

Lipase catalysed resolution of (*R*)- and (*S*)-1-trimethylsilyl-1-alkyn-3-ols: useful intermediates for the synthesis of optically active γlactones

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Abstract: Various (R)- and (S)-1-trimethylsilyl-1-alkyn-3-ols, chiral building units useful for the synthesis of biologically active compounds, have been efficiently resolved by enantioselective acetylation mediated by immobilized lipase PS. The resolution is applied to the synthesis of (R)- and (S)-5-octyl-2-(5H)-furanones. \bigcirc 1997, Elsevier Science Ltd. All rights reserved.

Optically active 1-trimethylsilyl-1-alkyn-3-ols 1 or 1-alkyn-3-ols 2 are of synthetic value as precursors of many biologically active natural products.¹ In fact, by carboxylation of alkynols 2 (Z)-4-hydroxy-2-alkenyl acids are obtained, which are key intermediates in the total synthesis of such naturally occurring compounds as unsaturated or saturated lactones and bislactones,² leucotrienes, prostaglandins, vitamin E and steroids.³



For preparing enantiomerically enriched alkynols 1 and 2, various approaches have been reported including enantioface-differentiating addition of lithium acetylide to aliphatic aldehydes in the presence of a chiral ligand,⁴ chemical reduction with chiral reducing agents,^{5a-d} enzymatic reduction of the corresponding ketone⁶ and resolution of the acetates by fermenting *Bacillus subtilis*⁷ or by lyophilized cells of baker's yeast.⁸ However, many of these approaches are not completely satisfactory because of low or moderate chemical yields and/or enantiomeric excess, unavailability of both enantiomers, high substrate specificity or other restrictions that limit their application. On the other hand attempted enantioselective hydrolysis of the carboxylic esters of alcohols 2, mediated by lipase (from Candida cylindracea, Pseudomonas sp. and Mucor sp.) failed to discriminate^{8a} between enantiomers or showed unsatisfactory enantioselection.⁹ Lipases are more effective in the asymmetric hydrolysis of some acyl derivatives of 1-trimethylsilyl-1-alkyn-3-ols 1, even if the utility of the reaction is strictly dependent on the nature of the acyl group. In fact good results were obtained in the hydrolysis of 3-propionyloxy-1-trimethylsilyl-1-octyne, mediated by Porcine Pancreatic Lipase ($E^{10}=13$) or by Lipase Amano PS (E=63),^{9a} while the corresponding acetate was resolved only with moderate enantioselectivity (E=5.3). Finally the possibility of efficiently obtaining alkynols 1 by lipase catalysed transesterification appeared limited to only 4-trimethylsilyl-3-butyn-2-ol, having a small (methyl) and a relatively large substituent at the hydroxymethine carbon.¹¹

For these reasons, when we needed to prepare and test some enantiomerically pure unsaturated lactones 3 as possible new Michael acceptors able to induce the synthesis of heat shock proteins in

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Table 1. Enzymatic resolution of racemates 1a-h



R	conv (%) ^s (time, h)	(S)-1 ee % ^b (yield, %) ^c	(R)-4 ce % ^d (yield, %) ^c	E
	51 (20)	89 (45)	85 (46)	
b	30 (20)	40 (68)	94 (25)	50
	49 (300)	87 (46)	90 (42)	
с	30 (16)	42 (67)	97 (26)	>100
	52 (300)	>98 (44)	92 (47)	
d	39 (48)	59 (58)	90 (35)	50
	54 (300)	98 (41)	81 (50)	
e	37 (16)	57 (59)	96 (34)	>100
	51 (260)	>98 (46)	94 (48)	
f	30 (14)	42 (66)	97 (25)	>100
	52 (240)	>98 (47)	91 (49)	
g	36 (16)	55 (60)	96 (31)	>100
	51 (250)	>98 (45)	93 (48)	
h	41 (90)	55 (56)	80 (36) S	16
	51 (320)	78 (45)	73 (46)	

^a Determined by GLC after silylation of the reaction mixture. ^b Enantiomeric excess were determined by ¹H-NMR analysis of the Mosher esters. ^c After flash chromatography. ^d Enantiomeric excess were determined by ¹H-NMR using 0.2-0.4 equiv of Eu(hcf)₃ and by GLC an a chiral column for the compound (S)-1d, (R)-4a and (R)-4b.

cells,¹² we searched for a new and efficient enzymatic resolution of 1-trimethylsilyl-1-alkyn-3-ols 1 which, in enantiomerically pure form, were useful starting material for obtaining the desired lactones.

As a result, in the present paper we describe an excellent¹³ biocatalytic resolution, mediated by immobilized lipase PS,¹⁴ of seven 1-trimethylsilyl-1-alkyn-3-ols (**1a-g**, Table 1) with alkyl chains of various lengths. Many of the alkynols are obtained in an enantiomerically enriched form suitable for the preparation of (R)- and (S)-5-alkyl-2-(5H)-furanones 3 of high enantiomeric purity here exemplified by the preparation of both (R)- and (S)- enantiomers of 3e (Scheme 1).



(i): K2CO3, McOH/H2O, rt, 2h. (ii): nBuLi, THF, -40°C, then CO2. (iii): H2, Lindlar, quinoline, McOH. (iv): CF3CO2H.

Scheme 1.

Results and discussion

Eight racemic 1-trimethylsilyl-1-alkyn-3-ols 1a-h with alkyl or aryl chains of different lengths, prepared by reaction of trimethylsilyl acetylide with the appropriate aldehyde, were selected in order to explore their possible lipase catalysed resolution, the scope and the limitations of the process. Preliminary experiments were performed in order to screen enzymes, solvents, acylating agents and to optimise other reaction conditions. With this aim, the enantioselective transesterification of 1trimethylsilyl-1-undecyn-3-ol 1e was carried out in the presence of six commercial lipases,¹⁴ using vinyl acetate or trifluoroethyl butyrate as acyl donors and diisopropyl ether or tert-butylmethyl ether as solvents. In these experiments, it was observed that lipase from porcine pancreas (PPL), lipase from Candida cylindracea and lipase from Candida antarctica were completely inactive, lipase from Pseudomonas fluorescens showed high selectivity (E >30) but poor activity, while free and immobilized¹⁵ Lipase PS were able to catalyse the transesterification with higher and comparable enantioselectivity (E > 30). Since supported lipase PS was much more active than the free enzyme, it was selected and tested for the catalysis of the enantioselective transesterification of the other substrates, using vinyl acetate and disopropyl ether in combination with the enzyme. The results obtained show that the enantioselectivity of the reactions is always excellent for all substrates having an aliphatic chain bonded to the hydroxymethyne centre, independent of its length (Table 1). In fact, the lipase catalysed resolutions of the racemic 1-trimethylsilyl-1-alkyn-3-ols 1a-g yield the acetylated (R)-compounds in enantiomeric excesses often suitable for their use in the enantioselective synthesis of natural products.

Similarly unreacted (S)-alcohols were obtained with high enantiomeric excess when the extent of conversion was allowed to proceed a little over 50%.¹⁰ However, the presence of an aryl substituent on the hydroxymethine centre as in **1h** resulted in a significant decrease in the enantioselectivity of the reaction (E=16). The favourable results obtained in the enantioselective transesterification of the alcohols **1a-g**, prompted us to test the possibility of using the same enzyme for the reverse reaction, the enantioselective hydrolysis of the acetates. Thus we found that the enzymatic hydrolysis of the racemic acetates **4c** and **4e** in a phosphate buffer afforded the (R)-alcohols with excellent enantioselection (E >30).

The enantiomeric excesses and the absolute configuration of the (S)-1-trimethylsilyl-1-alkyn-3ols **1a-g** were determined by ¹H-NMR analysis of their (*R*)- and (*S*)-Mosher esters, ¹⁶ while the enantiomeric excess of the acetates (*R*)-**4a-h** were evaluated by ¹H-NMR using Eu(hcf)₃.¹⁷ For the compounds (*S*)-**1d**, (*R*)-**4a** and (*R*)-**4b** (see Experimental section) the enantiomeric excesses were also evaluated by GLC analysis on a chiral column. In the case of the aromatic compound **1h** the absolute configuration was also confirmed by ¹H-NMR analysis of the (*R*)- and (*S*)-Mosher esters of the corresponding alcohols **2h**, prepared in order to have a diagnostic proton near to the stereogenic center.¹⁸

The utility of the above results was then demonstrated in the synthesis of (R)- and (S)-5-octyl-2-(5H)-furanones **3e** of high enantiomeric purity (Scheme 1) starting with the acetate (R)-**4e** and the

alcohol (S)-1e. Each of them was separately treated with methanolic potassium carbonate which caused the elimination of the silyl group and, in the case of 4e, the simultaneous saponification of the acetoxy group. The obtained alkynols were then transformed, according to a known reaction sequence,^{4,19} into the lactones 3e without loss of stereochemical integrity; in fact the final lactones were obtained with the same enantiomeric excess as the starting materials. The route includes the carboxylation of the terminal acetylenic carbon, the hydrogenation on Lindlar catalyst of the formed acetylenic acid 5 and a brief exposure to acid to accelerate the lactonization.

Thus we may conclude that 1-trimethylsilyl-1-alkyn-3-ols can be obtained, by biochemical kinetic resolution, with an enantiomeric enrichment suitable for the synthesis of enantiomerically pure natural products.

Experimental section

The ¹H-NMR spectra (500.13 MHz) were recorded in CDCl₃ at 303 K and were referenced to CHCl₃ at 7.24 ppm. HPLC analyses were carried out on a Merck superspher 100 RP-18 column, 4 mm×25 cm, with a flow rate of 1 mL/min and a detection performed at 241 nm. Optical rotations were measured for 1% CHCl₃ solutions if not otherwise reported. TLC was carried out on silica gel 60 F₂₅₄ microplates. Column chromatography refers to flash chromatography.²⁰ (*S*)- and (*R*)- α -Methoxy- α -trifluoromethylphenyl acetates [(*S*)- and (*R*)-MTPA] were prepared from the appropriate (*R*)- and (*S*)-MTPA chlorides.¹⁶ All organic solvents were dried before use. Usual workup refers to washing the organic layer with water, drying it over Na₂SO₄, and evaporating the solvent under reduced pressure. The progress of all reactions, the column chromatography and compound purity were monitored by GLC, TLC and/or HPLC. Chiral GLC analyses were performed on dimethylpentylcyclodextrin- β column (50 m, 0.25 mm ID, film thickness 0.25 µm, Mega). The 1-trimethylsilyl-1-alkyn-3-ols **1a-h** were prepared (with the reported yields) on a 2 mmol scale according to G. Traverso *et al.*²¹ and, after rapid chromatography (eluting with hexane–ethyl acetate, 95:5, v:v), showed the following properties.

1-Trimethylsilyl-1-pentyn-3-ol **1a**; (76% yield): an oil. ¹H-NMR δ 4.28 (1H, dt, J 4.9, 6.3 Hz, H-3). Anal. Calcd for C₈H₁₆OSi: C, 61.49; H, 10.32. Found: C, 61.41; H, 10.27.

1-Trimethylsilyl-5-methyl-1-hexyn-3-ol **1b**; (87% yield): an oil. ¹H-NMR δ 4.35 (1H, dt, J 5.6, 7.0 Hz, H-3). Anal. Calcd for C₁₀H₂₀OSi: C, 65.15; H, 10.94. Found: C, 65.27; H, 10.79.

1-Trimethylsilyl-1-octyn-3-ol 1c; (86% yield): an oil. ¹H-NMR δ 4.33 (1H, dt, J 5.6, 6.3 Hz, H-3). Anal. Calcd for C₁₁H₂₂OSi: C, 66.60; H, 11.18. Found: C, 66.53; H, 11.26.

(*E*)-1-Trimethylsilyl-4-octen-1-yn-3-ol 1d; (87% yield): an oil. ¹H-NMR δ 5.86 (1H, dt, J 7.0, 14.7 Hz, H-5), 5.57 (1H, dd, J 5.6, 14.7, H-4), 4.80 (1H, dd, J 4.9, 5.6 Hz, H-3). Anal. Calcd for C₁₁H₂₀OSi: C, 67.28; H, 10.27. Found: C, 67.39; H, 10.34.

1-Trimethylsilyl-1-undecyn-3-ol 1e; (85% yield): an oil. ¹H-NMR δ 4.33 (1H, dt, J 5.6, 6.3 Hz, H-3). Anal. Calcd for C₁₄H₂₈OSi: C, 69.93; H, 11.74. Found: C, 69.84; H, 11.81.

1-Trimethylsilyl-1-tetradecyn-3-ol **1f**; (87% yield): an oil. ¹H-NMR δ 4.33 (1H, dt, J 6.3, 7.0 Hz, H-3). Anal. Calcd for C₁₇H₃₄OSi: C, 72.27; H, 12.13. Found: C, 72.18; H, 12.25.

1-Trimethylsilyl-1-esadecyn-3-ol **1g**; (78% yield): an oil. ¹H-NMR δ 4.32 (1H, dt, J 5.6, 6.3 Hz, H-3). Anal. Calcd for C₁₉H₃₈OSi: C, 73.47; H, 12.33. Found: C, 73.39; H, 12.37.

3-Trimethylsilyl-1-phenylpropyn-1-ol **1h**; (75% yield; eluted with hexane–ethyl acetate 20:80, v:v): an oil. ¹H-NMR δ 5.44 (1H, d, J 6.3 Hz, H-1). Anal. Calcd for C₁₂H₁₆OSi: C, 70.53; H, 7.89. Found: C, 70.85; H, 7.62.

Lipase-mediated acylation of racemic 1-trimethylsilyl-1-alkyn-3-ols (1a-h) in organic solvents. General procedure

Vinyl acetate (2 mmol) and immobilized Lipase PS (250 mg; 30% on Hyflo Super Cell) were added to a solution of each racemic 1-trimethylsilyl-1-alkyn-3-ol (**1a-h**; 1 mmol) in diisopropyl ether (10 mL). The resulting suspension was then shaken at 25°C and monitored by GLC, after silylation (pyridine-trimethylchlorosilane-hexamethyldisilazane, 10:4:2, v:v:v, rt, 1 h) of the hydroxy group of

the unreacted alcohol. The final conversion extent was however established by ¹H-NMR analysis of the isolated crude mixture. When the desired conversion was reached, the enzyme was recovered by filtration and the solvent was evaporated under reduced pressure. The unreacted alcohols (S)-1**a-h** and the acetate (R)-4**a-h** were then separated by flash chromatography, eluting with hexane-ethyl acetate (100:5; v:v), and were obtained with the yields and the enantiomeric excess shown in Table 1. When the reactions were stopped at a conversion higher than 50% (see Table 1), the unreacted (S)-alcohols 1**a-h** (purity >98% by GLC after silvlation) showed correct physicochemical properties, identical to those observed for their racemates, and the following optical rotations: $[\alpha]_D^{25} - 4.1$ for (S)-1**a**, -10.2 for (S)-1**b**, -1.8 for (S)-1**c**, +38.5 for (S)-1**d**, +1.1 for (S)-1**e**, +1.2 for (S)-1**f**, +2.4 for (S)-1**g**, -13.3 for (S)-1**h**.

Similarly when the reactions were terminated at the conversions lower than 50% (see Table 1), the acetates (*R*)-4**a**-**h** showed ¹H-NMR spectra with appropriate proton signals and the following optical rotations: $[\alpha]_D^{25}$ +123.3 for (*R*)-4**a**, +77.6 for (*R*)-4**b**, +86.8 for (*R*)-4**c**, -13.9 for (*R*)-4**d**, +71.4 for (*R*)-4**e**, +67.7 for (*R*)-4**f**, +63.8 for (*R*)-4**g**, +28.6 for (*R*)-4**h**.

Lipase-mediated hydrolysis of racemic 1-trimethylsilyl-1-alkyn-3-ol acetates (4c and 4e). General procedure

Lipase PS (75 mg) and each racemic 1-trimethylsilyl-1-alkyn-3-ol acetate (**4c** and **4e**; 0.5 mmol) dissolved in acetone (0.5 ml) were shaken at 25°C in phosphate buffer (5 mL; 0.1 M; pH 7) and the reaction was monitored by GLC, after the derivatization described above for the transesterification procedure. After about 150 h a satisfactory hydrolysis was reached in both cases and the mixture was extracted and worked up to afford respectively the alcohol (*R*)-**1c** (conv. 50%; ee 90%; 45% yield; $[\alpha]_D^{25} - 1.3$) and (*R*)-**1e** (conv. 51%; ee 94%; 45% yield; $[\alpha]_D^{25} - 1.0$).

Synthesis of (R)- and (S)-5-octyl-2-(5H)-furanones [(R)- and (S)-3e]

i) Desilylation. (S)-1-Trimethylsilyl-1-undecyn-3-ol [(S)-1e; 315 mg; 1.3 mmol] was dissolved in a saturated solution of K₂CO₃ in moist methanol (15 mL containing 1 mL of H₂O) and stirred at room temperature for 2 h. The solvent then was evaporated under reduced pressure, the residue was recovered with diethyl ether and worked-up to afford crude (S)-1-undecyn-3-ol (96% yield). A sample was purified by rapid chromatography and showed $[\alpha]_{D}^{25} - 10.1$ (dioxane, c 2).^{8a} The crude compound was then used for the successive carboxylation without purification. The enantiomeric (R)-alkynol was obtained (95% yield) by reacting in similar conditions the 3-acetoxy-1-trimethylsilyl-1-undecyne [(R)-4e] and the purified sample showed $[\alpha]_{D}^{25} + 9.9$ (dioxane, c 2).¹⁹

ii) Carboxylation. To a stirred solution of the (S)-1-undecyn-3-ol (186 mg; 1.1 mmol) in THF (5 mL), *n*-BuLi (1.65 mL of a 1.6 M solution in hexane; 2.64 mmol) was added dropwise at -40° C and, after 0.5 h, CO₂ was bubbled for 1 h. The mixture was then treated with a saturated cold aqueous solution of NaCl and extracted with hexane. The aqueous solution was then acidified to pH 3 by addition of cold diluted HCl and extracted with diethyl ether to afford, after usual work-up, the crude (S)-4-hydroxy-2-dodecynoic acid [(S)-5] which was used for the next reaction without purification. A sample was purified from pentanoic acid contaminant by rapid chromatography and showed: m.p. 78°C; $[\alpha]_{D}^{25}$ –13.0 (dioxane, c 2); ¹H-NMR δ 5.70 (2H, broad signal, -COOH and OH), 4.51 (1H, t, J 7.5 Hz, H-4), 1.75 (2H, m, H₂-5), 1.44 (2H, m, H₂-6), 1.27 (10H, m, 5×CH₂), 0.86 (3H, t, J 7.5 Hz, H₃-12).

Starting from (R)-3-hydroxyundec-1-yne the enantiomeric (R)-5 was obtained. A purified sample showed: m.p. 78°C; $[\alpha]_{\rm D}^{25}$ +12.7 (dioxane, c 2).¹⁹ The crude product was then directly reached.

iii) Hydrogenation and lactonization. The above crude acid (S)-5 (190 mg) was hydrogenated in methanol (30 mL) under a hydrogen atmosphere in the presence of Lindlar catalyst (50 mg) and pure quinoline (10 μ L). After filtration of the catalyst, the solution was evaporated under reduced pressure to leave a crude hydroxyacid (S)-6 which was treated with trifluoroacetic acid (10 μ L) in dichloromethane (2 mL) at room temperature for 10 min. After usual work-up, the crude compound

was purified by distillation under reduced pressure to afford the (S)-5-octyl-2-(5H)-furanone [(S)-3e]: b.p. 100°C (Buchi GKR-50, 0.2 mmHg); $[\alpha]_{D}^{25}$: +76.2 (dioxane, c 2); ¹H-NMR δ 7.62 (1H, dd, J 6.1, 1.5 Hz, H-4), 6.24 (1H, dd, J 6.1, 2.3 Hz, H-3), 5.20 (1H, dddd, J 6.9, 6.9, 2.3, 1.5 Hz, H-5), 0.88 (5H, t, J 7.3 Hz, CH₃-).

A similar reaction sequence afforded the (R)-5-octyl-2-(5H)-furanone [(R)-3e]: $[\alpha]_{D}^{25} -73.3$ (dioxane, c 2).¹⁹

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