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Identification of Fluorinated (*R*)-(-)-Aporphine Derivatives as Potent and Selective Ligands at Serotonin 5-HT_{2C} Receptor

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Abstract: A series of novel aporphine derivatives were synthesized for initial screening at the 5-HT₂ receptor subtypes. Among them, Compounds **11a** and **11b** were identified as potent 5-HT_{2C} hit ligands with highly selectivity over other 5-HT₂ receptor subtypes. Molecular docking study revealed that compounds **11a** and **11b** formed two key interactions with the binding site of 5-HT_{2C} receptor, including a saltbridge to D3.32 and a H-bond interaction with N6.55.

Keywords: Aporphine derivatives; 5-HT_{2C} receptor; Hit ligands; Molecular docking

The serotonin (5-hydroxytryptamine, 5-HT) system is known to play a critical role in the regulation of numerous neurological functions.^{1,2} The current classification of the serotonin receptor family includes at least fourteen receptor subtypes classified into seven major classes (5-HT₁₋₇) based upon the conjunction of genetic and

molecular structure, intracellular transduction mechanisms, and pharmacological criteria.³ All of these serotonin receptor subtypes are G-protein-coupled receptors (GPCR) with the exception of the ligand-gated ion channel 5-HT₃.⁴ The 5-HT₂ receptor subtype family is composed of three GPCR members (5-HT_{2A}, 5-HT_{2B}, 5-HT_{2C}) which share a high level of amino acid sequence homology within transmembrane regions.⁵ In particular, the 5- HT_{2C} receptor is predominantly expressed in the central nervous system (CNS), and has been identified as a promising therapeutic target for the treatment of obesity and other CNS disorders, such as depression, schizophrenia and drug addiction.⁶⁻⁹ To date, a number of 5-HT_{2C} ligands have been disclosed (Fig. 1). Among them, lorcaserin was approved by the FDA as a first-in-class antiobesity drug in 2012.¹⁰ Its efficacy in treatment of nicotine addiction is currently being evaluated in clinical trials.¹¹ Despite dramatic advancements in 5-HT_{2C} ligands development, attaining high 5-HT_{2C} receptor selectivity remains an attractive research area, as non-specific interaction with the other two 5-HT₂ receptors, 5-HT_{2A} and 5-HT_{2B} receptors, results in hallucinations and potentially fatal fibrotic cardiac valvulopathy, respectively.¹²⁻¹⁴ A major challenge to develop selective 5-HT_{2C} ligands involves the highly conserved ligand recognition transmembrane region of the 5-HT₂ receptor subtypes.⁴ Therefore, it is difficult to design a ligand exhibiting high specificity for the 5-HT_{2C} receptor.



Fig. 1. Structures of some representative 5-HT_{2C} receptor ligands.

Aporphines are a group of tetrahydroisoquinoline alkaloids which exhibit a wide range of pharmacological activities.¹⁵⁻²⁰ In the CNS, aporphines have been extensively explored as ligands for serotonin, adrenergic and dopamine receptors.²¹⁻²⁴ Recently, Liu and co-workers identified several aporphine derivatives as 5-HT_{2C} receptor hit ligands by screening a focused target-specific natural compound library, indicating that aporphines can be a viable source of novel ligands for 5-HT_{2C} receptor.²⁵ Previously reported structure-activity relationship (SAR) studies on aporphines indicated that introducing a more lipophilic group at C11 position leads to the compound possessing high affinity toward the serotonin receptors with a complete loss of affinity for the dopamine receptors.²⁶ Inspired by these valuable results and aiming to develop potent 5-HT_{2C} ligands with highly selectivity, a series of novel aporphine derivatives with fluorinated alkyl, benzyl or benzoyl groups attached to C11 through an O-linkage were synthesized for initial screening at the 5-HT₂ receptor subtypes (Fig. 2). Generally, introduction of a fluorine atom into bioactive molecules is well-established strategy for improving the target affinity.^{27,28} Herein, we wish to report the synthetic and pharmacological experiments of these novel aporphine derivatives.



Fig. 2. Fluorinated aporphine derivatives for initial screening at the 5-HT₂ receptor

subtypes.

Aporphine derivatives **8a-11a** and **8b-11b** were synthesized from natural alkaloid thebaine (1) as depicted in Scheme 1, and the detailed protocols are found in Supporting Information. (*R*)-(-)-2-Methoxy-11-hydroxyaporphine (**7a**) and (*R*)-(-)-*N*-propyl-2-methoxy-11-hydroxynoraporphine (**7b**) were used as key intermediates, and were obtained as we have detailed in previous publication.²⁹ Deprotonation of intermediate **7a** or **7b** with potassium carbonate in dry DMF followed by alkylation with 1-fluoro-2-iodoethane, 1-fluoro-3-iodopropane or 4-fluorobenzyl bromide provided aporphine derivatives **8a-10a** or **8b-10b**. In addition, esters **11a** and **11b** were prepared by facile coupling 4-fluorobenzoyl chloride with intermediates **7a** and **7b** in the presence of 4-(Dimethylamino)pyridine, respectively. Spectral data (¹H and ¹³C NMR) and elemental analysis for these target compounds were consistent with their proposed structures.

Scheme 1. Synthesis of novel aporphine derivatives 8a-11a and 8b-11b^a



^a Reagents and conditions: (i): 1. DIAD, Benzene, reflux; 2. Pyridine hydrochloride, MeOH, rt, yield 90%. (ii): 1-Iodopropane, K₂CO₃, EtOH, reflux, yield 65%. (iii): L-Selectride, THF, reflux, yields 22-35%. (iv): N-Phenyl-bis(trifluoromethanesulfonimide), Et₃N, CH₂Cl₂, rt, yields 71-72%. (v): MeSO₃H, 90
^oC, yields 55-59%. (vi): Pd/C, Mg, NH₄OAc, MeOH, rt, yields 62-87%. (vii): 1-Fluoro-2-iodoethane or 1-fluoro-3-iodopropane, K₂CO₃, DMF, 80 °C, yields 56-89%. (viii): 4-Fluorobenzyl bromide, K₂CO₃, DMF, 80 °C, yields 74-76%. (ix): 4-Fluorobenzyl chloride, 4-(Dimethylamino)pyridine, Et₃N, THF, rt, yields 77-86%.

The binding affinities of aporphine derivatives 8a-11a and 8b-11b for the 5-HT₂

receptor subtypes were examined by the National Institute of Mental Health (NIMH) Psychoactive Drug Screening Program (PDSP), using competitive binding assays with membrane preparations obtained from transiently transfected HEK293 T cells (for 5- HT_{2A}) and stably transfected HEK293 (for 5- HT_{2B}), HEK293 T cells (for 5- HT_{2C}).³⁰ Data from these evaluations are presented in Table 1.

Among these newly synthesized aporphine derivatives, compounds 8a, 9a, 11a and 11b were identified as potential hits for 5-HT_{2C} receptor. Compounds 11a and 11b showed more significant binding property to 5-HT_{2C} receptor, with inhibitory constant (K_i) value of 32 nM and 24 nM, respectively. The other two aporphine derivatives, 8a and 9a, exhibited moderate binding affinity ($K_i = 343$ and 338 nM, respectively). These results indicated that the carbonyl moiety of C11 fluorinated benzoyl group in compounds 11a and 11b might form a strong interaction with the binding site of 5-HT_{2C} receptor, while N- substituent might not play a large role here. To our delight, compounds 11a and 11b also possessed good 5-HT₂ receptor subtype selectivity. Compound **11a** showed 14-fold selectivity over the 5-HT_{2A} receptor ($K_i =$ 450 nM) and 3.2-fold selectivity versus the 5-HT_{2B} receptor ($K_i = 102$ nM). Compound **11b** showed 17-fold selectivity over the 5-HT_{2A} receptor ($K_i = 414$ nM) and 4.4-fold selectivity versus the 5-HT_{2B} receptor ($K_i = 106$ nM). Notably, in comparison with the reference drug lorcaserin, compounds 11a and 11b exhibited slightly lower affinity at the 5-HT_{2C} receptor [11a ($K_i = 32 \text{ nM}$) and 11b ($K_i = 24 \text{ nM}$) vs. Lorcaserin ($K_i = 15 \text{ nM}$)] and lower selectivity over the 5-HT_{2B} receptor, but

improved selectivity over the 5-HT_{2A} receptor.

reference for the 5-HT ₂ receptor subtypes ^{<i>a</i>}							
Compound -	$K_{\rm i} \pm { m SEM} \ ({ m nM})^b$			Selectivity			
	5-HT _{2A}	5-HT _{2B}	5-HT _{2C}	5-HT _{2A} /5-HT _{2C}	5-НТ _{2В} /5-НТ _{2С}		
8a	1138 ± 205	552 ± 95	343 ± 41	3.3	1.6		
9a	434 ± 62	422 ± 67	338 ± 36	1.3	1.2		
10a	2313 ± 389	2681 ± 451	>10000		-		
11a	450 ± 61	102 ± 13	32 ± 3	14	3.2		
8b	>10000	3044 ± 589	>10000	-	-		
9b	>10000	>10000	>10000	-	-		
10b	>10000	>10000	>10000	-	-		
11b	414 ± 51	106 ± 13	24 ± 3	17	4.4		
Lorcaserin	112 ^c	174 ^c	15 ^c	7.5	12		

 Table 1. Binding affinities of aporphine derivatives 8a-11a and 8b-11b and the reference for the 5-HT₂ receptor subtypes^a

^{*a*} All compounds were tested as their HCl salts. The following tritiated radioligands were used: [³H]Ketanserin (5-HT_{2A}), [³H]LSD (5-HT_{2B}), [³H]Mesulergine (5-HT_{2C}). ^{*b*} The K_i values are means ± standard errors of 2-3 experiments. ^{*c*} Data from ref. 31.

To explain the higher binding affinity of compounds **11a** and **11b** at the 5- HT_{2C} receptor and explore a potential binding mode, molecular docking study on compound **11b** using the GoldSuite was performed based on 5- HT_{2C} receptor model which was disclosed very recently (PDB 6BQG).³² As shown in Fig. 3, compound **11b** is

properly stabilized into the 5-HT_{2C} receptor binding site through a key salt-bridge between the positively charged nitrogen atom of compound **11b** and the carboxylate of D3.32, which is fully conserved in 5-HT and other monoamine receptors.³³ Compound **11b** also forms a H-bond interaction with N6.55 through the carbonyl oxygen atom of C11 fluorinated benzoyl group. This observation provided a potential explanation as to why compounds **11a** and **11b** possessed higher binding affinity than other newly synthesized aporphine derivatives, and supported our previous prediction. *N-n*-Propyl substituent of compound **11b** orients toward the vacant binding site of 5-HT_{2C} receptor, thereby explained the comparable binding affinity between compounds **11a** and **11b** at the 5-HT_{2C} receptor. In addition, the diphenyl moiety of aporphine structure may elicit π -alkyl interactions with some residues on TMH3, TMH6 and TMH7 (transmembrane helix) to further stabilize the ligand binding.



Fig. 3. Binding mode of compound **11b** in the binding site of 5-HT_{2C} receptor. **Left** is 3D view of the receptor-ligand interactions. Amino acid residues engaged in ligand binding are shown as sticks. Dotted yellow lines represent H-bonds with polar residues. **Right** is 2D view of the receptor-ligand interactions.

In summary, a series of novel aporphine derivatives were synthesized for initial screening at the 5-HT₂ receptor subtypes. Compounds **11a** and **11b** were identified as potent 5-HT_{2C} hit ligands with highly selectivity over other 5-HT₂ receptor subtypes. Molecular docking study revealed that compounds **11a** and **11b** formed two key interactions with the binding site of 5-HT_{2C} receptor, including a salt-bridge to D3.32 and a H-bond interaction with N6.55, which result in higher binding affinity than other newly synthesized aporphine derivatives. Compounds **11a** and **11b** are useful starting points for further SAR exploration and optimization studies. We are continuing in this vein and will report our findings in due course.

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Supplementary data

Supplementary data associated with this article can be found, in the online

version.

References

- 1. Gaspar P, Cases O, Maroteaux L. Nat. Rev. Neurosci. 2003;4:1002.
- 2. Gross C, Zhuang X, Stark K, Ramboz S, Oosting R, Kirby L, Santarelli L, Beck S,

Hen R. Nature 2002;416:396.

- 3. Saxena PR. Pharmacol. Ther. 1995;66:339.
- 4. McCorvy JD, Roth BL. Pharmacol. Ther. 2015;150:129.
- 5. Roth BL, Willins DL, Kristiansen K, Kroeze WK. Pharmacol. Ther. 1998;79:231.
- 6. Jacobson KA. Biochem. Pharmacol. 2015;98:541.
- 7. Meltzer HY, Roth BL. J. Clin. Invest. 2013;123:4986.
- 8. Smith BM, Thomsen WJ, Grottick AJ. Expert Opin. Invest. Drugs 2006;15:257.
- 9. Rosenzweig-Lipson S, Comery TA, Marquis KL, Gross J, Dunlop J. Handb. Exp. Pharmacol. 2012;213:147.
- 10. Narayanaswami V, Dwoskin LP. Pharmacol. Ther. 2016;170:116.

11. Zeeb FD, Higgins GA, Fletcher PJ. ACS Chem. Neurosci. 2015;6:1231.

- 12. Nichols DE. Pharmacol. Ther. 2004;101:131.
- Rothman RB, Baumann MH, Savage JE, Rauser L, McBride A, Hufeisen SJ, Roth
 BL. *Circulation* 2000;102:2836.
- Fitzgerald LW, Burn TC, Brown BS, Patterson JP, Corjay MH, Valentine PA, Sun JH, Link JR, Abbaszade I, Hollis JM, Largent BL, Hartig PR, Hollis GF, Meunier PC, Robichaud AJ, Robertson DW. *Mol. Pharmacol.* 2000;57:75.

15. Jantan I, Raweh SM, Yasin YH, Murad S. Phytother. Res. 2006;20:493.

16. Mohamed SM, Hassan EM, Ibrahim NA. Nat. Prod. Res. 2010;24:1395.

17. Graziose R, Rathinasabapathy T, Lategan C, Poulev A, Smith PJ, Grace M, Lila

MA, Raskin I. J. Ethnopharmacol. 2011;133:26.

18. Makarasen A, Sirithana W, Mogkhuntod S, Khunnawutmanotham N, Chimnoi N,

Techasakul S. Planta Med. 2011;77:1519.

19. Liu CM, Kao CL, Wu HM, Li WJ, Huang CT, Li HT, Chen CY. *Molecules* 2014;19:17829.

20. Nordin N, Majid NA, Hashim NM, Rahman MA, Hassan Z, Ali HM. Drug Des. Devel. Ther. 2015;9:1437.

21. Ivorra MD, Valiente M, Martinez S, Madrero Y, Noguera MA, Cassels BK, Sobarzo EM, D'Ocon P. *Planta Med.* 2005;71:897.

22. Zhang A, Zhang Y, Branfman AR, Baldessarini RJ, Neumeyer JL. J. Med. Chem.2007;50:171.

23. Munusamy V, Yap BK, Buckle MJ, Doughty SW, Chung LY. Chem. Biol. Drug Des. 2013;81:250.

24. Zhao R, Lu W, Fang X, Guo L, Yang Z, Ye N, Zhao J, Liu Z, Jia J, Zheng L, Zhao B, Zhang A, Zhen X. *Pharmacol. Biochem. Behav.* 2014;124:204.

25. Peng Y, Zhao S, Wu Y, Cao H, Xu Y, Liu X, Shui W, Cheng J, Zhao S, Shen L,

Ma J, Quinn RJ, Stevens RC, Zhong G, Liu Z. Biophys Rep. 2018;4:50.

26. Kapadia N, Harding Wayne. Med. chem. (Los Angeles) 2016;6:241.

27. Zhou Y, Wang J, Gu Z, Wang S, Zhu W, Aceña JL, Soloshonok VA, Izawa K,

Liu H. Chem. Rev. 2016;116:422.

28. Meanwell NA. J. Med. Chem. 2018; Article ASAP.

29. Si Y, Gardner MP, Tarazi FI, Baldessarini RJ, Neumeyer JL. J. Med. Chem. 2008;51:983.

30. Besnard J, Ruda GF, Setola V, Abecassis K, Rodriguiz RM, Huang XP, Norval S, Sassano MF, Shin AI, Webster LA, Simeons FRC, Stojanovski L, Prat A, Seidah NG, Constam DB, Bickerton GR, Read KD, Wetsel WC, Gilbert IH, Roth BL, Hopkins AL. *Nature* 2012;492:215.

31. Thomsen WJ, Grottick AJ, Menzaghi F, Reyes-Saldana H, Espitia S, Yuskin D, Whelan K, Martin M, Morgan M, Chen W, Al-Shamma H, Smith B, Chalmers D, Behan D. J. Pharmacol. Exp. Ther. 2008;325:577.

32. Peng Y, McCorvy JD, Harpsøe K, Lansu K, Yuan S, Popov P, Qu L, Pu M, Che T, Nikolajsen LF, Huang X, Wu Y, Shen L, Bjørn-Yoshimoto WE, Ding K, Wacker D, Han GW, Cheng J, Katritch V, Jensen AA, Hanson MA, Zhao S, Gloriam DE, Roth BL, Stevens RC, Liu Z. *Cell* 2018;172:719.

33. Venkatakrishnan AJ, Deupi X, Lebon G, Tate CG, Schertler GF, Babu MM. *Nature* 2013;494:185.