-methyl pyrrole-2-carboxylic \quad acid ^{25}$ (25, $X = CO_2H$) (200 mg, 0.7 mmol) in dichloromethane (10 cm³) and a solution of sodium hydrogen carbonate (119 mg, 1.4 mmol) in water (8 cm³) was stirred vigorously under argon and an aqueous solution (7.1 cm³) of iodine (0.1 mol dm⁻³) and potassium iodide (0.2 mol dm⁻³) was added over 5 min. The resulting mixture was stirred for a further 2 min and then solid sodium metabisulfite was added to destroy the excess iodine. The organic layer was separated and the aqueous layer was extracted with dichloromethane $(3 \times 15 \text{ cm}^3)$. The combined organic layers were dried and evaporated under reduced pressure to give the crude iodopyrrole 12. To this was added a solution of acetoxymethylpyrrole 11³³ (305 mg, 0.7 mmol) in anhydrous dichloromethane (10 cm³) and the resulting solution was cooled to -78 °C under argon. Stannic chloride (83 mm³, 0.7 mmol) was added dropwise and the stirring continued for 3 h, during which time the solution was allowed to warm up to 0 °C. The solution was then re-cooled to -78 °C and a solution of samarium(II) iodide in THF (0.1 mol dm⁻³; 14 cm³, 1.4 mmol) was added, followed by water (100 mm³). The solution was allowed to warm to room temperature over 1 h and then evaporated under reduced pressure. Purification by PLC, eluting with ethyl acetate—hexane (1:1) and then ethyl acetate, gave lactam 21 as an oil (238 mg, 54%), with physical characteristics identical to those reported.8

9-Benzyloxycarbonyl-1-chloro-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin

A suspension of triphosgene (50 mg, 160 μ mol), DMAP (920 mg, 160 μ mol) and lactam **21** (50 mg, 80 μ mol) in anhydrous

dichloromethane (2 cm³) was heated at reflux under argon for 1 h. The mixture was *rapidly* cooled in ice, filtered through a plug of Celite and evaporated under reduced pressure to give the crude 5-*chloropyrrolenine* 13 as an oil (52 mg), which was generally used without further purification.

TLC of this material (diethyl ether-methanol, 20:1) showed that it contained, in addition to 13, two minor components. Their relative proportions were roughly equal after equilibration at room temperature for a few hours but strongly favoured 13 after heating at reflux. The three very air- and moisture-sensitive components were separated by PLC, eluting with diethyl ether-methanol (20:1) in the dark under argon, to give: (i) a compound thought to be O-chloroformylimidate 15 $(R_f 0.6)$; $\delta_H(CDCl_3, 400 \text{ MHz}) 1.61 (3 \text{ H, s, 4-Me}), 2.44–2.94$ $(8 \text{ H}, 2 \times \text{CH}_2\text{CH}_2), 2.79 \text{ and } 3.41 \text{ (each } 1 \text{ H}, d, J 16, 5-H_2), 3.44$ (2 H, s, CH₂CO₂), 3.53 and 3.84 (each 1 H, d, J 18, CH₂CO₂), 3.59, 3.62, 3.63 and 3.77 (each 3 H, s, OMe), 5.17 and 5.31 (each 1 H, d, J 12.5, CH₂Ph), 7.25–7.40 (5 H, m, Ph) and 10.02 (1 H, br s, NH); (ii) chloropyrrolenine 13 (R_f 0.8); ¹H NMR data identical to those previously reported; (iii) a compound thought to be enolised N-chloroformyl lactam 24 $(R_f \ 0.9)$; $\lambda_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{nm}$ 277 and 332; $\nu_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$ 3443, 3338, 3029, 2954, 1734, 1583, 1453 and 1175; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 2.00 (3 H, s, 4-Me), 1.93-2.01 (1 H, m), 2.10-2.17 (1 H, m), 2.33-2.52 (5 H, m) and 2.86 (2 H, t, J 8, 2 × CH₂CH₂CO₂ and OH), 3.38 (2 H, s, CH₂CO₂), 3.58, 3.62, 3.64 and 3.73 (each 3 H, s, OMe), 3.73 and 4.12 (each 1 H, d, J 15.5, 5-H₂), 5.21 and 5.30 (each 1 H, d, J 12.5, CH₂Ph), 5.59 (1 H, s, C=CHCO₂), 7.33-7.36 (5 H, m, Ph) and 8.61 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100$ MHz) 19.4 and 20.7 (2 × $CH_2CH_2CO_2$), 22.1 (4-Me), 29.3, 30.6, 31.6 and 34.6 (3 × CH_2CO_2 and C-5), 51.5, 51.9, 52.0 and 52.0 (OMe), 66.0 (CH₂Ph), 78.5 (C-4), 105.7 (C=CHCO₂), 116.6, 118.3, 125.0, 129.4, 130.5, 136.4, 136.5 and 142.5 (C=C), 128.3 and 128.7 (C=CH) and 158.7, 160.1, 166.4, 172.0, 172.4 and 173.5 (C=O); m/z (+FAB) 689 (MH⁺, 4%) and 372 (C₂₀H₂₂NO₆, 100).

9-Benzyloxycarbonyl-1-iodo-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin

A solution of acetoxymethylpyrrole 11^{33} (75 mg, 175 µmol) and iodopyrrole 12^{8} (64 mg, 175 µmol) in dry dichloromethane (3 cm³) was stirred with boron trifluoride–diethyl ether (50 mg, 175 µmol) at 0 °C under argon for 1 h. Saturated aqueous sodium hydrogen carbonate (3 cm³) was added and then, after 5 min, the organic layer was separated and the aqueous layer extracted with dichloromethane (3 × 5 cm³). The combined organic layers were dried and evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether, gave *iodopyrrolenine* 14 (19 mg, 13%) as a moisture-sensitive gum; physical data were as previously 8 described; m/z (FD) 736 (M⁺, 100%).

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis-(methoxycarbonylmethyl)-4-methyl-1-phenylseleno-4,5-dihydrodipyrrin 16

A solution of chloropyrrolenine **13** (400 mg, 0.62 mmol) in dry degassed dichloromethane (10 cm³) was added under argon to selenophenol ¹⁶ (471 mg, 3.0 mmol), stirred under argon for 30 min, poured into saturated aqueous sodium carbonate (50 cm³) and extracted with dichloromethane (4 × 25 cm³). The combined extracts were dried and evaporated under reduced pressure. Purification by column chromatography, eluting with diethyl ether, gave *phenylselenopyrrolenine* **16** (452 mg, 95%) as a gum (Found: M⁺, 766.2015. $C_{38}H_{42}N_2O_{10}^{80}Se$ requires M, 766.2005); $v_{max}(CH_2Cl_2)/cm^{-1}$ 3350, 1720, 1700, 1160 and 1070; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 1.10 (3 H, s, 4-Me), 2.33 and 3.09 (each 1 H, d, J 15, 5-H₂), 2.47–2.58 and 2.92–3.01 (8 H, m, 2 × CH_2CH_2), 3.30 and 3.38 (each 1 H, d, J 16, CH_2CO_2), 3.43 (2 H, s, CH_2CO_2), 3.59, 3.62, 3.65 and 3.69 (each 3 H, s, OMe), 5.27

 $(2 \text{ H, s, } CH_2\text{Ph}), 7.28-7.37 \text{ and } 7.53-7.57 (10 \text{ H, m, } 2 \times \text{Ph}) \text{ and}$ 9.94 (1 H, m, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm MHz}) 20.30, 20.58, 20.65,$ 29.59, 31.37, 32.44 and 32.81 (7 × CH_2 and 4-Me), 51.40, 51.76, 51.89 and 52.52 (OMe), 65.38 (OCH₂Ph), 83.37 (C-4), 115.28, 117.19, 125.75, 127.92, 128.04, 128.44, 128.60, 129.50, 131.19, 134.86, 136.55 and 138.72 (C=C) and 157.05, 160.33, 167.99, 170.40, 172.32, 172.86 and 173.74 (C=O and N=C-C=C); m/z (FD) 766 (M⁺ for ⁸⁰Se, 100%).

Attempted reduction of phenylselenopyrrolenine 16

A solution of phenylselenopyrrolenine 16 (31 mg, 42 μmol) in dry degassed toluene (5 cm³) was added dropwise via a syringe pump to a solution of triphenyltin hydride (100 mg, 0.27 mmol) in dry degassed toluene (10 cm³), which was being irradiated by UV light. After 4 h the solution was evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether-hexane (1:1), gave at lower R_f methylpyrrole **27** (13 mg, 84%), identical with authentic material, 12 and at higher R_f phenylselenopyrrole **26** (10.5 mg, 67%) (Found: M^+ , 395.0639. $C_{18}H_{21}NO_4^{80}Se$ requires M, 395.0636); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3400, 2975, 1725, 1260 and 1170; $\delta_{\rm H}({\rm CDCl_3}, 400~{\rm MHz})$ 2.21 (3 H, s, C-Me), 2.45 and 2.83 (each 2 H, t, J 7, CH₂CH₂), 3.46 (2 H, s, CH₂CO₂), 3.61 and 3.68 (each 3 H, s, OMe), 7.07-7.25 (5 H, m, Ph) and 7.93 (1 H, br s, NH); m/z (FD) 395 (M⁺ for ⁸⁰Se, 100%).

Reduction was also attempted by dropwise addition of 16 to a solution of triphenyltin hydride (4 equiv.) and AIBN in refluxing dry, degassed benzene. Again 27 (56%) and 26 (59%) were isolated.

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7bis(methoxycarbonylmethyl)-4-methyl-1-phenylthio-4,5dihydrodipyrrin 17

A solution of chloropyrrolenine 13 (62 mg, 96 µmol) in dry dichloromethane (3 cm³) was stirred with thiophenol (110 mg, 1 mmol) at room temperature for 90 min, then added to saturated aqueous sodium hydrogen carbonate (10 cm³) and extracted with dichloromethane $(3 \times 10 \text{ cm}^3)$. The combined extracts were dried and evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether, gave the phenylthiopyrrolenine 17 (56 mg, 81%) as a gum (Found: M⁺, 718.2562. $C_{38}H_{42}N_2O_{10}S$ requires M, 718.2560); $v_{max}(CH_2Cl_2)/cm^{-1}$ 3300, 1720, 1700 and 1170; $\delta_{\rm H}({\rm CDCl_3}, 400~{\rm MHz})$ 1.06 (3 H, s, 4-Me), 2.28 and 3.04 (each 1 H, d, J 15, 5-H₂), 2.45-2.64 and 2.90-2.99 (8 H, m, $2 \times CH_2CH_2$), 3.29 and 3.37 (each 1 H, d, J 16, CH₂CO₂), 3.36 and 3.43 (each 1 H, d, J 16, CH₂CO₂), 3.59, 3.61, 3.66 and 3.70 (each 3 H, s, OMe), 5.24 (2 H, s, CH₂Ph), 7.29–7.38 and 7.48–7.50 (10 H, m, $2 \times Ph$) and 9.91 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 \text{ MHz}) 20.02, 20.26, 20.55, 29.66, 31.41,$ 32.56, 32.67 and 34.86 (7 × CH₂ and 4-Me), 50.75, 51.28, 51.64 and 52.41 (OMe), 65.38 (OCH₂Ph), 81.71 (C-4), 115.20, 117.23, 127.90, 128.34, 128.61, 129.25, 129.56, 131.40, 133.77, 136.62 and 137.72 (C=C) and 157.49, 160.36, 169.57, 170.42, 172.35, 173.03 and 173.81 (C=O and N=C-C=C); m/z (FD) 718 (M⁺, 100%).

9-Benzyloxycarbonyl-1-methoxy-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin 18

A mixture of the lactam 21 (1.44 g, 2.3 mmol), trimethyloxonium tetrafluoroborate (340 mg, 2.3 mmol) and 1,8-bis-(dimethylamino)naphthalene (492 mg, 2.3 mmol) was stirred in dry dichloromethane (20 cm³) under argon for 28 h and then evaporated under reduced pressure. Purification on a flash chromatography column (25 × 2 cm), eluting with diethyl ether, gave methoxypyrrolenine 18 (1.22 g, 83%) as a gum (Found: M⁺ 640.2634. $C_{33}H_{40}N_2O_{11}$ requires M, 640.2632); $v_{max}(CH_2Cl_2)$ cm⁻¹ 3300, 2950, 1730s, 1690, 1450, 1380, 1250, 1170 and 1100; $\delta_{\rm H}({\rm CDCl_3},\,250~{\rm MHz})$ 1.03 (3 H, s, 4-Me), 2.15 and 2.96 (each 1 H, d, J 15, 5-H₂), 2.46–2.59 and 3.00–3.04 (8 H, m, $2 \times \text{CH}_2$ -CH₂), 3.38 and 3.44 (each 2 H, s, CH₂CO₂), 3.61, 3.63, 3.64,

3.67 and 3.69 (each 3 H, s, OMe), 5.18 and 5.29 (each 1 H, d, J 12, CH_2Ph), 7.31–7.40 (5 H, m, Ph) and 10.61 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3},\ 100\ {\rm MHz})\ 20.5\ (4{\rm -Me}),\ 19.51,\ 20.5,\ 29.56,\ 31.26,$ 31.93, 32.85 and 34.68 (7 × CH_2), 51.23, 51.46, 51.73, 52.18 and 54.74 (OMe), 65.63 (OCH₂Ph), 74.51 (C-4), 128.00 and 128.36 (C=CH), 115.02, 116.74, 130.00, 131.12, 131.88 and 136.18 (C=C) and 158.30, 160.26, 169.86, 170.46, 172.08, 172.98 and 173.61 (C=O and N=C-C=C); m/z (FD) 640 (M⁺, 100%).

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7bis(methoxycarbonylmethyl)-4-methyl-1-(4-nitrobenzyloxy)-4,5dihydrodipyrrin 19‡

A solution of 18-crown-6 (42 mg, 0.16 mmol) in benzene (2 cm³) was stirred with silver carbonate (400 mg, 1.5 mmol) at room temperature for 30 min and then lactam 21 (100 mg, 0.16 mmol) and p-nitrobenzyl bromide (340 mg, 1.6 mmol) were added. The mixture was stirred overnight and then evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether, gave imino ether 19 (37 mg, 72%) (Found: M+, 761.2799. $C_{39}H_{43}N_3O_{13}$ requires M, 761.2796); $\delta_H(CDCl_3, 400)$ MHz) 1.03 (s, 4-Me), 2.16 (1 H, d, J 15, 5-H_A), 2.50–2.60 and 2.97-3.02 (9 H total, $2 \times m$, $2 \times CH_2CH_2$ and $5-H_B$), 3.41 and 3.43 (each 2 H, s, CH₂CO₂), 3.60, 3.62, 3.63 and 3.71 (each 3 H, s, OMe), 5.24 and 5.31 (each 1 H, d, J 12, OC H_2 Ar) and 5.28 (2 H, s, OCH₂Ar), 7.23–7.46 (5 H, m, Ph), 8.15 and 8.17 (each 2 H, d, Ar) and 10.41 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3})$ 19.67 (4-Me), 20.57, 20.65, 29.73, 31.47, 32.18, 32.85 and 34.92 ($7 \times CH_2$), 51.44, 51.72, 51.94 and 52.46 (OMe), 65.77 (OCH₂Ph), 68.16 (OCH₂Ar), 75.11 (C-4), 123.76, 128.18, 128.42 and 128.58 (C=CH), 115.36, 117.13, 122.75, 127.02, 127.71, 130.03, 131.10, 131.93, 136.38, 143.60, 147.69 (C=C) and 159.38, 160.69, 169.44, 169.96, 172.22, 173.01 and 173.74 (C=O and N=C-C=C); m/z (FD) 761 (M⁺, 100%).

9-Benzyloxycarbonyl-1-(4-bromobenzyloxy)-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5dihydrodipyrrin 20‡

A solution of the lactam 21 (238 mg, 0.38 mmol) in dry benzene (4 cm³) was stirred with silver carbonate (315 mg, 1.14 mmol) and 18-crown-6 (99 mg, 0.38 mmol) at room temperature for 1.5 h and then p-bromobenzyl bromide (946 mg, 3.8 mmol) was added. The mixture was stirred for 3 h, then heated at reflux for 1.5 h and then evaporated under reduced pressure. Purification by PLC, eluting with 2% methanol in diethyl ether, gave the imino ether **20** (187 mg, 62%) as an oil (Found: M⁺, 794.2039. $C_{39}H_{43}^{79}BrN_2O_{11}$ requires M, 794.2050); $\delta_H(CDCl_3, 400 \text{ MHz})$ 1.07 (3 H, s, 4-Me), 2.22-2.25 (7 H, m), 2.68 (2 H, t, J 7) and 2.92 (1 H, d, J 15, 2 × CH₂CH₂ and 5-H₂), 3.37–3.45 (4 H, m, $2 \times \text{CH}_2\text{CO}_2$, 3.59, 3.60, 3.60 and 3.66 (each 3 H, s, OMe), 5.01 and 5.06 (each 1 H, d, J 12, OCH₂Ar), 5.22-5.26 (2 H, m, OCH₂Ar), 6.95 and 7.46 (each 2 H, d, J 8, Ar), 7.20–7.30 (5 H, m, Ph) and 10.50 (1 H, br s, NH); m/z (FD) 794 and 796 (1:1; M^+ , 100%).

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10H)-one N,N'-boronate

A solution of crude 5-chloropyrrolenine 13 (52 mg, 80 μmol; from reaction of lactam 21 with triphosgene) in dry THF (3 cm³) was stirred with a solution of lithium triethylborohydride in THF (1.0 mol dm⁻³; 160 mm³, 160 μmol) initially at 0 °C and then at room temperature for 1 h. Saturated aqueous ammonium chloride (2 cm³) and ethyl acetate (10 cm³) were added and the organic phase was separated, dried and evaporated under reduced pressure. Purification by flash chromatography on silica, eluting with diethyl ether, gave the N,N'boronate of lactam 21 (28 mg, 53%) as an oil (Found: MH+, 653.2490. $C_{32}H_{37}BN_2O_{12}$ requires MH, 653.2518); $\lambda_{max}(CH_2Cl_2)/2$

nm 282; $v_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$ 3691, 2954, 1778, 1736, 1438, 1345, 1173 and 1117; $\delta_{\text{H}}(\text{CDCl}_3, 400 \text{ MHz})$ 1.23 (3 H, s, 4-Me), 2.42–2.56 (2 H, m), 2.63 (4 H, m) and 2.71–2.87 (2 H, m, 2 × CH₂CH₂), 2.63 and 3.11 (each 1 H, d, J 16, 5-H₂), 3.39 (2 H, s, CH₂CO₂), 3.52 and 3.57 (each 1 H, d, J 16, CH₂CO₂), 3.60, 3.63, 3.66 and 3.74 (each 3 H, s, OMe), 5.17 and 5.43 (each 1 H, d, J 12.5, OCH₂Ph) and 7.28–7.41 (5 H, m, Ph); $\delta_{\text{C}}(\text{CDCl}_3, 100 \text{ MHz})$ 19.5 and 19.9 (CH₂CH₂CO₂), 21.8 (4-Me), 29.4, 30.9, 31.1, 31.1 and 34.5 (5 × CH₂), 51.6, 51.7, 52.3 and 52.8 (OMe), 65.0 (OCH₂Ph), 67.2 (C-4), 116.0, 121.6, 130.1, 134.4, 135.8, 141.3 and 153.7 (C=C), 128.2, 128.5 and 128.6 (C=CH), 160.8 (CO₂Bn), 162.2, 166.8, 170.8, 173.1 and 173.2 (C=O); m/z (+FAB) 653 (MH⁺, 55%) and 545 (MH – BnOH, 100).

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis-(methoxycarbonylmethyl)-4,10-dimethyl-4,5-dihydrodipyrrin-1(10*H*)-one 22

A solution of crude 5-chloropyrrolenine 13 (60 mg, 90 μmol; produced, as described above, by the action of triphosgene on lactam 21) in dry THF (2 cm³) was stirred with a solution of zinc borohydride³⁴ in diethyl ether (0.14 mol dm⁻³; 1.32 cm³, 180 µmol) at room temperature for 3 h. Saturated aqueous ammonium chloride (1 cm³) was added and the phases were separated. The organic phase was dried and evaporated under reduced pressure. Purification by flash chromatography on silica, eluting with diethyl ether-methanol (20:1), gave Nmethyl lactam 22 (27 mg, 44%) as an oil (Found: MH+, 641.2715. $C_{33}H_{40}N_2O_{11}$ requires MH, 641.2710); $\lambda_{max}(CH_2Cl_2)/CH_2Cl_3$ nm 277; v_{max}(CHCl₃)/cm⁻¹ 3319, 2954, 2930, 1731, 1689, 1438, 1252 and 1174; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 1.30 (3 H, s, 4-Me), 2.33– 3.01 (11 H, $2 \times CH_2CH_2$, $2 \times CH_AH_BCO_2$ and $5-H_AH_B$), 2.68 (3 H, s, NMe), 3.30 (1 H, d, J 19.5, CH_AH_BCO₂), 3.37 (1 H, d, J 18, $CH_AH_BCO_2$), 3.50 (1 H, d, J 16, 5- H_AH_B), 3.59, 3.60, 3.64 and 3.69 (each 3 H, s, OMe), 5.17 and 5.33 (each 1 H, d, J 12.5, OCH_2Ph), 7.25–7.42 (5 H, m, Ph) and 9.70 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm ~MHz})$ 20.1 and 20.4 (2 × $C{\rm H_2CH_2CO_2}$), 22.4 (4-Me), 24.5, 29.7, 29.8, 30.4, 31.0 and 34.5 (5 × CH₂ and NMe), 51.4, 51.6, 52.1 and 52.9 (OMe), 65.7 (OCH₂Ph), 66.1 (C-4), 128.0 and 128.4 (C=CH), 115.2, 116.4, 128.2, 129.8, 136.2, 136.5 and 146.5 (C=C), 160.1 (CO₂Bn) and 169.3, 171.1, 172.1, 173.6 and 173.8 (C=O); m/z (+FAB) 641 (M⁺, 40%) and $372 (C_{20}H_{22}NO_6, 100).$

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis-(methoxycarbonylmethyl)-4,10-dimethyl-1,4,5,10-tetrahydrodipyrrin 23

A solution of crude 5-chloropyrrolenine 13 (140 mg, 0.2 mmol; produced, as described above, by the action of triphosgene on lactam 21) in dry THF (2.5 cm³) was stirred with a solution of zinc borohydride ³⁴ in diethyl ether (0.15 mol dm⁻³; 1.45 cm³, 0.2 mmol) at room temperature for 12 h, then cooled to 0 °C. Glacial acetic acid (0.5 cm³) was dripped in over a period of 30 min, then saturated aqueous disodium EDTA (1 cm³) was added and the phases were separated. The organic phase was dried and evaporated under reduced pressure. Purification by PLC, eluting with ethyl acetate, gave N-methyl amine 23 (37 mg, 25%) as an oil; $\lambda_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{nm}$ 281; $\nu_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$ 3030, 2953, 1732, 1696, 1437, 1248 and 1171; $\delta_{\text{H}}(\text{CDCl}_3, 400 \text{ MHz})$ 0.96 (3 H, s, 4-Me), 2.18-2.34 and 2.49-2.62 (each 4 H, m, $2 \times CH_2CH_2$), 2.30 (3 H, s, NMe), 2.66 and 2.88 (each 1 H, d, J 16), 2.86–2.91 and 3.00–3.04 (each 1 H, m), 3.19, 3.37 and 3.43 (each 1 H, d, J 13) and 3.50–3.63 (1 H, obscured, $4 \times CH_2$), 3.50, 3.57, 3.60 and 3.63 (each 3 H, s, OMe), 5.18 and 5.24 (each 1 H, d, J 12.5, CH₂Ph), 7.30-7.39 (5 H, m, Ph) and 11.02 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm MHz})$ 18.5 (4-Me), 20.4 and 22.4 (CH₂CH₂CO₂), 29.4, 29.9, 30.6, 31.7 and 38.7 (5 × CH₂), 34.8 (NMe), 51.4, 51.6, 52.0 and 52.0 (OMe), 59.9 (CH₂N), 65.4 (OCH₂Ph), 71.6 (C-4), 113.5, 116.4, 129.4, 132.6, 133.1, 136.2 and 136.4 (C=C), 128.0, 128.1 and 128.4 (C=CH), 160.3

(CO_2Bn) and 171.1, 172.3, 173.1 and 173.7 (C=O); m/z (+FAB) 627 (MH⁺, 40%), 372 ($C_{20}H_{22}NO_6$, 10) and 254 (100).

4-[5-Benzyloxycarbonyl-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-1-chloro-2,8-bis-(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-9-(2,2,2-tribromoethoxycarbonyl)-4,5-dihydrodipyrrin 30

A stirred solution of dipyrrolic lactam 288 (250 mg, 215 μmol) and 4-(dimethylamino)pyridine (80 mg, 650 µmol) in dry dichloromethane (15 cm³) was treated dropwise with a solution of phosgene (0.65 mmol) in toluene (0.5 cm³) at room temperature under argon, then stirred for 2 h and the solvent evaporated under a stream of argon. Purification by PLC, eluting with diethyl ether, gave chloropyrrolenine 30 (200 mg, 86% based on unrecovered starting material) as a water-sensitive gum; $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3350, 2950, 1740, 1700, 1245 and 1150; $\delta_{\rm H}({\rm CDCl_3}, 400 \,{\rm MHz}) \, 2.31 - 2.62 \, (12 \,{\rm H, m}), \, 3.03 \, (2 \,{\rm H, t}, J \, 8)$ and 3.16 and 3.19 (each 1 H, d, J 15, 3 × CH₂CH₂ and CH₂CCH₂), 3.34 and 3.45 (each 1 H, d, J 17, CH₂CO₂), 3.57 (2 H, s, CH₂CO₂), 3.58, 3.60, 3.60, 3.62, 3.63 and 3.79 (each 3 H, s, OMe), 3.71 and 3.80 (each 1 H, d, J 17, CH₂CO₂), 5.01 and 5.13 (each 1 H, d, J 12, OCH₂CBr₃), 5.21 and 5.29 (each 1 H, d, J 12, OCH_2Ph), 7.30–7.41 (5 H, m, Ph) and 9.80 and 10.10 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3},\ 100\ {\rm MHz})\ 19.01,\ 20.05,\ 20.27,\ 29.28,$ 29.66, 30.21, 30.42, 31.39, 31.52, 34.68, 34.83 and 36.72 (11 × CH₂ and CBr₃), 51.39, 51.53, 51.75, 51.92 and 53.09 (OMe), 65.59 (OCH₂Ph), 76.49 (OCH₂CBr₃), 83.38 (C-4), 116.07, 116.34, 118.29, 122.04, 122.33, 127.84, 130.49, 131.20, 136.11 and 138.80 (C=C) and 158.10, 160.11, 161.81, 170.82, 171.12, 172.12, 172.45, 173.39 and 173.47 (C=O and C=N); m/z (FD) 1173, 1175, 1177, 1179 and 1181 (M⁺, 100%).

4-[5-Benzyloxycarbonyl-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-1-phenyl-

seleno-9-(2,2,2-tribromoethoxycarbonyl)-4,5-dihydrodipyrrin 31 A solution of chloropyrrolenine 30 (210 mg, 178 μmol) in dry degassed dichloromethane (5 cm³) was stirred with a solution of selenophenol¹⁶ (314 mg, 2.0 mmol) in dichloromethane (3 cm³) at room temperature under argon for 30 min, then added to saturated aqueous sodium hydrogen carbonate (20 cm³) and extracted with dichloromethane $(4 \times 15 \text{ cm}^3)$. The combined extracts were dried and evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether, gave phenylselenopyrrolenine 31 (78 mg, 34%) as a gum (Found: C, 47.7; H, 4.55; N, 3.3. C₅₂H₅₆Br₃N₃O₁₆Se requires C, 48.1; H, 4.35; N, 3.25%); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 1715, 1695, 1160 and 1070; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 2.28–2.58 (14 H, m, 3 × CH₂CH₂ and $CH_AH_BCCH_AH_B$), 3.09-3.17 (6 H, m, 2 × CH_2CO_2 and $CH_AH_BCCH_AH_B$), 3.53, 3.58, 3.62 and 3.76 (18 H, 4×s, $6 \times OMe$), 3.66 and 3.82 (each 1 H, d, J 17, CH_2CO_2), 5.07 and 5.12 (each 1 H, d, J 12, CH₂CBr₃), 5.22 and 5.28 (each 1 H, d, J 12, CH_2Ph), 7.26–7.37 and 7.41–7.43 (10 H, m, 2 × Ph) and 9.67 and 9.96 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 \,{\rm MHz})$ 18.98, 20.43, 20.55, 29.19, 30.08, 30.33, 30.61, 31.07, 32.22, 34.83, $35.00 (11 \times CH_2)$, 51.39, 51.49, 51.72, 53.02 (OMe), 65.41(OCH₂Ph), 76.68 (OCH₂CBr₃), 85.72 (C-4), 115.88, 116.09, 118.17, 121.73, 122.07, 127.93, 128.10, 128.35, 129.00, 129.65, 130.53, 131.46, 135.32, 136.36, 141.50 and 154.90 (C=C) and 158.53, 160.15, 171.28, 171.86, 172.11, 172.57, 173.30 and 173.54 (C=O and C=N); m/z (FD) 1295, 1296, 1297, 1298, 1299, 1300, 1301, 1302, 1303 and 1304 (ratio 4:11:5:14:12:20:14: 9:7:4, M⁺, 100%).

9-Benzyloxycarbonyl-4-[5-benzyloxycarbonyl-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-1-phenylseleno-4,5-dihydrodipyrrin 32
Dipyrrolic lactam 29 ¹¹ (80 mg, 80 μmol) was reacted with phos-

gene, as described above for the synthesis of 30, followed by selenophenol, as described above for the synthesis of 31. Purification by PLC, eluting with methanol–diethyl ether (1:40), gave the phenylselenopyrrolenine 32 (42 mg, 49%) as a gum (Found: M^+ , 1123.3248. $C_{57}H_{61}N_3O_{16}^{80}Se$ requires M, 1123.3217); $\nu_{\rm max}({\rm CH_2Cl_2})/{\rm cm}^{-1}$ 3300, 1715, 1700, 1170 and 1065; $\delta_{\rm H}({\rm CDCl_3})$, 400 MHz) 2.26-2.35, 2.40-2.52 and 2.80-3.22 (18 H, m, $3 \times \text{CH}_2\text{CH}_2$, CH₂CCH₂ and CH₂CO₂), 3.47 (2 H, s, CH₂CO₂), 3.53, 3.54, 3.57, 3.59, 3.61 and 3.70 (each 3 H, s, OMe), 3.65 and 3.83 (each 1 H, d, J 17, CH₂CO₂), 5.22 and 5.28 (each 1 H, d, J 13, CH₂Ph), 5.22 and 5.31 (each 1 H, d, J 13, CH₂Ph), 7.27-7.39 (15 H, m, 3 × Ph) and 9.68 and 9.71 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm \ MHz})$ 19.00, 20.50, 29.20, 30.18, 30.31, 30.62, 31.07, 32.17, 34.69 and 34.84 (CH₂), 51.32, 51.44, 51.68, 51.74 and 52.87 (OMe), 65.38 and 65.45 (OCH₂Ph), 85.85 (C-4), 115.49, 117.16, 118.16, 121.72, 122.11, 125.00, 127.91, 128.09, 128.36, 128.38, 129.65, 129.70, 129.80, 135.34, 136.43 and 141.51 (C=C) and 154.89, 160.12, 160.18, 171.27, 171.88, 172.17, 172.60, 173.29 and 173.61 (C=O and C=N); m/z (FD) 1123 (M⁺ for ⁸⁰Se, 100%).

9-Benzyloxycarbonyl-4-[5-benzyloxycarbonyl-4-(2-methoxycarbonylethyl)-3-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10H)-thione 36

A solution of lactam 35⁸ (52 mg, 53 μmol) in benzene (3 cm³) was heated at reflux with Lawesson's reagent (12 mg, 29 µmol) under argon for 45 min and then evaporated under reduced pressure. Purification on a column of silica gel PF_{254} (5 × 1 cm), eluting with dichloromethane and then diethyl ether, gave thiolactam 36 (43 mg, 81%) as an oil (Found: M+, 999.3428. $C_{51}H_{57}N_3O_{16}S$ requires M, 999.3459); $\lambda_{max}(EtOH)/nm$ 281; v_{max} (CHCl₃)/cm⁻¹ 3430, 3300, 2950, 1720s, 1690, 1570, 1435, 1180 and 1075; $\delta_{\rm H}({\rm CDCl_3},\,400~{\rm MHz})$ 2.45–2.51 (8 H, m) and 2.89-3.01 (4 H, m, 3 × CH₂CH₂), 2.82 and 3.06 (each 2 H, d, J 15, 2 × 4-CH₂), 3.32 and 3.50 (each 2 H, d, J 17, CH₂CO₂), 3.49 (2 H, s, CH₂CO₂), 3.55 and 3.72 (each 3 H, s, OMe), 3.60 and 3.64 (each 6 H, s, 2 × OMe), 5.20 and 5.27 (each 2 H, d, J 12, CH₂Ph), 7.28–7.39 (10 H, m, Ph), 9.46 (1 H, br s, NH) and 9.62 (2 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3},\ 100.57\ {\rm MHz},\ {\rm DEPT})\ 20.68$ (3 C), 29.61 (2 C), 31.01 (3 C), 31.67 and 34.64 (2 C, CH₂), 51.45 (2 C), 52.27 (2 C) and 52.99 (1 C, OMe), 66.00 ($2 \times OCH_2$), 74.16 (C-4), 115.90, 118.33, 128.90 (br) and 130.24 (each 2 C, pyrrole-C), 128.00, 128.15 and 128.40 (phenyl-CH), 135.98 (phenyl-C), 143.23 and 147.84 (C=C), 160.65 (2 C), 171.05, 172.80 (2 C), 173.44 and 173.57 (2 C, C=O) and 197.50 (C=S); m/z (FD) 999 (M⁺, 100%).

4-[4-(2-Methoxycarbonylethyl)-3-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis-(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10H)-one 38

A solution of lactam dibenzyl ester 35⁸ (100 mg) in methanol (5 cm³) was stirred with 10% palladium-on-charcoal (10 mg) under an atmosphere of hydrogen until uptake of gas ceased (45 min), then filtered through Celite and evaporated. A solution of the residue in trifluoroacetic acid (5 cm³) was allowed to stand for 7 h at room temperature under argon, then added to water (50 cm³) and extracted with dichloromethane (3 \times 10 cm³). The extract was washed with 5% aqueous sodium hydrogen carbonate (30 cm³), dried and evaporated. Purification on a short column of silica gel PF_{254} (5 × 2.5 cm diam.), eluting with diethyl ether-methanol (19:1), gave lactam 38 (59 mg, 81%) as a gum (Found: M^+ , 715.2950. $C_{35}H_{45}N_3O_{13}$ requires M, 715.2952); λ_{max} end absorption only; $v_{\text{max}}(\text{CHCl}_3)/\text{cm}^{-1}$ 3450, 3350br, 2950, 1720, 1685, 1440, 1175 and 1010; $\delta_{H}(CDCl_3, 400)$ MHz) 2.41 and 2.46 (each 2 H, m, 2-CH₂CH₂), 2.49 and 2.66 (each 4 H, m, $2 \times CH_2CH_2CO_2$), 2.72 and 2.95 (each 2 H, d, J 15, CH₂CCH₂), 3.25 and 3.38 (each 2 H, d, J 16, $2 \times$ CH₂CO₂), 3.48 (2 H, s, 3-CH₂), 3.62 and 3.79 (each 3 H, s, OMe), 3.64 and 3.66 (each 6 H, s, 2 × OMe), 6.33 (2 H, d, J 3,

 $2 \times \alpha$ -H), 7.57 (1 H, br s, lactam-NH) and 8.75 (2 H, br s, $2 \times \text{pyrrole-NH}$; m/z (FD) 715 (M⁺, 100%).

4-[5-Benzyloxycarbonyl-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-9-(2,2,2-tribromoethoxycarbonyl)-4,5-dihydrodipyrrin-1(10H)-thione 40

A solution of lactam 28⁸ (120 mg, 104 µmol) and Davy's reagent ²⁰ 44 (16.3 mg, 57 μmol) in dry 1,2-dimethoxyethane (1 cm³) was stirred at room temperature for 20 min and then evaporated under reduced pressure. Purification by PLC, eluting with methanol-diethyl ether (1:40), gave the thiolactam 40 (114 mg, 94%) as a gum (Found: C, 47.3; H, 4.65; N, 3.55. $C_{46}H_{52}Br_3N_3O_{16}S$ requires C, 47.0; H, 4.5, N, 3.6%); $v_{max}(CH_2-V_3)$ Cl_2 /cm⁻¹ 3400, 2960, 1730, 1700, 1430 and 1170; $\delta_{\text{H}}(\text{CDCl}_3,$ 400 MHz) 2.46–2.58 and 2.71–2.76 (12 H, m, $3 \times CH_2CH_2$), 2.89 and 3.20 (each 1 H, d, J 15) and 3.06 and 3.12 (each 1 H, d, J 15, CH₂CCH₂), 3.21 and 3.45 (each 1 H, d, J 17, CH₂CO₂), 3.57, 3.58, 3.59, 3.60, 3.61 and 3.81 (each 3 H, s, OMe), 3.57-3.81 (4 H, obscured, $2 \times \text{CH}_2\text{CO}_2$), 5.04 and 5.12 (each 1 H, d, J 12, CH₂CBr₃), 5.19 and 5.24 (each 1 H, d, J 12, CH₂Ph), 7.27– 7.39 (5 H, m, Ph) and 9.29, 9.44 and 10.26 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm ~MHz})$ 19.16, 20.34, 20.85, 29.20, 30.34, 30.54, 31.42, 31.79, 34.59 and 34.93 (CH₂ and CBr₃), 51.48, 51.86, 52.23, 52.53 and 53.31 (OMe), 65.64 (OCH₂Ph), 73.49 (C-4) and 76.67 (CH₂CBr₃), 115.62, 119.46, 122.14, 122.51, 129.53, 131.11, 135.25, 144.07 and 147.29 (C=C), 160.16, 161.99, 171.70, 171.78, 171.95, 172.05 and 173.70 (C=O) and 197.73 (C=S); *m*/*z* (FD) 1171, 1173, 1175, 1177 (1:3:3:1, M⁺, 100%).

4-[5-Benzyloxycarbonyl-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10H)-one 41

A solution of tribromoethyl ester 288 (100 mg, 86 μmol) in glacial acetic acid (2 cm³) was stirred with zinc dust (200 mg) under argon at room temperature for 20 min and then filtered. The filtrate was added to aqueous sodium carbonate (15 cm³) and extracted with dichloromethane $(4 \times 10 \text{ cm}^3)$. The combined extracts were dried and evaporated under reduced pressure. A solution of the residue in freshly distilled trifluoroacetic acid (1 cm³) was stirred at room temperature under argon for 7 h, then poured into water (20 cm³) and extracted with dichloromethane (4 × 15 cm³). The combined extracts were dried and evaporated under reduced pressure. Purification by PLC, eluting with 5% methanol in diethyl ether, gave the α -free pyrrolic lactam 41 (62 mg, 85%) as a foam (Found: C, 60.8; H, 6.0; N, 4.8. $C_{43}H_{51}N_3O_{15}$ requires C, 60.8; H, 6.05; N, 4.9%); $\nu_{max}(CH_2Cl_2)/cm^{-1}$ 3400, 2950, 1720, 1685, 1435 and 1175; $\delta_{\rm H}({\rm CDCl_3}, 400 \,{\rm MHz}) \, 2.39 - 2.50 \, (8 \,{\rm H, m}) \, {\rm and} \, 2.63 - 2.68 \, (4 \,{\rm H, m})$ $3 \times \text{CH}_2\text{CH}_2$), 2.74 and 3.11 (each 1 H, d, J 15) and 2.81 and 2.99 (each 1 H, d, J 15, CH₂CCH₂), 3.20 and 3.38 (each 1 H, d, J 16, CH₂CO₂), 3.47 and 3.55 (each 1 H, d, J 16, CH₂CO₂), 3.57, 3.59, 3.60, 3.63, 3.64 and 3.77 (each $3 \text{ H}, \text{ s}, 6 \times \text{OMe}$), 3.57-3.77(2 H, obscured, CH₂CO₂), 5.17 and 5.27 (each 1 H, d, J 12, CH_2Ph), 6.34 (1 H, d, J 2, α -H), 7.28–7.38 (5 H, m, Ph) and 7.41, 8.91 and 9.54 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100$ MHz) 19.14, 19.62, 20.61, 29.80, 30.43, 30.59, 30.74, 31.62, 34.69 and 34.74 (CH₂), 51.45, 51.54, 51.75, 52.04 and 52.95 (OMe), 65.68 and 66.68 (C-4 and OCH₂Ph), 112.25, 114.61, 118.99, 120.87, 121.87, 122.28, 123.25, 127.98, 128.19, 128.33, 136.00, 137.21 and 149.83 (C=C) and 160.48, 171.86, 172.38, 173.29, 173.40, 173.51 and 173.62 (C=O); m/z (FD) 849 (M⁺, 100%).

General procedure for the conversion of lactams into thiolactams

A solution of the lactam (0.44 mmol) in dry benzene (24 cm³) was heated at reflux under argon with Lawesson's reagent (95 mg, 0.23 mmol) for 45 min and then evaporated under reduced pressure. Purification by chromatography gave the thiolactam.

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10*H*)-thione 46

Method A. Lactam 21 was reacted with Lawesson's reagent according to the above general procedure. Purification by flash chromatography, eluting with diethyl ether, gave thiolactam 46 (72%) which crystallised from aqueous methanol, mp 114-116 °C (Found: C, 60.2; H, 6.2; N, 4.3%; M⁺, 642.2251. C₃₂H₃₈- $N_2O_{10}S$ requires C, 59.8; H, 6.0; N, 4.35%; M, 642.2247); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 2950, 1740s, 1710, 1680, 1585, 1490, 1450, 1200 and 1065; $\delta_{\rm H}({\rm CDCl_3}, 250~{\rm MHz})$ 1.38 (3 H, s, 4-Me), 2.45-2.60 (6 H, m) and 2.68-2.80 (2 H, m, $2 \times CH_2CH_2$), 2.81and 3.00 (each 1 H, d, J 15, 5-H₂), 3.32 and 3.70 (each 1 H, d, J 17, CH₂CO₂), 3.35 and 3.58 (each 1 H, d, J 17, CH₂CO₂), 3.59, 3.62, 3.71 and 3.77 (each 3 H, s, OMe), 5.19 and 5.30 (each 1 H, d, J 12, CH₂Ph), 7.27-7.41 (5 H, m, Ph) and 9.13 and 10.17 (each 1 H, br s, NH); $\delta_{\rm C}$ (CDCl₃, 62.5 MHz) 22.43 (4-Me), 20.47, 20.53, 29.75, 31.92, 31.98, 32.72 and 34.73 (CH₂), 51.29, 51.39, 52.56 and 52.80 (OMe), 65.75 (PhCH₂), 70.86 (C-4), 127.90, 128.11 and 128.34 (phenyl-CH), 115.12, 118.24, 128.80, 130.00, 136.12 and 149.95 (C=C), 160.53, 170.94 and 173.55 (C=O) and 196.52 (C=S); *m/z* (FD) 642 (M⁺, 100%).

Method B. Dichloromethane (3 cm³) at 0 °C was saturated with hydrogen sulfide and then stirred with a solution of the halopyrrolenines 13/14 (36 mg, 50 μ mol) in dichloromethane (2 cm³) for 1 h at room temperature. The solvent was evaporated in a stream of nitrogen. Purification by chromatography gave the thiolactam 46 (25 mg, 80%).

General procedure for desulfurisation with nickel boride

A stirred solution of nickel(II) chloride hexahydrate (1.19 g, 5 mmol) and boric acid (4.0 g) in methanol (70 ml) was cooled in ice and a solution of sodium borohydride (380 mg, 10 mmol) in water (5 ml) was added over 1 min. After 5 min the black precipitate was allowed to settle and the supernatant was decanted. The precipitate of nickel boride was washed with methanol (3 \times 20 ml) decanting excess methanol after each washing and used immediately without drying.

A solution of the thiolactam (0.2 mmol) in methanol (6 cm³) and acetic acid (0.6 cm³) was added to a suspension of the above nickel boride in methanol (15 cm³). The mixture was stirred under hydrogen at room temperature for 30 min, then filtered, washing the residue with methanol (15 cm³). The filtrate was mixed with 5% aqueous sodium hydrogen carbonate (200 cm³), and extracted with dichloromethane (4 × 50 cm³). The combined extracts were dried, evaporated under reduced pressure and purified by chromatography as described.

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis-(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin 49

The thiolactam 46 was reduced with nickel boride according to the general procedure above. Purification by PLC, eluting with ethanol-diethyl ether (1:9), gave pyrrolenine 49 (45%) as a gum (Found: M^+ , 610.2535. $C_{32}H_{38}N_2O_{10}$ requires M, 610.2527); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3320, 2960, 2850, 1730s, 1700, 1430, 1210 and 1160; $\delta_{\rm H}({\rm CDCl_3}, 400 {\rm ~MHz})$ 1.10 (3 H, s, 4-Me), 2.23 and 3.10 (each 1 H, d, J 15, 5-H₂), 2.46–2.53 and 2.96–3.02 (6 H, m) and 2.61 (2 H, t, J 7, 2 × CH₂CH₂), 3.37 and 3.47 (each 1 H, d, J 17, CH₂CO₂), 3.38 and 3.48 (each 1 H, d, J 16, CH₂CO₂), 3.60, 3.64 and 3.69 (12 H, each s, $4 \times OMe$), 5.23 and 5.32 (each 1 H, d, J 12, CH₂Ph), 7.30–7.42 (5 H, m, Ph), 7.92 (1 H, s, 2-H) and 10.20 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm MHz})$ 19.60 (4-Me), 20.43, 20.53, 29.61, 31.12, 32.15, 32.68 and 34.72 (CH₂), 51.23, 51.61, 51.76 and 52.29 (OMe), 65.46 (PhCH₂), 83.30 (C-4), 115.22, 117.15, 129.72, 131.20, 137.41 and 157.85 (C=C), 127.89, 128.07 and 128.38 (C=CH), 160.46, 172.20 and 173.66 (C=O) and 164.78 (C-2); m/z (FD) 610 (M⁺, 100%).

Amine 47 (ca. 5%) was also found at lower $R_{\rm f}$. When the reaction time was extended to 15 h, amine 47 (51%) became the only product isolated (Found: M⁺, 612.2682. $C_{32}H_{40}N_2O_{10}$

requires M, 612.2683); $v_{\rm max}({\rm CH_2CI_2})/{\rm cm}^{-1}$ 3280, 2940, 1740s, 1700, 1450, 1260, 1160 and 1090; $\delta_{\rm H}({\rm CDCI_3}, 250~{\rm MHz})$ 1.16 (3 H, s, 4-Me), 2.29–2.36 (4 H, m) and 2.51 and 2.99 (each 2 H, t, J 7, 2 × CH₂CH₂), 2.48 and 2.58 (each 1 H, d, J 15, 5-H₂), 3.09–3.15 (3 H, m, CH₂CO₂ and NCH_AH_B), 3.37 and 3.49 (each 1 H, d, J 16, CH₂CO₂), 3.52 (1 H, d, J 15, NCH_AH_B), 3.59, 3.61, 3.64 and 3.65 (each 3 H, s, OMe), 5.21 and 5.32 (each 1 H, d, J 12, CH₂Ph), 7.27–7.42 (5 H, m, Ph) and 10.74 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCI_3}, 100~{\rm MHz})$ 20.38 (4-Me), 22.33, 25.74, 29.33, 29.97, 31.56, 33.86 and 34.61 (CH₂), 50.82, 51.09, 51.37 and 51.72 (OMe), 53.07 (C-2), 65.15 (PhCH₂), 70.88 (C-4), 114.80, 116.93, 132.44, 132.55, 136.43 and 138.99 (C=C), 127.65, 128.15 and 129.60 (C=CH) and 160.28, 171.75, 172.03, 172.74 and 173.23 (C=O); mlz (FD) 612 (M⁺, 100%).

9-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-1-methylthio-4,5-dihydrodipyrrin 48

Thiolactam 46 (375 mg, 0.584 mmol) was dissolved in redistilled trifluoroacetic acid (10 cm³) and distilled trimethyl orthoformate (10 cm³) was immediately added. The solution was stirred in the dark under argon for 30 min and then evaporated under reduced pressure. The residue was twice dissolved in dichloromethane (20 cm³) and evaporated under reduced pressure and then purified by flash chromatography (40×2.5 cm), eluting with diethyl ether, to give the thioimidate 48 (365 mg, 95%) as a gum (Found: M+, 656.2404. C₃₃H₄₀N₂O₁₀S requires M, 656.2403); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 2950, 1720s, 1690, 1440, 1190 and 1075; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 1.05 (3 H, s, 4-Me), 2.28 (3 H, s, S-Me), 2.30 and 2.99 (each 1 H, d, J 15, 5-H₂), 2.53–2.58 and 2.98-3.00 (8 H, m, 2 × CH₂CH₂), 3.39 and 3.44 (each 2 H, s, CH₂CO₂), 3.61, 3.64, 3.64 and 3.69 (each 3 H, s, OMe), 5.22 and 5.29 (each 1 H, d, J 12, CH_2Ph), 7.33–7.40 (5 H, m, Ph) and 10.67 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 \,{\rm MHz})$ 12.75 (SMe), 19.9 (4-Me), 19.9, 20.55, 29.70, 31.42, 32.55, 32.77 and 34.81 (CH₂), 51.21, 51.54, 51.72 and 52.19 (OMe), 65.70 (CH₂Ph), 81.40 (C-4), 128.08 and 128.50 (C=CH), 115.21, 130.22, 136.38, 136.44 and 157.82 (C=C) and 160.39, 169.76, 172.06, 172.74 and 173.61 (C=O and C-2); m/z (FD) 656 (M⁺, 100%).

9-Benzyloxycarbonyl-4-[5-benzyloxycarbonyl-4-(2-methoxycarbonylethyl)-3-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin 51

Thiolactam 36 was reduced with nickel boride following the general procedure described above. Purification by PLC, eluting with diethyl ether-methanol (19:1), gave pyrrolenine 51 (33 mg, 46%) as a gum; $\lambda_{\text{max}}(\text{EtOH})/\text{nm}$ 281; $\nu_{\text{max}}(\text{CHCl}_3)$ 3420, 3320, 2950, 1725, 1685, 1445, 1435, 1255, 1175 and 1080; $\delta_{\rm H}({\rm CDCl_3},$ 400 MHz) 2.33 (2 H, m), 2.48 (6 H, m) and 2.87–3.00 (4 H, m, $3 \times \text{CH}_2\text{CH}_2$), 2.40 and 3.10 (each 2 H, d, J 15, 2 × 4-CH₂), 3.32 and 3.37 (each 2 H, d, J 16, 2 × CH₂CO₂), 3.45 (2 H, s, CH_2CO_2), 3.58 and 3.59 (each 6 H, s, 2 × OMe), 3.58 and 3.71 (each 3 H, s, OMe), 5.26 and 5.30 (each 2 H, d, J 12, CH₂Ph), 7.30–7.42 (10 H, m, 2 × Ph), 7.80 (1 H, s, 2-H) and 10.03 (2 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3},\ 100.57\ {\rm MHz})\ 20.60,\ 20.65\ (2\ {\rm C}),\ 29.56$ (2 C), 30.14, 31.13, 32.42 (2 C) and 34.92 (2 C, CH₂), 51.50 (2 C), 51.87, 51.98 (2 C) and 52.99 (OMe), 65.71 (2 × OCH₂), 86.13 (C-4), 115.62, 117.37, 129.85 and 130.11 (each 2 C pyrrole-C), 128.14, 128.34, 128.57 and 136.48 (2 × Ph), 140.47 and 156.04 (C=C), 166.33 (C-2) and 160.42 (2 C), 171.36, 172.42 (2 C), 172.77 and 173.79 (2 C, C=O); m/z (FD) 967 (M⁺, 100%).

2,7-Bis(2-methoxycarbonylethyl)-3,8-bis(methoxycarbonyl-methyl)-4-methyl-9-(2,2,2-tribromoethoxycarbonyl)-4,5-dihydrodipyrrin-1(10*H*)-one 54

Iodopyrrole 12 and acetoxymethylpyrrole 53⁸ were reacted by the procedure described ¹¹ for the synthesis of 29 except that the hydrolysis of the halopyrrolenines required 15 h. Purification by flash chromatography, eluting with 0–3% ethanol in diethyl

ether, gave lactam 54 (29%) as a gum (Found: M+, 797.9630. $C_{27}H_{33}^{79}Br_3N_2O_{11}$ requires M, 797.9635); $v_{max}(CH_2Cl_2)/cm^{-1}$ 3250, 2960, 1720s, 1430, 1210 and 1040; $\delta_{\rm H}({\rm CDCl_3}, 400~{\rm MHz})$ 1.40 (3 H, s, 4-Me), 2.46–2.67 (6 H, m) and 2.72 (2 H, t, J 7, $2 \times CH_2CH_2$, 2.78 and 3.14 (each 1 H, d, J 15, 5-H₂), 3.36 and 3.72 (each 1 H, d, J 17, CH₂CO₂), 3.63, 3.65, 3.68 and 3.79 (each 3 H, s, OMe), 3.63-3.79 (1 H, obscured) and 4.09 (1 H, d, J 17, CH₂CO₂), 4.99 and 5.13 (each 1 H, d, J 17, CH₂CBr₃), 6.54 (1 H, br s, lactam-NH) and 10.08 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 \,{\rm MHz}) \, 24.27 \, (4-{\rm Me}), \, 19.22, \, 19.82, \, 30.66, \, 30.93,$ 31.33, 33.29, 34.42 and 36.11 (CH₂ and CBr₃), 51.52, 51.85, 51.91 and 52.93 (OMe), 63.11 (C-4), 76.68 (CH₂O), 117.93, 122.60, 123.55, 129.96, 135.89 and 150.89 (C=C) and 158.78, 170.82, 171.44, 171.62 and 174.07 (C=O); m/z (FD) 798, 800, 802 and 804 (1:3:3:1, M⁺, 100%).

2,7-Bis(2-methoxycarbonylethyl)-3,8-bis(methoxycarbonylmethyl)-4-methyl-9-(2,2,2-tribromoethoxycarbonyl)-4,5-dihydrodipyrrin-1(10H)-thione 55

Lactam 54 was reacted with Lawesson's reagent according to the above general procedure. The residue was purified by flash chromatography, eluting with diethyl ether, to give thiolactam **55** as a foam (Found: M^+ , 813.9374. $C_{27}H_{33}^{79}Br_3N_2O_{10}S$ requires M, 813.9406); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 2950, 1725s, 1430, 1200 and 1160; $\delta_{H}(CDCl_{3}, 400 \text{ MHz})$ 1.45 (3 H, s, 4-Me), 2.57-2.80 (8 H, m, 2 × CH₂CH₂), 2.83 and 3.24 (each 1 H, d, J 15, 5-H₂), 3.38 and 3.72 (each 1 H, d, J 17, CH₂CO₂), 3.64, 3.65, 3.74 and 3.79 (each 3 H, s, OMe), 3.75 and 4.08 (each 1 H, d, J 17, CH₂CO₂), 5.01 and 5.11 (each 1 H, d, J 17, CH₂CBr₃), 8.74 (1 H, br s, thiolactam-NH) and 10.12 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 \text{ MHz}) 23.04 (4-Me), 19.11, 21.08, 31.02,$ 31.92, 32.76, 34.13 and 36.17 (CH₂ and CBr₃), 51.51, 51.93, 52.21 and 53.02 (OMe), 70.87 (C-4), 76.74 (CH₂O), 118.14, 122.61, 123.80, 129.27, 141.96 and 149.44 (C=C), 158.80, 170.79, 171.52, 173.55 and 174.77 (C=O) and 196.55 (C=S); m/z (FD) 814, 816, 818 and 820 (1:3:3:1, M⁺, 100%).

 $9\hbox{-}Carboxy\hbox{-}2,7\hbox{-}bis (2\hbox{-}methoxy carbonylethyl)\hbox{-}3,8\hbox{-}bis (methoxy-arbonylethyl)\hbox{-}3,8\hbox{-}bis (methoxy-arbonylethyl)\hbox{-}3,8\hbox{-}5,8\hbox$ carbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10H)-thione 56 A solution of thiolactam 55 (500 mg, 0.61 mmol) in glacial acetic acid (30 cm³) was stirred with zinc dust (1.30 g) for 20 min, then filtered, mixed with water (50 cm³) and extracted with dichloromethane $(4 \times 50 \text{ cm}^3)$. The combined extracts were washed with water (100 cm³), dried and evaporated under reduced pressure. Purification by flash chromatography (20 × 2 cm), eluting with ethanol-diethyl ether (1:9), gave acid 56 (302 mg, 90%) as a gum (Found: M⁺, 552.1772. C₂₅H₃₂N₂O₁₀S requires M, 552.1776); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 3400–2700, 1730s, 1650, 1450 and 1160; $\delta_{H}(CDCl_{3}, 400 \text{ MHz})$ 1.23 (3 H, s, 4-Me), 2.45–2.52 and 2.67–2.72 (8 H, m, $2 \times CH_2CH_2$), 2.71 and 3.19 (each 1 H, d, J 15, 5-H₂), 3.52 and 3.58 (each 1 H, d, J 16, CH₂CO₂), 3.64, 3.64, 3.68 and 3.74 (each 3 H, s, OMe), 3.83 (2 H, s, CH₂CO₂) and 9.99 and 10.49 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3},\ 100\ {\rm MHz})\ 20.70\ (4{\rm -Me}),\ 19.37,\ 20.92,\ 30.94,\ 31.42,$ 32.11, 32.31 and 34.69 (CH₂), 51.65, 51.82, 52.12, 52.73 (OMe), 72.09 (C-4), 119.42, 123.13, 123.98, 129.66, 140.35, 151.99 (C=C), 164.35, 169.83, 172.55 and 173.64 (C=O) and 196.19 (C=S); m/z (FD) 552 (M⁺, 100%).

2,7-Bis(2-methoxycarbonylethyl)-3,8-bis(methoxycarbonylmethyl)-4-methyl-1-methylthio-9-(2,2,2-tribromoethoxycarbonyl)-4,5-dihydrodipyrrin 57

Thiolactam 55 was treated with trifluoroacetic acid and trimethyl orthoformate according to the procedure described above for the synthesis of 48. Purification by flash chromatography, eluting with diethyl ether, gave thioimidate 57 (92%) as a gum (Found: M^+ , 827.9551. $C_{28}H_{35}^{79}Br_3N_2O_{10}S$ requires M, 827.9562); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3350, 2970, 1725 and 1180; $\delta_{\rm H}({\rm CDCl_3}, 400 \ {\rm MHz}) \ 1.50 \ (3 \ {\rm H, \ s, \ 4-Me}), \ 2.27-2.63 \ (9 \ {\rm H, \ m, \ m})$

 $2 \times CH_2CH_2$ and 5-H_A), 2.81 (3 H, s, SMe), 3.43 (1 H, d, J 15, 5-H_B), 3.55 and 3.66 (each 1 H, d, J 16, CH₂CO₂), 3.64, 3.64, 3.65 and 3.74 (each 3 H, s, OMe), 3.74 and 3.93 (each 1 H, d, J 17, CH₂CO₂), 5.02 and 5.14 (each 1 H, d, J 12, CH₂CBr₃) and 10.05 (1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100.57 \ {\rm MHz})$ 14.71 (SMe), 19.18, 19.36, 20.19, 30.54, 31.55, 31.81, 33.04, 34.91 and 36.20 $(7 \times CH_2, 4$ -Me and $CBr_3)$, 51.71, 51.80, 51.89 and 52.87 (OMe), 80.10 (C-4), 118.19, 122.52, 123.34, 128.22, 136.95 and 158.59 (C=C) and 159.76, 168.31, 171.42, 172.14, 173.28 and 180.03 (C=O and C-2); m/z (FD) 828, 830, 832 and 834 (1:3:3:1, M⁺, 100%).

9-Carboxy-2,7-bis(2-methoxycarbonylethyl)-3,8-bis(methoxycarbonylmethyl)-4-methyl-1-methylthio-4,5-dihydrodipyrrin 58

Tribromoethyl ester 57 was cleaved using zinc and acetic acid following the same procedure as for the preparation of 56. Purification by PLC, eluting with 5% methanol in diethyl ether, gave the carboxylic acid 58 (95%) as a gum (Found: M+, 566.1851. $C_{26}H_{34}N_2O_{10}S$ requires M, 566.1822); $v_{max}(CH_2Cl_2)/CH_2Cl_2$ cm⁻¹ 3400–2700, 3300, 1730, 1650 and 1180; $\delta_{\rm H}({\rm CDC1_3}, 400$ MHz) 1.13 (3 H, s, 4-Me), 2.26 and 3.06 (each 1 H, d, J 15, 5-H₂), 2.37-2.39 (2 H, m), 2.50-2.58 (4 H, m) and 2.68 (2 H, t, J 8, 2 × CH₂CH₂), 2.56 (3 H, s, SMe), 3.44 (2 H, s, CH₂CO₂), 3.63, 3.65, 3.66 and 3.71 (each 3 H, s, OMe), 3.75 and 3.89 (each 1 H, d, J 17, CH₂CO₂) and 10.70 (1 H, br s, NH); m/z (FD) 566 (M⁺, 100%).

3-(2-Methoxycarbonylethyl)-4-methoxycarbonylmethyl-5methyl-2-methylthiopyrrole 62

A solution of carboxylic acid 61²⁵ (1.00 g, 3.53 mmol) and thiourea (270 mg, 3.55 mmol) in ethanol-water (1:1; 20 cm³) was stirred at room temperature under argon and a solution of iodine (0.1 mol dm⁻³) in aqueous potassium iodide (0.2 mol dm⁻³; 36 cm³) was added dropwise. After a further 30 min, sodium hydrogen carbonate (2.0 g) was added and the mixture was extracted with dichloromethane $(4 \times 15 \text{ cm}^3)$. The combined extracts were washed with water (30 ml), dried and evaporated to dryness. Without further purification, the intermediate thiourea derivative was dissolved in methanol-water (1:1) and sodium hydroxide (150 mg, 3.75 mmol) was added. The solution was stirred for 30 min at room temperature under argon and then methyl iodide was added. The solution was stirred for 15 h and then extracted with chloroform (4 × 15 cm³). The combined extracts were washed with water (30 cm³), dried and evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether, gave the methylthiopyrrole 62 (275 mg, 27%) as an oil (Found: M⁺, 285.1038. C₁₃H₁₉NO₄S requires M, 285.1035); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3450, 2940, 1730, 1435, 1300, 1270 and 1165; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 2.17 and 2.21 (each 3 H, s, SMe and 5-Me), 2.53 and 2.82 (each 2 H, t, J 8, CH₂CH₂), 3.38 (2 H, s, CH₂CO₂), 3.66 and 3.67 (each 3 H, s, OMe) and 7.88 (1 H, br s, NH); m/z (EI) 285 (M⁺, 100%), 238 (55), 226 (35) and 212 (55).

9-Carboxy-1-methoxy-2,8-bis(2-methoxycarbonylethyl)-3,7bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin 63

A solution of benzyl ester 18 (126 mg, 197 μmol) in methanol (10 cm³) was stirred with 10% palladium-on-charcoal (20 mg) under hydrogen until uptake of gas ceased (ca. 1.5 h), then filtered and evaporated under reduced pressure. Purification by PLC, eluting with 5% ethanol in diethyl ether, gave acid 63 (94 mg, 87%) as a gum (Found: M⁺, 550.2149. $C_{26}H_{34}N_2O_{11}$ requires M, 550.2163); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3400–2400, 2940, 1730, 1660, 1435, 1260 and 1170; $\delta_{\rm H}({\rm CDCl_3}, 400~{\rm MHz})$ 1.11 (3 H, s, 4-Me), 2.30 and 3.02 (each 1 H, d, J 15, 5-H₂), 2.44-2.61 (6 H, m) and 2.99–3.03 (2 H, m, $2 \times CH_2CH_2$), 3.40 (2 H, s, CH₂CO₂), 3.42 and 3.47 (each 1 H, d, J 17, CH₂CO₂), 3.63, 3.63, 3.64, 3.70 and 3.95 (each 3 H, s, OMe) and 10.54 (1 H, br s, NH); m/z (FD) 550 (M⁺, 100%).

1-Methoxy-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin 64

A solution of carboxylic acid **63** (77 mg, 0.14 mmol) in dry dichloromethane (5 cm³) was stirred with toluene-*p*-sulfonic acid (39 mg, 0.21 mmol) under argon at room temperature for 24 h, then washed with water (3 × 5 cm³), dried and evaporated under reduced pressure. Purification by PLC, eluting with 10% ethanol in diethyl ether, gave the *methoxypyrrolenine* **64** (62 mg, 88%) as a gum (Found: M⁺, 506.2354. C₂₅H₃₄N₂O₉ requires *M*, 506.2264); $\nu_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3400, 2950, 1725s, 1680, 1440, 1265 and 1165; $\delta_{\text{H}}(\text{CDCl}_3$, 250 MHz) 1.02 (3 H, s, 4-Me), 2.18 and 2.96 (each 1 H, d, *J* 15, 5-H₂), 2.41–2.57 and 2.70–2.76 (8 H, m, 2 × CH₂CH₂), 3.37 (4 H, s, 2 × CH₂CO₂), 3.62, 3.63, 3.68 and 3.90 (each 3 H, s, OMe), 6.41 (1 H, d, *J* 2, 9-H) and 9.44 (1 H, br s, NH); *m/z* (FD) 506 (M⁺, 100%).

9-Benzyloxycarbonyl-4-[3-(2-methoxycarbonylethyl)-4-methoxycarbonylmethyl-5-(2,2,2-tribromoethoxycarbonyl)pyrrol-2-yl-methyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10*H*)-thione 66

Lactam 658 was reacted with Lawesson's reagent as described in the general procedure above. Purification by flash chromatography, eluting with 2% ethanol in diethyl ether, gave thiolactam 66 (71%) as a gum (Found: C, 47.1; H, 4.5; N, 3.5. $C_{46}H_{52}Br_3N_3O_{16}S$ requires C, 47.0; H, 4.5; N, 3.6%); v_{max} $(CH_2Cl_2)/cm^{-1}$ 3440, 3300, 2960, 1730s, 1700, 1430, 1250 and 1170; $\delta_{\rm H}({\rm CDCl_3}, 400 {\rm \, MHz}) \, 2.44 - 2.52 \, (9 {\rm \, H, \, m}), \, 2.68 - 2.72 \, (2 {\rm \, H, \, m})$ m) and 2.89–2.94 (2 H, m, 4-C H_AH_B and 2 × C H_2CH_2), 2.91 and 3.11 (each 1 H, d, J 15, 4-CH₂), 3.24 (1 H, d, J 15, 4-CH_AH_B), 3.41 (1 H, d, J 17, CH_AH_BCO₂), 3.58, 3.58, 3.64, 3.65, 3.70 and 3.74 (each 3 H, s, OMe), 3.58-3.74 (3 H, obscured, CH_AH_BCO₂ and CH₂CO₂), 3.87 and 3.90 (each 1 H, d, J 17, CH₂CO₂), 5.00–5.03 (2 H, m, CH₂CBr₃), 5.18 and 5.26 (each 1 H, d, J 12, CH₂Ph), 7.27-7.37 (5 H, m, Ph) and 9.35, 10.00 and 10.20 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 {\rm ~MHz})$ 19.15, 20.62, 29.26, 29.52, 29.58, 30.68, 30.92, 31.00, 31.18, 31.51, 34.50, 34.61 and 35.93 (CH₂ and CBr₃), 51.93, 51.48, 51.77, 51.93, 52.42 and 53.03 (OMe), 66.06 (CH₂Ph), 73.99 and 76.76 (C-4 and OCH₂CBr₃), 115.63, 118.11, 118.33, 122.72, 123.54, 128.79, 130.10, 135.82, 143.41 and 147.56 (C=C), 127.94, 128.09 and 128.32 (phenyl-CH), 158.52, 171.49, 173.36, 173.46 and 173.54 (C=O) and 197.40 (C=S); *m/z* (FD) 1171, 1173, 1175 and 1177 (1:3:3:1, M⁺, 100%).

Acid-catalysed fragmentation of the thiolactam acid 67 derived from tribromoethyl ester 66

A solution of the tribromoethyl ester **66** (126 mg, 0.107 mmol) in acetic acid (3 cm³) was stirred with zinc dust (260 mg) for 20 min and then filtered. The filtrate was diluted with water (10 cm³) and extracted with dichloromethane ($5 \times 10 \text{ cm}^3$). The combined extracts were washed with water ($2 \times 10 \text{ cm}^3$), dried and evaporated under reduced pressure. Purification by PLC, using methanol–diethyl ether (3:17), gave the acid **67** (87 mg, 89%); m/z (FD) 909.

A solution of acid **67** (50 mg) in dry dichloromethane (5 ml) was stirred at room temperature with toluene-*p*-sulfonic acid (13.6 mg) under argon in the dark for 20 h. TLC then showed essentially one product in addition to baseline material. The product was isolated by PLC, eluting with ethanol–diethyl ether (1:9), and identified as the dipyrromethane **68** by comparison with an authentic sample synthesised as below.

1-Benzyloxycarbonyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis-(methoxycarbonylmethyl)dipyrromethane 68

A solution of iodide 69^{11} (112 mg, 155 µmol) in methanol (10 cm³) was stirred with Adams catalyst (10 mg) and sodium acetate (100 mg) under hydrogen until uptake of gas ceased (*ca.* 2 h). The catalyst was then filtered off and the filtrate evaporated under reduced pressure. The residue was partitioned between dichloromethane (20 cm³) and water (20 cm³) and the

aqueous layer was further extracted with dichloromethane $(3 \times 10 \text{ cm}^3)$. The combined organic layers were dried and evaporated under reduced pressure. Purification by PLC, eluting with diethyl ether, gave the α-free dipyrromethane **68** (62 mg, 67%) as a light-sensitive gum (Found: M⁺, 596.2351. C₃₁H₃₆N₂O₁₀ requires M, 596.2370); $\nu_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3350, 2950, 1730, 1690, 1430, 1250 and 1170; $\delta_{\text{H}}(\text{CDCl}_3$, 400 MHz) 2.47–2.53 (4 H, m), 2.72 (2 H, t, J 8) and 3.00 (2 H, t, J 8, 2 × CH₂CH₂), 3.49 and 3.64 (each 2 H, s, CH₂CO₂), 3.57, 3.57, 3.60 and 3.74 (each 3 H, s, OMe), 3.79 (2 H, s, 5-H₂), 5.23 (2 H, s, CH₂Ph), 6.42 (1 H, d, J 2, 9-H), 7.26–7.40 (5 H, m, Ph) and 9.35 and 10.20 (each 1 H, br s, NH); m/z (FD) 596 (M⁺, 100%).

9-Carboxy-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10*H*)-one 73

A solution of benzyl ester 21 (3.95 g, 6.31 mmol) in methanol (50 cm³) was stirred with 10% palladium-on-charcoal (100 mg) in the dark under hydrogen until uptake of gas ceased (4 h), then filtered through Celite, washing with methanol (20 cm³), and evaporated under reduced pressure to give the acid 73 (2.80 g, 83%), mp 177-178 °C (from methanol) (Found: C, 55.8; H, 6.0; N, 5.1. $C_{25}H_{32}N_2O_{11}$ requires C, 56.0; H, 6.0; N, 5.2%); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 3200–2400, 2950, 1730s, 1680 and 1420; $\delta_{\rm H}({\rm CDCl_3}, 250 \ {\rm MHz})$ 1.01 (3 H, s, 4-Me), 2.59–2.64 and 3.04-3.07 (8 H, m, 2 × CH₂CH₂), 2.35 and 3.17 (each 1 H, d, J 15, 5-H₂), 3.47 (2 H, s, CH₂CO₂), 3.47 and 3.55 (each 1 H, d, J 16, CH₂CO₂), 3.62, 3.64, 3.65 and 3.72 (each 3 H, s, OMe) and 9.45 and 10.30 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 {\rm MHz})$ 19.99 (4-Me), 19.56, 20.72, 30.01, 31.01, 31.57, 34.23 and 34.77 (CH₂), 51.33, 51.54, 51.94 and 52.50 (OMe), 65.01 (C-4), 117.42, 120.34, 129.92, 130.90, 133.86 and 154.67 (C=C) and 165.44, 169.34, 171.95, 173.16, 173.71 and 174.25 (C=O); m/z (FD) 536 (M⁺, 100%).

9-Formyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10*H*)-one 74

A solution of carboxylic acid 73 (950 mg, 1.77 mmol) in redistilled trifluoroacetic acid (20 cm³) under argon in the dark was stirred at room temperature for 75 min, then cooled to 0 °C and treated with trimethyl orthoformate (20 cm³). After a further 40 min at 0 °C the solution was poured into 10% aqueous sodium carbonate (200 cm³) and extracted with dichloromethane $(4 \times 75 \text{ cm}^3)$. The combined extracts were dried and evaporated under reduced pressure. Purification by flash chromatography $(25 \times 2 \text{ cm})$, eluting with 0-4% ethanol in diethyl ether, gave aldehyde 74 (664 mg, 72%), which crystallised from dichloromethane-diethyl ether, mp 84-85 °C (Found: C, 57.4; H, 6.45; N, 5.1. $C_{25}H_{32}N_2O_{10}$ requires C, 57.7; H, 6.2; N, 5.4%); ν_{max} -(CH₂Cl₂)/cm⁻¹ 3300, 2950, 1740s, 1690, 1440 and 1100; $\delta_{\rm H}({\rm CDCl_3}, 250 {\rm ~MHz}) 1.31 (3 {\rm ~H, s, 4-Me}), 2.49-2.57 (6 {\rm ~H, m})$ and 2.99 (2 H, t, J 8, 2 × CH₂CH₂), 2.76 and 3.05 (each 1 H, d, J 15, 5-H₂), 3.37 and 3.49 (each 1 H, d, J 16, CH₂CO₂), 3.40 and 3.58 (each 1 H, d, J 17, CH₂CO₂), 3.63, 3.72 and 3.77 (12 H, each s, 4 × OMe), 7.41 (1 H, br s, lactam-NH), 9.51 (1 H, s, CHO) and 10.62 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 \,{\rm MHz})$ 23.25 (4-Me), 19.25, 19.67, 29.65, 30.74, 31.37, 33.85 and 35.77 (CH₂), 51.46, 51.52, 52.27 and 52.66 (OMe), 63.44 (C-4), 116.15, 128.75, 133.39, 133.85, 135.38 and 151.36 (C=C), 170.39, 171.69, 172.50, 172.78 and 173.39 (C=O) and 177.38 (CHO); m/z (FD) 520 (M⁺, 100%).

Reaction of formyl lactam 74 with Lawesson's reagent

Formyl lactam **74** (27 mg, 52 µmol) was reacted with Lawesson's reagent following the general procedure described above. The residue was purified by PLC, eluting with ethanolether (1:9), to give two higher $R_{\rm f}$ compounds: (i) at lower $R_{\rm f}$, thioformyl lactam **75** (15 mg, 53%) as an oil; $\nu_{\rm max}({\rm CH_2Cl_2})/{\rm cm}^{-1}$ 3300, 2950, 1735s, 1700, 1440 and 1150; $\delta_{\rm H}({\rm CDCl_3}, 250~{\rm MHz})$ 1.38 (3 H, s, 4-Me), 2.46–2.60 (6 H, m) and 2.97 (2 H, t, J 7, 2 × CH₂CH₂), 2.81 and 3.00 (each 1 H, d, J 15, 5-H₂), 3.33 and

3.70 (each 1 H, d, J 17, CH₂CO₂), 3.38 and 3.54 (each 1 H, d, J 16, CH₂CO₂), 3.64, 3.75 and 3.80 (12 H, each s, $4 \times$ OMe), 7.15 (1 H, br s, lactam-NH), 10.21 (1 H, br s, pyrrole-NH) and 10.57 (1 H, s, CHS); m/z (FD) 536 (M⁺, 100%); (ii) at higher $R_{\rm f}$, thioformyl thiolactam **79** (7 mg, 24%) as a gum (Found: M⁺, 522.1543. C₂₅H₃₂N₂O₈S₂ requires M, 552.1600); $\nu_{\rm max}$ (CH₂Cl₂)/cm⁻¹ 3250, 2950, 1730s, 1450, 1300, 1140 and 990; $\delta_{\rm H}$ (CDCl₃, 250 MHz) 1.43 (3 H, s, 4-Me), 2.49–2.82 and 2.93–2.98 (8 H, m, 2 × CH₂CH₂), 2.87 and 3.03 (each 1 H, d, J 15, 5-H₂), 3.34 and 3.58 (each 1 H, d, J 17, CH₂CO₂), 3.39 and 3.79 (each 1 H, d, J 16, CH₂CO₂), 3.65, 3.81 and 3.82 (12 H, each s, J 4 × OMe), 9.03 and 10.23 (each 1 H, br s, NH) and 10.60 (1 H, s, CHS); m/z (FD) 552 (M⁺, 100%).

Reaction of formyl lactam **74** (114 mg, 220 μmol) with Lawesson's reagent (91 mg, 220 μmol) as above gave thioformyl thiolactam **79** (51%).

$\hbox{$3$-(2-Methoxycarbonylethyl)-4-methoxycarbonylmethyl-5-methyl-2-thioformylpyrrole } 60$

A solution of aldehyde 59¹³ (260 mg, 0.97 mmol) in dry benzene (10 ml) was heated at reflux with Lawesson's reagent (215 mg, 0.53 mmol) under argon in the dark for 45 min and then evaporated under reduced pressure. Purification by flash chromatography $(20 \times 2 \text{ cm})$, eluting with dichloromethane, gave thioaldehyde 60 (198 mg, 72%) as red blocks, mp 112-113 °C (from diethyl ether-hexane) (Found: C, 55.0; H, 6.1; N, 4.85; S, 11.3%; M⁺, 283.0874. C₁₃H₁₇NO₄S requires C, 55.1; H, 6.05; N, 4.9; S, 11.3%; M, 283.0878); $\lambda_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{nm}$ 399 (strong) and 310; $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 2940, 1740s, 1490, 1430 and 1160; $\delta_{\rm H}({\rm CDCl_3}, 400~{\rm MHz})$ 2.24 (3 H, s, 5-Me), 2.48 and 3.00 (each 2 H, m, CH₂CH₂), 3.44 (2 H, s, CH₂CO₂), 3.68 and 3.71 (each 3 H, s, OMe), 9.35 (1 H, br s, NH) and 10.50 (1 H, s, CHS); $\delta_{\rm C}$ (CDCl₃, 62.5 MHz) 12.23 (5-Me), 19.60, 29.66 and 35.49 (CH₂), 51.67 and 52.06 (OMe), 117.60, 133.04, 140.10 and 142.10 (pyrrole-C), 171.24 and 172.54 (C=O) and 194.42 (CH=S); m/z 283 (M⁺, 100%), 250, 224, 209, 164 and 150.

9-Formyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10H)-thione 80

A solution of thioformyl thiolactam 79 (51 mg, 92 µmol) in dimethylformamide (2 cm³) was stirred with morpholine (2 drops) and water (10 drops) under argon in the dark for 20 h, then evaporated under reduced pressure. Purification by PLC, eluting with 7% ethanol in diethyl ether, gave formyl thiolactam **80** (26 mg, 53%) as a gum (Found: M^+ , 536.1835. $C_{25}H_{32}N_2O_9S$ requires M, 536.1829); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 2950, 1730s, 1695, 1440, 1175 and 1090; $\delta_{\rm H}({\rm CDCl_3},\,400~{\rm MHz})$ 1.36 (3 H, s, 4-Me), 2.54-2.74 (6 H, m) and 3.01 (2 H, t, J 7, 2 × CH, CH,), 2.81 and 3.07 (each 1 H, d, J 15, 5-H₂), 3.42 and 3.52 (each 1 H, d, J 16, CH₂CO₂), 3.46 and 3.62 (each 1 H, d, J 18, CH₂CO₂), 3.64, 3.65, 3.77 and 3.78 (each 3 H, s, OMe), 9.43 (1 H, br s, thiolactam-NH), 9.47 (1 H, s, CHO) and 10.68 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 \, {\rm MHz}) \, 22.05 \, (4-{\rm Me}), \, 19.19, \, 20.90,$ 29.80, 31.08, 31.93, 33.13 and 35.97 (CH₂), 51.54, 51.63, 52.64 and 52.86 (OMe), 71.08 (C-4), 115.88, 128.86, 133.43, 133.63, 141.43 and 149.79 (C=C), 170.56, 172.80, 172.91 and 173.73 (C=O), 177.31 (CHO) and 196.44 (C=S); m/z (FD) 536 (M⁺,

9-Formyl-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin 82

A solution of thiolactam **80** (45 mg, 84 µmol) in methanol (3 cm³) was heated at reflux with freshly prepared Raney nickel (one small spatula) for 20 min, then filtered, washing with methanol, and evaporated under reduced pressure. The residue was purified by PLC to give the *pyrrolenine* **82** (15 mg, 52% based on unrecovered **80**) as a gum (Found: M^+ , 504.2110. $C_{25}H_{32}N_2O_9$ requires M, 504.2108); $\nu_{max}(CH_2Cl_2)/cm^{-1}$ 3300, 2950, 1730s, 1640, 1570, 1435, 1260, 1200 and 1170; $\delta_H(CDCl_3)$

400 MHz) 1.09 (3 H, s, 4-Me), 2.21 and 3.14 (each 1 H, d, J 15, 5-H₂), 2.48–2.52 and 2.56–2.65 (8 H, m, 2 × CH₂CH₂), 3.39 and 3.45 (each 1 H, d, J 16, CH₂CO₂), 3.43 and 3.48 (each 1 H, d, J 16, CH₂CO₂), 3.64, 3.65, 3.67 and 3.73 (each 3 H, s, OMe), 7.94 (1 H, s, 2-H), 9.56 (1 H, s, CHO) and 10.44 (1 H, br s, NH); m/z (FD) 504 (M⁺, 100%).

9-Cyano-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10*H*)-one 78

A solution of formyl lactam **74** (191 mg, 0.367 mmol) in methanol (10 cm³) was treated with hydroxylamine hydrochloride (45 mg, 0.65 mmol), sodium acetate (54 mg, 0.66 mmol) and water (10 drops), heated at reflux under argon for 45 min and then evaporated under reduced pressure. The residue was partitioned between dichloromethane (20 cm³) and water (20 cm³) and the aqueous layer was extracted with dichloromethane (3 × 20 cm³). The combined organic layers were dried and evaporated under reduced pressure to give crude oximes **76**.

Without further purification, the oximes 76 were dissolved in dry acetonitrile (3 cm³) and added to a mixture of dimethylformamide (71 mg, 75 µl, 0.97 mmol) and freshly distilled phosphorus oxychloride (120 mg, 0.80 mmol) in dry acetonitrile (2 cm³) at -23 °C. The solution was stirred at -23 °C for 20 min and then at room temperature for 1 h, poured into 5% aqueous sodium hydrogen carbonate (25 cm³) and extracted with dichloromethane (4 × 20 cm³). The combined extracts were dried and evaporated under reduced pressure. Purification by PLC, eluting with 10% ethanol in diethyl ether, gave the nitrile 78 (142 mg, 74%) as a gum (Found: M+, 517.2065. $C_{25}H_{31}N_3O_9$ requires M, 517.2060); $v_{max}(CH_2Cl_2)/Cl_2$ cm⁻¹ 3300, 2960, 2210, 1725s, 1690, 1435, 1100 and 1065; $\delta_{\rm H}({\rm CDCl_3}, 400 {\rm \ MHz})$ 1.29 (3 H, s, 4-Me), 2.50–2.66 and 2.81– 2.85 (8 H, m, $2 \times \text{CH}_2\text{CH}_2$), 2.77 and 3.02 (each 1 H, d, J 15, 5-H₂), 3.33 and 3.47 (each 1 H, d, J 16, CH₂CO₂), 3.40 and 3.59 (each 1 H, d, J 17, CH₂CO₂), 3.64, 3.65, 3.70 and 3.77 (each 3 H, s, OMe), 7.53 (1 H, br s, lactam-NH) and 10.43 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3}, 100 \text{ MHz}) 23.15 (4-Me), 19.53, 20.12,$ 29.62, 30.59, 31.15, 33.36 and 34.27 (CH₂), 51.55, 51.64, 52.24 and 52.74 (OMe), 63.63 (C-4), 98.93, 113.95, 114.36, 130.29, 132.67, 135.39 and 151.55 (C=C and C=N) and 170.74, 171.89, 172.28, 172.81 and 173.70 (C=O); m/z (FD) 517 (M⁺, 100%).

9-Cyano-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin-1(10*H*)-thione 81

Lactam 78 was reacted with Lawesson's reagent according to the above general procedure. The residue was purified by flash chromatography, eluting with diethyl ether, to give thiolactam **81** (88%) as a gum (Found: M^+ , 533.1820. $C_{25}H_{31}N_3O_8S$ requires M, 533.1832); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3300, 2960, 2220, 1730s, 1450 and 1000; $\delta_{H}(CDCl_3, 400 \text{ MHz})$ 1.41 (3 H, s, 4-Me), 2.52-2.81 (6 H, m) and 2.84 (2 H, t, J 8, $2 \times CH_2CH_2$), 2.91 and 3.05 (each 1 H, d, J 15, 5-H₂), 3.33 and 3.53 (each 1 H, d, J 16, CH₂CO₂), 3.39 (1 H, d, J 17, CH_AH_BCO₂), 3.64, 3.68, 3.78 and 3.80 (each 3 H, s, $4 \times OMe$), 3.64–3.80 (1 H, obscured, CH_A- H_BCO_2), 8.95 (1 H, br s, lactam-NH) and 10.18 (1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3},\ 100\ {\rm MHz})\ 23.10\ ({\rm C-4}),\ 20.16,\ 21.14,$ 29.77, 30.84, 31.45, 32.72 and 34.53 (CH₂), 51.79, 51.94, 52.93 and 53.16 (OMe), 70.96 (C-4), 98.81, 113.79, 129.97, 132.92, 142.2 and 148.97 (C=C and C=N), 171.67, 172.94, 173.42 and 174.52 (C=O) and 196.19 (C=S); m/z (FD) 533 (M⁺, 100%).

9-Cyano-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4-methyl-4,5-dihydrodipyrrin 83

Thiolactam **81** was reduced with nickel boride according to the general procedure above. Purification by PLC, eluting with ethanol–diethyl ether (1:9), gave *pyrrolenine* **83** (40% based on unrecovered **81**) as a gum (Found: M⁺, 501.2108. $C_{25}H_{31}N_3O_8$ requires M, 501.2111); $v_{max}(CH_2Cl_2)/cm^{-1}$ 3350, 2960, 2220, 1725s, 1690, 1440 and 1150; $\delta_H(CDCl_3, 400 \text{ MHz})$ 1.07 (3 H, s,

4-Me), 2.14 and 3.10 (each 1 H, d, J 15, 5-H₂), 2.51 and 2.66 (6 H, m) and 2.86 (2 H, t, J 8, 2 × CH₂CH₂), 3.40–3.43 (4 H, m, 2 × CH₂CO₂), 3.66, 3.67 and 3.73 (12 H, each s, 4 × OMe), 7.95 (1 H, s, 2-H) and 10.45 (1 H, br s, NH); $\delta_{\rm C}$ (CDCl₃, 100 MHz) 19.39 (4-Me), 20.28, 20.51, 29.71, 31.21, 32.24, 32.66 and 34.47 (CH₂), 51.50, 51.70, 51.87 and 52.35 (OMe), 83.3 (C-4), 97.88, 114.04, 114.7, 132.62, 132.92, 137.6 and 158.0 (C=C and C=N), 164.9 (C-2), 170.08, 171.65, 171.71 and 172.71 (C=O), m/z (FD) 501 (M⁺, 100%).

9-Formyl-4-[5-formyl-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10*H*)-one 86

Hydrogenation of dibenzyl ester 29 and decarboxylationformylation of the resulting diacid 85 was carried out using similar procedures to those used for $21 \rightarrow 73 \rightarrow 74$ except that the hydrogenation required 3 h and decarboxylation required 6.5 h. Purification by flash chromatography, eluting with 0-5% ethanol in diethyl ether, gave the dialdehyde 86 (72% over both steps) as a gum (Found: M⁺, 771.2862. C₃₇H₄₅N₃O₁₅ requires M, 771.2851); $v_{\text{max}}(\text{CH}_2\text{Cl}_2)/\text{cm}^{-1}$ 3280, 2950, 1730, 1640, 1460, 1250 and 1070; $\delta_{\rm H}({\rm CDCl_3},\,400~{\rm MHz})$ 2.41–2.55 (6 H, m), 2.72 (2 H, t, J7) and 2.91-2.98 $(4 \text{ H}, \text{ m}, 3 \times \text{CH}_2\text{CH}_2)$, 2.52 and 3.18(each 1 H, d, J 15, 4-CH₂), 2.85 and 3.08 (each 1 H, d, J 15, 4-CH₂), 3.25 and 3.47 (each 1 H, d, J 16, CH₂CO₂), 3.62, 3.63, $3.64, 3.67, 3.68 \text{ and } 3.83 \text{ (each 3 H, s, 6} \times \text{OMe)}, 3.62-3.83 \text{ (4 H, s, 6} \times \text{OMe)}$ obscured, 2 × CH₂CO₂), 7.63 (1 H, br s, lactam-NH), 9.50 (2 H, s, $2 \times CHO$) and 10.14 and 10.46 (each 1 H, br s, pyrrole-NH); $\delta_{\rm C}({\rm CDCl_3}, 62.5 {\rm \ MHz})$ 19.10, 19.26, 19.68, 29.60, 30.93, 31.03, 31.73, 31.96, 32.10 and 34.64 (CH₂), 51.56, 51.68, 52.19, 52.29 and 53.01 (OMe), 66.66 (C-4), 116.30, 122.96, 129.05, 129.75, 132.03, 132.73, 137.40 and 149.83 (C=C), 170.46, 171.11, 172.29, 172.87, 173.31 and 173.38 (C=O) and 177.60 and 177.79 (CHO); m/z (FD) 771 (M⁺, 100%).

9-Cyano-4-[5-cyano-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10*H*)-one 87

Dialdehyde 86 was reacted with hydroxylamine hydrochloride and then phosphorus oxychloride following an analogous procedure to that employed for 74-76-78. Purification by flash chromatography, eluting with 2% ethanol in diethyl ether, gave the dinitrile 87 (50%) as a gum (Found: M⁺, 765.2845. $C_{37}H_{43}N_5O_{13}$ requires M, 765.2857); $v_{\text{max}}(CH_2Cl_2)/cm^{-1}$ 3300, 2960, 2220, 1725s, 1695, 1435, 1200 and 1170; $\delta_{\rm H}({\rm CDCl_3}, 400$ MHz) 2.39–2.60 (8 H, m), 2.67 (2 H, t, J 7) and 2.78 (2 H, t, J 8, $3 \times \text{CH}_2\text{CH}_2$), 2.71 and 3.15 (each 1 H, d, J 15, 4-CH₂), 2.82 and 3.00 (each 1 H, d, J 15, 4-CH₂), 3.18 and 3.43 (each 1 H, d, J 16, CH₂CO₂), 3.55 (2 H, s, CH₂CO₂), 3.64, 3.64, 3.68, 3.69, 3.69 and 3.83 (each 3 H, s, $6 \times OMe$), 3.64-3.83 (2 H, obscured, CH₂CO₂) and 7.74, 9.97 and 10.21 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100 \ {\rm MHz})$ 19.25, 19.90, 20.20, 29.30, 30.63, 30.68, 30.87, 32.24 and 34.33 (CH₂), 51.69, 51.83, 52.06, 52.22, 52.55 and 53.31 (OMe), 66.38 (C-4), 99.17, 100.72, 113.66, 113.91, 114.65, 121.25, 125.89, 128.29, 128.92, 129.68, 132.87, 137.76 and 149.90 (C=C and C=N), 170.83, 171.70, 172.15, 172.87, 172.89, 173.59 and 174.21 (C=O); m/z (FD) 765 (M⁺, 100%).

9-Cyano-4-[5-cyano-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin-1(10*H*)-thione 88

Lactam **87** was reacted with Lawesson's reagent following the general procedure described above. Purification by flash chromatography, eluting with 2% ethanol in diethyl ether, gave the *thiolactam* **88** (67%) as a gum (Found: M⁺, 781.2632. $C_{37}H_{43}N_5O_{12}S$ requires M, 781.2629); $v_{max}(CH_2Cl_2)/cm^{-1}$ 3300,

2950, 2220, 1730s, 1440, 1270 and 1000; $\delta_{\rm H}({\rm CDCl_3}, 400~{\rm MHz})$ 2.49–2.82 (13 H, m, 3 × CH₂CH₂ and 4-CH_AH_B), 2.89 and 3.10 (each 1 H, d, *J* 15, 4-CH₂), 3.22 and 3.50 (each 1 H, d, *J* 16, CH₂CO₂), 3.29 (1 H, d, *J* 15, 4-CH_AH_B), 3.56 (2 H, s, CH₂CO₂), 3.64, 3.67, 3.70, 3.73, 3.75 and 3.86 (each 3 H, s, 6 × OMe), 3.64–3.86 (2 H, obscured, CH₂CO₂), 9.31, 9.61 and 10.07 (each 1 H, br s, NH); $\delta_{\rm C}({\rm CDCl_3}, 100~{\rm MHz})$ 19.17, 20.17, 21.25, 29.28, 30.34, 30.72, 30.85, 31.11, 31.74, 34.15 and 34.47 (CH₂), 51.79, 52.25, 52.32, 52.87 and 53.49 (OMe), 73.48 (C-4), 99.15, 100.90, 113.58, 113.82, 114.31, 121.33, 126.03, 128.46, 129.26, 133.01, 143.81 and 147.91 (C=C and C=N), 170.84, 172.06, 172.86, 173.36, 173.93 and 174.85 (C=O) and 197.20 (C=S); *m/z* (FD) 781 (M⁺, 100%).

9-Cyano-4-[5-cyano-3-(2-methoxycarbonylethyl)-4-(methoxycarbonylmethyl)pyrrol-2-ylmethyl]-2,8-bis(2-methoxycarbonylethyl)-3,7-bis(methoxycarbonylmethyl)-4,5-dihydrodipyrrin 89

Thiolactam **88** was reduced with nickel boride following the general procedure described above, except that the reaction time was increased to 2.5 h. Purification by normal phase HPLC on a Kontron S5W column, eluting with hexane–ethyl acetate (1:1), gave the *pyrrolenine* **89** (14%) as a gum (Found: M^+ , 749.2908. $C_{37}H_{43}N_5O_{12}$ requires M, 749.2908); $\nu_{max}(CH_2-Cl_2)/cm^{-1}$ 3300, 2950, 2220, 1730s, 1695, 1440, 1200 and 1175; $\delta_H(CDCl_3, 400 \text{ MHz})$ 2.29 and 3.19 (each 1 H, d, J 15, 4-CH_AH_B), 2.41–2.50, 2.55–2.65 and 2.86 (13 H, m, 3 × CH₂CH₂ and 4-CH_AH_B), 3.07 (1 H, d, J 15, 4-CH_AH_B), 3.29 and 3.40 (each 1 H, d, J 16, CH₂CO₂), 3.49 and 3.58 (each 1 H, d, J 17, CH₂CO₂), 3.59 (2 H, s, CH₂CO₂), 3.63, 3.65, 3.67, 3.70, 3.74 and 3.83 (each 3 H, s, OMe), 7.81 (1 H, s, N=C-H) and 10.16 and 10.35 (each 1 H, br s, NH); m/z (FD) 749 (M⁺, 100%).

Hydrolysis of the ester groups of pyrrolenines 83 and 89

A solution of pyrrolenine **83** (44 mg, 88 µmol) in dry degassed methanol (1 cm³) was treated with degassed aqueous potassium hydroxide (4 mol dm⁻³; 1 cm³) and then shaken under argon in the dark at room temperature for 15 h. A sample was then removed and prepared for NMR analysis by four times evaporating *in vacuo* and redissolving the residue in D₂O (1 cm³); $\delta_{\rm H}({\rm D_2O}, 400 \ {\rm MHz}, {\rm relative to HOD signal at 4.77 ppm) 1.14 (3 H, s, 4-Me), 2.12, 2.25, 2.43, 2.62 (each 2 H, t,$ *J*8, 2 × CH₂CH₂), 2.60 and 3.01 (each 1 H, d,*J*15, 5-H₂), 3.18 and 3.27 (each 2 H, d,*J*16, 2 × CH₂CO₂) and 7.79 (1 H, s, 2-H).

The remainder of the solution was then washed with diethyl ether $(2 \times 2 \text{ cm}^3)$ and evaporated *in vacuo*. The residue was dissolved in acetic acid (400 µl), and methanol (4 cm^3) and diethyl ether (10 cm^3) were added. The mixture was treated with diazomethane at room temperature until the yellow colour persisted, stirred for a further hour and then evaporated under reduced pressure. Purification by PLC gave the starting pyrrolenine **83** as the major product (20 mg, 46% recovery).

For enzymatic studies, the pyrrolenine **89** was hydrolysed under exactly the same conditions.

Crystal data for 60§

C₁₃H₁₇NO₄S, M = 283.34. Monoclinic, a = 8.367(4), b = 19.764(8), c = 18.672(4) Å, $\beta = 95.16(3)^\circ$, V = 1428.2(11) ų [from centring angles for 25 reflections $(40 \le 2\theta \le 50^\circ$, $\lambda = 1.541$ 84 Å, T = 290 K)], space group $P2_1/a$ (Alt. $P2_1/c$, No. 14), Z = 4, $D_x = 1.318$ g cm⁻³, red block, $0.38 \times 0.35 \times 0.19$ mm, μ (Cu-Kα) = 2.111 mm⁻¹.

§ Full crystallographic details, excluding structure factor tables, have been deposited at the Cambridge Crystallographic Data Centre (CCDC). For details of the deposition scheme, see 'Instructions for Authors', *J. Chem. Soc.*, *Perkin Trans. 1*, available *via* the RSC Web page (http://www.rsc.org/authors). Any request to the CCDC for this material should quote the full literature citation and the reference number 207/200.

Data collection and processing

Siemens P3 diffractometer, $\omega/2\theta$ scans, graphite-monochromated Cu-Ka X-radiation; 2255 reflections measured $(9 \le 2\theta \le 115^{\circ})$, 1959 unique [merging R = 0.018], giving 1809 with $F \ge 4\sigma(F)$ and 1959 which were retained in all calculations. No crystal decay was observed and no corrections were applied for absorption.

Structure solution and refinement

Automatic direct methods 35 (all non-H atoms). Full-matrix least-squares refinement 35 with all non-H atoms anisotropic; H atoms were included at geometrically calculated positions and thereafter allowed to ride on their respective parent atoms at a distance of 0.98 Å. The weighting scheme selected gave satisfactory agreement analyses. Final R_f $[F \ge 4\sigma(F)] = 0.0459$, wR_2 [all data] = 0.1310, $S[F^2]$ = 1.03 for 187 refined parameters. An extinction correction³⁵ refined to 0.0065(10) and the final ΔF synthesis showed no peaks above 0.33 e Å^{-3} .

Crystal data for 77 §

 $C_{27}H_{35}N_3O_{11}$, M = 577.58. Monoclinic, a = 7.517(3), b =18.551(5), c = 20.760(8) Å, $\beta = 94.30(3)^{\circ}$, $V = 2887(2) \text{ Å}^3$ [from centring angles for 25 reflections ($40 \le 2\theta \le 50^{\circ}$, $\lambda = 1.541 84$ Å, T = 290 K], space group $P2_1/c$ (No. 14), Z = 4, $D_x = 1.329 \text{ g}$ cm⁻³, colourless block, $0.41 \times 0.38 \times 0.25$ mm, μ (Cu-K α) = 0.875 mm^{-1} .

Data collection and processing

Siemens P3 diffractometer, $\omega/2\theta$ scans, graphite-monochromated Cu-Ka X-radiation; 4195 reflections measured $(6 \le 2\theta \le 115^{\circ})$, 3938 unique [merging R = 0.013], giving 3441 with $F \ge 4\sigma(F)$ and 3938 which were retained in all calculations. No crystal decay was observed and no corrections were applied for absorption.

Structure solution and refinement

Automatic direct methods³⁵ (all non-H atoms). Full-matrix least-squares refinement 35 with all non-H atoms anisotropic; H atoms were included at geometrically calculated positions and thereafter allowed to ride on their respective parent atoms at a distance of 0.98 Å. The weighting scheme selected gave satisfactory agreement analyses. Final R $[F \ge 4\sigma(F)] = 0.0437$, wR_2 [all data] = 0.1284, $S[F^2] = 1.03$ for 391 refined parameters. The final ΔF synthesis showed no peaks above 0.38 e Å⁻³.

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References

1 The following paper is regarded as Part 46: N. P. J. Stamford, J. Crouzet, B. Cameron, A. I. D. Alanine, A. R. Pitt, A. A. Yeliseev and A. R. Battersby, Biochem. J., 1996, 313, 335.

- 2 Preliminary account of part of this work: C. J. Hawker, W. M. Stark and A. R. Battersby, J. Chem. Soc., Chem. Commun., 1987, 1313.
- 3 F. J. Leeper and A. R. Battersby, Chem. Rev., 1990, 90, 1261; F. J. Leeper, Nat. Prod. Rep., 1989, 6, 171.
- 4 A. R. Battersby, G. L. Hodgson, E. Hunt, E. McDonald and J. Saunders, J. Chem. Soc., Perkin Trans. 1, 1976, 273
- 5 A. R. Battersby, C. J. R. Fookes, M. J. Meegan, E. McDonald and H. K. W. Wurziger, J. Chem. Soc., Perkin Trans. 1, 1982, 2786.
- 6 A. R. Battersby, M. G. Baker, H. A. Broadbent, C. J. R. Fookes and F. J. Leeper, J. Chem. Soc., Perkin Trans. 1, 1987, 2027.
- 7 J. H. Mathewson and A. H. Corwin, J. Am. Chem. Soc., 1961, 83,
- 8 W. M. Stark, C. J. Hawker, G. J. Hart, A. Philippides, P. M. Petersen, J. D. Lewis, F. J. Leeper and A. R. Battersby, J. Chem. Soc., Perkin Trans. 1, 1993, 2875; W. M. Stark, G. J. Hart and A. R. Battersby, J. Chem. Soc., Chem. Commun., 1986, 465.
- 9 M. A. Cassidy, N. Crockett, F. J. Leeper and A. R. Battersby, J. Chem. Soc., Perkin Trans. 1, 1996, 2079.

 10 A. C. Spivey, A. Capretta, C. S. Frampton, F. J. Leeper and A. R.
- Battersby, J. Chem. Soc., Perkin Trans. 1, 1996, 2091.
- 11 C. J. Hawker, A. C. Spivey, F. J. Leeper and A. R. Battersby, J. Chem. Soc., Perkin Trans. 1, 1998, 1509.
- 12 A. R. Battersby, C. J. R. Fookes, K. E. Gustafson-Potter, E. McDonald and G. W. J. Matcham, J. Chem. Soc., Perkin Trans. 1, 1982, 2427.
- 13 W. M. Stark, M. G. Baker, F. J. Leeper, P. R. Raithby and A. R. Battersby, J. Chem. Soc., Perkin Trans. 1, 1988, 1187.
- 14 D. Liotta, Organoselenium Chemistry, Wiley, Chichester, 1987,
- p. 325. 15 S. Karady, J. S. Amato, L. M. Weinstock and M. Sletzinger, Tetrahedron Lett., 1978, 407.
- 16 H. J. Reich and M. L. Cohen, J. Org. Chem., 1979, 44, 3148.
- 17 C. J. Hawker, A. Philippides and A. R. Battersby, J. Chem. Soc., Perkin Trans. 1, 1991, 1833.
- 18 G. R. Pettit and E. E. Van Tamelen, Org. React., 1962, 12, 356; J. S. Pizey, Synthetic Reagents, Ellis Horwood, Chichester, 1974, vol. 2, p. 175.
- 19 B. S. Petersen and S.-O. Lawesson, Tetrahedron, 1979, 35, 2433.
- 20 H. Davy, J. Chem. Soc., Chem. Commun., 1982, 457.
- 21 R. Paul, P. Buisson and N. Joseph, Ind. Eng. Chem., 1952, 44, 1006.
- 22 I. Ojima, S. Inaba and K. Nakatsugawa, Synthesis, 1976, 207.
- 23 A. R. Battersby, S. Kishimoto, E. McDonald, F. Satoh and H. K. W. Wurziger, J. Chem. Soc., Perkin Trans. 1, 1979, 1927.
- 24 R. L. N. Harris, *Aust. J. Chem.*, 1970, **23**, 1199; 1972, **25**, 985. 25 A. R. Battersby, E. Hunt, E. McDonald, J. B. Paine III and J. Saunders, J. Chem. Soc., Perkin Trans. 1, 1976, 1008.
- 26 D. Mauzerall, J. Am. Chem. Soc., 1960, 82, 2601, 2605.
- 27 P. Wipf, C. Jenny and H. Heinzgartner, Helv. Chim. Acta, 1987, 70,
- 28 R. Mayer, J. Morgensten and J. Fabian, Angew. Chem., Int. Ed. Engl., 1964, 3, 277
- 29 M. G. Baker, PhD Thesis, Cambridge, 1985.
- 30 G. P. Arsenault and S. F. MacDonald, Can. J. Chem., 1961, 39, 2043.
- 31 G. J. Hart and A. R. Battersby, Biochem. J., 1985, 232, 151.
- 32 A. D. Miller, F. J. Leeper and A. R. Battersby, J. Chem. Soc., Perkin Trans. 1, 1989, 1943
- 33 A. R. Battersby, M. Ihara, E. McDonald, J. Saunders and R. J. Wells, J. Chem. Soc., Perkin Trans. 1, 1976, 283. 34 W. J. Gensler, F. A. Johnson and D. B. Sloan, J. Am. Chem. Soc.,
- 1960, 82, 6074.
- 35 SHELXTL-Plus Release 5.0. Siemens Analytical X-ray Instruments, Inc., Madison, WI, USA, 1993.

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