Total Synthesis of Granditropone, Grandirubrine, Imerubrine, and Isoimerubrine

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Abstract: Concise total syntheses of the naturally occurring tropoloisoquinolines grandirubrine (1), imerubrine (2), and isoimerubrine (3) are detailed. The regioselective total synthesis of grandirubrine (1) is based on the [4 + 2] cycloaddition reaction of the α -pyrone 44 with the cyclopropenone ketal 18. Subsequent retro-Diels-Alder loss of CO₂, norcaradiene rearrangement to the cycloheptatrienone ketal, and ketal hydrolysis provided the tropone 7 (granditropone). Regioselective hydroxylation of granditropone (NH₂NH₂; KOH) provided grandirubrine (1) and O-methylation of 1 provided both imerubrine (2) and isoimerubrine (3).

Grandirubrine $(1)^1$ and imerubrine $(2)^2$ constitute the initial members of a rare class of naturally occurring tropoloisoguinolines³ now including isoimerubrine (3),⁴ pareirubrines A and B (4 and 5),⁴ and pareitropone (6),⁴ which are structurally similar to colchicine⁵ and its naturally occurring congeners and biosynthetically related to the more common azafluoranthene alkaloids (Figure 1).6-8 The tropolones 1, 4, and 5 each appear to exist preferentially in the 6-keto tautomer in solution, but both 4 and 5 preferentially crystallize in the alternative 5-keto tautomer shown in Figure 1. Despite their intriguing structures which for 2,2 4,4 and 54 were unambiguously established in single crystal X-ray structure determinations, their cytotoxic properties,⁴ and their structural similarity to the mitotic inhibitor colchicine, little progress has been made on their synthesis. Only one recent total synthesis of grandirubrine and imerubrine has been reported⁹ and few related efforts have been dislcosed.¹⁰

In an extension of our early efforts on the divergent total syntheses of the azafluoranthene alkaloids which resulted in the total syntheses of rufescine (8) and imeluteine (10), herein we report the total syntheses of grandirubrine, imerubrine, and

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Figure 1.

isoimerubrine which proceed through the intermediate tropone 7 (granditropone). The approach underscores the ease with which a single precursor may be utilized in the divergent preparation of related agents through implementation of a series of α -pyrone inverse electron demand Diels—Alder reactions (Scheme 1).^{11,12}

In the initial development of the approach, 8 ring D introduction for construction of the azafluoranthene alkaloids was successfully realized through use of the electron-rich dienophiles, 1,1-dimethoxyethylene or 1,1,2-trimethoxyethylene, and the participation of the strained olefin of a cyclopropenone ketal 13 in a Diels—Alder reaction with the electron-deficient α -pyrone was expected to provide a direct introduction of the seven-membered tropone ring of the tropoloisoquinolines. $^{14-16}$ This cyclopropenone ketal [4+2] cycloaddition approach to a tropone annulation is complementary to its thermal [3+4] cycloaddition reaction that proceeds by a reversibly generated π -delocalized singlet vinylcarbene which we have utilized

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Scheme 2

successfully in an approach to colchicine (Scheme 2).¹⁷ In this latter work, the [3 + 4] cycloadduct was readily converted into the corresponding tropone while that of the corresponding [4 + 2] cycloadduct was not. Provided the Diels-Alder adduct could be effectively converted to the corresponding tropone, the tropone C5 carbonyl introduction with the [4 + 2]cycloaddition approach was anticipated to permit the regioselective tropolone introduction required of 1-3 while this would not appear as accessible to the more symmetrical C6 tropone derived from a [3 + 4] cycloaddition. Although the initial stages of this work culminating in the synthesis of the azafluoanthene alkaloids were completed nearly 10 years ago, the recent reports of useful levels of cytotoxic activity with the tropoloisoquinolines^{4,5} and azafluoranthene alkaloids⁶ provided the incentive for us to renew our efforts. In particular, the reported potent cytotoxic activity of the tropone 6 (pareitropone, $IC_{50} = 0.0008$ μg/mL, P388) relative to that of the naturally occurring tropolones $(0.2-1 \mu g/mL)$ suggested that the intermediate granditropone (7) may prove more interesting than the natural products themselves.

Tropolone Annulation. Prior to implementing the [4 + 2]cycloaddition route to the tropone/tropolone annulation, we first

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Scheme 3

examined this process in detail with a readily available system closely related to the tropoloisoguinolone nucleus. Treatment of 1-acenaphthenone¹⁸ (11) with 1.1 equiv of t-BuOK (t-BuOH-THF) followed by the addition of 12¹⁹ (1 equiv, 0 °C, 45 min) provided 14, which was immediately treated with 6 N HCl (3 equiv. 25 °C, 12 h) without workup or isolation. Without purification, the resulting crude acid 1520 was treated with TFAA-CH₂Cl₂ (1:1, 25 °C, 14 h) to provide the α-pyrone 13 in 74% overall yield (Scheme 3). Although the intermediates could be isolated, purified, and characterized, some loss due to instability was experienced and the overall yields improved if the three steps were conducted without the intermediate isolations. Stoichiometric LDA could be employed in place of t-BuOK, but the overall conversions were lower and the isolation of 14 (65%) was more problematic. Alternatively, treatment of 14 with acid (TFA, HOAc, HCO₂H) under nonaqueous conditions led to the clean generation of 16.20 Hydrolysis of 16 (6 equiv of LiOH, THF-H₂O, 25 °C, 6 h) followed by 1 N HCl neutralization similarly provided 15,20 which upon treatment with Ac₂O (25 °C, 48 h, 88-90% overall) provided the α-pyrone 13. Although this latter approach required more manipulations, the sequence could be effectively conducted without the intermediate isolations and provided 13 in superb overall yield (88-90% from 11). While these two complementary approaches to 13 appear deceptively simple, the subjection of 14 or its derived products to a range of alternative reaction conditions resulted in their reversion back to 11 and they proved sensitive to air oxidation in the presence of traces

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⁽¹⁸⁾ PDC oxidation (2 equiv, CH₂Cl₂, 25 °C, 6 h, 77%) of commercially available 1-acenaphthenol (Aldrich) provided 11.

⁽²⁰⁾ For 14: ¹H NMR (CDCl₃, 250 MHz) δ 8.52 (1H, s), 8.13 (1H, d, J = 7.2 Hz), 8.09 (1H, d, J = 7.2 Hz), 7.73 (1H, d, J = 7.2 Hz), 7.63 (1H, t, J = 7.2 Hz), 7.56 (1H, t, J = 7.2 Hz), 3.52 (1H, s), 1.85 (6H, s). For 15: ¹H NMR (CDCl₃ 250 MHz) δ 8.30–7.40 (6H, m), 7.22 (0.6H, t, J = 7.5 Hz), 7.54 (10.8H, t), J = 7.8 Hz), 4.24 (0.8H, t), J = 7.8 Hz), 4.24 (0.8H, t), J = 7.8 Hz), 4.24 (1.8H, t), 4.24 (1. d, J = 7.8 Hz), 3.84 (1.2H, d, J = 7.5 Hz). For **16**: ¹H NMR (CDCl₃, 400 MHz) δ 9.17 (1H, s), 8.24 (1H, d, J = 7.1 Hz), 8.22 (1H, d, J = 8.1 Hz), 7.99 (1H, d, J = 8.4 Hz), 7.97 (1H, d, J = 6.9 Hz), 7.82 (1H, dd, J = 7.1, 8.1 Hz), 7.73 (1H, dd, J = 6.9, 8.4 Hz).

of base. Moreover, while the former three-step synthesis of 13 from 11 proved satisfactory for this simple system, the latter approach proved more successful when applied to 1-3.

The key Diels-Alder reaction of 13 with the cyclopropenone ketal 18²¹ proceeded well at room temperature under pressurepromoted reaction conditions (12 kbar, CHCl₃, 25 °C, 20 h, 73%) and cleanly provided exclusively the stable exo adduct 20 (Scheme 4). The adduct 20 was surprisingly stable to conventional chromatographic purification but losses were incurred during such purification. The stereochemistry of the Diels-Alder reaction was inferred from the typical thermal stability of the exo (≥ 100 °C) versus endo (< 25 °C) adducts, ^{14,15} confirmed upon observation of the diagnostically small C4a-H/C5-H ¹H NMR coupling constant (J = 3.8 Hz; exo calcd J= 4.1 Hz, endo calcd J = 4.9 Hz), and unambiguously established in a single crystal X-ray structure determination (Scheme 4).²² The [4 + 2] cycloaddition of 13 with the cyclopropenone ketal 18 proved surprisingly more effective in CHCl₃ than most other solvents, and the clean generation of the single exo adduct was unusual. 14,15,17 This potentially may be attributed to reaction through the cyclopropenium cation 19 with enhanced dipole, secondary orbital overlap, and stabilizing electrostatic interactions preferentially achieved through the exo transition state. Presumably trace acid in CHCl₃ serves as the adventious catalyst. Consistent with this interpretation, the comparable reaction in THF (13 kbar, 25 °C, 64 h) provided a 1:1 mixture of 20 and 23 (86%), albeit requiring longer reaction times and higher reaction pressures. In this reaction, the tropone 23 is derived from the thermally unstable endo [4 + 2]

Scheme 5

cycloadduct (<25 °C) that suffers retro-Diels—Alder loss of CO₂ and an ensuing electrocyclic rearrangment to the cycloheptatrienone ketal which was detected in the crude 1H NMR. Subsequent hydrolysis of the labile ketal upon chromatographic purification provided 23. The alternative thermal reaction of the cyclopropenone ketal 18 with 13 (75 °C, C₆H₆ or CH₃CN, 1–48 h), which would be expected to proceed through reversible π -delocalized singlet vinylcarbene generation and subsequent [3 + 4] cycloaddition, 14,17 provided only recovered 13 with no evidence of reaction.

Thermal retro-Diels-Alder loss of CO₂ from exo-20, the ensuing low-temperature electrocyclic norcaradiene rearrangement,²³ and hydrolysis of the labile tropone ketal to provide 23 was effectively and directly accomplished upon warming at 90 °C (pyridine, 48 h, 80%) without detection of the intermediate diene 21 or tropone ketal 22 (Scheme 5). Presumably the labile ketal was hydrolyzed upon concentration and chromatographic assay or purification. The use of higher reaction temperatures (100-120 °C) led to progressively diminished conversions (65-41%), indicating further thermal consumption of the product.¹⁷ Alternative attempts to employ more conventional solvents for the initial thermal loss of CO₂ proved more problematic. The reaction proved slower in both toluene (110 °C, 3 d) or xylene (135 °C, 24 h) and the generation of the product tropone 23 was accompanied by a variable but substantial amount of the benzoate ester 28²⁴ (Scheme 6). Presumably, 28 is derived from adventious acid-catalyzed rearrangement of 20 or the norcaradiene 21 which is in equilibrium with the tropone ketal under

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the reaction conditions. Consistent with this, the addition of base (Et₃N) to the reaction mixtures improved the **23:28** ratio and the use of pyridine as the reaction solvent surpressed its generation altogether. Deliberate acid treatment of **20** with HOAc-THF-H₂O (6:5:2, 25 °C, 72 h) or 5% aqueous HCl-THF (1:1, 25 °C, 4 h) did cleanly provide **28**²⁴ (43-55%). However, treatment of **20** with NaOCH₃ (25 °C, 24 h) also provided a mixture of the esters **29**²⁴ in better than 70% combined conversion presumably via methanolysis of the lactone, subsequent elimination of H₂O with generation of a norcaradiene and its similar rearrangement to **29** illustrating that this conversion may not be limited to acidic reaction conditions.

The final conversion of tropone 23 to the tropolone 25 proceeded uneventfully. Without optimization, treatment of 23 with hydrazine (THF-EtOH, 25 °C, 4 d, 65-76%) cleanly and regioselectively provided the 6-aminotropone 24 as the exclusive addition product (Scheme 5). Importantly, the isomeric 4-aminotropone was not detected under these reaction conditions, suggesting that the steric or electronic requirements for nucleophilic addition at C4 are more demanding. Hydrolysis of 24 (2 N KOH-EtOH 1:1, 100 °C, 48 h, 66%) provided the tropolone 25. Methylation of 25 under a variety of conditions provided a 3:1 to 1:1 mixture of O-methyl tropolones 26 and 27 in which 26 often predominated.

In the course of these studies, we also examined a number of alternatives to the 13 + 18 diene—dienophile combination. The dienes 31-33, 26 readily prepared by addition of the potassium enolate of 11^{27} (1.1 equiv of t-BuOK, 0 °C, 30 min) to methyl 3-methoxyacylate (30) and subsequent O-acylation or silation, failed to react with the cyclopropenone ketal 18 or cyclopropenone (34)^{13,21} under thermal or pressure-promoted reaction conditions (Scheme 7). Presumably, this may be attributed to preferential adoption of the unreactive transoid versus cisoid diene conformation. Similarly, the α -pyrone 35^{11}

Scheme 7

failed to react productively with 18 under thermal or pressurepromoted reaction conditions and both 35 and 13 failed to react productively with cyclopropenone. These observations proved instrumental in defining our approach to 1-3 detailed below.

Total Synthesis of 1-3. The ketone 438 was prepared from 5,6,7-trimethoxyisoquinoline (36)²⁸ as previously detailed with notable improvements (Scheme 8). First, the bromide 37 was prepared by low-temperature, acid-catalyzed bromination of 36 (1.4 equiv of NBS, cat. H₂SO₄, THF, 25 °C, 1 h, 80%)⁸ rather than constructed from 2-bromo-3,4,5-trimethoxybenzyl bromide.²⁸ Trap of the aryllithium reagent derived from 37 with DMF provided the aldehyde 38 directly in good yields (61%), and in prior efforts, this was accomplished in three indirect steps (CO(OCH₃)₂, 83%; Dibal-H, MnO₂, 52%). In addition, the conversion of 39 to 41²⁹ without intermediate purification of 40 provided 41 in higher overall conversions. Finally, the yield for hydrolysis and decarboxylation of 42 was improved by subsequent treatment with KOH which drives the reaction to completion and precluded the isolation of small amounts of the reaction intermediates.

Conversion of 43 to the α -pyrone 44 (52%) was accomplished by the five-step, one-pot sequence detailed for 13 without purification of the reaction intermediates. Treatment of 43 with t-BuOK (1.5 equiv, t-BuOH-THF) followed by the addition of 12 (1.5 equiv, 0 °C, 15 min) provided 45. Treatment of 45 with TFA under nonaqueous conditions (25 °C, 14 h) led to clean generation of 46. Hydrolysis of 46 (LiOH, H_2O , 0 °C, 15 min) followed by 1 N HCl neutralization and acid-catalyzed decarboxylation provided 48, which upon treatment with Ac_2O (25 °C, 48 h, 52% overall) provided the α -pyrone 44 (Scheme 9). Notably, the hydrolysis of 46 conducted in THF- H_2O

⁽²⁴⁾ For **28**: ¹H NMR (CDCl₃, 250 MHz) δ 8.58 (1H, d, J = 1.5 Hz), 8.11 (1H, dd, J = 1.5, 7.9 Hz), 8.05 (2H, d, J = 6.9 Hz), 7.98 (1H, d, J = 7.9 Hz), 7.94 (1H, d, J = 8.2 Hz), 7.92 (1H, d, J = 8.2 Hz), 7.70 (2H, dd, J = 6.9, 8.2 Hz), 4.27 (2H, s), 3.44 (2H, s), 1.07 (6H, s). For **29** (R = COOMe, R¹ = CH₂CMe₂CH₂OH): ¹H NMR (CDCl₃, 400 MHz) δ 8.31 (1H, s), 8.20 (1H, s), 8.05 (1H, d, J = 6.9 Hz), 8.04 (1H, d, J = 6.9 Hz), 7.96 (1H, d, J = 8.1 Hz), 7.95 (1H, d, J = 8.1 Hz), 7.70 (2H, dd, J = 6.9, 8.1 Hz), 4.21 (2H, s), 3.98 (3H, s), 3.39 (2H, s), 0.99 (6H, s). For **29** (R = COOMe, R¹ = Me): ¹H NMR (CDCl₃, 250 MHz) δ 8.28 (2H, s), 8.06 (2H, d, J = 7.0 Hz), 7.97 (2H, d, J = 8.0 Hz), 7.73 (2H, dd, J = 7.0, 8.0 Hz), 3.97 (6H, s).

⁽²⁵⁾ When the reaction was conducted in THF alone without the EtOH cosolvent, **24** (76%) and the corresponding 4-aminotropone (17%) were produced. For 5-oxo-4-amino-9*H*-cycloheptatrieno[a]acenaphthylene: 1 H NMR (CDCl₃, 400 MHz) δ 8.02 (1H, d, J = 7.0 Hz), 7.97 (1H, d, J = 7.2 Hz), 7.93 (1H, d, J = 8.0 Hz), 7.89 (1H, d, J = 8.2 Hz), 7.70 (1H, dd, J = 7.2, 8.2 Hz), 7.69 (1H, dd, 7.0, 8.0 Hz), 7.61 (1H, dd, 1.0, 8.9 Hz), 7.40 (1H, dd, J = 8.9, 11.7 Hz), 7.27 (1H, dd, J = 1.0, 11.7 Hz), 6.72 (2H, br s).

⁽²⁶⁾ For 31: ¹H NMR (CDCl₃, 250 MHz) δ 7.94 (1H, d, J = 7.2 Hz), 7.87 (1H, dd, J = 0.8, 7.9 Hz), 7.82 (1H, d, J = 8.3 Hz), 7.81 (1H, d, J = 16.2 Hz), 7.70–7.50 (3H, m), 6.65 (1H, d, J = 16.2 Hz), 3.83 (3H, s), 2.50 (3H, s); IR (KBr) ν_{max} 1769, 1707, 1630, 1612, 1428, 1309, 1177, 1117, 821, 775 cm⁻¹. For 32: ¹H NMR (CDCl₃, 250 MHz) δ 7.97 (1H, d, J = 16 Hz), 7.85 (1H, d, J = 8 Hz), 7.82 (1H, d, J = 7 Hz), 7.71 (1H, d, J = 8 Hz), 7.66 (1H, d, J = 7 Hz), 7.54 (1H, dd, J = 7, 8 Hz), 7.51 (1H, dd, J = 7, 8 Hz), 6.50 (1H, d, J = 16 Hz), 3.82 (3H, s), 1.13 (9H, s), 0.31 (6H, s). For 33: ¹H NMR (CDCl₃, 250 MHz) δ 8.20 (1H, d, J = 8.1 Hz), 8.09 (1H, d, J = 8.5 Hz), 8.00 (1H, d, J = 7.3 Hz), 7.79 (1H, dd, J = 7.3, 8.1 Hz), 7.76 (1H, dd, J = 7.1, 8.5 Hz), 7.46 (1H, d, J = 15.8 Hz), 6.44 (1H, d, J = 15.8 Hz), 3.76 (3H, s), 3.19 (3H, s).

⁽²⁷⁾ The lithium enolate generated with LDA (1.4 equiv, -78 °C THF or THF-HMPA) was unreactive toward 30.

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Scheme 9

provided lower conversions (25-40%) due to its insolubility and was improved significantly by using H_2O alone as the reaction solvent.

The key Diels—Alder reaction of 44 with the cyclopropenone ketal 18 could be accomplished under a variety of conditions and in a range of solvents and, after purification, provided a mixture of the stable adduct exo-49, granditropone (7) derived from the thermally unstable endo-49, and small amounts of 50 (Scheme 10). The stereochemistry of exo-49 could be inferred from the typical thermal stability of the exo (≥ 100 °C) versus endo (≤ 25 °C) adducts^{14,15} and their direct analogy to relative

stability of the adducts derived from the Diels-Alder reactions leading to 20 and 23, and was confirmed upon observation of the diagnostic C4a-H/C5-H 1 H NMR coupling constant (J =4.0 Hz, exo calcd J = 4.1 Hz). Hydrolysis of the tropone ketal, derived from decarboxylation and electrocyclic rearrangement of endo-49, to provide 7 and rearrangement of exo-49 to 50³⁰ both were determined to occur upon chromatographic purification of the reaction mixture. Consequently, treatment of the crude Diels-Alder reaction mixture with 3.6 N HCl-EtOAc/ THF to promote the conversion of exo-49 to the corresponding tropone ketal followed by aqueous workup with concurrent tropone ketal hydrolysis provided 7 directly in yields as high as 40-60% accompanied by small amounts of **50** (10-20%) Scheme 11. In contrast to exo-20, which cleanly provided the tropone upon thermal decarboxylation (Scheme 6), exo-49 provided 5130 presumably derived from rearrangement of the intermediate norcardiene under the reaction conditions. Moreover, in contrast to exo-20, which provided predominately rearrangement products upon acid treatment (Scheme 6), exo-49 cleanly and predominately provided granditropone (7) upon aqueous acid treatment (Scheme 10).

The final conversions of granditropone (7) to 1–3 proved uneventful. Treatment of 7 with hydrazine (THF, 25 °C, 18 h, 78%) cleanly provided the C6-amine 52 with no trace of the isomeric C4-amine (Scheme 11). Hydrolysis of 52 (2 N KOH—CH₃OH 1:4, 85 °C, 30 h, 70%) cleanly provided grandirubrine (1) identical in all respects with the properties reported for authentic material.^{1,2} Final O-methylation of 1 with TMSCHN₂ (THF—CH₃OH, 25 °C, 20 h, 72—76%) provided a 1:1 (2:1 CH₃OH—THF) to 2:1 (1:1 CH₃OH—THF) mixture of imerubrine (2) and isoimerubrine (3), which were readily separable by SiO₂ chromatography, and both proved identical in all respects with the properties reported²⁻⁴ for authentic material.³¹

Experimental Section

4-Oxa-5-oxobenz[a]acenaphthylene (13). Method A. A solution of 11¹⁸ (3.0 g, 17.9 mmol) in 15 mL of anhydrous THF was added dropwise to a stirred solution of 20 mL of t-BuOK (1 M in t-BuOH, 20 mmol, 1.1 equiv) in 15 mL of THF at −78 °C under Ar, and the resulting dark purple solution was stirred for 30 min. The reaction mixture was allowed to warm to 0 °C, and 1219 (3.3 g, 17.9 mmol) was added. The red reaction mixture was stirred at 0 °C for 45 min, treated with 9 mL of 6 N HCl (3 × 17.9 mmol) at 25 °C overnight (12 h), poured onto H_2O , and extracted with $CH_3OH-CH_2Cl_2$ (1:9, 5 × 30 mL). The combined organic phases were dried (MgSO₄) and concentrated. The crude intermediate 15, a yellow-orange solid, was dissolved in 5 mL of CH₂Cl₂ and 5 mL of (CF₃CO)₂O, and the reaction mixture was stirred at 25 °C overnight (14 h). Evaporation of the solvent and chromatography (SiO₂, 25% EtOAc-hexane) afforded 2.9 g (13.2 mmol, 74%) of pure 13 as a dark orange solid: mp 139-139.5 °C (EtOAc-hexane); ¹H NMR (CDCl₃, 400 MHz) δ 7.90 (1H, dd, J = 0.4, 8.1 Hz), 7.85 (1H, br d, J = 6.9 Hz), 7.79 (1H, d, J = 9.4

(30) For **50**: ¹H NMR (CDCl₃, 400 MHz) δ 8.73 (1H, d, J = 1.2 Hz), 8.65 (1H, d, J = 5.9 Hz), 8.17 (1H, dd, J = 1.2, 7.9 Hz), 8.02 (1H, d, J = 7.9 Hz), 7.69 (1H, d, J = 5.9 Hz), 4.23 (2H, s), 4.18 (3H, s), 4.14 (3H, s), 4.03 (3H, s), 3.50 (1H, br s), 3.40 (2H, br s), 1.03 (6H, s). For **51**: ¹H NMR (CDCl₃, 400 MHz) δ 8.61 (1H, d, J = 5.9 Hz), 8.09 (1H, dd, J = 1.2, 7.6 Hz), 7.70 (1H, d, J = 5.9 Hz), 7.54 (1H, dd, J = 1.2, 7.6 Hz), 7.48 (1H, t, J = 7.6 Hz), 4.32 (2H, s), 4.15 (3H, s), 4.11 (3H, s), 4.03 (3H, s), 3.47 (2H, br s), 3.35 (2H, br s), 0.97 (6H, s).

(31) L1210 cytotoxic testing results (IC₅₀) are provided in the supporting information. In contrast to the reported potent cytotoxic activity of pareitropone (6), IC₅₀ = 0.0008 μ g/mL (P388), the L1210 cytotoxic activities of granditropone (7, IC₅₀ = 1 μ g/mL) and 20 (IC₅₀ = 16 μ g/mL) proved comparable or less potent than those of the corresponding tropolones: 1 (0.2 μ g/mL), 2 (4 μ g/mL), 3 (1 μ g/mL), and 25 (0.1 μ g/mL). The most interesting observation was the equivalent cytotoxicities (IC₅₀ = 0.1-0.2 μ g/mL) of 25 and grandirubrine (1), indicating that the three methoxy substituents and the quinoline nitrogen of the natural products may not be essential to the observed properties.

Scheme 11

Hz), 7.76 (1H, dd, J=0.4, 8.2 Hz), 7.62 (1H, dd, J=0.4, 6.9 Hz), 7.56 (1H, dd, J=6.9, 8.1 Hz), 7.51 (1H, dd, J=6.9, 8.2 Hz), 6.22 (1H, d, J=9.4 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 162.1, 161.3, 138.7, 131.0, 130.8, 128.7, 128.6, 128.0, 127.3, 127.9, 126.5, 123.4, 121.9, 116.0, 111.4; IR (KBr) $\nu_{\rm max}$ 3050, 1725, 1572, 1517, 1356, 1210, 1172, 1065, 1035, 820, 770 cm⁻¹; FABHRMS (NBA) m/e 221.0601 (M⁺ + H, C₁₅H₈O₂ requires m/e 221.0603). Anal. Calcd for C₁₅H₈O₂: C, 81.81; H, 3.66. Found: C, 81.59; H, 3.64.

Method B. A solution of 1-acenaphthenone 11 (40 mg, 0.24 mmol) in 2 mL of anhydrous THF was added dropwise to a stirred solution of 0.26 mL of t-BuOK (1 M in t-BuOH, 0.26 mmol, 1.1 equiv) in 3 mL of THF at -78 °C under N₂, and the resulting dark purple solution was stirred for 30 min. The reaction mixture was allowed to warm to 0 °C, and 12 (45 mg, 0.24 mmol) was added. The red reaction mixture was stirred at 0 °C for 45 min before being quenched with the addition of saturated aqueous NH₄Cl (1 mL). The mixture was extracted with CH₃OH-CHCl₃ (1:9, 7 mL × 5). The combined organic phases were dried (MgSO₄) and concentrated in vacuo. The crude intermediate 14 was dissolved in 1 mL of CF₃COOH, and the reaction mixture was stirred at 25 °C for 14 h. After evaporation of CF₃COOH, the crude intermediate 16 was dissolved in 5 mL of THF. The mixture was treated with 1 mL of aqueous 1 N LiOH under N₂ for 6 h at 25 °C, then treated with 1 mL of aqueous 1 N HCl. The reaction mixture was extracted with CH2Cl2 (5 mL × 3), dried (MgSO4), and concentrated in vacuo. The crude intermediate 15 was treated with 2 mL of Ac_2O at 25 °C for 48 h. Evaporation of the solvent and chromatography (SiO₂, 25% EtOAc-hexane) afforded 13 (46 mg, 0.21 mmol. 88%).

3b-Hydroxy-5-carboxyl-4-oxo-3b,3c,4a,5-tetrahydro-4H-cyclopropano[f]benz[a]acenaphthylene Lactone 2,2-Dimethyl-1,3-propylene Ketal (20). Method A. a-Pyrone 13 (20 mg, 0.09 mmol), cyclopropenone ketal 1821 (127 mg, 0.91 mmol, 10 equiv), and 0.2 mL of CHCl3 were combined in a Teflon tube. The tube was sealed on both ends with brass clamps and placed under pressure (12 kbar)32 for 20 h at 25 °C. Chromatography (SiO₂, 15% EtOAc-hexane eluent) afforded 23.9 mg (0.066 mmol, 73%) of 20 as a glassy material: mp 158-159 °C (dec, EtOAc-hexane, colorless prisms); ¹H NMR (CDCl₃, 400 MHz) δ 7.88 (1H, dd, J = 0.6, 8.2 Hz), 7.80-7.77 (2H, m), 7.66 (1H, dd, J = 7.0, 8.2 Hz), 7.61-7.57 (2H, m), 6.88 (1H, d, J = 6.2)Hz), 3.99 (1H, ddd, J = 0.6, 3.8, 6.2 Hz), 3.83 (1H, d, J = 11.0 Hz), 3.75 (1H, d, J = 11.0 Hz), 3.57 (2H, s), 2.16 (1H, dd, J = 3.8, 10.5)Hz), 1.72 (1H, dd, J = 0.6, 10.5 Hz), 1.16 (3H, s), 1.04 (3H, s); ¹³C NMR (CDCl₃, 100 MHz) δ 172.2, 151.8, 139.3, 136.6, 132.0, 131.1, 128.3, 128.1, 126.4, 125.7, 121.0, 119.6, 118.7, 97.6, 86.1, 77.0, 76.2, 41.3, 31.8, 31.5, 30.5, 22.6, 22.1; IR (KBr) ν_{max} 2958, 2868, 1762, 1392, 1190, 1084, 997, 787 cm⁻¹.

A single crystal X-ray structure determination was conducted on **20** with colorless cubes grown from EtOAc-hexane.²² Crystal data: $C_{23}H_{20}O_4$, MW = 360.4, monoclinic, space group C2/c (No. 15, $C_{2,h}6$), a = 16.096(3) Å, b = 11.620(4) Å, c = 20.200(2) Å, $\beta = 109.33(1)^\circ$, V = 3564(1) Å³, F(000) = 1520, Z = 8, De = 1.343 mg/m³, $\mu = 0.700$ mm⁻¹ (Cu K α).

Method B. α -Pyrone 13 (20 mg, 0.091 mmol), cyclopropenone ketal 18 (38 mg, 0.27 mmol, 3 equiv), and 0.9 mL of THF were combined in Teflon tube. The tube was sealed on both ends with brass clamps and placed under pressure (12 kbar) for 64 h at 25 °C. Chromatography (SiO₂, 20–60% EtOAc—hexane eluent) afforded 14 mg (0.039 mmol, 43%) of 20 and 9 mg (0.039 mmol, 43%) of 23.

5-Oxo-6-amino-5*H***-cycloheptatrieno[a]acenaphthylene (23).** A solution of **20** (236 mg, 0.66 mmol) in 3 mL of pyridine was warmed at 90 °C with stirring for 2 d. The yellow reaction mixture was cooled to 25 °C and concentrated in vacuo. Chromatography (SiO₂, 5–40% EtOAc-CH₂Cl₂ gradient elution) afforded 120 mg (0.52 mmol, 80%) of **23** as a dark yellow wax: 1 H NMR (CDCl₃, 400 MHz) δ 8.00 (1H, d, J = 8.2 Hz), 8.00 (1H, d, J = 7.2 Hz), 7.96 (1H, d, J = 8.2 Hz), 7.95 (1H, d, J = 7.0 Hz), 7.88 (1H, dd, J = 0.6, 2.8 Hz), 7.80–7.60 (3H, m), 7.30 (1H, dd, J = 8.4, 12.1 Hz), 7.16 (1H, ddd, J = 0.6, 2.8, 12.1 Hz); 13 C NMR (CDCl₃, 100 MHz) δ 186.4, 146.7, 145.8, 140.6, 136.2, 136.1, 135.4, 134.8, 132.3, 130.3, 128.5, 128.4, 128.3, 127.5, 124.8, 119.9, 118.9; IR (KBr) ν_{max} 3422, 1642, 1560, 1444, 1221, 827, 772 cm⁻¹; FABHRMS (NBA) m/e 231.0814 (M⁺ + H, C₁₇H₁₀O requires m/e 231.0810).

5-Oxo-6-amino-5H-cycloheptatrieno[a]acenaphthylene (24). A solution of 23 (22 mg, 0.1 mmol) in 8 mL of THF-EtOH (1:1) was treated with 10 drops of hydrazine hydrate at 0 °C. The solution was allowed to warm to 25 °C and was stirred for 4 d before the reaction mixture was concentrated in vacuo. Chromatography (SiO₂, 75%

⁽³²⁾ The pressure-promoted Diels-Alder reactions were carried out in a AGP-10002 pressure generator manufactured by Leco Corp., Tem-Press Division, Bellefonte, PA 16823.

EtOAc—hexane) afforded 18 mg (0.73 mmol, 76%) of **24** as a red solid: mp 240–242 °C (CH₃OH—CH₂Cl₂); ¹H NMR (CDCl₃, 400 MHz) δ 8.04 (1H, d, J = 7.1 Hz), 8.00 (1H, s), 7.94 (1H, d, J = 8.2 Hz), 7.83 (1H, d, J = 10.1 Hz), 7.82 (1H, d, J = 7.0 Hz), 7.81 (1H, d, J = 8.1 Hz), 7.69 (1H, dd, J = 7.1, 8.1 Hz), 7.63 (1H, dd, J = 7.0, 8.2 Hz), 6.94 (1H, d, J = 10.1 Hz), 6.00 (2H, br s); ¹³C NMR (CDCl₃, 100 MHz) δ 175.3, 155.8, 146.7, 138.2, 137.7, 134.7, 133.4, 130.1, 128.5, 128.4, 128.3, 127.6, 125.7, 123.4, 120.5, 117.7, 111.0; IR (KBr) ν_{max} 3414, 3294, 1595, 1491, 1460, 1424, 819, 769 cm⁻¹; FABHRMS (NBA) m/e 246.0925 (M⁺ + H, C₁₇H₁₁NO requires m/e 246.0919).

6-Oxo-5-hydroxy-6H-cycloheptatrieno[a]acenaphthylene (25). A solution of **24** (18 mg, 0.074 mmol) in 1:1 EtOH-2 N aqueous KOH (2 mL) was warmed at 100 °C under N₂ for 2 d. After cooling to 25 °C, the crude reaction mixture was diluted with CH₂Cl₂, acidified with 10% aqueous HCl, and extracted with CH₂Cl₂ (3 × 6 mL). The combined organic layers were dried (MgSO₄) and concentrated in vacuo to afford 12 mg (0.049 mmol, 66%) of **25** as a purple solid: mp 259-260 °C (CH₃OH-CH₂Cl₂); ¹H NMR (DMSO- d_6 , 400 MHz) δ 8.47 (1H, d, J = 7.0 Hz), 8.29 (1H, d, J = 10.7 Hz), 8.23 (1H, s), 8.23 (1H, d, J = 7.0 Hz), 8.16 (1H, d, J = 8.0 Hz), 8.04 (1H, d, J = 8.0 Hz), 7.90-7.70 (2H, m), 7.33 (1H, d, J = 10.7 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 171.9, 168.7, 147.1, 138.0, 137.2, 136.8, 132.4, 129.9, 129.4, 129.0, 128.7, 128.5, 127.3, 121.7, 119.8, 119.7, 119.0; IR (KBr) ν_{max} 3419, 3205, 1607, 1451, 1426, 1256, 818, 768 cm⁻¹; FABHRMS (NBA) m/e 247.0768 (M⁺ + H, C₁₇H₁₀O₂ requires m/e 247.0759).

5-Oxo-6-methoxy-5*H*-cycloheptatrieno[*a*]acenaphthylene (26) and 6-Oxo-5-methoxy-6*H*-cycloheptatrieno[*a*]acenaphthylene (27). A solution of 25 (3.0 mg, 0.012 mmol) in 1 mL of CH₃OH-CH₂Cl₂ (1: 1) was treated with excess diazomethane at 25 °C for 3 h before being concentrated in vacuo. Chromatography (SiO₂, 2% CH₃OH-CH₂Cl₂) afforded 1.0 mg (0.0038 mmol, 32%) of 26 and 27 as a mixture in a ratio of 3:2 determined by ¹H NMR.

A solution of 25 (2.0 mg, 0.008 mmol) in 1 mL of THF and 0.5 mL of CH₃OH was treated with trimethylsilyldiazomethane (0.2 mL of 1 M in hexane) at 25 °C for 4 d before being concentrated in vacuo. Chromatography (SiO₂, 2% CH₃OH—CH₂Cl₂) afforded 1.6 mg (0.0062 mmol, 77%) of a mixture of 26 and 27 in a ratio of 1:1 determined by $^{\rm I}$ H NMR

A mixture of **25** (2.0 mg, 0.008 mmol), K_2CO_3 (3.7 mg, 0.025 mmol), and CH_3I (0.2 mL) in 1 mL of acetone was warmed at reflux for 2 d. The reaction mixture was poured onto H_2O (2 mL) and extracted with CH_2Cl_2 (3 × 3 mL), dried (MgSO₄), and concentrated in vacuo. Chromatography (SiO₂, 2% $CH_3OH-CH_2Cl_2$) afforded 1.0 mg (0.0038 mmol, 43%) of a mixture of **26** and **27** in a ratio of 3:1 determined by 1H NMR.

A mixture of **25** (2.0 mg, 0.008 mmol), K_2CO_3 (3.7 mg, 0.025 mmol), and CH_3I (0.5 mL) in 2 mL of DMF was stirred at 25 °C for 12 h. The reaction mixture was poured onto H_2O (2 mL) and extracted with CH_2 - Cl_2 (3 × 3 mL), dried (MgSO₄), and concentrated in vacuo. Chromatography (SiO₂, 2% $CH_3OH-CH_2Cl_2$) afforded 1.5 mg (0.0057 mmol, 72%) of a mixture of **26** and **27** in a ratio of 2:1 determined by ¹H NMR. For **26** and **27** (2:1) mixture: ¹H NMR (CDCl₃, 400 MHz) δ 8.10 (0.5H, d, J = 7.0 Hz), 8.05 (1H, s), 8.02 (0.5H, d, J = 8.2 Hz), 8.01 (1H, d, J = 7.0 Hz), 8.00 (0.5H, d, J = 12 Hz), 7.99 (0.5H, d, J = 7.1 Hz), 7.98 (1H, d, J = 8.2 Hz), 7.94 (0.5 H, d, J = 8.2 Hz), 7.90 (2H, d, J = 7.6 Hz), 7.79 (1H, d, J = 10.0 Hz), 7.77–7.63 (3H, m), 7.60 (0.5H, s), 7.37 (0.5H, d, J = 12.0 Hz), 6.89 (1H, d, J = 10.0 Hz), 4.17 (1.5H, s), 4.03 (3H, s).

8-Bromo-5,6,7-trimethoxyisoquinoline (37). 5,6,7-Trimethoxyisoquinoline (36, 1.67 g, 7.6 mmol), NBS (1.43 g, 8.0 mmol, 1.05 equiv), and 3 drops of concentrated H_2SO_4 were combined in 75 mL of anhydrous THF, and the mixture was stirred under N_2 at 25 °C for 2 h. The reaction mixture was neutralized with the addition of 5% aqueous NaHCO₃, poured onto H_2O (50 mL), extracted with CH₂Cl₂ (3 × 60 mL), dried (MgSO₄), and concentrated in vacuo. Chromatography (SiO₂, 30% EtOAc—hexane) afforded 1.99 g (87%) of 37 as a white solid identical in all respects to authentic material:²⁸ mp 64—66 °C (lit. mp 64—67 °C).²⁸

5,6,7-Trimethoxyisoquinoline-8-carboxaldehyde (38). A stirred solution of 37 (3.96 g, 13.3 mmol) in 66 mL of anhydrous Et₂O under Ar at 0 °C was treated with *n*-BuLi (5.85 mL of 2.5 M in hexane, 14.6 mmol, 1.1 equiv), and the reaction mixture was stirred at 0 °C for 30

min before the addition of DMF (5.15 mL, 66.5 mmol, 5 equiv). After the mixture was stirred for 30 min at 0 °C, $\rm H_2O$ was added, and the reaction mixture was allowed to warm to 25 °C. The mixture was poured onto $\rm H_2O$ and extracted with $\rm Et_2O$ (50 mL) and $\rm CH_2Cl_2$ (2 × 50 mL). The combined organic extracts were dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, 30–70% EtOAchexane gradient elution) afforded 2.04 g (8.26 mmol, 61%) of 38 as a white solid identical in all respects to authentic material (mp 92–92.5 °C, lit. mp 92–92.5 °C) and 580 mg (2.65 mmol, 20%) of 5,6,7-trimethoxyisoquinoline (36).

8-((1,3-Dithiane-2-ylidene)methyl)-5,6,7-trimethoxyisoquinoline (39). A stirred solution of 2-(trimethylsilyl)-1,3-dithiane (5.97 g, 31.1 mmol, 1.3 equiv) in 40 mL of anhydrous THF at 0 °C under Ar was treated with *n*-BuLi (11.5 mL of 2.5 M in hexane, 28.7 mmol, 1.2 equiv). The reaction mixture was stirred at 0 °C for 15 min and allowed to cool to -78 °C before the dropwise addition of a solution of **38** (5.91 g, 23.9 mmol) in 20 mL of anhydrous THF. After stirring for 30 min, the reaction mixture was treated with H₂O, poured onto H₂O, and extracted with CH₂Cl₂ (2 × 80 mL). The combined organic phases were dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, CH₂Cl₂–EtOAc–hexane (3:1:1)) afforded 9.29 g (26.6 mmol, 89%) of **39** as a pale yellow oil identical in all respects with authentic material.⁸

Methyl 2-(5,6,7-Trimethoxyisoquinolyl)acetate (40). A solution of 39 (9.29 g, 26.6 mmol) and HgCl₂ (15.9 g, 58.5 mmol, 2.2 equiv) in 1000 mL of 9:1 CH₃OH-H₂O was warmed at reflux under N₂ overnight. The white precipitate was filtered and washed with 9:1 CH₂-Cl₂-CH₃OH. The combined filtrates were concentrated in vacuo. The residue was dissolved in CH₂Cl₂, washed with 5% aqueous NaHCO₃, dried (MgSO₄), and concentrated in vacuo to afford crude 40 identical to authentic material⁸ which was used directly in the next reaction.

Methyl 2-(N-Tosyl-1-cyano-1,2-dihydro-5,6,7-trimethoxy-8-isoquinolyl)acetate (41). p-Toluenesulfonyl chloride (7.62 g, 39.3 mmol, 1.5 equiv) was added to a vigorously stirred solution of crude 40 (26.6 mmol) and KCN (5.19 g, 79.8 mmol, 3 equiv) in 500 mL of 1:1 CH₂-Cl₂-H₂O, and the reaction mixture was stirred at 25 °C overnight. The reaction mixture was poured onto H₂O and extracted with CH₂Cl₂ (3 × 100 mL). The combined organic phases were dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, 20% EtOAc-hexane) afforded 10.4 g (24.3 mmol, 91%) of 41 as a colorless oil identical in all respects with authentic material.⁸

Methyl 8-Amino-4,5,6-trimethoxycyclopentadieno[2,1,5-ij]isoquinoline-7-carboxylate (42). A stirred solution of 41 (10.4 g, 24.3 mmol) in 250 mL of anhydrous THF under N₂ was treated with t-BuOK (73 mL of 1 M in t-BuOH, 73 mmol, 3 equiv). The resulting purple solution was stirred at 25 °C for 6 h before the addition of saturated aqueous NH₄Cl. The red mixture was poured onto saturated aqueous NH₄Cl and extracted with EtOAc. The combined organic phases were dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, CH₂-Cl₂-EtOAc—hexane (1:1:1)) afforded 6.31 g (20.0 mmol, 82%; typically 80–98%) of 42 as a red crimson solid identical in all respects with authentic material (mp 145–146 °C, lit. mp 155–156 °C).8

8-Oxo-4,5,6-trimethoxycyclopenteno[1,2,3-ij]isoquinoline (43). A purple solution of **42** (118 mg, 0.37 mmol) in 10 mL of 9:1 dioxane-1 N aqueous HCl was warmed at 110 °C under N₂ with vigorous stirring for 10 min before 1 N aqueous KOH (1 mL) was added. The resulting mixture was stirred at 110 °C for 3 h. The yellow reaction mixture was cooled to 25 °C, poured onto 5% aqueous NaHCO₃, and extracted with EtOAc (3×). The combined organic phases were dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, 50% EtOAchexane) afforded 86 mg (0.33 mmol, 89%) of **43** identical in all respects with authentic material (mp 124 °C, lit. mp 124-125 °C).

3-Aza-9,10,11-trimethoxy-4-oxa-5-oxobenz[a]acenaphthylene (44). A solution of 43 (80 mg, 0.31 mmol) in anhydrous THF (7 mL) was added dropwise to a stirred solution of t-BuOK (0.46 mL, 1 M in t-BuOH, 0.46 mmol, 1.5 equiv) in THF (3 mL) at -78 °C under N_2 , and the resulting dark purple solution was stirred for 30 min. The reaction mixture was allowed to warm to 0 °C before 12 (86 mg, 0.46 mmol, 1.5 equiv) was added. The red reaction mixture was stirred at 0 °C for 15 min, quenched with the addition of saturated aqueous NH₄-Cl (2 mL). The mixture was extracted with CH₃OH—CHCl₃ (1:9, 7 × 10 mL). The combined organic phases were dried (MgSO₄) and

concentrated in vacuo. The crude intermediate 45 was dissolved in CF₃COOH (4 mL), and the reaction mixture was stirred at 25 °C for 14 h. After evaporation of CF₃COOH, the crude intermediate 46 was dissolved in 20 mL of H₂O. The mixture was treated with 2 mL of 1 N aqueous LiOH under N2 for 15 min at 0 °C before neutralization through addition of 2 mL of 1 N aqueous HCl. The reaction mixture was extracted with EtOAc (60 mL), washed with H_2O (3 × 10 mL), dried (MgSO₄), and concentrated in vacuo. The crude intermediate 48 was treated with 4 mL of Ac₂O at 25 °C for 48 h. Evaporation of the solvent and chromatography (SiO₂, 20-40% EtOAc-hexane) afforded 50 mg of 44 (0.16 mmol, 52%). For 45: ¹H NMR (CD₃OD, 400 MHz) δ 8.52 (1H, d, J = 5.6 Hz), 8.20 (1H, s), 7.83 (1H, d, J =5.6 Hz), 4.06 (3H, s), 4.02 (3H, s), 3.84 (3H, s), 1.77 (6H, s). For 46: ¹H NMR (CDCl₃, 400 MHz) δ 9.15 (1H, s), 8.88 (1H, d, J = 5.6 Hz), 7.95 (1H, d, J = 5.6 Hz), 4.27 (3H, s), 4.18 (3H, s), 4.00 (3H, s). For 44: recrystallized from EtOAc-hexane, mp 179-180 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.66 (1H, d, J = 5.6 Hz), 7.91 (1H, d, J = 9.4Hz), 7.71 (1H, d, J = 5.6 Hz), 6.31 (1H, d, J = 9.4 Hz), 4.15 (3H, s), 4.11 (3H, s), 3.99 (3H, s); 13 C NMR (CDCl₃, 100 MHz) δ 161.0, 158.6, $153.6,\ 150.5,\ 150.3,\ 148.7,\ 145.8,\ 145.4,\ 139.5,\ 126.2,\ 118.7,\ 117.7,$ 116.1, 114.0, 62.2, 62.1, 61.4; IR (KBr) ν_{max} 3447, 2950, 1730, 1629, 1466, 1414, 1341, 1146, 1014 cm⁻¹. Anal. Calcd for C₁₇H₁₃NO₅: C, 65.59; H, 4.21; N, 4.50. Found: C, 65.46; H, 4.24; N, 4.54.

3-Aza-3b-hydroxy-5-carboxy-4-oxo-7,8,9-trimethoxy-3b,3c,4a,5tetrahydro-4H-cyclopropano[f]benz[a]acenaphthylene Lactone 2,2-Dimethyl-1,3-propylene Ketal (49) and 3-Aza-9,10,11-trimethoxy-5-oxo-5*H*-cycloheptatrieno[*a*]acenaphthylene (7, Granditropone). α-Pyrone 44 (40 mg, 0.13 mmol), cyclopropenone ketal 18 (55 mg, 0.39 mmol, 3 equiv), pyridine (31 mg, 0.39 mmol, 3 equiv), and 1.3 mL of CHCl3 were combined in Teflon tube. The tube was sealed on both ends with brass clamps and placed under pressure (12 kbar)³² for 2.5 h at 25 °C. Chromatography (Al₂O₃, 10-100% EtOAc-hexane gradient elution) afforded 21 mg (0.047 mmol, 36%) of exo-49, 8.0 mg (0.024 mmol, 20%) of granditropone (7), and 5.0 mg (0.012 mmol, 10%) of **50**.³⁰ For *exo-***49**: ¹H NMR (CDCl₃, 400 MHz) δ 8.58 (1H, d, J = 6.0 Hz), 7.69 (1H, d, J = 6.0 Hz), 6.90 (1H, d, J = 6.1 Hz), 4.17 (1H, d, J = 10.8 Hz), 4.08 (3H, s), 4.03 (3H, s), 4.02 (3H, s), 4.00 (1H, dd, J = 4.0, 6.1 Hz), 3.76 (1H, dd, J = 1.8, 10.8 Hz), 3.65(1H, d, J = 10.7 Hz), 3.54 (1H, dd, J = 1.8, 10.7 Hz), 2.22 (1H, dd, J = 1.8, 10.7 Hz)J = 4.0, 10.3 Hz), 1.69 (1H, d, J = 10.3 Hz), 1.23 (3H, s), 0.97 (3H, s); ¹³C NMR (CDCl₃, 100 MHz) δ 172.1, 158.7, 150.4, 149.5, 148.4, 147.4, 145.2, 130.1, 127.1, 122.1, 119.1, 114.2, 97.2, 85.7, 77.2, 77.2, 61.8, 61.5, 61.0, 41.7, 32.3, 30.8, 30.2, 23.3, 22.1; IR (KBr) ν_{max} 3447, 2948, 1768, 1536, 1559, 1411, 1364, 1084 cm⁻¹; FABHRMS (NBA) m/e 584.0667 (M⁺ + H, C₂₅H₂₅NO₇ requires 584.0685).

For 7: purple solid, mp 155–157 °C (EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 8.66 (1H, d, J = 5.8 Hz), 8.12 (1H, dd, J = 0.8, 2.8 Hz), 7.97 (1H, dt, J = 8.5, 0.8 Hz), 7.75 (1H, d, J = 5.8 Hz), 7.24 (1H, dd, J = 8.5, 12.3 Hz), 7.07 (1H, ddd, J = 0.8, 2.8, 12.3 Hz), 4.15 (3H, s), 4.14 (3H, s), 4.04 (3H, s); ¹³C NMR (CDCl₃, 100 MHz) δ 186.8, 157.3, 152.0, 150.0, 145.5, 145.4, 143.4, 140.7, 135.7, 133.1, 128.0, 127.0, 126.2, 122.9, 121.4, 115.1, 61.9, 61.5, 61.3; IR (KBr) ν_{max} 3447, 2923, 1617, 1569, 1458, 1410, 1365, 1291, 1143, 1010 cm⁻¹; FABHRMS (NBA) m/e 322.1072 (M⁺ + H, C₁₉H₁₅NO₄ requires 322.1079).

A solution of 49 (11.4 mg, 0.025 mmol) in 10 mL of THF was treated with 1 mL of 3.6 M HCl-EtOAc at 25 °C for 6 d. The reaction mixture was poured onto 20 mL of saturated NaHCO₃ and extracted with CH_2Cl_2 (3 × 10 mL). The combined organic phase was dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, 25-80% EtOAc-hexane) afforded 2.7 mg (0.006 mmol, 24%) of recovered 49 and 3.3 mg (0.01 mmol, 41%, 54% based on recovered 49) of 7.

A solution of **44** (40 mg, 0.13 mmol), **18** (55 mg, 0.39 mmol, 3 equiv), and pyridine (31 mg, 0.39 mmol, 3 equiv) in CHCl₃ (1.3 mL) in a Teflon tube sealed with brass clamps was placed under 12 kbar pressure³² for 2.5 h. Following depressurization and removal of the solvent in vacuo, the crude product was placed in 30 mL of THF and treated with 3.6 M HCl-EtOAc (3 mL). The resulting mixture was stirred at 25 °C for 4 d before being poured onto 20 mL of saturated aqueous NaHCO₃ and extracted with EtOAc (3 × 30 mL). The combined organic phase was dried (MgSO₄) and concentrated in vacuo. Chromatography (SiO₂, 25–80% EtOAc-hexane) afforded **7** (16–24 mg, 0.072 mmol, 40–60%) and **50** (5.0 mg, 0.012 mmol, 10%).

3-Aza-9,10,11-trimethoxy-5-oxo-6-amino-5H-cycloheptatrieno[a]-acenaphthylene (52). A solution of 7 (6.1 mg, 0.019 mmol) in 8 mL of THF was treated with 8 drops of hydrazine hydrate at 0 °C. The solution was allowed to warm to 25 °C and was stirred for 18 h. The reaction mixture was poured onto 5 mL of aqueous NaHCO₃, extracted with EtOAc (10 mL), and washed with saturated aqueous NaCl. The organic phase was dried (Na₂SO₄) and concentrated in vacuo. Chromatography (Al₂O₃, 75–100% EtOAc—hexane, 10–20% CH₃OH—CH₂Cl₂) afforded 5.0 mg (0.014 mmol, 78%) of **52** as a red glassy material: ¹H NMR (CDCl₃, 400 MHz) δ 8.66 (1H, d, J = 5.8 Hz), 8.24 (1H, s), 8.07 (1H, d, J = 10.3 Hz), 7.72 (1H, d, J = 5.8 Hz), 6.90 (1H, d, J = 10.3 Hz), 4.11 (3H, s), 4.10 (3H, s), 4.04 (3H, s); IR (KBr) ν_{max} 3421, 2939, 1599, 1507, 1458, 1411, 1087, 1016, 976 cm⁻¹; FABHRMS (NBA) m/e 337.1173 (M⁺ + H, C₁₉H₁₆N₂O₄ requires m/e 337.1188).

Grandirubrine (1). A solution of **52** (5.0 mg, 0.014 mmol) in 5 mL of CH₃OH and 1.6 mL of 2 N aqueous KOH was warmed at 85 °C under N₂ for 30 h. After cooling to 25 °C, the crude reaction mixture was diluted with 8 mL of H₂O, washed with CH₂Cl₂, acidified with the addition of 3.2 mL of 1 N aqueous HCl, and extracted with CH₂-Cl₂ (3 × 4 mL). The combined organic layers were dried (Na₂SO₄) and concentrated in vacuo to afford 3.3 mg (0.0098 mmol, 70%) of 1: mp 201-202 °C (lit.⁴ mp 201-203 °C); ¹H NMR (CDCl₃, 400 MHz) δ 8.73 (1H, d, J = 5.7 Hz), 8.42 (1H, s), 8.33 (1H, d, J = 10.6 Hz), 7.80 (1H, d, J = 5.7 Hz), 7.42 (1H, d, J = 10.6 Hz), 4.17 (3H, s), 4.14 (3H, s), 4.04 (3H, s); ¹³C NMR (CDCl₃, 100 MHz) δ 173.5, 167.7, 157.5, 152.4, 150.0, 149.8, 146.2, 145.6, 137.2, 131.3, 126.2, 124.4, 121.7, 120.2, 119.9, 115.8, 62.0, 61.5, 61.5; IR (KBr) ν_{max} 3442, 1617, 1559, 1458, 1406, 1261 cm⁻¹; FABHRMS (NBA) m/e 338.1080 (M⁺ + H, C₁₉H₁₅NO₅ requires 338.1087).

Imerubrine (2) and Isoimerubrine (3). A solution of 1 (3.1 mg, 0.0092 mmol) in 3 mL of CH₃OH-THF (2:1) was treated with 0.5 mL of a solution containing TMSCHN₂ (2 M in hexane solution) at 25 °C for 20 h before being concentrated in vacuo. Chromatography (SiO₂, 60–100% EtOAc—hexane) afforded 1.32 mg (0.0038 mmol, 41%) of 2 and 1.11 mg (0.0032 mmol, 35%) of 3. For 2: R_f 0.24 (EtOAc); mp 181–183 °C (lit. ^{1,2} mp 183–185 °C); ¹H NMR (CDCl₃, 400 MHz) δ 8.67 (1H, d, J = 5.7 Hz), 8.28 (1H, s), 8.05 (1H, d, J = 10 Hz), 7.74 (1H, d, J = 5.7 Hz), 6.86 (1H, d, J = 10 Hz), 4.14 (3H, s), 4.12 (3H, s), 4.04 (3H, s), 4.00 (3H, s); ¹³C NMR (CDCl₃, 100 MHz) δ 179.4, 164.1, 157.6, 151.1, 150.2, 148.8, 145.6, 145.3, 136.7, 128.4, 126.3, 126.2, 126.1, 121.9, 115.0, 112.0, 61.9, 61.4, 61.2, 56.4; FABHRMS (NBA) m/e 352.1241 (M⁺ + H, C₂₀H₁₇NO₅ requires 352.1247).

For 3: R_f 0.18 (EtOAc); mp 183–184 °C (lit.⁴ mp 183–185 °C); 1H NMR (CDCl₃, 400 MHz) δ 8.72 (1H, d, J = 5.7 Hz), 8.27 (1H, d, J = 12.2 Hz), 7.90 (1H, s), 7.79 (1H, d, J = 5.7 Hz), 7.39 (1H, d, J = 12.2 Hz), 4.19 (3H, s), 4.17 (3H, s), 4.16 (3H, s), 4.01 (3H, s); 13 C NMR (CDCl₃, 100 MHz) δ 180.2, 164.4, 158.5, 154.4, 151.3, 149.5, 145.2, 139.0, 136.9, 136.5, 132.5, 125.9, 121.3, 121.0, 115.8, 106.8, 62.2, 62.0, 61.5, 56.9; FABHRMS (NBA) m/e 352.1258 (M⁺ + H, C₂₀H₁₇NO₅ requires 352.1247).

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Supporting Information Available: The results of the cytotoxic evaluation (L1210) of 1-3, 7, 8, 10, 44, 49, 52, 12, 20, and 23-25 (1 table) and details of the X-ray structure determination of 20 are provided (20 pages). This material is contained in many libraries on microfiche, immediately follows this article in the microfilm version of the journal, can be ordered from the ACS, and can be downloaded from the Internet; see any current masthead page for ordering information and Internet access instructions.

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