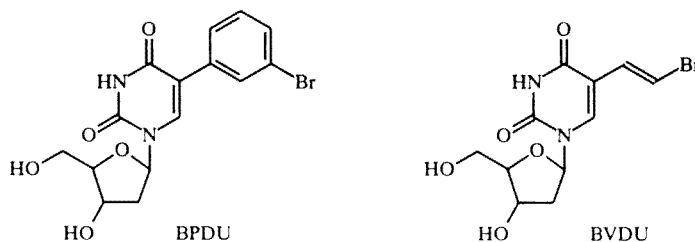


## SYNTHESIS OF VARIOUS 5-(3-SUBSTITUTED PHENYL)-2'-DEOXYURIDINES\*

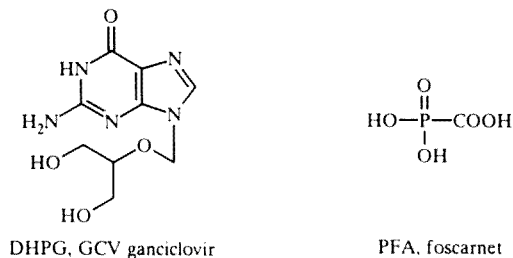
Ulf Wellmar, Anna-Britta Hörnfeldt, Salo Gronowitz,  
and Nils Gunnar Johansson

*A series of 5-aryl-2'-deoxyuridines has been prepared and evaluated as antiviral agents. The following substituents have been used in position 3 of the phenyl ring: chloro, iodo, amino, azido, methylthio, and vinyl. None of the new compounds showed any significant activity when tested against human immunodeficiency virus 1 (HIV-1), herpes simplex virus 1 (HSV-1), or human cytomegalovirus (HCMV).*

5-(3-Bromophenyl)-2'-deoxyuridine (BPDU) is structurally reminiscent of (*E*)-5-(2-bromovinyl)-2'-deoxyuridine (BVDU). BVDU has potent activity against herpes simplex virus type 1 (HSV-1), for a review see [1]. We have previously reported that BPDU has some activity against HSV-1, and in contrast to BVDU also a weak activity against human cytomegalovirus (HCMV) [2].



5-(Heteroaryl)-substituted 2'-deoxyuridines have earlier been reported to possess potent anti-HSV-1 activities [3-6]. However, we have found that in contrast to BVDU, the anti-HSV-1 activity of this class of compounds is highly dependent on the cells and virus strains used in the assay and differ by a factor 500 or more [2]. HCMV infections often lead to severe diseases in immunocompromised individuals, such as acquired immunodeficiency syndrome (AIDS) patients and transplant recipients [7-11].

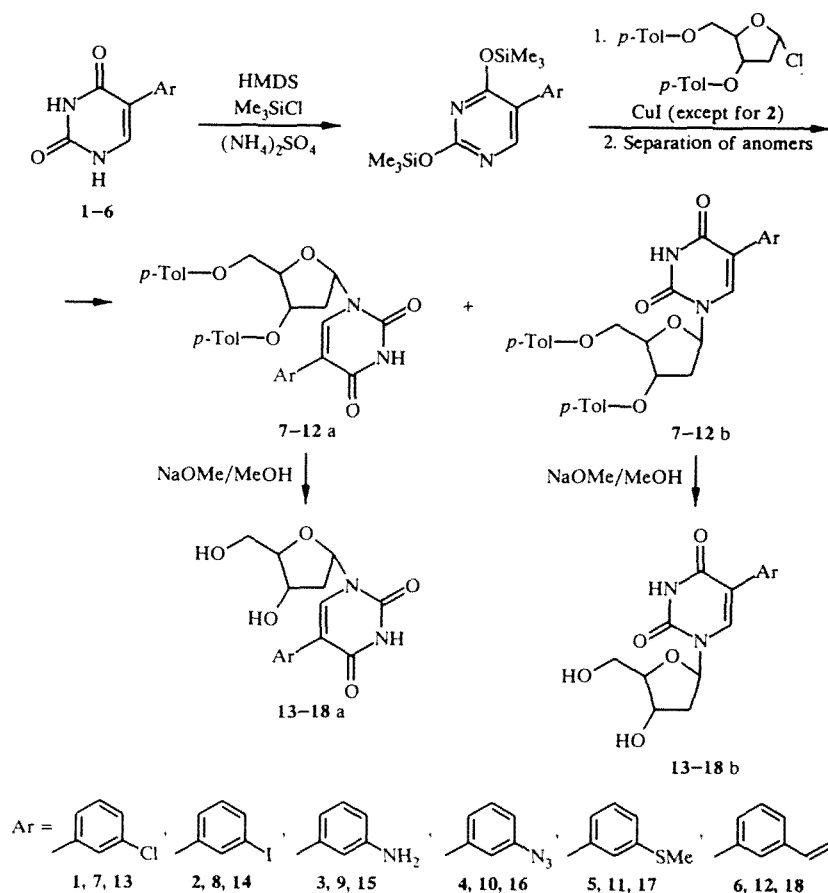


\*Dedicated to Professor Édmund Lukevits on the occasion of his 60th birthday.

The two drugs currently of choice for the treatment of HCMV, 9-(1,3-dihydroxy-2-propoxymethyl)guanine (DHPG, GCV ganciclovir) and phosphonoformic acid (PFA, foscarnet) [12-15], both result in serious side effects as well as the emergence of drug-resistant virus [15, 16].

In view of this we have studied, and earlier reported, antiviral activities of structural variations on the lead compound BPDU. A second *meta*-substituent on the 5-phenyl function does not alter the activity against HSV-1 and HCMV [17]. Bromo substituted 5-furyl- and 5-thienyl-2'-deoxyuridines also have about the same anti-HCMV activities and slightly differing anti-HSV-1 activities [2]. The ribofuranosyl, arabinofuranosyl, and 2'-deoxy-2'-fluoroarabinofuranosyl analogues of BPDU were slightly more active against HCMV but inactive against HSV-1 [18]. In continuing this structure-activity investigation we decided to prepare some 5-(3-substituted phenyl)-2'-deoxyuridines with 3-substituents that would be able to closely mimic the bromo group. The considerations in choosing the appropriate 3-substituents as well as the preparation of the corresponding uracils has been reported elsewhere [19]. In the present work these uracils have been coupled with 2-deoxy-3,5-di-O-*p*-toluoylribofuranosyl chloride. The  $\alpha$ - and  $\beta$ -anomers were separated and the sugar moiety deprotected, giving the desired 5-(3-substituted phenyl)-2'-deoxyuridines.

5-(3-Chlorophenyl)-, 5-(3-iodophenyl)-, 5-(3-aminophenyl)-, 5-(3-azidophenyl)-, 5-(3-methylthiophenyl)-, and 5-(3-styryl)uracils **1-6** (Scheme 1) were prepared as previously described [18]. The coupling between 2,4-di-(trimethylsiloxy)pyrimidines and 2-deoxy-3,5-di-O-*p*-toluoyl-*D*-erythropentose chloride [20] usually results in a mixture of  $\alpha$ - and  $\beta$ -anomers. By performing the reaction in anhydrous chloroform [21] with copper(I) iodide catalysis [22], the  $\alpha/\beta$ -ratio can be lowered due to less anomerization of the chloro sugar and faster coupling. Since it has been shown that the unnatural  $\alpha$ -anomers of certain nucleosides show biological activity [23-25], we were also interested in obtaining the  $\alpha$ -anomers of the 5-(3-substituted phenyl)-2'-deoxyuridines presented in this paper.



Scheme 1. Preparation of 5-(3-substituted phenyl)-2'-deoxyuridines.

The  $\beta$ -optimized conditions gave satisfactory yields (55-92%) except for the coupling with 2,4-di-(trimethylsiloxy)-5-(3-iodophenyl)pyrimidine. This could be due to Ullmann homo-coupling, and when performing the reaction without copper (I) iodide catalysis, 90% of the desired product (**8**) was isolated. For the coupling with 2,4-di-(trimethylsiloxy)-5-(3'-aminophenyl)-

TABLE 1. Antiviral Activity

Compound	HCMV ( $\mu\text{g/ml}$ )		HSV-1 ( $\mu\text{g/ml}$ )
	$ED_{50}^a$	$CD_{50}$	$ED_{50}^b$
13a	120	•	†
13b	120	•	†
14a	75	200	†
14b	85	200	†
15b	•	•	†
16a	140	•	†
16b	160	•	†
17a	•	•	†
17b	•	•	†
18a	•	•	†
18b	•	•	†
BPDU	60(10 <sup>c</sup> )	150	50(7 <sup>d</sup> )

• > 200  $\mu\text{g/ml}$ .

† 0-5% inhibition at 100  $\mu\text{g/ml}$ .

<sup>a</sup>ELISA assay; HEL cells.

<sup>b</sup>XTT assay; vero cells; C42 strain.

<sup>c</sup>CMV cytopathic assay (see text).

<sup>d</sup>HSV-1 plaque assay (see text).

pyrimidine no  $\alpha$ -anomer could be detected, while for the other five products the  $\alpha/\beta$ -ratio varied from 0.21 to 1.17. This allowed the isolation of sufficient amounts of both the  $\alpha$ - and the  $\beta$ -anomers.

The anomers of compounds **7**, **8**, and **10-12** could easily be separated by column chromatography using dichloromethane/ethyl acetate as eluent. Compound **9** was purified using dichloromethane/ethyl acetate/triethylamine as eluent.

The protecting *p*-toluoyl groups on the sugar unit were easily removed with a dilute solution of sodium methoxide in methanol. After being washed through a short column of silica gel 60 with methanol in order to remove the excess of sodium methoxide, the 5-(3-substituted phenyl)-2'-deoxyuridines **13-18** (Scheme 1) were obtained pure after column chromatography using dichloromethane/methanol as eluent.

## ANTIVIRAL ACTIVITY

### Results and Discussion

The anomers of compounds **13-18** were tested for inhibition of HCMV and HSV-1 multiplication in cell culture using ELISA and XTT assay conditions, respectively. The results are listed in Table 1. As can be seen, both  $\alpha$ - and  $\beta$ -anomers of the *meta*-iodo analogue (**14**) of BPDU were about equally active to BPDU in inhibition of HCMV, and both anomers of the *meta*-chloro (**13**) and *meta*-azido (**16**) analogues had diminished activities. All other analogues were inactive against HCMV, and compounds **13-18** did not inhibit HSV-1 under the assay conditions used.

Compounds **13-18** were also tested for inhibition of HIV-1 in an XTT assay and all were inactive at 100  $\mu\text{g/ml}$ . The mode of action for some of the weak anti-HCMV compounds is not known. Usually nucleoside analogues are active as triphosphates. However, HCMV does not encode a virus-specific nucleoside kinase enzyme capable of monophosphorylation, in contrast to HSV and varicella zoster virus [26, 27]. A protein kinase encoded by the HCMV UL97 gene is, however, capable of phosphorylating the acyclic nucleoside analogue ganciclovir [28, 29]. If this is the case also for the weakly active anti-HCMV compounds discussed here or if some other mechanism at hand is not known.

Results on anti-HSV-1 activities for this class of compounds obtained by using different cell lines and virus strains, and referred to in the introduction above, will be reported [30].

## EXPERIMENTAL

Melting points were recorded on a Lietz Wetzlar Microscope Heating Stage 350 Melting Point Apparatus and are uncorrected. The  $^1\text{H}$  NMR spectra were recorded on a Varian XL-300 spectrometer. The mass spectra were recorded on a JEOL-SX 102 spectrometer. The  $^1\text{H}$  NMR data for the four groups of compounds presented in this paper ( $\alpha$ - and  $\beta$ -anomers of protected and unprotected 2'-deoxyuridines) are very similar within the groups. Complete  $^1\text{H}$  NMR data are therefore only presented for one representative compound out of each class (**7a**, **7b**, **13a**, and **13b**). For compounds **14-18**, only data for the 5-substituent are presented. Elemental analyses were only performed on the  $\beta$ -anomers.

**General Procedure for the Coupling Between 5-(3-Substituted Phenyl)-2,4-di-(trimethylsiloxy)pyrimidines and 2-Deoxy-3,5-di-O-*p*-toluoyl-D-erythropentosyl Chloride.** A 250 ml flask equipped with a magnetic stirrer, a condenser and a nitrogen inlet was charged with 13.48 mmol of the 5-aryluracil (**1-6**), 70 ml of 1,1,1,3,3,3-hexamethyldisilazane, 1 ml of trimethylchlorosilane, and 0.5 g of ammonium sulfate. The reaction mixture was refluxed with stirring for 12 h. After cooling to room temperature, the excess of 1,1,1,3,3,3-hexamethyldisilazane was evaporated *in vacuo*. The residue was dissolved in 120 ml of anhydrous chloroform and added at 0°C to a solution of 4.76 g (12.26 mmol) of 2-deoxy-3,5-di-O-*p*-toluoyl-D-erythropentosyl chloride dissolved in 180 ml of anhydrous chloroform in a 500 ml flask equipped with a magnetic stirrer and a drying tube. Then 2.34 g (12.26 mmol) of copper (I) iodide was added, the ice bath was removed and the reaction mixture was allowed to stand with stirring at room temperature for 2 h. The chloroform was evaporated *in vacuo* and the residue was dissolved in 10 ml of chloroform. This solution was applied onto a column of silica gel 60 and the anomers were eluted with dichloromethane/ethyl acetate (**7**, **8**, **10-12**) or dichloromethane/ethyl acetate/triethyl amine (**9**).

**5-(3-Iodophenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (**8**)** was obtained as described above but without adding copper (I) iodide.

**5-(3-Chlorophenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (**7a** and **7b**).** Column chromatography using dichloromethane/ethyl acetate 9:1 as eluent gave 1.27 g (18%) of **7a** as a syrup and 4.72 g (67%) of **7b** as a white solid.

**7a:**  $^1\text{H}$  NMR (deuteriochloroform):  $\delta$  2.33 (3H, s, Ph-CH<sub>3</sub>), 2.41 (3H, s, Ph-CH<sub>3</sub>), 2.56 (1H, d,  $J$  = 15.5 Hz, H2'b), 2.99 (1H, dt,  $J$  = 6.6, 15.6 Hz, H2'a), 4.49 (1H, dd,  $J$  = 4.3, 11.9 Hz, H5'b), 4.56 (1H, dd,  $J$  = 4.2, 12.0 Hz, H5'a), 4.93 (1H, t,  $J$  = 4.0 Hz, H4'), 5.60 (1H, d,  $J$  = 5.9 Hz, H3'), 6.39 (1H, d,  $T$  = 5.8 Hz, H1'), 6.98-7.95 (17H, m, aromatic), 9.36 (1H, s, N-H<sub>3</sub>). HRMS: Found: 575.1577 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>28</sub>ClN<sub>2</sub>O<sub>7</sub>: 575.1585.

**7b:** mp 186-188°C;  $^1\text{H}$  NMR (deuteriochloroform):  $\delta$  2.26-2.43 (1H, m, H2'a), 2.34 (3H, s, Ph-CH<sub>3</sub>), 2.43 (3H, s, Ph-CH<sub>3</sub>), 2.81 (1H, ddd,  $J$  = 1.3, 5.4, 14.2 Hz, H2'b), 4.58 (1H, q,  $J$  = 2.7 Hz, H4'), 4.68 (1H, dd,  $J$  = 3.3, 12.3 Hz, H5'b), 4.77 (1H, dd,  $J$  = 2.8, 12.3 Hz, H5'a), 5.62 (1H, dt,  $J$  = 1.4, 6.5 Hz, H3'), 6.47 (1H, dd,  $J$  = 5.3, 8.8 Hz, H1'), 7.02-7.97 (17H, m, aromatic), 8.75 (1H, s, 3-NH). HRMS: Found: 575.1584 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>28</sub>ClN<sub>2</sub>O<sub>7</sub>: 575.1585. Found, %: C 64.66; H 4.73; N 4.93. C<sub>31</sub>H<sub>28</sub>ClN<sub>2</sub>O<sub>7</sub>. Calculated, %: C 64.76; H 4.73; N 4.8.

**5-(3-Iodophenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (**8a** and **8b**).** Column chromatography using dichloromethane/ethyl acetate 9:1 as eluent gave 0.93 g (12%, with copper(I) iodide) or 2.21 g (27%, without copper(I) iodide) of **8a** and 1.19 g (14%, with copper (I) iodide) or 5.14 g (63%, without copper(I) iodide) of **8b**, both as white solids.

**8a:** mp 65-69°C. HRMS: Found: 667.0925 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>28</sub>IN<sub>2</sub>O<sub>7</sub>: 667.0941.

**8b:** mp 190-193°C. HRMS: Found: 667.0926 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>28</sub>IN<sub>2</sub>O<sub>7</sub>: 667.0941.

Found, %: C 55.66; H 4.09; N 4.24. C<sub>31</sub>H<sub>27</sub>IN<sub>2</sub>O<sub>7</sub>. Calculated, %: C 55.87; H 4.08; N 4.20.

**5-(3-Aminophenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (**9b**).** Column chromatography using dichloromethane/ethyl acetate/triethylamine 23:1:1 as eluent gave 3.75 g (55%) of **9b** as a white solid. Mp 81-86°C. HRMS: Found: 556.2078 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>30</sub>N<sub>3</sub>O<sub>7</sub>: 556.2084.

Found, %: C 65.69; H 6.13; N 7.97. C<sub>31</sub>H<sub>29</sub>N<sub>3</sub>O<sub>7</sub>. Calculated, %: C 67.02; H 5.26; N 7.56 (better EA could not be obtained).

**5-(3-Azidophenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (**10a** and **10b**).** Column chromatography using dichloromethane/ethyl acetate 9:1 as eluent gave 2.14 g (30%) of **10a** as a syrup and 3.56 g (50%) of **10b** as a white solid.

**10a:** HRMS: Found: 582.1987 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>28</sub>N<sub>5</sub>O<sub>7</sub>: 582.1989.

**10b:** mp 162-164°C. HRMS: Found: 582.1987 (MH<sup>+</sup>). Calculated for C<sub>31</sub>H<sub>28</sub>N<sub>5</sub>O<sub>7</sub>: 582.1989.

Found, %: C 63.85; H 4.75; N 12.10. C<sub>31</sub>H<sub>27</sub>N<sub>5</sub>O<sub>7</sub>. Calculated, %: C 64.02; H 4.68; N 12.04.

**5-(3-Methylthiophenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (**11a** and **11b**).** Column chromatography using dichloromethane/ethyl acetate 9:1 as eluent gave 1.15 g (16%) of **11a** as a syrup and 5.47 g (76%) of **11b** as a white solid.

**11a:** HRMS: Found: 587.1851 (MH<sup>+</sup>). Calculated for C<sub>32</sub>H<sub>31</sub>N<sub>5</sub>O<sub>7</sub>S: 587.1852.

**11b**: mp 89-94°C. HRMS: Found: 587.1860 (MH<sup>+</sup>). Calculated for C<sub>32</sub>H<sub>31</sub>N<sub>5</sub>O<sub>7</sub>S: 587.1852.

Found, %: C 65.44; H 5.22; N 4.82. C<sub>32</sub>H<sub>30</sub>N<sub>5</sub>O<sub>7</sub>S. Calculated, %: C 65.52; H 5.15; N 4.78.

**5-(3-Styryl)-2'-deoxy-3',5'-di-O-*p*-toluoyluridine (12a and 12b)**. Column chromatography using dichloromethane/ethyl acetate 9:1 as eluent gave 2.36 g (34%) of **12a** as a syrup and 2.09 g (30%) of **12b** as a white solid.

**12a**: HRMS: Found: 567.2128 (MH<sup>+</sup>). Calculated for C<sub>33</sub>H<sub>31</sub>N<sub>2</sub>O<sub>7</sub>: 567.2131.

**12b**: mp 163-165°C. HRMS: Found: 567.2131 (MH<sup>+</sup>). Calculated for C<sub>33</sub>H<sub>31</sub>N<sub>2</sub>O<sub>7</sub>: 567.2131.

Found, %: C 69.99; H 5.44; N 4.92. C<sub>33</sub>H<sub>30</sub>N<sub>2</sub>O<sub>7</sub>. Calculated, %: C 69.96; H 5.34; N 4.94.

**General Procedure for the Deprotection of 5-(3-Substituted Phenyl)-2'-deoxy-3,5-di-O-*p*-toluoyl Uridines**. A 250 ml flask equipped with a magnetic stirrer and a drying tube was charged with 1.74 mmol of the 5-(3-substituted phenyl)-2'-deoxy-3',5'-di-O-*p*-toluoyl uridines and 165 ml of methanol. Then 26 ml (5.20 mmol) of 0.20 M sodium methoxide in methanol was added dropwise. The reaction mixture was allowed to stand at room temperature for 24 h and was then poured onto a short column of silica gel 60. The silica was washed with methanol and after the addition of 2.5 g of coarse silica gel, the methanol was evaporated *in vacuo*. The coarse silica gel was applied onto a column of silica gel 60 and the product was eluted with dichloromethane/methanol.

**5-(3-Chlorophenyl)-α-2'-deoxyuridine (13a)**.

Yield: 481 mg (63%); mp 203-207°C; <sup>1</sup>H NMR (deuteriomethanol): δ 2.14 (1H, d, J = 14.8 Hz, H2'b), 2.72 (1H, ddd, J = 5.9, 7.7, 14.8 Hz, H2'a), 3.56 (1H, dd, J = 4.6, 12.0 Hz, H5'b), 3.61 (1H, dd, J = 4.5, 12.0 Hz, H5'a), 4.34-4.41 (2H, m, H3' and H4"), 6.31 (1H, dd, J = 2.1, 7.7 Hz, H1"), 7.31 (1H, dt, J = 1.8, 8.2 Hz, H6"), 7.36 (1H, t, J = 7.4 Hz, H5"), 7.47 (1H, dt, J = 1.6, 7.3 Hz, H4"), 7.63 (1H, t, J = 1.4 Hz, H2"), 8.20 (1H, s, H6). HRMS: Found: 339.0747 (MH<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>16</sub>ClN<sub>2</sub>O<sub>5</sub>: 339.0748.

**5-(3-Chlorophenyl)-β-2'-deoxyuridine (13b)**.

Yield: 578 mg (98%); mp 67-72°C; <sup>1</sup>H NMR (deuteriomethanol): δ 2.32-2.36 (2H, m, H2'a and H2'b), 3.75 (1H, dd, J = 3.1, 12.0 Hz, H5'b), 3.83 (1H, dd, J = 2.9, 12.0 Hz, H5'a), 3.96 (1H, q, J = 3.2 Hz, H4'), 4.45 (1H, q, J = 4.6 Hz, H3'), 6.34 (1H, t, J = 6.4 Hz, H1'), 7.30 (1H, dt, J = 1.8, 8.2 Hz, H6"), 7.35 (1H, t, J = 7.9 Hz, H5"), 7.48 (1H, dt, J = 1.6, 7.3 Hz, H4"), 7.66 (1H, t, J = 1.9 Hz, H2"), 8.37 (1H, s, H6). HRMS: Found: 339.0756 (MH<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>16</sub>ClN<sub>2</sub>O<sub>5</sub>: 339.748.

Found, %: C 52.89; H 4.54; N 8.23. C<sub>15</sub>H<sub>15</sub>ClN<sub>2</sub>O<sub>5</sub>. Calculated, %: C 53.19; H 4.46; N 8.27.

**5-(3-Iodophenyl)-α-2'-deoxyuridine (14a)**.

Yield: 576 mg (77%); mp 322-325°C (starts to decompose at ~230°C); <sup>1</sup>H NMR (deuteriomethanol): δ 7.14 (1H, t, J = 7.8 Hz, H5"), 7.54 (1H, dt, J = 1.6, 7.8 Hz, H6"), 7.66 (1H, dt, J = 1.6, 7.9 Hz, H4"), 7.96 (1H, t, J = 1.6 Hz, H2"). HRMS: Found: 431.0107 (MH<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>16</sub>IN<sub>2</sub>O<sub>5</sub>: 431.0104.

**5-(3-Iodophenyl)-β-2'-deoxyuridine (14b)**.

Yield: 524 mg (70%) as a syrup; <sup>1</sup>H NMR (deuteriomethanol): δ 7.14 (1H, t, J = 7.8 Hz, H5"), 7.53 (1H, ddd, J = 1.0, 1.6, 7.9 Hz, H6"), 7.65 (1H, ddd, J = 1.0, 1.7, 7.9 Hz, H4"), 8.00 (1H, t, J = 1.7 Hz, H2"). HRMS: Found: 431.0096 (MH<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>16</sub>IN<sub>2</sub>O<sub>5</sub>: 431.0104.

Found, %: C 41.65; H 3.82; N 6.21. C<sub>15</sub>H<sub>15</sub>IN<sub>2</sub>O<sub>5</sub>. Calculated, %: C 41.88; H 3.51; N 6.51.

**5-(3-Aminophenyl)-β-2'-deoxyuridine (15b)**.

Yield: 539 mg (97%); mp 93-98°C; <sup>1</sup>H NMR (deuteriomethanol): δ 6.69 (1H, ddd, J = 1.0, 2.3, 8.0 Hz, H6"), 6.87 (1H, ddd, J = 1.0, 1.6, 7.7 Hz, H4"), 7.91 (1H, t, J = 1.8 Hz, H2"), 7.10 (1H, t, J = 7.7 Hz, H5"); HRMS: Found: 320.1249 (MH<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>17</sub>N<sub>3</sub>O<sub>5</sub>: 320.1246.

Found, %: C 55.84; H 5.85; N 12.15. C<sub>15</sub>H<sub>16</sub>N<sub>3</sub>O<sub>5</sub>. Calculated, %: C 56.42; H 5.37; N 13.16 (better EA could not be obtained).

**5-(3-Azidophenyl)-α-2'-deoxyuridine (16a)**.

Yield: 547 mg (91%); mp 97-102°C; <sup>1</sup>H NMR (deuteriomethanol): δ 7.01 (1H, ddd, J = 1.4, 2.3, 7.6 Hz, H6"), 7.31 (1H, m, H4"), 7.34 (1H, t, J = 1.5 Hz, H2"), 7.39 (1H, dt, J = 0.8, 7.5 Hz, H5"). HRMS: Found: 346.1157 (MH<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>16</sub>N<sub>5</sub>O<sub>5</sub>: 346.1151.

**5-(3-Azidophenyl)-β-2'-deoxyuridine (16b)**.

Yield: 589 mg (98%); mp 160-163°C; <sup>1</sup>H NMR (deuteriomethanol): δ 7.01 (1H, dt, J = 2.2, 6.9 Hz, H6"), 7.02-7.38 (2H, m, H2" and H4"), 7.39 (1H, t, J = 8.1 Hz, H6"). HRMS: Found: 368.0957 (MNa<sup>+</sup>). Calculated for C<sub>15</sub>H<sub>15</sub>N<sub>5</sub>NaO<sub>5</sub>: 368.0971.

Found, %: C 51.94; H 4.39; N 20.33. C<sub>15</sub>H<sub>15</sub>N<sub>5</sub>O<sub>5</sub>. Calculated: C 52.17%, H 4.38%, N 20.28%.

#### **5-(3-Methylthiophenyl)- $\alpha$ -2'-deoxyuridine (17a).**

Yield: 597 mg (98%); mp 156-160°C;  $^1\text{H}$  NMR (deuteriomethanol):  $\delta$  2.48 (1H, s,  $\text{SCH}_3$ ), 7.18-7.33 (3H, m,  $\text{H4''}$ ,  $\text{H5''}$  and  $\text{H6''}$ ), 7.46 (1H, m,  $\text{H2''}$ ). HRMS: Found: 351.1007 ( $\text{MH}^+$ ). Calculated for  $\text{C}_{16}\text{H}_{19}\text{N}_2\text{O}_5\text{S}$ : 351.1015.

#### **5-(3-Methylthiophenyl)- $\beta$ -2'-deoxyuridine (17b).**

Yield: 597 mg (98%); mp 157-160°C;  $^1\text{H}$  NMR (deuteriomethanol):  $\delta$  2.49 (1H, s,  $\text{SCH}_3$ ), 7.18-7.34 (3H, m,  $\text{H4''}$ ,  $\text{H5''}$  and  $\text{H6''}$ ), 7.48 (1H, m,  $\text{H2''}$ ). HRMS: Found: 351.1025 ( $\text{MH}^+$ ). Calculated for  $\text{C}_{16}\text{H}_{19}\text{N}_2\text{O}_5\text{S}$ : 351.1015.

Found, %: C 54.79; H 5.24; N 8.23.  $\text{C}_{16}\text{H}_{18}\text{N}_2\text{O}_5\text{S}$ . Calculated, %: C 54.85; H 5.18; N 8.00.

#### **5-(3-Styryl)- $\alpha$ -2'-deoxyuridine (18a).**

Yield: 546 mg (95%) as a syrup;  $^1\text{H}$  NMR (deuteriomethanol):  $\delta$  5.25 (1H, dd,  $J = 1.0, 11.0$  Hz,  $\text{CH}=\text{CH}_2$  (*cis*)), 5.81 (1H, dd,  $J = 1.0, 17.6$  Hz,  $\text{CH}=\text{CH}_2$  (*trans*)), 6.75 (1H, dd,  $J = 10.8, 17.7$  Hz,  $\text{CH}=\text{CH}_2$ ), 7.33 (1H, t,  $J = 7.8$  Hz,  $\text{H5''}$ ), 7.39 (1H, dt,  $J = 1.6, 7.8$  Hz,  $\text{H6''}$ ), 7.44 (1H, dt,  $J = 1.6, 7.3$  Hz,  $\text{H4''}$ ), 7.62 (1H, t,  $J = 1.8$  Hz,  $\text{H2''}$ ). HRMS: Found: 331.1287 ( $\text{MH}^+$ ). Calculated for  $\text{C}_{17}\text{H}_{19}\text{N}_2\text{O}_5$ : 331.1294.

#### **5-(3-Styryl)- $\beta$ -2'-deoxyuridine (18b).**

Yield: 552 mg (96%); mp 104-107°C;  $^1\text{H}$  NMR (deuteriomethanol):  $\delta$  5.24 (1H, dd,  $J = 1.0, 10.9$  Hz,  $\text{CH}=\text{CH}_2$  (*cis*)), 5.81 (1H, dd,  $J = 1.0, 17.6$  Hz,  $\text{CH}=\text{CH}_2$  (*trans*)), 6.75 (1H, dd,  $J = 10.9, 17.7$  Hz,  $\text{CH}=\text{CH}_2$ ), 7.33 (1H, t,  $J = 7.6$  Hz,  $\text{H5''}$ ), 7.39 (1H, dt,  $J = 1.5, 7.6$  Hz,  $\text{H6''}$ ), 7.44 (1H, dt,  $J = 1.7, 7.4$  Hz,  $\text{H4''}$ ), 7.63 (1H, t,  $J = 1.7$  Hz,  $\text{H2''}$ ). HRMS: Found: 331.1292 ( $\text{MH}^+$ ). Calculated for  $\text{C}_{17}\text{H}_{19}\text{N}_2\text{O}_5$ : 331.1294.

Found, %: C 61.47; H 5.54; N 8.28.  $\text{C}_{17}\text{H}_{18}\text{N}_2\text{O}_5$ . Calculated, %: C 61.81; H 5.49; N 8.48.

### **Materials and Experimental Procedures: Virology**

Inhibition of HIV-1 and HSV-1 (C42 strain) were performed as XTT assays in MT4 cells (human T cell line) and Vero cells, respectively. Effect on cell growth was determined as an XTT assay on non-confluent HEL cells without presence of any virus. In the CMV assay, reduction in cytopathic effect caused by the virus was determined in MRC-5 cells (human embryonic lung cells). These assays were performed as previously described [31]. The CMV ELISA was an *in situ* determination of viral antigens in HEL cells and performed essentially as described [32].

### **ACKNOWLEDGEMENTS**

The authors wish to thank Mr. Jan Glans for the HPLC separations, Mr. Einar Nilsson for the mass spectrometer analyses, and Dr. Lotta Vrang for the antiviral testing. Grants from Swedish National Board for Industrial and Technical Development and Medivir AB to S. G. and A.-B. H. are gratefully acknowledged.

### **REFERENCES**

1. E. De Clercq, Targets for the Design of Antiviral Agents, Proc. of the NATO Advanced Study Institute, Les Arcs, France, June 19-July 28, E. De Clercq and R. T. Walker (eds.), Plenum Press, New York—London (1984), p. 203.
2. U. Wellmar, A.-B. Hörnfeldt, S. Gronowitz, and N. G. Johansson, Antiviral Chemistry & Chemotherapy (Manuscript).
3. P. Wigerinck, R. Snoeck, P. Claes, E. De Clercq, and P. Herdewijn, J. Med. Chem., **34**, 1767 (1991).
4. P. Wigerinck, C. Pannecouque, R. Snoeck, P. Claes, E. DeClercq, and P. Herdewijn, J. Med. Chem., **34**, 2383 (1991).
5. P. Wigerinck, L. Kerremans, P. Claes, R. Snoeck, P. Maudgal, E. DeClercq, and P. Herdewijn, J. Med. Chem., **36**, 538 (1993).
6. J. Liu, A. Van Aerschot, I. Luyten, P. Wigerinck, C. Pannecouque, J. Balzarini, E. De Clercq, and P. Herdewijn, Nucleosides Nucleotides, **14**, 525 (1995).
7. R. Betts, Prog. Med. Virol., **24**, 44 (1982).

8. A. Macher, C. Reichert, S. Straus, D. Longo, J. Parillo, H. Lane, A. Fauci, A. Rook, J. Manischewitz, and G. Quinnan, *N. Eng. J. Med.*, **309**, 1454 (1983).
9. A. Tyms, D. Taylor, and J. Parkin, *J. Antimicrob. Chemother.*, **23**, Suppl. A, 89 (1989).
10. R. H. Rubin, *Rev. Infect. Dis.*, **12**, Suppl. 7, 754 (1990).
11. G. Schmidt, *Transpl. Proc.*, **23**, Suppl. 3, 126 (1991).
12. Collaborative DHPG Treatment Study Group, *Eng. J. Med.*, **314**, 801 (1986).
13. O. L. Laskin, C. M. Stahl-Bayliss, C. M. Kalman, and L. R. Rosecan, *J. Infect.*, **155**, 323 (1987).
14. J. P. H. Verhyden, *Rev. Infect. Dis.*, **10**, Suppl. 3, 5477 (1988).
15. D. Faulds and R. C. Heel, *Drugs*, **39**, 597 (1990).
16. K. Knox, W. Dobryski, and D. Carrigan, *Lancet*, **337**, 1292 (1991).
17. T. Persson, A.-B. Hörnfeldt, S. Gronowitz, and N. G. Johansson, *Antiviral Chem. Chemotherapy*, **7**, 94 (1996).
18. U. Wellmar, A.-B. Hörnfeldt, S. Gronowitz, and N. G. Johansson, *Antiviral Chem. Chemotherapy* (Manuscript).
19. U. Wellmar, A.-B. Hörnfeldt, and S. Gronowitz, *J. Heterocycl. Chem.*, **33**, 409 (1996).
20. M. Hoffer, *Chem. Ber.*, **93**, 2777 (1960).
21. A. J. Hubbard, A. S. Jones, and R. T. Walker, *Nucleic Acids Res.*, **12**, 6827 (1984).
22. J. N. Frescos, *Nucleosides Nucleotides*, **8**, 549 (1989).
23. E. M. Acton, R. N. Goerner, H. S. Uh, K. J. Ryan, and D. W. Henry, *J. Med. Chem.*, **22**, 518 (1979).
24. S. Ya. Mel'nik, A. A. Bakhmedova, T. D. Miniker, I. V. Yartseva, M. N. Preobrazhenskaya, O. A. Zagulyaeva, V. P. Mamaev, E. V. Chekunova, and S. S. Marennikova, *Bioorg. Khim.*, **10**, 1645 (1981).
25. T. Yamaguchi and M. Saneyoshi, *Chem. Pharm. Bull.*, **32**, 1441 (1984).
26. S. Kit, *Microbiol. Sci.*, **2**, 369 (1985).
27. G. A. Gentry, *Pharmacol. Ther.*, **54**, 319 (1992).
28. E. Littler, A. D. Stuart, and M. S. Chee, *Nature*, **358**, 160 (1992).
29. V. Sullivan, C. L. Talarico, S. C. Stanat, M. Davis, D. M. Coen, and K. K. Biron, *Nature*, **358**, 162 (1992).
30. N. G. Johansson and L. Vrang, Unpublished data.
31. A. Popescu, A.-B. Hörnfeldt, S. Gronowitz, and N. G. Johansson, *Nucleosides Nucleotides*, **14**, 1233 (1995).
32. W. A. Tatarowicz, N. S. Lurain, and K. D. Thompson, *J. Virological Methods*, **35**, 207 (1991).