FULL PAPER

Thieno–Pyrrole-Fused BODIPY Intermediate as a Platform to **Multifunctional NIR Agents**

Samuel G. Awuah,^[a] Sushanta K. Das,^[b] Francis D'Souza,^[b] and Youngjae You^{*[a]}

Abstract: We report the synthesis, photophysical and electrochemical properties, and in vivo fluorescence imaging of a series of new thieno-pyrrole-fused near-infrared (NIR) BODIPY agents by using a versatile intermediate as a building block. The versatile thienopyrrole-fused BODIPY intermediate was rationally designed to bear bromosubstituents and absorb in the mid-red region (635 nm) to act as an organic electrophile for the development of NIR multifunctional agents. The use of subsequent palladium-catalyzed and nucleophilic substitution reactions afforded highly conjugated NIR BODI-PYs. The novel BODIPYs exhibit longwavelength absorptions in the NIR region (650-840 nm). The agents produce sharp fluorescence bands, and most of them display respectable quantum yields of fluorescence (0.05-0.87) useful for biomedical imaging, as demonstrated by in vivo imaging with SBDPiR740. Interestingly, a number of

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agents in the series that are non-halogenated were reactive to O_2 at the triplet photo-excited state coupled with a favorable redox potential and decent fluorescence, and hence could be potential candidates for use as photosensitizers in fluorescence-guided photodynamic therapy. Furthermore, the synthetic approach allows further functionalization of the highly conjugated NIR BODIPYs to tune the excited states (PET, ICT) and to conjugate targeting moieties for enhanced biological applications.

Introduction

Near infrared (NIR) dyes are the subject of intense research as a result of their useful applications, which extend beyond the limited applications of UV and visible light. These dyes have been used in photovoltaic and advanced optoelectronics,^[1] photodynamic therapy (PDT), and non-invasive bioimaging^[2] as well as in laser printers and digital copy machines.^[3] In PDT and bioimaging, the long-wavelength absorbing chromophores may have the valuable advantage of deeper tissue penetration.^[4,5] However, NIR dye development generally requires tedious multi-step synthesis, often resulting in low reaction yields, reactive intermediates, poor solubility, poor photostability, and limited functionalization.[6]

The 4,4-difluoro-4-bora-3a,4a-diaza-s-indacenes (BODI-PYs) provide synthetic advantages in addition to their pho-

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tostability, sharp absorption and emission, high molar absorptivity properties, and facile tuning of excited states.^[7,8] Chemical modifications to the dipyrromethene core as well as functionalization of the boron center and the meso positions make BODIPYs adaptable for numerous applications as protein labels, fluorophores,^[9] photosensitizers for PDT including activatable forms,^[10, 11] energy concentrators,^[12] and chemosensors.^[13] Despite the wide use of BODIPYs and their excellent photophysical properties, the development of NIR BODIPYs with absorption >700 nm is still challenging,^[14] especially within the region of 800–900 nm. Hence, it is critical to develop a versatile but robust synthetic route to attain desirable functionalized NIR BODIPY analogues. Our work reported herein puts BODIPY dyes with unique NIR absorption and emission within reach, thus opening up new opportunities for their applications.

The search for building blocks for NIR BODIPYs has been a center of research activity in recent years. A classical but relevant route is to obtain a halogenated BODIPY framework for further reaction toward extended conjugation (Figure 1). Approaches by Dehaen and Boen et al.^[15-17] who synthesized meso-halogenated BODIPYs, and 3,5-dichloroand 1,7-dibromo-substituted BODIPYs, respectively, and by Hao et al. who developed 3-chloro and 3,5-diiodo-substituted BODIPYs for S_NAr and palladium-catalyzed coupling reactions^[18-24] pushed the new BODIPYs to the visible/far red region of the electromagnetic spectrum. Churchill and coworkers, Hao et al., and Ravikanth et al. addressed the issue of halogen regioselectivity in BODIPYs for use as major building blocks.^[25-27] Ravikanth and co-workers synthesized

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Figure 1. Chemical structures of BODIPY building blocks.

polyarylated BODIPYs using hexabromo-substituted BODIPY as synthons, while Hao et al. used bromo-substituted BODIPYs for regioselective substitution and Suzuki coupling reactions. Work by Arbeloa et al. demonstrated the selective functionalization of polyiodinated BODIPY precursors.^[28] So far, three routes to obtaining halogenated BODIPY precursors for further functionalization have been established: 1) Total synthesis-type reactions via halogenations of the pyrrole ring, 2) halogenation of the dipyrromethane precursor, and 3) electrophilic aromatic substitution of the BODIPY framework. Heterocycle-fused [3,2-b] pyrrole-derived BODIPY features a long excitation wavelength, high extinction coefficient, high photostability as NIR chromophores, insensitivity to different solvent polarities as well as high brightness as a fluorophore.^[29] Our earlier work demonstrated the use of thieno-fused BODIPYs as useful photosensitizers upon halogenation as they possessed a decent fluorescent potential.^[30] Zhao et al. followed up with interesting work to develop non-symmetric thienofused BODIPYs as bright and tunable fluorescent dyes.[31] Recently, Shen et al. synthesized thieno-pyrroles as potential photosensitizers for PDT.^[32] Overall, thieno-fused [3,2-b] pyrrole-derived BODIPYs show promise for functional use; therefore, versatile but robust synthetic approaches are needed to overcome the rigid synthetic protocol currently in practice with less varied synthetic outcomes. In addition, there are only few reports studying this class of BODIPYs as well as its behavior in vivo.

We report the synthesis of multi-functional NIR BODIPY agents based on the thieno-fused [3,2-b] pyrrole framework and their versatility as fluorophores for diagnostics, including small-animal non-invasive imaging and as photosensitizers, generating singlet oxygen via a novel thieno-pyrrolefused BODIPY intermediate. The strategy to access the NIR agents involved 1) building a reactive BODIPY electrophile (**BDP635**) bearing bromo-substituents at the 2,8-positions with absorption in the visible region (635 nm) as an intermediate and 2) functionalizing the reactive BODIPY intermediate through transition-metal-catalyzed coupling and nucleophilic substitution reactions.

Results and Discussion

Synthesis

The essence of the strategy is that the red-emitting intermediate with halogen substitution at the 2,8-positions provides an excellent starting point and offers the use of mild reaction conditions to obtain NIR BODIPY agents, herein referred to as **SBDPiRs**, in moderate to high yield. The use of the thieno-pyrrole fusion creates flexibility in balancing the excited-state properties of these novel NIR BODIPY dyes.

The rational design of the reactive thieno–pyrrole-fused BODIPY intermediate comprises a heteroaryl ring fusion of the traditional BODIPY core with thiophene, trifluoromethyl substitution at the *meso* position (as depicted in our previous work),^[30] and an additional dibromo substitution positioned at the periphery of the core (Scheme 1). Compound



Scheme 1. Rational approach to the synthesis of rare multifunctional NIR BODIPY agents.

4, 2-bromo-4H-thieno[3,2-b]pyrrole-5-carboxylic acid, was obtained following the established Hemetsberger-Knittel synthesis using 5-bromo-2-thiophene carbaldehyde (1) and ethylazidoacetate (2) with subsequent hydrolysis (Scheme 2). The versatile BODIPY building block (BDP635) was then synthesized through a TFA-mediated condensation with TFA anhydride to obtain the CF3 mesosubstituted dipyrromethene unit (a proposed mechanism is presented in Scheme S1 and the ESI mass spectrum of decarboxylated acid 4 is given in Figure S3 in the Supporting Information) followed by boron difluoride chelation. Our next objective was to use this halide-substituted thieno-pyrrole-fused BODIPY (BDP635) as a platform to obtain NIR dyes, the SBDPiRs. The use of the Suzuki, Stille, and Heck coupling reactions with nucleophilic substitution reactions aided aryl functionalization (Scheme 3).

First, **BDP635** was subjected to Suzuki cross-coupling conditions for a relatively short reaction time of 2–3 hours

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Scheme 2. Synthesis of the BODIPY intermediate BDP635.

with various aryl boronic acids (3–4 equiv) to afford diarylated SBDPiRs in 20–54% yield. The relatively short reaction time gives insight into the reactivity of **BDP635** in comparison to the reported hexabromo BODIPY.^[24] The used boronic acids possessed different degrees of donor character from the dimethylphenyl, methoxyphenyl and trimethoxyphenyl to thiophenyl units and the functionalized hydroxyphenyl boronic acids. The Suzuki coupling reaction offered an opportunity to develop highly conjugated systems for the desired applications (Scheme 3). In addition, the functional group tolerance displayed by Suzuki reactions offers an additional advantage.

Second, we provide a useful alternative to the organoborane-based reactions by applying the Stille coupling using the organotin reagent tetraphenyltin. In this approach, the reaction was run for 1 hour to afford the diarylated **SBDPiR690** in 60% yield.

Third, we applied the Heck reaction to generate extended π -conjugated **SBDPiRs** in an attempt to induce a further redshift. The strategy involved connecting the aryl system to the BODIPY chromophore via an extended π -system. To demonstrate the reactivity of **BDP635** towards the Heck re-

action, styrene and the donor-effective 4-methoxystyrene were applied as reagents. Owing to the relatively high reactivity of **BDP635**, the reactions could be performed under less harsh conditions than those used with other BODIPY analogs.^[31] This may be due to the fact that **BDP635** possesses a strong electron-withdrawing CF₃ group in addition to the halide substituents. We also used sodium bicarbonate as a base in this reaction instead of the commonly used triethylamine. The Heck reaction required a relatively longer reaction time (>6 h) among the **SBDPiR** series using about 3–6 equivalents of the vinyl substrates to afford the diarylated **SBDPiR755** and **SBDPiR790** in approximately 25% yield.

Finally, **BDP635** was reactive towards sulfur and oxygen nucleophiles, and the desired di-substituted products were formed within 0.5–1 h in yields of 25–70%. The reaction conditions to produce the ethers (**SBDPiR650** and **SBDPiR650_Ome**) and thioether (**SBPDiR700**) were similar to those reported for the 3,5-dihalogenated BODIPY dyes.^[34]

X-ray Structure Analysis

Since the incorporation of heavy atoms with large atomic radii into chromophores may result in a deformation of the planar structure, we sought to gain insight into this structural effect, if any, on our versatile scaffold, **BDP635**, and indirectly on our new NIR BODIPY dyes, the **SBDPiRs**. To this end, **BDP635** was crystallized by the slow evaporation of its solution in dichloromethane at room temperature (Figure 2). The conjugated nature of the structure was confirmed by the comparable bond lengths for bridging (C6) to both pyrrole rings (C14, C15) of 1.397(3) Å and 1.406(4) Å, respectively, and by the comparable bond lengths of



Scheme 3. Palladium-catalyzed coupling and S_NAr reactions of BDP635.

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Figure 2. ORTEP view of the X-ray crystal structures of **BDP635**. Displacement ellipsoids are drawn at the 50% probability level.

1.545(4) Å and 1.549(4) Å, respectively, between the pyrrole nitrogen (N11, N18) and boron. Furthermore, the sterically crowded boron center maintained a geometry that was almost tetrahedral with an N-B-N angle of $105.1(2)^{\circ}$ and an F-B-F angle of about $108.9(2)^{\circ}$. Despite the incorporation of the bromine atom, the angle of intersection of Br1 and Br2 to the thienopyrrole ring remained similar with C-C-Br angles of $125.7(2)^{\circ}$ and $126.1(2)^{\circ}$ respectively, thus indicating the preserved planarity of the central 18-atom core of **BDP635**.

Theoretical Insights into the Intermediate

To understand the electronic and excited-state behaviors of **BDP635**, density functional theory (DFT) calculations were performed in vacuo. Geometry optimization was carried out using the 6-311G* basis set, and the absorption-related excitation was calculated by using time-dependent DFT (TD-DFT), the CPCM model, and dichloromethane as solvent. The optimized structure also had a preserved planarity, as no significant change was found in the conformation (Figure 3). In addition, the frontier molecular orbital displayed an even electron distribution within the BODIPY chromophore in the HOMO and an electron pull effect in

the LUMO by the trifluoromethyl group. The lack of donor groups influenced this HOMO-LUMO behavior and the energy gap of 2.02 eV, thus inducing a strong S0 \rightarrow S1 transition in the visible region. The deviation between the predicted and the experimental absorption spectra was about 20 nm, thus demonstrating that DFT calculation is a useful tool for predicting UV/Vis spectra of these dyes, taking into account solvent effects.

Optical Properties

The spectroscopic characterization of the NIR BODIPY dyes was performed by studying their absorption and emission behavior in CHCl₃ (Figure 4). The absorption spectra of all SBDPiRs were comparable in shape to those of the previously reported boron dipyrromethene dyes, with the maximal absorption ascribed to the S0-S1 transition and the 'shoulder' peak at a shorter wavelength assigned to the $S0 \rightarrow S2$ transition. The **SBDPiRs** absorbed in a wide range of wavelengths (650-900 nm), presumably due to the combinational effects from the strengths of the π -electron donor and acceptor, extended π -conjugation, and effects of the sulfur atoms. Solutions of SBDPiR650, SBDPiR650_OMe, and SBDPiR700 in chloroform were blue and green, respectively. All three dyes exhibited fluorescence emission in the NIR region (660, 660, and 730 nm, respectively). SBDPiR650 and SBDPiR650_OMe absorbed at the shortest wavelengths in this series owing to the disruption of the π conjugation by the O atom between the core and peripheral phenyl groups. On the other hand, SBDPiR700 was less affected due to the effect imparted by the S atom. The impact of substituents at the phenyl groups on the spectral properties was clearly seen in SBDPiR690, SBDPiR730, SBDPiR731, SBDPiR735, SBDPiR740, and SBDPiR840 (Table 1). These dyes displayed a vivid green to greenish brown color in organic solvents. The absorption shifted towards the red by about 40 nm from SBDPiR690 when methoxyphenyl, trimethoxyphenyl, and hydroxyphenyl donor groups were used. The use of thiophene donor substituents is well known to impart significant red-shifting;^[35] this was further confirmed in SBDPiR735 by its absorption pattern. Interestingly, the use of dimethylaminophenyl moieties as donors induced an absorption >840 nm, a rare absorption observed for the BODIPY chromophore. We attribute the large bathochromic shift (>200 nm) from the starting BDP635 to an internal charge transfer (ICT).^[36] When the donor moieties were conjugated to the BODIPY chromophore via a vinylic bond, dark blue and greenish brown colors were observed for solutions of SBDPiR755 and **SBDPiR790**, respectively. The extended π -conjugation by



Figure 3. Quantum chemical calculations for BDP635.

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Figure 4. Absorption and emission spectra of **BDP635** and **SBDPiRs** in CHCl₃ (**SBDPiR740** in EtOH).

separating the donor moieties from the BODIPY chromophore by a vinylic bond imparted a bathochromic shift of >50 nm (SBDiR690 \rightarrow SBDPiR755; SBDPiR731 \rightarrow SBDPiR790). The emission bands of the two dyes were in the NIR region of >800 nm.

Table 1. Summary of the spectroscopic data.

	$\lambda_{\max(abs)}$ [nm]	$\lambda_{\max(\mathrm{flu})}$ [nm]	$arepsilon^{[a]} [M^{-1} cm^{-1}]$	$arPsi_{ m flu}{}^{[b]}$	τ ^[c] [ns]	$arPsi_{\Delta}^{[\mathrm{d}]}$	$E_{1/2} (\text{Ag/AgCl})^{[e]}$	Oxidation
Agent							Reduction	
BDP635	635	650	150000	0.05	< 0.1	0.52	-0.18, -0.97	1.59
SBDPiR650	649	660	118000	0.86	4.14	0.02	-0.39, -1.07	1.16, 1.43, 1.99
SBDPiR650_OMe	649	660	121000	0.87	4.40	-	-0.56, -1.25	1.01, 1.42
SBDPiR690	688	700	120000	0.22	1.55	0.42	-0.93, -1.65	0.79, 1.04, 1.52
SBDPiR700	694	715	105000	0.28	3.28	0.05	-0.24, -0.86	0.97, 1.18
SBDPiR730	728	761	140 000	0.23	2.60	_	-0.25, -0.92	1.17, 1.65
SBDPiR731	731	755	185000	0.38	3.22	-	-0.45, -1.13	0.98, 1.17, 1.36
SBDPiR735	733	763	125000	0.28	3.09	-	-0.35, -1.03	1.06
SBDPiR740	738	763	160 000	0.10	1.20	-	-0.32	1.02, 1.26
SBDPiR755	753	785	110000	0.07	0.93	_	-0.23, -0.90	1.17, 1.5
SBDPiR790	786	810	85000	0.01	1.30	_	-0.26, -0.91	1.0, 1.16, 1.62
SBDPiR840	841	860	30000	0.002	0.90	_	-0.42	0.50

[a] Molar absorptivity measurements were carried out in CHCl₃ except for **SBDPiR740** for which measurements were performed in EtOH. [b] The fluorescence quantum yield was determined in CHCl₃ (MeOH for **SBDPiR740**) at 298 K with aza-BODIPY (**BDP635–SBDPiR700**) and **KFL4** (**SBDPiR730–SBDPiR840**) as reference ($\Phi_{flu} = 0.34$ and 0.56 in CHCl₃, respectively). [c] Lifetimes are denoted as τ for all agents. [d] Singlet oxygen quantum yields were determined in CHCl₃ with [Ru(bipy)₃]Cl₂ as reference ($\Phi_{\Delta} = 0.77$ in MeOH). [e] All potentials were determined from DPV plots of the agents.

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Singlet Oxygen Generation

We anticipated a degree of intersystem crossing (ISC) efficiency by these SBDPiRs due to the heavy-atom effect that may be potentially induced by the thieno fusion. The ISC efficiency was further supported by monitoring the capability of singlet oxygen (1O2) generation by the photoexcited SBDPiRs, as detected by the phosphorescence spectra of ${}^{1}O_{2}$ at 1270 nm (Figure 5). The highly reactive ${}^{1}O_{2}$ is a cytotoxic species in PDT, which is generated by dyes called photosensitizers upon light illumination. The 1O2 generation efficiency (Φ_{Δ}) decreased in the order of **BDP635**> SBDPiR690 > SBDPiR700 > SBDPiR650. The observation for BDP635 was readily attributed to the additional bromosubstituents at the 2,8-positions. The capabilities of these agents to generate ${}^{1}O_{2}$ coupled with their reasonable fluorescence emission ($\Phi_{\rm F}$ =0.05, 0.22, 0.28, and 0.86, respectively) suggest a possible application as NIR-active theranostic drugs for dual-functioning PDT and imaging.



Figure 5. Phosphorescence spectra of singlet oxygen generated by **BDP635** and **SBDPiRs** in CHCl₃.

The electrochemical behaviors of all agents were also examined in CH_2Cl_2 containing 0.1 M TBAP. Cyclic voltammograms are shown in Figure 6 and the redox potentials for each oxidation and reduction are listed in Table 1. The investigated compounds with the exception of SBDPiR840 underwent two reversible reductions at $E_{1/2} =$ -0.18 to -0.93 V and $E_{1/2} =$ -0.66 to -1.65 V, to form the respective radical anion and dianion. Two-to-three oxidations were detected at $E_{1/2} =$ 0.44 to 1.59 V, $E_{1/2} = 0.95$ to 1.77 V, and $E_{1/2} = 1.36$ to 1.99 V,

Electrochemistry

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Figure 6. Cyclic voltammograms of **BDP635** and **SBDPiRs** in CH_2Cl_2 containing 0.1 M TBAP at the scan rate of 0.1 V s⁻¹.

giving rise to dications and to trications, respectively. In addition, the first oxidation was largely reversible while the second and third oxidations were mostly quasi-reversible or irreversible, possibly because of the instability of the dication and trication generated. The multiple electrochemical waves might correspond to multiple electron transfers in the extensively conjugated BODIPY compounds, thus giving rise to radical anions and dianions on reduction and radical cations and dications on oxidation. The separation between oxidation waves, about 0.2-0.6 V, is within that of polycyclic aromatic compounds.[37] BDP635 showed three reductive waves with a separation of 0.3 V between the second and third waves, possibly due to the instability of the dication generated. The HOMO-LUMO gap estimated from the first oxidation and first reduction potentials varied from 1.77 to 0.90 V, tracking the red-shift of the absorption and emission peak of the derivatives in Table 1.

In Vivo Imaging

To demonstrate the use of these extraordinary NIR BODIPY dyes for in vivo fluorescence imaging, the time-dependent biodistribution of **SBDPiR740** in a living mouse was examined (Figure 7). This study, although not extensive, offers insight of the property of such fluorophores in biological tissue. **SBDPiR740** was selected due to its hydroxy groups that introduce a hydrophilic character to the lipophilic **SBDPiRs**, the availability of the phenolic groups for further modifications, and its compatibility with several imaging instruments with relative ease. A $2.5 \,\mu$ molkg⁻¹ drug dose was injected into the intraperitoneal cavity of a female nude mouse. The mouse was imaged using an IVIS imaging system with excitation and emission filters set at 745 nm and 800 nm, respectively. In the first 10 min, **SBDPiR740**



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Figure 7. Mouse in vivo imaging with **SBDPiR740** (n=3).

mained largely within the peritoneal cavity. At 30 min, it seemed that the dye had absorbed into the blood circulation, as fluorescence signals spread beyond the cavity towards the head region. Fluorescence intensities at the skin reached maximum levels between 3 h and 6 h. Kidneys and liver were observed as intense bright fluorescence spots. After 24 h, **SBDPiR740** was considerably cleared with a degree of fluorescence signal in the kidneys.

Conclusions

In summary, the synthesis of a series of novel thieno-fused [3,2-b] pyrrole-derived NIR BODIPYs was achieved. Through a rational synthetic design of a versatile intermediate that possesses bromo-substituents at the 2,8-positions of the expanded BODIPY chromophore, palladium-catalyzed cross-coupling and nucleophilic substitution was readily accessible.

The spectroscopic data indicated that the incorporation of various donor moieties resulted in a rare bathochromic shift for BODIPYs, with absorbance maxima reaching 840 nm. The use of thiophene with low-lying excited states demonstrated an unprecedented singlet oxygen generation by these non-halogenated NIR BODIPYs. The non-halogenated SBDPiR photosensitizers have an added benefit of fluorescence emission, that is, they hold promise for fluorescenceguided PDT. Our study provides a useful electrochemical insight into these thieno-pyrrole-derived BODIPYs. The versatility of our approach is also demonstrated in the incorporation of functionalities for further conjugation to other materials, particularly, biomolecules for cellular and tissuetargeting. We demonstrated the potential of the SBDPiRs for in vivo imaging using SBDPiR740. The time-dependent biodistribution of this class of BODIPYs at a low dose $(2.5 \,\mu\text{mol}\,\text{kg}^{-1})$ indicate that they may find use as dyes for in vivo imaging. Biological evaluation of these novel BODI-PYs is currently under investigation and will be reported elsewhere. Further functionalization of these NIR dyes for specific biomedical applications in fluorescence optical imaging and PDT is in progress.

Experimental Section

General Methods

Chemical reagents and solvents of analytical grade were purchased from commercial suppliers (Sigma–Aldrich, USA; VWR, USA) and were used without purification. Air-sensitive reactions were performed under an atmosphere of nitrogen. NMR spectra were recorded in $CDCl_3$, CD_2Cl_2 , $[D_6]acetone, or <math>[D_6]DMSO$ by using a Varian 300 MHz spectrometer. Chemical shifts are given in parts per million relative to TMS or 7.23 ppm for residual CHCl₃. Coupling constants were measured in Hz. High-resolution mass spectrometry measurements were performed using electron impact at an ionized voltage of 70 eV and electrospray (ESI) ionization mass spectrometry was done with a QTOF-1 analyzer.

Theoretical Calculations

Geometry optimization was performed with Gaussian 09^[38] using density functional theory (DFT). Electronic excitation was computed using the time-dependent (TD)-DFT calculation at the B3LYP/PBE PBE^[39] level using the CPCM model in dichloromethane as solvent.

2-Bromo-4H-thieno [3, 2-b] pyrrole-5-carboxylic acid (4)

Compound **3** (0.30 g, 1.1 mmol) was dissolved in EtOH (10 mL). Subsequently, NaOH (0.62 g, 15.5 mmol) in water (5 mL) was added, and the mixture was refluxed for 1 h. The reaction was then cooled to room temperature and chilled in an ice bath to acidify the mixture with concentrated HCl. The precipitate was filtered, washed with water, and dried under vacuum to afford **4** as a gray solid (0.22 mg, 70%). ¹H NMR ([D₆]DMSO, 300 MHz): δ =12.76 (s, 1H), 12.15 (s, 1H), 7.21 (s, 1H), 6.99 ppm (s, 1H); HRMS ESI (*m*/*z*): calcd for C₇H₄BrNO₂S: 246.0812; found: 245.9224 [*M*+H]⁺. We should note that **4** has been previously reported and was prepared by the published method with slight modifications.^[28]

2,8-Di(bromo)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5-difluoro-5bora-3a,4a-dithio-s-indacene (**BDP635**)

Compound 4 (1.0 g, 1.8 mmol) was dissolved in TFA (25 mL) and heated to 40 °C for 15 min, upon an intense red color appeared. Subsequently, trifluoroacetic anhydride (9 mL) was added and the temperature was then raised to 80°C with continued stirring for 4 h. The resulting deep blue reaction solution was then allowed to coolto room temperature and poured into an aqueous NaHCO3 solution containing crushed ice. The resultant precipitate was then filtered and dried in vacuo. The dry solid was then dissolved in CH2Cl2 (250 mL) and stirred for 5 min at room temperature under a nitrogen atmosphere. Next, boron trifluoride dietherate (4 mL) and triethylamine (3 mL) were added and the reaction mixture was stirred at room temperature for 1 h. Following evaporation of the solvent, the crude product was purified by flash chromatography on silica gel (CH₂Cl₂) to afford BDP635 as a bluish-green metallic solid(160 mg, 16%). ¹H NMR (CDCl₃, 300 MHz): $\delta = 7.31$ (s, 2H), 7.24 ppm (s, 2H); ¹³C NMR: $\delta = 159.4$, 137.5, 136.0, 131.5, 119.3, 117.2, 114.6, 112.0 ppm; HRMS EI (m/z): calcd for C₁₄H₄BBr₂F₅N₂S₂: 529.8200; found: 529.8170 $[M]^+$.

2,8-Di(4-hydroxyphenyl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2 g]-5,5difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR740**)

To a solution of BDP635 (0.19 g, 0.36 mmol) in toluene/THF/H₂O (30 mL, 1:1:1) was added 4-hydroxyboronic acid (0.20 g, 1.43 mmol) and Na₂CO₃ (0.11 g 1.05 mmol). The reaction solution was purged by bubbling nitrogen gas through for 10 min. A catalytic amount of [Pd(PPh₃)₄] (22 mg, ~5 mol%) was added and the reaction was heated to 80°C for 2 h. After completion of the reaction as judged by TLC, the reaction was diluted with 5 mL water and extracted with diethyl ether. The combined organic layer was washed with water and brine (100 mL each), respectively, and dried over anhydrous Na2SO4. The dried mixture was purified by silica-gel column chromatography (EtOAc/n-hexane 1:1) to afford SBDPiR740 as a dark green solid (102 mg, 54%, purity: 99.3% based on HPLC, see Figure S7 in the Supporting Information). ¹H NMR ([D₆]acetone, 300 MHz): $\delta = 7.84$ (d, J = 9.0 Hz, 4H), 7.52 (s, 2H), 7.37 (s, 2H) 7.03 ppm (d, J=9.0 Hz, 4H); ¹³C NMR: $\delta=150.3$, 134.4, 131.7, 130.2, 129.2, 128.4, 119.3, 115.9, 115.6, 115.1 ppm; HRMS EI (m/z): calcd for C₂₆H₁₄BF₅N₂O₂S₂: 556.0510; found: 556.0520 [M]⁺.

2,8-Di(4-methoxyphenyl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5difluoro-5-bora-3a,4a-diaza-s-indacene (SBDPiR731)

The method described for **SBDPiR740** was followed using **BDP635** (0.10 g, 0.2 mmol), 4-methoxyphenylboronic acid (0.12 g, 0.8 mmol), and Na₂CO₃ (0.06 g 0.6 mmol). The dried mixture was purified by silica-gel column chromatography (EtOAc/toluene 5:95) to afford **SBDPiR731** as a dark green solid (53 mg, 50%). ¹H NMR (CD₂Cl₂, 400 MHz): δ = 7.76 (d, *J*=8.0 Hz, 4H), 7.34 (s, 2H), 7.32 (s, 2H), 7.03 (d, *J*=8.0 Hz, 4H), 3.91 ppm (s, 6H); HRMS EI (*m*/*z*): calcd for C₂₈H₁₈BF₅N₂O₂S₂: 584.0823; found: 584.0825 [*M*]⁺.

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2,8-Di(thiophenyl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5-difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR735**)

The method described for **SBDPiR740** was followed using **BDP635** (0.10 g, 0.2 mmol), 2-thiopheneboronic acid (0.10 g, 0.8 mmol.) and Na₂CO₃ (0.06 g 0.6 mmol). The dried mixture was purified by silica-gel column chromatography (EtOAc/toluene 5:95) to afford **SBDPiR735** as a dark green solid (51 mg, 50%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =7.24 (m, 2H), 7.53 (s, 2H), 7.54 (s, 2H), 7.34 (s, 2H), 7.57 ppm (m, 4H); ¹³C NMR: δ =134.6, 134.3, 130.8, 130.5, 129.6, 128.7, 128.1, 128.0, 127.9, 118.2, 108.6 ppm; HRMS EI (*m*/*z*): calcd for C₂₂H₁₀BF₅N₂S₄: 535.9740; found: 535.9725 [*M*]⁺.

2,8-Di(3,4,5-trimethoxyphenyl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5-difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR730**)

The method described for **SBDPiR740** was followed using **BDP635** (0.12 g, 0.2 mmol), 3,4,5-trimethoxyphenylboronic acid (0.19 g, 0.9 mmol), and Na₂CO₃ (0.07 g 0.7 mmol). The reaction solution was purged by bubbling nitrogen gas through for 10 min. The dried mixture was purified by silica-gel column chromatography (EtOAc/toluene 5:95) to afford **SBDPiR730** as a dark green solid (54 mg, 45 %). ¹H NMR (CD₂Cl₂, 300 MHz): δ = 7.36 (s, 2H), 7.32 (s, 2H), 6.98 (s, 4H), 3.93 (s, 12H), 3.87 ppm (s, 6H); ¹³C NMR: δ = 163.0, 160.9, 153.7, 140.9, 138.4, 134.3, 129.1, 118.4, 108.5, 104.2, 60.7, 56.2 ppm; HRMS EI (*m*/*z*): calcd for C₃₂H₂₆BF₃N₂O₆S₂: 704.1245, found: 704.1226 [*M*]⁺.

2,8-Di(4-N,N,dimethylaminophenyl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5-difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR840**)

The method described for **SBDPiR740** was followed using **BDP635** (0.12 g, 0.2 mmol), 4-*N*,*N*-dimethylaminophenylboronic acid (0.11 g, 0.7 mmol), and Na₂CO₃ (0.07 g 0.7 mmol). The dried mixture was purified by silica-gel column chromatography (toluene) to afford **SBDPiR840** as a dark green solid (24 mg, 20%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =8.15 (d, *J*=6.0 Hz, 4 H), 7.70–7.50 (m, 3 H), 6.70 (s, 1 H), 6.85 (d, *J*=6.0 Hz, 4 H), 2.95 (s, 12 H); ¹³C NMR: δ =153.4, 136.8, 136.3, 136.0, 134.0, 131.9, 130.0, 128.5, 126.9, 116.4, 112.4, 111.0, 40.3, 39.8 ppm; HRMS EI (*m/z*): calcd for C₃₀H₂₄BF₅N₄S₂: 610.1456, found: 610.1470 [*M*]⁺.

2,8-Di(styryl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2 g]-5,5-difluoro-5bora-3 a,4 a-diaza-s-indacene (**SBDPiR755**)

BDP635 (0.08 g, 0.2 mmol), styrene (0.04 g, 0.4 mmol), Pd(OAc)₂ (20 mol%), triphenylphosphine (20 mol%), and Na₂CO₃ (0.06 g 0.6 mmol) were dissolved in anhydrous toluene (3 mL). The reaction mixture was then heated to 80°C with stirring under nitrogen for 6 h. After completion (monitored by TLC), the mixture was diluted with 10 mL toluene, washed with water and brine (50 mL each), and dried over anhydrous Na₂SO₄. The dried mixture was purified by silica-gel column chromatography (toluene) to afford **SBDPiR755** as a black solid (17 mg, 22%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =7.59 (d, *J*=8.0 Hz, 4H), 7.39 (d, *J*=8.0 Hz, 4H), 7.26 (s, 3H), 7.20 (s, 1H), 7.12 ppm (s, 2H); ¹³C NMR: δ =161.3, 158.9, 135.6, 129.6, 128.9, 127.3, 122.9, 111.8 ppm; HRMS EI (*m/z*): calcd for C₃₀H₁₈BF₅N₂S₂: 576.0925; found: 576.0906 [*M*]⁺.

2,8-Di(4-(methoxy-phenyl)vinyl)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5-difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR790**)

The method described for **SBDPiR755** was followed using **BDP635** (0.08 g, 0.2 mmol), 4-methoxystyrene (0.04 g, 0.4 mmol), Pd(OAc)₂ (20 mol%), triphenylphosphine (20 mol%), and Na₂CO₃ (0.06 g 0.6 mmol) dissolved in anhydrous toluene (3 mL). The dried mixture was purified by silica-gel column chromatography (toluene) to afford **SBDPiR790** as a dark brown solid (20 mg, 25%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =7.44 (d, *J*=9.0 Hz, 4H), 7.17 (s, 2H), 7.16 (s, 2H), 7.15 (s, 2H), 6.99 (s, 2H), 6.88 ppm (d, *J*=9.0 Hz, 4H); ¹³C NMR: δ =161.5, 135.2, 134.9, 134.3, 129.0, 128.7, 128.1, 127.3, 120.6, 117.2, 114.6, 114.1, 113.6, 111.3, 55.4 ppm; HRMS EI (*m*/*z*): calcd for C₃₂H₂₂BF₅N₂O₂S₂: 636.1136; found: 636.1120 [*M*]⁺.

BDP635 (0.03 g, 0.1 mmol), tetraphenyltin (0.03 g, 0.1 mmol), $[Pd(PPh_3)_4]$ (20 mol%), triphenylphosphine (20 mol%), and Na₂CO₃ (2 mL, 1 M aq.) were dissolved in toluene (3 mL). The reaction solution was heated to 80 °C with stirring under nitrogen for 1 h. After completion (monitored by TLC), the reaction was diluted with 10 mL toluene, washed with water and brine (50 mL each) and dried over anhydrous Na₂SO₄. The dried mixture was purified by silica-gel column chromatography (toluene) to afford as **SBDPiR690** as a green solid (10 mg, 60%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =7.80 (s, 1H), 7.70 (s, 1H), 7.60 (d, *J*=8.0 Hz, 4H), 7.50 (s, 1H), 7.41 ppm (peaks overlap, 7H); ¹³C NMR: δ =163.3, 161. 3, 137.9, 137.3, 137.1, 136.9, 130.7, 129.8, 129.2, 128.9, 128.6, 128.2, 126.7 ppm; HRMS EI (*m*/*z*): calcd for C₂₆H₁₄BF₅N₂S₂: 524.0612; found: 524.0599 [*M*]⁺.

2,8-Di(thiophenoxy)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5difluoro-5-bora-3 a,4 a-diaza-s-indacene (SBDPiR700)

BDP635 (0.03 g, 0.06 mmol), thiophenol (0.02 g, 0.14 mmol), and K₂CO₃ (0.02 g, 0.14 mmol) were dissolved in anhydrous CH₃CN (3 mL). The reaction solution was heated to 80 °C with stirring under nitrogen for 30 min, upon which the reaction turned red. After completion (monitored by TLC), the mixture was diluted with 10 mL Et₂O, washed with 1 M M₂CO₃ (aq.) (50 mL) and dried over anhydrous Na₂SO₄. The dried mixture was purified by silica-gel column chromatography (toluene) to afford **SBDPiR700** as a green solid (10 mg, 30%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =7.62 (d, *J*=6.0 Hz, 4H), 7.47 (d, *J*=6.0 Hz, 4H), 7.24 (s, 2H), 7.15 (s, 2H), 6.91 ppm (s, 2H); ¹³C NMR: δ =161.9, 159.6, 134.7, 133.5, 133.9, 133.0, 132.9, 131.1, 129.9, 129.8, 129.7, 129.6, 117.6, 111.8, 113.3 ppm; HRMS EI (*m*/z): calcd for C₂₆H₁₄BF₅N₂S₄: 588.0053; found: 588.0037 [*M*]⁺.

2,8-Di(4-methoxyphenoxy)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR650_OMe**)

BDP635 (0.06 g, 0.1 mmol), 4-methoxyphenol (0.06 g, 0.3 mmol), and K_2CO_3 (0.05 g, 0.4 mmol) were dissolved in anhydrous CH₃CN (3 mL). The reaction mixture was then heated to 80 °C with stirring under nitrogen for 30 min. After completion of the reaction (monitored by TLC), the mixture was dried, dissolved in CH₂Cl₂, washed with water and brine (50 mL each) and dried over anhydrous Na₂SO₄. The dried mixture was purified by silica-gel column chromatography (*n*-hexane) to afford **SBDPiR650_OMe** as a blue solid (15 mg, 25%). ¹H NMR (CD₂Cl₂, 300 MHz): δ =7.22 (d, *J*=8.0 Hz, 4H) 7.20 (s, 2H), 6.90 (d, *J*=8.0 Hz, 4H), 6.21 (s, 2H), 3.77 ppm (s, 6H); ¹³C NMR: δ =179.3, 158.1, 157.7, 149.3, 135.0, 125.7, 121.1, 118.6, 114.8, 94.9, 55.6 ppm; HRMS EI (*m*/*z*): calcd for C₂₈H₁₈BF₃N₂O₄S₂: 616.0721; found: 616.0724 [*M*]⁺.

2,8-Di(biphenyl-4-yloxy)-11-trifluoromethyl-dithieno[2,3-b]-[3,2g]-5,5difluoro-5-bora-3a,4a-diaza-s-indacene (**SBDPiR650**)

BDP635 (0.06 g, 0.1 mmol), 4-phenylphenol (0.06 g, 0.3 mmol), and K_2CO_3 (0.05 g, 0.4 mmol) were dissolved in anhydrous CH₃CN (3 mL). The reaction mixture was then heated to 80 °C with stirring under nitrogen for 30 min. After completion of the reaction (monitored by TLC), the mixture was diluted with 10 mL toluene, washed with water and brine (50 mL each) and dried over anhydrous Na₂SO₄. The dried mixture was purified by silica-gel column chromatography (EtOAc/*n*-hexane 1:1) to afford **SBDPiR650** as a reddish-brown solid (42 mg, 70%). ¹H NMR (CDCl₃, 300 MHz): δ =7.52 (m, 3H), 7.49 (m, 5H) 7.43 (m, 5H), 7.30 (d, J=9.0 Hz, 3H), 7.20 (s, 1H), 6.80 (d, J=9.0 Hz, 2H), 6.39 ppm (s, 1H); ¹³C NMR: δ =178.0, 157.7, 155.0, 139.7, 133.7, 128.8, 128.6, 128.2, 127.6, 126.9, 126.6, 126.5, 121.3, 120.1, 118.6, 115.5, 95.9 ppm; HRMS EI (*m*/*z*): calcd for C₃₈H₂₂BF₃N₂O₂S₂: 708.1136; found: 708.1121 [*M*]⁺.

Electrochemical Measurements

Cyclic voltammetry was performed with a homemade three-electrode cell that consisted of a platinum button or glassy carbon working electrode, a platinum wire counter-electrode, and a saturated calomel reference electrode (SCE). The SCE was separated from the bulk of the solution

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by a fritted-glass bridge of low porosity which contained the solvent/supporting electrolyte mixture (tetra-*n*-butylammonium perchlorate, TBAP in *o*-DCB).

Photophysical Measurements

Absorption spectra were recorded on a Perkin Elmer lambda 25 UV/Vis spectrometer. Steady-state and lifetime fluorescence were measured using a Horiba Jobin Yvon UV/Vis/NIR fluorimeter attached with a Data Station v2.4 equipped with a Horiba NanoLED. Relative fluorescence quantum yields ($\Phi_{\rm F}$) were estimated using KFL4 in chloroform $(\Phi_{\rm F}=0.56)$ as a reference. Fluorescence lifetimes were determined by a single exponential curve fit. Singlet oxygen luminescence spectra at 1270 nm were recorded by using Horiba Jobin Yvon UV-Vis/NIR fluorimeter equipped with a FluorEssence v3.5 system. To measure the extinction coefficient, an SBDPiR (ca. 1 mg) was dissolved in chloroform (50 mL), and serially diluted to yield different dye concentrations (10⁻⁷- 10^{-6} M) from the stock solution. A graph of the absorbance (A) versus concentration (C) was plotted and the extinction coefficient was determined by using the Lambert-Beer law. The fluorescence emission spectra of SBDPiRs using KFL4 and aza-BODIPY as reference standards were recorded at 25 °C. Dilute solutions of all dyes were prepared to compare their fluorescence emission spectra after excitation at 710 nm. The quantum yields were calculated following standard protocols.^[9,40] Five serial dilutions were made with chloroform as a solvent, and the absorbance (A) of these solutions was less than 0.1. The integrated fluorescence intensity (F) at each concentration was recorded after excitation at 710 nm. A graph of F versus A was plotted to generate the gradient (G). The quantum yield was calculated using the equation:

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m std}}iggr)iggl(rac{\eta_{
m x}^2}{\eta_{
m std}^2}iggr)$$

Where the subscripts std and x denote the standard and test sample respectively, Φ is the fluorescence quantum yield, G is the gradient from the plot of integrated fluorescence intensity vs. absorbance, and η is the refractive index of the solvent. The quantum yield of **KFL4** was 0.56 in chloroform.

In Vivo Optical Imaging

Optical imaging was performed with an IVIS Spectrum small-animal in vivo imaging system. The Living Image Software v3.0 was used to analyze images and measurements of fluorescent signals. Excitation and emission wavelengths of 745 nm and 800 nm, respectively, were used to acquire in-vivo fluorescent images of **SBDPiR740** (MW = 556.05 gmol⁻¹). All images were attained using a 0.5 s exposure time and an f/stop of 1, with a sampling of multiple angles with the animal under Isoflurane an aesthesia. Animals were injected with 2.5 μ mol kg⁻¹ of **SBDPiR740** via i.p. administration. The injection solution was prepared by solubilizing **SBDPiR740** in ethanol (4 mM) followed by dilution with PBS containing 1% Tween 80 and 5% dextrose. The solution was filtered through a 0.2 μ m sterile syringe filter before i.p. injection.

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FULL PAPER

Something to PET: We have rationally designed a versatile thieno–pyrrole-fused BODIPY intermediate useful for the development of near-infrared (NIR) multifunctional agents. The NIR excitation range is 650–840 nm. The agents produce bright fluores-cence useful for biomedical imaging (see image), generate singlet oxygen (non-halogenated BODIPY) with decent fluorescence for dual therapy and imaging, and were functionalized with donor moieties to induce photoin-duced electron transfer/internal charge transfer (PET/ICT).



Fluorescent Probes

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Thieno–Pyrrole-Fused BODIPY Inter-

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