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Tetrahydrofuran as a Scaffold for Peptidomimetics. Application to the Design and Synthesis of Conformationally Constrained Metalloproteinase Inhibitors

Stephen Hanessian,* Nicolas Moitessier and Serge Wilmouth

Department of Chemistry, Université de Montréal, P.O. Box 6128, Succursale Centre-Ville, Montréal, Quebec, Canada, H3C 3J7

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Abstract—Enantiopure functionalized tetrahydrofurans were synthesized as prototypical conformationally constrained analogs of acyclic succinic acid-based inhibitors of metalloproteinases. While preliminary docking study showed good congruence with a potent inhibitor, the biological results were disappointing. Molecular dynamics simulation revealed the weakness of the H-bond interactions. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

Matrix metalloproteinases (MMPs), such as collagenases, stromelysins and gelatinases, represent a family of zincdependent proteinases that are key enzymes implicated in matrix degradation and reconstruction.¹ While the formation of MMPs in tissues is naturally regulated, certain pathological conditions can lead to their overproduction which results in a number of debilitating physiological effects such as tumor progression,² arthritis³ and sclerosis.⁴ Consequently, the exploration and development of inhibitors of MMPs have become important areas of research during the past decade, and several inhibitors have been reported with promising clinical profiles.⁵ Initially, inhibitors consisted mainly of succinic acid derivatives such as 1 (Batimastat, British Biotech.),⁶ 2 (Marimastat, British Biotech.),⁷ 3 (Ro32-3555, Roche),⁸ 4 (Dupont-Merck),⁹ or 5 (D-2163, Chiroscience)¹⁰ (Fig. 1).

A number of sulfonamides or sulfones were also reported with great potential, such as **6** (CGS 27023A, Novartis),¹¹**7** (AG3340, Agouron)¹² and **8** (RS 130830, Roche Bioscience).¹³ Our own work¹⁴ in this area led to a novel series of sulfonamides **9** and **10** which exhibited low nanomolar inhibitory activity against a number of MMPs (Fig. 2).

The X-ray crystal structures of stromelysin-1 (MMP-3) complexed with a number of natural or synthetic inhibitors^{6c,9,15} at resolutions ranging from 1.7 to 2.5 Å, have revealed unique modes of binding. NMR studies of

^{*} Corresponding author. Tel.: +514-343-6738; fax: +514-343-5728; e-mail: hanessia@ere.umontreal.ca



Figure 1. Selected structures of succinic-type MMP inhibitors.

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Figure 2. Selected arylsulfone or arylsulfonamide containing MMP inhibitors.

enzyme-inhibitor complexes have also been instrumental in the design of new inhibitors.¹⁶ The three-dimensional view of the active site of MMP-3 and complexes with inhibitors has opened opportunities in the design and synthesis of molecules capable of adopting conformations that are conducive to optimal binding. In principle, this should be a relatively easy task, in view of the obviously accessible regions, and predictably advantageous interactions. For example, the active site of MMP-3 offers a zinc binding site as an anchoring point for an inhibitor harboring appropriately deployed functional groups such as P_1' , P_2' and P_1 that can interact with the long narrow S_1' pocket, a shallow hydrophobic S_2' pocket and an exposed promiscious S_1 pocket, respectively (Fig. 3).

Restricting a molecule in its bioactive conformation by imposing rigidity can dramatically enhance its activity.¹⁷ Numerous examples have been reported in which acyclic peptide-like inhibitors of enzymes were modified to nonpeptidic counterparts by utilizing heterocyclic scaffolds such as lactams,¹⁸ some of which include an *N*-sulfonyl moiety.^{15f,19} In contrast, the synthesis of conformationally constrained succinic-like inhibitors of MMPs have been limited, probably due to the greater stereochemical challenge of assembling cyclic scaffolds with vicinal appendages having specific orientations. Cyclopropane^{20a,b} and cyclohexene^{20a} derivatives have been disclosed with little or no activity, suggesting that the enzyme has strict



Figure 3. Bioactive conformation of Batimastat 1/MMP-3 complex, and structures of proposed constrained inhibitors 11a,b and 12a,b showing simulated interactions with pertinent sites.

conformational requirements. In contrast, macrocyclic analogs have been reported with excellent inhibitory properties towards several MMPs.²¹

We report herein, our studies on the computer-aided design and synthesis of enantiopure substituted tetrahydrofuran motifs as representative prototypical inhibitors of stromelysin-1 and related MMPs.

Results and Discussion

Molecular modeling and design of a constrained analog

Modeling of Batimastat **1** within the active site of stromelysin-1 was based on the previously reported crystal structure of the catalytic domain of this enzyme bound to the inhibitor L-764,004.^{15c} Batimastat was manually docked by analogy to its binding mode in MMP-8.^{6c} In order to explore the free energy surface and qualitatively evaluate the stability of the complex, a 100 ps molecular dynamics (MD) trajectory at 300 K was performed in aqueous medium. This resulted in a stable structure along the trajectory.

The results indicated that Batimastat, as with other inhibitors,^{9,15} was bound to the catalytic domain with a strong chelation of the zinc atom by the hydroxamic acid moiety and with an hydrogen bond network between the linear main chain of the inhibitor and the surrounding backbone of the enzyme. Furthermore, the two aromatic moieties of the inhibitor acted as a wall, thus excluding water and stabilizing the complex. Indeed, the phenyl ring wedged in between the two lipophilic Val¹⁶³ and Leu²²² side-chains could be involved in strong hydrophobic interactions.

With this model in hand, we turned our attention to the design of two constrained analogs built around a tetrasubstituted tetrahydrofuran with a substitution pattern that maximizes interactions with the enzyme (Fig. 3).

We considered the 'normal' amide analog **11a**, the so-called 'reverse' amide analog **12a** and their anomeric variants. In the former case, we opted for a D-Phe rather than an L-Phe moiety (as found in Batimastat) with an amide orientation that could interact favorably to form two hydrogen bonds with Tyr^{223} (Fig. 3). The 'reverse' amide analog **12a** incorporated the L-Phe moiety for better H-bond alignment. Thus, the D- and L-Phe and isobutyl moieties in **11a** and **12a** were chosen to represent the P_2' and P_1' sites, and we opted for an appendage containing heteroatoms, such as the methoxyethoxy group to simulate interaction at the P_1 site which is in part solvent exposed.¹⁵

Analogs **11** and **12** were prepared as individual 2,3-*cis* and 2,3-*trans* anomers (tetrahydrofuran numbering), in order to probe the effect of spatial disposition at the P_1 site. The results of a preliminary docking study of **11** and **12** in the active site of stromelysin were promising. Having reached this conclusion, we undertook the synthesis of **11a** and **12a** (and their respective anomers) in order to validate these predictions for potential activity. During the course of our



Figure 4. Stereoselective construction of tetrahydrofuran glyconsides.

studies, an excellent study based on molecular modeling of the S_1 ' subsite of MMP-3 was reported.²²

Synthesis

The synthesis is based on the stereocontrolled functionalization of a suitable chiron **13** readily prepared from D-mannitol²³ or L-glutamic acid²⁴ (Fig. 4). The use of a bulky protecting group would allow the stereoselective functionalization of the positions 2 and 3 in a stepwise fashion to afford the all *trans* isomer as exemplified in Fig. 4.

Conjugate addition of isobutyl magnesium bromide in the presence of cuprous iodide to 13 furnished 14 as a single isomer following well documented precedents.²⁴ The stereocontrolled introduction of a vinyl group as a carboxyl precursor took advantage of the steric bias presented by the isobutyl group. Thus, the lithium enolate prepared from 14 was alkylated with 2-phenylselenoacetaldehyde, using Kowalski's method.²⁵ The resulting aldol product was mesylated, and subjected to elimination to give the alltrans isomer 15, in good overall yield (d.r. 2.5:1). The minor 3-epi-isomer was separated by chromatography. Subjecting the enolate of 14 to trisyl azide²⁶ led to the expected 3-azido lactone 16 accompanied by its 3-epiisomer as a minor product (d.r. 2.7:1). Stereochemical assignments for 15 and 16 were ascertained by detailed NOESY and COSY experiments. A variety of other electrophiles, that could lead to suitable precursors of 11 and 12, were also investigated. Indeed, CNCO₂Me (d.r. 8:1, 67%), benzyloxymethyl chloride (d.r 4:1, 50%), methoxymethyl iodide (d.r. 5.6:1, 84%) and allyl iodide (d.r. 4:1, 96%) afforded the expected *trans* isomers as major products, following the anticipated mode of attack. Although these ratios constituted an improvement compared to that found for 15, conversion to the corresponding acids or alcohols was problematic or low yielding. Similarly, the electrophilic amination of the enolate derived from 14 employing di-tertbutylazodicarboxylate (DBAD) proceeded uneventfully (d.r. 3.6:1, 89%), but the subsequent cleavage of the intermediate hydrazine derivative met with failure. Reduction of the lactones 15 and 16 with Dibal-H under controlled conditions afforded the corresponding lactols 17 and 18 for



Scheme 1. (a) *i*-BuMgBr, CuI, DMS, THF, -78 to -40° C, then 13, -78 to 0° C, 98%, d.r.>50:1. (b) i: LiHMDS, PhSeCH₂CHO, THF, -78° C, ii: MsCl, Et₃N, CH₂Cl₂, 67% (2 steps), d.r.: 2.5:1. (c) i: LiHMDS, trisyl azide, THF, -78° C, ii: AcOH, 30° C, 95% (2 steps), d.r.: 2.7:1. (d) Dibal-H, CH₂Cl₂, -60° C, 92% (for 17); 73% (for 18, along with 20% of recovered s.m.).

further elaboration into the intended targets **11** and **12** (Scheme 1).

The introduction of the methoxyethoxy appendage was accomplished by conversion of the lactol **17** into the corresponding 2-thiopyridylcarbonate ester followed by addition of 2-methoxyethanol, according to the protocol previously reported from this laboratory.²⁷ A direct Fischer²⁸ glycosylation was low yielding, while application of Schmidt's²⁹ trichloroacetimidate method led to a mixture of products, possibly due to the greater reactivity of the furanosyl donor.

There remained to introduce the peptidic side chain and the hydroxamic acid unit at C-3 and C-5 respectively. In view of the tendency for intramolecular cyclizations resulting from the attack of the hydroxamate unit onto the appendage at C-3, particularly as the aldehyde or ester group, we opted to proceed with the *N*-Boc-*O*-trityl hydroxamate derivative. The inseparable anomeric mixture of products **19**

(2,3-trans/2,3-cis, 1.8:1), was subjected to desilylation to afford the separable alcohols 20a and 20b which were oxidized first with tetrapropylammonium perruthenate³⁰ to the aldehydes 21a and 21b, and finally to the acids 22a and **22b** using the Pinnick protocol.³¹ Conversion of each acid to the *O*-trityl hydroxamate³² afforded **23a** and **23b** which were protected as the N-Boc derivatives 24a and 24b respectively. Oxidative cleavage of the vinyl group led to the aldehydes 25a and 25b then to the acids 26a and 26b. Condensation with D-phenylalanine N-methyl amide in the presence of EDC, HOBT and N-methyl morpholine led to the expected amides 27a and 27b. The corresponding N-diphenylmethyl amide 28 was similarly prepared from the 2,3-trans-isomer 26a. Finally, treatment with TFA in dichloromethane gave the intended prototypes 11a and 11b and the analog 29 from their immediate precursors (Scheme 2). 33

Scheme 3 shows the conversion of the lactol **18** into the 'reverse' amide analogs **12a** and **12b**, essentially according to the protocol used for **11a** and **11b**. The anomeric 2-methoxyethoxy glycosides **30a** and **30b** (2.1:1 ratio) were separated by column chromatography and their stereochemical assignments were ascertained by detailed NOESY experiments.

To avoid lactam formation between the free amine on C-3 and the hydroxamic acid at position C-5, we introduced the peptidic unit first. Each anomer was then individually reduced to the corresponding amines **31a** and **31b** and acetylated with *N*-acetyl-D-phenylalanine using EDC and HOBT to afford **32a** and **32b** respectively. The *N*-*p*methoxyphenylsulfonyl derivative **33** was prepared from the 2,3-*trans* glycoside **31a** in the usual manner. Subsequent transformations followed the protocol used for the synthesis of **11a** and **11b**. Thus, after TBAF mediated deprotection, compounds **34a**, **34b** and **35** were converted to the carboxylic acids **36a**, **36b** and **37** respectively via the corresponding aldehydes. Introduction of the *O*-trityl



Scheme 2. (a) i: Di-(*S*-2-pyridyl) thiocarbonate, Et₃N, 4 Å MS, CH₂Cl₂, ii: MeOCH₂CH₂OH, AgOTf, 4 Å MS, 0°C, 81%, d.r. (*trans/cis*): 1.8:1. (b) TBAF, AcOH, THF, 96%. (c) TPAP, NMO, 4 Å MS, CH₂Cl₂, 61–72%. (d) NaClO₂, NaH₂PO₄, 2-methyl-2-butene, *t*-BuOH/H₂O, 86–98% (for **22a/b**), 94–94% (for **26a/b**). (e) TrONH₂, EDC, HOBT, NMM, THF, 70–66%. (f) Boc₂O, Et₃N, DMAP, CH₂Cl₂, 82–98%. (g) O₃, CH₂Cl₂, -78°C then DMS, 68–70%. (h) NH₂-D-NHMe, EDC, HOBT, NMM, THF, 68–93%. (i) Ph₂CHNH₂, EDC, HOBT, NMM, THF, 78%. (j) TFA, CH₂Cl₂, 83–57–83%.



Scheme 3. (a) i: Di-(S-2-pyridyl) thiocarbonate, Et₃N, 4 Å MS, CH₂Cl₂, ii: MeOCH₂CH₂OH, AgOTf, 4 Å MS, 0°C, 78%, d.r.(*trans/cis*): 2.1:1. (b) H₂, Pd/C, EtOH, 96–82%. (c) AcNH-L-Phe-OH, EDC, HOBT, NMM, THF, 66–70%. (d) *p*-methoxyphenyl sulfonyl chloride (PMPSO₂Cl), Et₃N, CH₂Cl₂, 88%. (e) TBAF, AcOH, THF, 87–81–91%. (f) i: TPAP, NMO, 4 Å MS, CH₂Cl₂, ii: NaClO₂, NaH₂PO₄, 2-methyl-2-butene, *t*-BuOH/H₂O, 56–69–74% (2 steps). (g) TrONH₂, EDC, HOBT, NMM, THF, 47–57–54%. (h) TFA, CH₂Cl₂, 78–90–90%.

hydroxamate moiety afforded the reverse amide analogs **38a** and **38b** as well as the sulfonamide **39** from the 2,3-*trans* isomer **37**. Detritylation then led to the desired final products **12a**, **12b** and **40**.

Biological evaluation and modeling

Compounds **11a**, **11b**, **29**, **12a**, **12b** and **40** were tested as inhibitors of MMP-2, MMP-3 and MMP-9 but were found to be inactive ($IC_{50} > 100 \mu M$).³⁴ In spite of our optimistic outlook based on the preliminary docking experiments discussed above, it was clear that one or more critical parameters were not satisfied. We then performed in depth molecular modeling studies with **11a** and **12a** in the binding site of MMP-3 which led to two possible conformations of each compound, differing in the orientation of the phenyl ring in relation to the peptidic chain (Fig. 5).

The results from the molecular dynamics study in aqueous medium of the 'reverse' amide analog **12a** clearly showed



12a, Conformation 2 (stable)

Figure 5. Refined conformations in vacuo of the analog 12a in the catalytic site of MMP-3.

that conformation 1 (Fig. 5) was not stable in presence of water. On the other hand, although exclusion of water and the attainment of a stable docked inhibitor/enzyme complex was indicated in the case of conformation 2, the H-bond network was weakened all along the dynamics simulation. Fig. 6 shows the proposed structures of the Batimastat/MMP-3 and **12a** (conformation 2) modeled in the active



Figure 6. Connolly map surface of: (Top) bioactive conformation of Batimastat in MMP-3; (Bottom) modeled conformation of **12a** in MMP-3. Color code for enzyme: red (carbonyl); white (amide NH); grey (Zn^{2+}) .

site of MMP-3. Although the interactions at the hydrophobic P_1' and P_2' sites appear to be favorable, there was considerable deviation of the amide backbone.

In the case of the normal amide analog **11a**, both conformers were unstable as the enzyme complex since water was only partly excluded from the complex.

The preliminary docking study performed at the outset of this project did not take into account the dynamic behavior of the inhibitor/enzyme complexes. On the other hand the molecular dynamics simulation was more rigorous, and revealed the weak H-bonded interactions of **12a**.

Conclusion

The synthesis of tetrahydrofuran derivatives as peptidomimetics was achieved using the stereocontrolled functionalization of a suitable chiron. Their elaboration into constrained hydroxamic acid derivatives as MMP inhibitors led to inactive compounds. Excluding undesired steric factors, it is clear that constraining part of the backbone of 1 into a tetrahydrofuran scaffold as in 11 and 12 perturbs the requisite alignment of crucial H-bonds as well as the hydrophobic interactions in the respective P and S subsites of 1 and MMP-3. Other studies have shown that minor deviations from the ideal bioactive conformation adopted by succinate-based inhibitors can result in substantial loss of activity.³⁵ Further studies are in progress to better understand the intricacies of scaffold design in our quest for effective metalloproteinase inhibitors.

Experimental

Molecular modeling

General. The molecular modeling was performed on Silicon Graphics Indigo2 workstation running IRIX (version 5.4) using AMBER³⁶ force field implemented in InsightII[®] version 95.0 program and Discover[®] package.³⁷ The force field was parameterized according to Guida³⁸ and Merz.³⁹ The atomic partial charges of the inhibitors were calculated using the semiempirical MNDO method implemented in the MOPAC program. The atomic partial charges of the enzyme and water molecules were calculated using the AMBER force field. A cutoff of 10 Å for the non-bonded interactions was used.

Complex building and refinement. The X-ray structure of L-764,004/stromelysin-1 complex was retrieved from the Brookhaven Protein Data Bank and used as a starting point^{15c} (code 1HFS). The hydrogens were added, visually inspected, then the model was refined by a relaxation process. Two water solvent layers of 8 and 15 Å around the inhibitor, and 12 Å around the enzyme were added and the system was allowed to relax following the procedure described below. The protein and the crystallographic water oxygen atoms were held fixed, the solvent water molecules being free to move. A preliminary minimization was performed to remove close atom contacts, then the system was more completely relaxed by 2000 cycles of minimiza-

tion using steepest descent followed by 5000 cycles using conjugate gradients. 6000 Steps of 1 fs of dynamics at 300 K were performed to allow the solvent molecules to orient themselves. At this point, the main chain of the protein, the heavy atoms of the inhibitor and crystallographic water oxygen atoms were tethered by a force constant. The relaxation started with a strong force constant to relax the side chains (1000 kcal/Å²) by 1000 cycles of minimization using steepest descent followed by 5000 cycles using conjugate gradients. The last 5000 steps were reiterated with the template force stepwise decreased (100, 50, 15 and finally 0 kcal/Å²). The inhibitor was removed and the solvated structure of the enzyme used in the following studies.

The X-ray structure of Batimastat co-crystallized in MMP-8^{6c} (code 1MMB from the Protein Data Bank) was used for modeling the Batimastat/MMP-3 complex. Batimastat was manually docked in the binding site of MMP-3 according to its conformation in MMP-8. The water was temporarily removed and the molecule was positioned inside the binding site by 300 steps of minimization using steepest descent followed by 3000 cycles using conjugate gradients while keeping the enzyme fixed. Water molecules were added and the system relaxed as described above. 100 000 Steps of 1 fs of MD simulation were performed (0.1 ns). For that purpose, a core of residues was defined (10 Å from any atoms of the inhibitor) and a shell (a layer of 5 Å) on which the calculations were performed. All the atoms within the core were unconstrained while the atoms within the shell and the second layer water molecule oxygens were restrained by an energy penalty of 50 kcal/Å². The rest of the enzyme and the water molecules around the enzyme were frozen and a structure was archived every 100 steps. The final structure was refined by energy minimization with the same constraints by 2000 cycles of minimization using steepest descent followed by 5000 cycles using conjugate gradients and finally unconstrained by 5000 cycles using conjugate gradients.

Inhibitors 11a and 12a/MMP-3 complexes: building and refinement. The starting structure of the inhibitors/MMP-3 complexes were manually built and refined by energy minimization then using a cooling procedure. To explore the conformational space, 5000 steps of MD at 900 K were performed followed by 5000 steps at 300 K and finally 200 cycles of minimization using steepest descent followed by 1000 cycles using conjugate gradients. For these calculations, the enzyme was held fixed and a distance-dependent dielectric constant (4 r) was used. The resulting model was saved and used as a starting point for the next loop. Fifty structures were obtained and compared. For the two modeled complexes, two main structures were extracted and studied. Water molecules were added and the system studied using a similar procedure as for the Batimastat/ MMP-3 complex.

Chemistry

General procedures. Solvents were distilled under positive pressure of dry nitrogen before use and dried by standard methods; THF and ether, from K/benzophenone; and CH_2Cl_2 and toluene, from $CaCl_2$. All commercially

available reagents were used without further purification. 4 Å molecular sieves were dried at 140°C prior to use. All reactions were performed under nitrogen atmosphere. NMR (¹H, ¹³C) spectra were recorded on AMX-300 and ARX-400 spectrometers in CDCl₃ or CD₃OD with tetramethylsilane as the internal standard. Low- and high-resolution mass spectra were recorded on VG Micromass, AEI-MS 902 or Kratos MS-50 spectrometers using fast atom bombardement (FAB) or electrospray techniques. Optical rotations were recorded on a Perkin-Elmer 241 polarimeter in a 1 dm cell at ambient temperature. Analytical thin-layer chromatography was performed on Merck 60F254 pre-coated silica gel plates. Visualization was performed by UV or by development using KMnO₄ or FeCl₃ solutions. Flash column chromatography were performed using (40-60 µm) silica gel at increased pressure. Melting points recorded were uncorrected.

5(*S*)-(*tert*-Butyl-diphenyl-silanyloxymethyl)-5*H*-furan-2one (13). This compound was prepared according to the reported procedure.^{23,24}

5(S)-(tert-Butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyldihydro-furan-2-one (14). Magnesium turnings (5.0 g, 206 mmol) were placed in an N₂-filled flask and covered with dry THF (10 mL). 1,2-Dibromoethane (0.15 mL) in dry THF (5 mL) was added dropwise until a vigourous reaction started and the solvent refluxed. iso-Butylbromide (10.6 mL, 100 mmol) in THF (100 mL) was added with a dropping funnel as to maintain a gentle reflux. The solution was stirred at 80°C for 2 h, cooled to room temperature, and immediately canulated in a N₂-filled bottle in which the solution started to crystallize (101 mL, titrated:⁴⁰ 0.74 M, 74%). To this solution (21.8 mL, 16.2 mmol) in dry THF (20 mL) was added a suspension of cuprous iodide (1.54 g, 8.09 mmol) in THF (20 mL) at -78° C. After stirring for 10 min, (0.3 mL) dimethylsulfide was added, and the greyish solution was stirred for 2 h, then allowed to warm to -40° C. After being re-cooled to -78° C, lactone 13 (949 mg, 2.70 mmol) was added in THF (20 mL), and the green solution was stirred at 0°C until it turned to a dark purple color. After stirring for a further 15 min, the mixture was quenched with saturated NH₄Cl, extracted with ether, washed with brine, dried over MgSO₄ and concentrated in vacuo. The resulting oil was triturated with CH₂Cl₂ and the insoluble salts were filtered off. The filtrate was dried over MgSO₄ and concentrated and the residue was chromatographed (hexanes/EtOAc, 9:1) to provide 14 (1.08 g, 2.63 mmol, 98%, d.r.>50:1) as a pale yellow oil; $R_{\rm f}$ =0.42 (hexanes/EtOAc, 4:1); $[\alpha]_D = +21.7$ (c 1.0, CHCl₃); IR (neat/NaCl) 1782.7 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.70-7.65 (m, 4H), 7.50-7.37 (m, 6H), 4.17 (m, 1H), 3.87 (dd, 1H, J=3.2, 11.5 Hz), 3.69 (dd, 1H, J=3.6, 11.5 Hz), 2.79 (dd, 1H, J=8.9, 17.4 Hz), 2.58 (m, 1H), 2.20 (dd, 1H, J=6.8, 17.4 Hz), 1.56 (m, 1H), 1.40-1.26 (m, 2H), 1.06 (s, 9H), 0.91 (d, 3H, J=5.8 Hz), 0.90 (d, 3H, J=5.8 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 176.7, 135.6, 135.5, 132.8, 132.5, 129.8, 127.7, 85.6, 64.3, 43.0, 35.3, 34.4, 26.7, 26.0, 22.9, 21.9, 19.1; LRMS: (FAB, NBA, m/z, %): 433 (1.5) (M+Na⁺), 411 (1.6) (M+H⁺), 409 (1.1), 353 (40), 333 (43), 199 (50), 135 (95); HRMS calcd for $C_{25}H_{34}O_3SiNa$ (M+Na⁺) 433.21750, found 433.21840.

5(S)-(tert-Butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-**3(S)-vinyl-dihydro-furan-2-one** (15). To a stirred solution of lactone 14 (2.00 g, 4.9 mmol) in dry THF (50 mL) was added at -78° C a solution of LiHMDS (5.84 mL, 5.84 mmol, 1 M solution in THF). After stirring for 1 h at -78°C, 2-phenylselenoacetaldehyde⁴¹ (1.26 g, 6.34 mmol) was added. After 30 min, saturated NH₄Cl was added and the mixture was extracted with ether, the organic phase was dried over Na₂SO₄ and concentrated in vacuo. The crude product was dissolved in CH₂Cl₂, triethylamine (3.4 mL, 23.8 mmol) was added at 0°C followed by mesyl chloride (1.5 mL, 19.5 mmol). After 30 min, cold water was added to the cloudy mixture which was diluted with CH₂Cl₂, washed with brine and concentrated in vacuo. Purification by flash chromatography (hexanes/EtOAc, 5:1) afforded the product 15 (1.03 g, 2.36 mmol, 48%) as a pale yellow oil, and the cis-epimer (0.41 g, 0.94 mmol, 19%) as pale yellow oil. The relative configuration of 15 was ascertained by a NOESY experiment; $R_{\rm f}$ =0.67 (hexanes/EtOAc, 4:1); $[\alpha]_{\rm D}$ =+10.3 (c 0.7, CHCl₃); IR (neat/NaCl) 1774.2, 1654.2 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.73-7.65 (m, 4H), 7.49-7.36 (m, 6H), 5.80 (ddd, 1H, J=7.8, 9.7, 17.7 Hz), 5.33 (m, 1H), 5.29 (m, 1H), 4.09 (m, 1H), 3.92 (dd, 1H, J=2.7, 11.8 Hz), 3.71 (dd, 1H, J=3.7, 11.8 Hz), 2.96 (dd, 1H, J=8.0, 9.0 Hz), 2.59 (m, 1H), 1.57 (m, 1H), 1.48–1.29 (m, 2H), 1.06 (s, 9H), 0.89 (2d, 6H, J=5.4 Hz); ¹³C NMR (300 MHz, CDCl₃) δ 176.1, 135.6, 135.4, 133.2, 132.9, 132.5, 129.8, 129.2, 127.7, 119.6, 84.5, 63.4, 52.1, 42.6, 39.1, 26.6, 25.3, 22.6, 22.4, 19.1; LRMS: (FAB, NBA, m/z, %): 873 (9) $(2 \times M + H^{+})$, 803 (75), 745 (85), 675 (9), 437 (12) (M+H⁺), 359 (22), 241 (26), 199 (60), 197 (68), 135 (100), 75 (41), 55 (25); HRMS calcd for $C_{27}H_{35}O_3Si$ (M-H⁺) 435.23553, found 435.23690.

3(S)-Azido-5(S)-(*tert*-butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-dihydro-furan-2-one (16). To a solution of LiHMDS (2.66 mL, 2.66 mmol, 1 M solution in THF) cooled to -78°C and diluted with dry THF (20 mL) was added dropwise the lactone 14 (840 mg, 2.05 mmol) in dry THF (30 mL). After stirring for 1 h, a solution of triisopropyl benzenesulfonyl azide (900 mg, 3.07 mmol) in dry THF (10 mL) was added at -78° C. The resulting mixture was stirred for a further 30 min. The solution was quenched with AcOH (0.47 mL, 8.19 mmol), warmed to 30°C and stirred for 30 min. The solution was diluted with ether and the organic phase was washed with saturated NaHCO₃ and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was flash chromatographed (hexanes/EtOAc, 9:1) to provide the expected 3(S)-trans isomer 16 (635 mg, 1.41 mmol, 69%, yellowish oil) and the corresponding 3(*R*)-*cis* isomer (240 mg, 0.53 mmol, 26%, yellowish oil); for 16: $R_f = 0.52$ (hexanes/EtOAc, 4:1); $[\alpha]_D = -44.2$ (c 1.2, CHCl₃); IR (neat/NaCl) 2109.2, 1784.2 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.70-7.65 (m, 4H), 7.50-7.38 (m, 6H), 4.08 (ddd, 1H, J=2.7, 3.5, 8.8 Hz), 3.94 (d, 1H, J=10.1 Hz), 3.92 (dd, 1H, J=2.7, 11.9 Hz), 3.71 (dd, 1H, J=3.5, 11.9 Hz), 2.62 (m, 1H), 1.72 (m, 1H), 1.46 (ddd, 1H, J=5.6, 8.8, 14.5 Hz), 1.33 (ddd, 1H, J=6.5, 8.8, 14.5 Hz), 1.07 (s, 9H), 0.95 (2d, 6H, J=6.8 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 172.6, 135.6, 135.5, 132.7, 132.4, 129.9, 127.8, 83.2, 63.8, 62.5, 41.0, 39.4, 26.6, 25.8, 23.2, 21.9, 19.2; LRMS: (FAB, NBA, m/z, %): 426 (23), 333 (8), 221 (27), 199 (47), 197 (40), 135 (100); HRMS

calcd for $C_{25}H_{33}O_3SiN_3K$ (M+K⁺) 490.19284, found 490.19420.

5(S)-(tert-Butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-3(S)-vinyl-tetrahydro-furan-2-ol (17). To a solution of lactone 15 (1.69 g, 3.88 mmol) in dry THF (40 mL) was added dropwise a solution of Dibal-H (4.65 mL, 4.65 mmol, 1 M solution in toluene) at -78° C. After stirring for 11 h at -60° C, the solution was quenched by adding EtOAc (5 mL) and allowing to warm to room temperature. A solution of NaHCO₃ was added and the resulting biphasic solution stirred for 1 h. The organic layer was then separated, extracted with ether, washed with saturated NaHCO₃, water and brine, dried over Na2SO4 and concentrated in vacuo. The residue was subjected to flash chromatography (hexanes/EtOAc, 9:1) to provide the lactol 17 (1.57 g, 3.58 mmol, 92%, colorless oil); $R_{\rm f}=0.46-0.67$ (hexanes/ EtOAc, 4:1); IR (neat/NaCl) 3417.7, 1640.7 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 7.74–7.70 (m, 4H), 7.51–7.37 (m, 6H), 5.97-5.71 (m, 1H), 5.27 (d, 0.3H, J=3.5 Hz), 5.22-5.18 (m, 2.4H), 5.07 (ddd, 0.3H, J=0.7, 1.9, 10.1 Hz), 3.96 (ddd, 0.3H, J=0.1, 3.1, 4.5 Hz), 3.90-3.84 (m, 1.7H), 3.81 (dd, 0.3H, J=3.0, 11.0 Hz), 3.68 (dd, 0.3H, J=4.5, 11.0 Hz), 3.62 (d, 0.7H, J=8.1 Hz), 3.50 (dd, 0.7H, J=3.4, 11.8 Hz), 2.40 (m, 1.7H), 2.22 (m, 0.3H), 1.60–1.17 (m, 3H), 1.12 (s, 6.3H), 1.08 (s, 2.7H), 0.84 (d, 3H, J=6.2 Hz), 0.78 (d, 3H, J=6.2 Hz); ¹³C NMR (75 MHz, CDCl₃) & 138.3, 136.1, 135.79, 135.66, 135.60, 132.39, 132.35, 129.98, 129.85, 129.59, 127.80, 127.77, 127.61, 117.4 (0.7C), 116.1 (0.3C), 102.8 (0.3C), 99.7 (0.7C), 86.6 (0.7C), 84.8 (0.3C), 65.6 (0.7C), 65.1 (0.3C), 58.5 (0.3C), 58.0 (0.7C), 42.74 (0.3C), 42.64 (0.7C), 42.57 (0.3C), 37.7 (0.7C), 26.84 (2.1C), 25.84 (0.9C), 25.73 (0.7C), 23.3 (0.7C), 22.9 (0.3C), 22.5 (0.3C), 22.0 (0.7C), 19.2 (0.3C), 19.1 (0.7C); LRMS: (FAB, NBA, m/z, %): 421 (38) (M-OH⁺), 351 (17), 199 (84), 197 (75), 135 (100); HRMS calcd for $C_{27}H_{37}O_2Si$ (M–OH⁺) 421.25629, found 421.25770.

3(S)-Azido-5(S)-(*tert*-butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-tetrahydro-furan-2-ol (18). Following the same procedure as for 17, lactone 16 (748 mg, 1.66 mmol) in dry THF (20 mL), Dibal-H (6.63 mL, 6.63 mmol, 1 M solution in toluene) afforded after flash chromatography (hexanes/EtOAc, 9:1) the lactol 18 (551 mg, 1.22 mmol, 73%) as a colorless oil together with unreacted starting material (149 mg, 0.27 mmol, 20%); R_f=0.50 (hexanes/ EtOAc, 4:1); IR (neat/NaCl) 3411.3, 2104.9 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 7.73-7.68 (m, 4H), 7.50-7.46 (m, 6H), 5.43 (d, 0.4H, J=1.8 Hz), 5.28 (m, 0.6H), 3.98 (ddd, 0.6H, J=2.7, 3.5, 8.8 Hz), 3.9 (m, 0.4H), 3.75-3.90 (m, 2H), 3.60 (dd, 0.4H, J=1.9, 4.8 Hz), 3.49 (dd, 0.6H, J=2.5, 11.0 Hz), 3.29 (dd, 0.6H, J=3.9, 10.2 Hz), 2.60 (m, 0.6H), 2.23 (m, 0.4H), 1.72 (m, 1H), 1.50 (m, 1H), 1.33 (m, 1H), 1.11 (s, 5.4H), 1.08 (s, 3.6H), 0.98-0.88 (m, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 135.7, 135.6, 135.5, 133.1, 133.0, 132.1, 130.0, 129.9, 129.7, 127.8, 127.6, 101.6 (0.4C), 97.1 (0.6C), 84.7 (0.6C), 84.5 (0.4C), 72.2 (0.4C), 68.2 (0.6C), 65.0 (0.6C), 64.8 (0.4C), 43.3 (0.4C), 42.2 (0.6C), 41.7 (0.4C), 36.0 (0.6C), 26.74 (1.8C), 26.69 (1.2C), 26.2 (0.4C), 26.0 (0.6C), 23.0 (0.4C), 22.8 (0.6C), 22.6 (0.6C), 22.1 (0.4C), 19.1 (0.4C), 19.0 (0.6C); LRMS: (FAB, NBA, *m/z*, %): 408 (2), 223 (6), 199 (67), 135 (100), 91 (30); HRMS calcd for $C_{27}H_{35}O_3SiN_3K$ (M+K⁺) 492.20847, found 492.21090.

tert-Butyl-[3(S)-isobutyl-5(S)-(2-methoxy-ethoxy)-4(S)vinyl-tetrahydro-furan-2(S)-ylmethoxy]-diphenyl-silane (19a) and tert-butyl-[3(S)-isobutyl-5(R)-(2-methoxyethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-ylmethoxy]diphenyl-silane (19b). To a mixture of lactol 17, (1.54 g, 3.51 mmol), were added di(S-2-pyridyl) thiocarbonate (3.9 g, 17.9 mmol) and 4 Å molecular sieves (200 mg) in CH₂Cl₂ (60 mL) followed by triethylamine (2.8 mL, 20.4 mmol). The resulting solution was stirred for 36 h at room temperature. The mixture of isomers obtained was quickly purified on silica gel and the crude was immediatly dissolved in CH_2Cl_2 (150 mL) in presence of powdered 4 Å molecular sieves. Methoxyethanol (0.80 mL, 10.5 mmol) was added and the mixture stirred 5 h at room temperature then cooled to 0° C. Freshly dried silver triflate (2.6 g, 10.5 mmol) was added and the stirring continued for 1 h at 0°C then 1 h at room temperature. The suspension was filtered, extracted with CH₂Cl₂ and the organic phase was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. Purification by flash chromatography (hexanes/EtOAc, 19:1) gave 19a and 19b as a colorless oil $(1.41 \text{ g}, 2.84 \text{ mmol}, 81\%, 1.8:1 \text{ ratio}): R_f=0.62$ (hexanes/ EtOAc, 9:1); IR (neat/NaCl) 1638.1 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 7.78-7.70 (m, 4H), 7.49-7.35 (m, 6H), 5.83 (ddd, 0.35H, J=9.0, 10.1, 19.5 Hz), 5.78 (ddd, 0.65H, J=9.1, 10.3, 19.5 Hz), 5.11 (ddd, 0.65H, J=0.9, 1.7, 19.5 Hz), 5.08 (ddd, 0.35H, J=0.8, 2.0, 19.5 Hz), 5.07 (d, 0.35H, J=10.5 Hz), 5.04 (ddd, 0.65H, J=0.7, 2.0, 10.5 Hz), 4.92 (d, 1H, J=4.8 Hz), 4.90 (d, 1H, J=2.5 Hz), 3.90-3.70 (m, 4H), 3.66-3.50 (m, 2H), 3.48 (m, 1H), 3.40 (s, 1.95H), 3.32 (s, 1.05H), 2.56 (ddd, 0.65H, J=2.3, 6.0, 8.7 Hz), 2.37 (ddd, 0.35H, J=4.5, 9.2, 10.7 Hz), 2.08–1.93 (m, 1H), 1.62–1.32 (m, 2H), 1.31–1.18 (m, 1H), 1.10 (s, 3.15H), 1.09 (s, 5.85H), 0.85 (d, 3.9H, J=5.8 Hz), 0.83 (d, 3.9H, J=5.8 Hz), 0.81 (d, 2.1H, J=6.5 Hz), 0.72 (d, 2.1H, J=6.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 138.9 (0.65C), 136.1 (0.35C), 135.68, 135.62, 133.6 (0.35C), 133.5 (0.65C), 129.6, 127.6, 117.0 (0.35C), 115.5 (0.65C), 108.6 (0.65C), 105.5 (0.35C), 87.2 (0.35C), 84.7 (0.65C), 71.9 (0.65C), 71.7 (0.35C), 68.3 (0.35C), 66.7 (0.65C), 66.4 (0.35C), 65.1 (0.65C), 59.0 (0.65C), 58.9 (0.35C), 57.0 (0.65C), 56.9 (0.35C), 43.1 (0.65C), 42.7 (0.35C), 42.4 (0.65C), 41.1 (0.35C), 26.81 (3.15C), 26.77 (5.85C), 25.9 (0.65C), 25.6 (0.35C), 23.2 (0.35C), 23.1 (0.65C), 22.3 (0.65C), 19.26 (0.35C); LRMS: (FAB, NBA, m/z, %): 495 (10), 421 (32) (M-OCH₂CH₂OMe⁺), 351 (20), 257 (49), 199 (59), 197 (60), 135 (88), 59 (100); HRMS calcd for $C_{30}H_{44}O_4SiNa (M+Na^+) 519.29065$, found 519.29190.

[3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-yl]-methanol (20a) and [3(S)-isobutyl-5(R)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-yl]-methanol (20b). To the mixture of anomers 19a and 19b (1.41 g, 2.84 mmol) in THF (100 mL) and AcOH (1 mL) was added portionwise TBAF (1.85 g, 7.11 mmol) at 0°C. The resulting mixture was stirred overnight, diluted with ether, and the organic phase was washed with water, brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash chromatography (hexanes/ EtOAc, 9:1 to 4:1) to afford 20a ((S): 451 mg, 1.75 mmol, 62%, colorless oil), and **20b** ((*R*): 251 mg, 0.97 mmol, 34%, colorless oil). The relative configuration of both 20a and 20b was determined by NOE spectroscopy; for 20a: $R_{\rm f}=0.54$ (hexanes/EtOAc, 1:1); $[\alpha]_{\rm D}=+76.0$ (c 0.8, CHCl₃); IR (neat/NaCl) 3467.8, 1641.7 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3) \delta 5.83 \text{ (ddd, 1H, } J=9.1, 10.1,$ 17.1 Hz), 5.09 (ddd, 1H, J=1.0, 1.7, 17.1 Hz), 5.02 (ddd, 1H, J=0.9, 1.7, 10.1 Hz), 4.85 (d, 1H, J=2.6 Hz), 3.85-3.73 (m, 3H), 3.60–3.51 (m, 4H), 3.36 (s, 3H), 2.83 (m, 1H), 1.88 (ddd, 1H, J=6.3, 8.0, 16.2 Hz), 1.61 (m, 1H), 1.40 (ddd, 1H, J=6.4, 8.0, 14.5 Hz), 1.24 (ddd, 1H, J=6.3, 7.8, 14.1 Hz), 0.85 (d, 3H, J=5.9 Hz), 0.82 (d, 3H, J=5.9 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 138.3, 115.7, 108.4, 84.5, 71.7, 66.7, 63.1, 58.8, 57.2, 42.5, 41.9, 25.8, 22.8, 22.3; LRMS: (FAB, NBA, m/z, %): 259 (7) (M+H⁺), 257 (7), 227 (7), 183 (100); HRMS calcd for $C_{14}H_{25}O_4$ [M-H⁺] 257.17529, found 257.17490; for **20b**: R_f =0.57 (hexanes/EtOAc, 1:1); $[\alpha]_{D} = -97.0$ (c 1.3, CHCl₃); IR (neat/NaCl) 3465.8, 1638.5 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 5.83 (ddd, 1H, J=9.1, 9.1, 18.5 Hz), 5.12 (m, 1H), 5.08 (m, 1H), 4.83 (d, 1H, J=4.4 Hz), 3.88-3.78 (m, 3H), 3.65 (ddd, 1H, J=3.4, 4.7, 11.3 Hz), 3.58-3.53 (m, 2H), 3.48 (ddd, 1H, J=0.5, 3.4, 13.0 Hz), 3.39 (s, 3H), 3.15 (m, 1H), 2.54–2.33 (m, 2H), 1.62 (m, 1H), 1.32 (ddd, 1H, J=4.2, 9.5, 13.5 Hz), 1.19 (ddd, 1H, J=5.6, 8.8, 13.5 Hz), 0.88 (d, 3H, J=6.4 Hz), 0.86 (d, 3H, J=6.4 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 135.6, 117.2, 104.9, 87.4, 71.6, 67.1, 63.7, 58.6, 57.1, 42.1, 37.3, 25.9, 23.5, 21.7; LRMS: (FAB, NBA, *m*/*z*, %): 289 (6) (M+Na⁺), 257 (20), 227 (6), 183 (100); HRMS calcd for $C_{14}H_{25}O_4$ (M-H⁺) 257.17529, found 257.17460.

3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-carbaldehyde (21a). To a solution of **20a** (414 mg, 1.60 mmol) in dry CH_2Cl_2 (50 mL) were added N-methyl morpholine (553 mg, 4.81 mmol) and 4 Å molecular sieves (300 mg). The suspension was stirred for 20 min. Then a catalytic amount of tetrapropylammonium perruthenate was added. The resulting mixture was stirred at room temperature for 90 min then filtered on a pad of silica gel. The residue was purified by flash chromatography (hexanes/EtOAc, 1:1 to 2:3) to provide 21a (251 mg, 0.98 mmol, 61%, brownish oil); $R_f=0.45-0.85$ (hexanes/ EtOAc, 2:3); $[\alpha]_{D} = +24.7$ (c 1.0, CHCl₃); IR (neat/NaCl) 1735.9, 1642.9 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.63 (d, 1H, J=2.6 Hz), 5.71 (ddd, 1H, J=8.9, 10.2, 17.1 Hz), 5.13 (ddd, 1H, J=1.0, 1.7, 17.1 Hz), 5.08 (ddd, 1H, J=0.9, 1.8, 10.1 Hz), 5.02 (d, 1H, J=2.5 Hz), 4.08 (dd, 1H, J=2.5, 8.2 Hz), 3.84 (m, 1H), 3.65-3.51 (m, 3H), 3.37 (s, 3H), 2.63 (ddd, 1H, J=2.1, 6.0, 8.4 Hz), 2.07 (m, 1H), 1.64 (m, 1H), 1.48 (m, 2H), 0.85 (d, 3H, J=6.5 Hz), 0.84 (d, 3H, J=6.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 200.5, 137.0, 116.6, 109.3, 87.1, 71.6, 67.1, 58.9, 56.1, 44.5, 41.9, 25.7, 22.5, 22.3; LRMS: (FAB, NBA, m/z, %): 257 (12) (M+H⁺), 227 (100), 197 (39), 181 (75); HRMS calcd for C₁₄H₂₅O₄ (M+H⁺) 257.17529, found 257.17630.

3(S)-Isobutyl-5(*R***)-(2-methoxy-ethoxy)-4(***S***)-vinyl-tetrahydro-furan-2(***S***)-carbaldehyde (21b).** Using the same procedure as for **21a**, compound **20b** (247 mg, 0.96 mmol) in dry CH₂Cl₂ (50 mL), NMO (330 mg 2.87 mmol), 4 Å molecular sieves (300 mg) and a catalytic amount of TPAP, led after flash chromatography (hexanes/EtOAc, 1:1 to 2:3) to **21b** (177 mg, 0.69 mmol, 72%, yellowish oil); $R_{\rm f}$ =0.24 (hexanes/EtOAc, 9:1); $[\alpha]_{\rm D}$ =-141.6 (*c* 0.7, CHCl₃); IR (neat/NaCl) 1733.6 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.61 (d, 1H, *J*=3.0 Hz), 5.80 (ddd, 1H, *J*=8.5, 9.1, 18.0 Hz), 5.16 (m, 1H), 5.12 (m, 1H), 5.05 (d, 1H, *J*=4.0 Hz), 4.01–3.89 (m, 2H), 3.67–3.52 (m, 3H), 3.37 (s, 3H), 2.45–2.28 (m, 2H), 1.68 (m, 1H), 1.43–1.27 (m, 2H), 0.89 (d, 3H, *J*=6.5 Hz), 0.82 (d, 3H, *J*=6.5 Hz); ¹³C NMR (300 MHz, CDCl₃) δ 202.3, 134.5, 118.2, 106.3, 88.7, 71.6, 67.1, 59.0, 56.3, 42.5, 41.0, 25.7, 23.5, 21.7; LRMS: (FAB, NBA, *m/z*, %): 257 (16) (M+H⁺), 227 (23), 197 (11), 181 (19), 151 (23), 59 (100); HRMS calcd for C₁₄H₂₅O₄ (M+H⁺) 257.17529, found 257.17460.

3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-carboxylic acid (22a). To 21a (201 mg, 0.78 mmol) in t-BuOH (5 mL) were successively added a solution of NaH₂PO₄ (184 mg, 1.18 mmol) in water (1 mL), 2-methyl-2-butene (0.4 mL) and sodium chlorite (213 mg, 2.36 mmol). The mixture was stirred for 3 h and the solvents were removed. The residue was taken up in CH₂Cl₂/CH₃OH and the salts were filtered off. The residue was purified by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 19:1) to provide the acid **22a** (183 mg, 0.67 mmol, 86%, colorless oil); $R_{\rm f}=0.50$ (CH₂Cl₂/CH₃OH, 9:1); $[\alpha]_{\rm D}=+74.0$ (c 0.9, CH₃OH); IR (neat/NaCl) 3462.5, 3192.7, 1732.2, 1643.7 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 5.79 (ddd, 1H, J=8.9, 10.2, 17.2 Hz), 5.13 (ddd, 1H, J=1.0, 1.7, 17.2 Hz), 5.08 (ddd, 1H, J=0.9, 1.7, 10.2 Hz), 4.98 (d, 1H, J=2.7 Hz), 4.91 (m, 1H), 4.16 (d, 1H, J=7.9 Hz), 3.79 (m, 1H), 3.62-3.53 (m, 3H), 3.35 (s, 3H), 2.51 (ddd, 1H, J=2.8, 6.0, 8.1 Hz), 2.13 (ddd, 1H, J=6.1, 8.0, 14.1 Hz), 1.73 (m, 1H), 1.70 (m, 1H), 0.85 (d, 3H, J=6.5 Hz), 0.84 (d, 3H, J=6.5 Hz); ¹³C NMR (75 MHz, CD₃OD) & 175.4, 139.1, 116.9, 110.6, 82.6, 72.8, 68.2, 59.1, 58.0, 43.9, 26.9, 23.3, 22.7; LRMS: (FAB, NBA, m/ z, %): 317 (8) (M-H+2Na⁺), 295 (40) (M+Na⁺), 273 (12) $(M+H^+)$, 227 (7), 197 (48), 183 (17), 151 (61), 59 (100); HRMS calcd for $C_{14}H_{25}O_5$ (M+H⁺) 273.17020, found 273.17110.

3(S)-Isobutyl-5(R)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-carboxylic acid (22b). To a solution of 21b (171 mg, 0.67 mmol) in *t*-BuOH (5 mL), were added NaH₂PO₄ (156 mg, 1.00 mmol) in water (0.9 mL), 2-methyl-2-butene (0.35 mL) and sodium chlorite (181 mg, Purification by flash chromatography 2.00 mmol). (CH₂Cl₂/CH₃OH, 1:0 to 19:1) afforded **22b** (178 mg, 0.67 mmol, 98%, colorless oil); $R_{\rm f}$ =0.61 (EtOAc); $[\alpha]_D = -121.1$ (c 0.8, CHCl₃); IR (neat/NaCl) 3166.8, 1754.3, 1642.5 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.1-8.5 (m, 1H), 5.73 (ddd, 1H, J=8.9, 9.9, 17.5 Hz), 5.15 (m, 1H), 5.10 (m, 1H), 4.98 (d, 1H, J=4.2 Hz), 4.14 (d, 1H, J=8.8 Hz), 3.88 (ddd, 1H, J=3.0, 5.8, 12.0 Hz), 3.75 (ddd, 1H, J=3.0, 6.4, 12.0 Hz), 3.62–3.50 (m, 2H), 3.38 (s, 3H), 2.48–2.31 (m, 2H), 1.88 (m, 1H), 1.47 (ddd, J=6.6, 6.6, 13.6 Hz), 1.36 (ddd, 1H, J=5.8, 8.0, 13.6 Hz), 0.89 (d, 3H, J=6.7 Hz), 0.82 (d, 3H, J=6.7 Hz); ¹³C NMR (75 MHz, $CDCl_3$) δ 173.8, 134.3, 118.4, 107.2, 83.4, 71.1, 68.7, 60.2, 58.5, 56.2, 43.42, 42.3, 25.0, 22.7, 22.3; LRMS: (FAB, NBA, m/z, %): 295 (6) (M+Na⁺), 273 (20) (M+H⁺), 227 (13), 197 (54), 151 (65), 137 (28), 59 (100); HRMS calcd for $C_{14}H_{25}O_5 (M+H^+)$ 273.17020, found 273.17000.

3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-carboxylic acid trityloxy-amide (23a). To 22a (172 mg, 0.63 mmol) in THF (3 mL) were successively added EDC (186 mg, 0.95 mmol), HOBT (128 mg, *N*-methyl morpholine 0.95 mmol) and (104 μL, 0.95 mmol). After stirring for 20 min, O-tritylhydroxylamine (260 mg, 0.95 mmol) was added, the resulting mixture was stirred overnight then diluted with ether, washed with 0.1N HCl, water, 0.5N NaHCO₃, brine, dried over Na₂SO₄ and concentrated in vacuo. Purification by flash chromatography (hexanes/EtOAc, 4:1) provided the title compound 23a (235 mg, 0.44 mmol, 70%, colorless oil); $R_{\rm f}=0.26$ (hexanes/EtOAc, 4:1); $[\alpha]_{\rm D}=+34.0$ (c 0.7, CHCl₃); IR (neat/NaCl) 3368.0, 1710.7 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 8.35 (s, 1H), 7.51-7.47 (m, 6H), 7.38-7.30 (m, 9H), 5.46 (ddd, 1H, J=8.9, 10.4, 17.0 Hz), 4.98 (d, 1H, J=17.0 Hz), 4.93 (d, 1H, J=7.2 Hz), 4.70 (d, 1H, J=2.2 Hz), 3.97 (d, 1H, J=7.8 Hz), 3.63 (m, 1H), 3.50-3.42 (m, 3H), 3.35 (s, 3H), 2.46 (ddd, 1H, J=2.3, 5.6, 8.3 Hz), 1.85 (m, 1H), 1.65 (m, 1H), 1.51 (ddd, 1H, J=6.1, 8.4, 13.5 Hz), 1.37 (ddd, 1H, J=5.8, 8.6, 13.5 Hz), 0.82 (d, 3H, J=6.5 Hz), 0.78 (d, 3H, J=6.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.6, 141.5, 137.4, 129.0, 128.8, 127.8, 127.7, 116.1, 108.8, 92.1, 82.2, 71.5, 66.9, 58.9, 55.6, 46.6, 43.2, 25.5, 23.0, 21.9; LRMS: (FAB, NBA, m/z, %): 528 (10), 484 (10), 454 (22), 437 (40), 243 (100); HRMS calcd for $C_{33}H_{39}O_5NNa$ (M+Na⁺) 552.27258, found 552.27500.

3(S)-Isobutyl-5(R)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-carboxylic acid trityloxy-amide (23b). This compound was prepared following the same procedure as for 23a. From 22b (172 mg, 0.63 mmol) in THF (5 mL), EDC (136 mg, 0.70 mmol), HOBt (94 mg, 0.70 mmol) N-methyl morpholine (76 µL, 0.70 mmol) and O-tritylhydroxylamine (191 mg, 0.70 mmol), was obtained after flash chromatography (hexanes/EtOAc, 4:1), 23b (221 mg, 0.42 mmol, 66%, colorless oil); $R_{\rm f}$ =0.36 (hexanes/EtOAc, 4:1); $[\alpha]_{D} = -36.4$ (c 0.7, CHCl₃); IR (neat/NaCl) 3384.2, 3275.5, 1708.7, 1643.9, 1596.2 cm⁻¹; ¹H NMR (75 MHz, CDCl₃) & 8.85 (s, 1H), 7.52–7.48 (m, 6H), 7.35–7.25 (m, 9H), 5.72 (ddd, 1H, J=9.1, 11.1, 15.5 Hz), 5.11 (m, 1H), 5.07 (m, 1H), 4.84 (d, 1H, J=4.8 Hz), 3.92 (d, 1H, J=8.8 Hz), 3.27 (s, 3H), 3.25 (m, 1H), 3.18 (m, 2H), 3.03 (ddd, 1H, J=3.0, 4.9, 10.8 Hz), 2.29 (m, 1H), 2.16 (m, 1H), 1.81 (m, 1H), 1.39 (ddd, J=6.8, 6.8, 13.5 Hz), 1.24 (ddd, 1H, J=6.4, 8.0, 13.5 Hz), 0.78 (d, 3H, J=6.5 Hz), 0.69 (d, 3H, J=6.5 Hz); ¹³C NMR (300 MHz, CDCl₃) δ 168.9, 141.7, 134.8, 129.1, 127.8, 127.6, 117.9, 106.7, 92.8, 84.4, 71.5, 67.6, 58.9, 56.3, 43.3, 42.9, 25.0, 22.8, 22.7; LRMS: (FAB, NBA, *m*/*z*, %): 552 (11) (M+Na⁺), 528 (11), 484 (8), 454 (9), 401 (10), 345 (18), 289 (14), 259 (76), 243 (100); HRMS calcd for $C_{33}H_{39}O_5NNa (M+Na^+) 552.27258$, found 552.27020.

3(*S*)-Isobutyl-5(*S*)-(2-methoxy-ethoxy)-4(*S*)-vinyl-tetrahydro-furan-2(*S*)-carboxylic acid, *tert*-butoxycarbonyltrityloxy-amide (24a). To 23a (228 mg, 0.43 mmol) in CH₂Cl₂ (10 mL) were added Boc₂O (187 μ L, 0.86 mmol), triethylamine (60 μ L, 0.43 mmol) and DMAP (53 mg, 0.43 mmol) at 0°C. After stirring for 16 h at room temperature, the solution was diluted with CH₂Cl₂, washed with 0.1N HCl, water, 0.5N NaHCO₃, water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash chromatography (hexanes/EtOAc, 9:1) to provide 24a (221 mg, 0.35 mmol, 82%, colorless oil); $R_{\rm f}=0.28$ (hexanes/EtOAc, 4:1); $[\alpha]_{\rm D}=-5.9$ (c 1.1, CHCl₃); IR (neat/NaCl) 1773.1, 1644.0 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) (2 rotamers 4:1) δ 7.41-7.36 (m, 6H), 7.32-7.27 (m, 9H), 5.76 (ddd, 0.2H, J=8.8, 10.0, 19.1 Hz), 5.72 (ddd, 0.8H, J=9.1, 10.1, 19.1 Hz), 5.26 (d, 0.2H, J=8.0 Hz), 5.13 (d, 0.2H, J=17.0 Hz), 5.17-5.00 (m, 2H), 4.95 (d, 0.2H, J=3.1 Hz), 4.88 (d, 0.8H, J=3.0 Hz), 4.22 (d, 0.8H, J=8.3 Hz), 3.90-3.75 (m, 1H), 3.63-3.48 (m, 3H), 3.79 (s, 0.6H), 3.36 (s, 2.4H), 2.61 (ddd, 0.2H, J=3.0, 6.6, 9.3 Hz), 2.48 (ddd, 0.8H, J=3.1, 6.6, 9.4 Hz), 2.21 (ddd, 0.2H, J=7.5, 7.5, 7.5 Hz), 2.15 (ddd, 0.8H, J=7.5, 7.5, 7.5 Hz), 1.53 (s, 7.2H), 1.47 (s, 1.8H), 1.50-1.32 (m, 1H), 1.30-1.11 (m, 2H), 0.82 (d, 3H, J=6.5 Hz), 0.69 (d, 0.6H, J=6.5 Hz), 0.68 (2d, 4.8H, J=6.5 Hz), 0.64 (d, 0.6H, J=6.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 156.1 (0.2C), 149.3 (0.2C), 147.9 (0.8C), 146.9 (0.8C), 143.65 (0.8C), 143.56 (0.2C), 137.7 (0.8C), 137.4 (0.2C), 129.1 (1.2C), 129.0 (4.8C), 127.5 (4.8C), 127.4 (1.2C), 127.2 (0.6C), 127.1 (2.4C), 116.6 (0.2C), 116.1 (0.8C), 109.5 (0.2C), 108.9 (0.8C), 92.3 (0.2C), 91.8 (0.8C), 84.1 (0.8C), 83.8 (0.2C), 80.9 (1C), 71.7 (1C), 67.4 (0.2C), 67.2 (0.8C), 58.91 (0.2C), 58.87 (0.8C), 56.5 (0.8C), 56.4 (0.2C), 46.1 (0.2C), 44.0 (0.8C), 41.9 (0.2C), 41.8 (0.8C), 27.44 (2.4C), 27.36 (0.6C), 25.5 (0.2C), 25.3 (0.8C), 22.8 (0.2C), 22.6 (0.8C), 22.3 (0.8C), 21.9 (0.2C); LRMS: (FAB, NBA, m/z, %): 532 (4), 307 (11), 289 (10), 259 (23), 243 (100), 165 (42).

3(S)-Isobutyl-5(R)-(2-methoxy-ethoxy)-4(S)-vinyl-tetrahydro-furan-2(S)-carboxylic acid, tert-butoxycarbonyltrityloxy-amide (24b). This compound was prepared following the same procedure as for 24a. From 23b (211 mg, 0.40 mmol) in CH₂Cl₂ (10 mL), Boc₂O (184 µL, 0.80 mmol), triethylamine (55 µL, 0.40 mmol), DMAP (49 mg, 0.40 mmol), was obtained, after flash chromatography (hexanes/EtOAc, 19:1), 24b (247 mg, 0.39 mmol, 98%, colorless oil); $R_{\rm f}$ =0.52 (hexanes/EtOAc, 4:1); $[\alpha]_{\rm D} = -75.2$ (c 0.8, CHCl₃); IR (neat/NaCl) 1770.7, 1641.8, 1600.8 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.40-7.32 (m, 6H), 7.31-7.27 (m, 9H), 5.72 (ddd, 1H, J=9.2, 11.2, 16.8 Hz), 5.10 (m, 1H,), 5.05 (dd, 1H, J=2.0, 8.0 Hz), 4.92 (d, 1H, J=4.5 Hz), 3.92 (d, 1H, J=9.2 Hz), 3.80 (m, 1H), 3.56–3.48 (m, 3H), 3.37 (s, 3H), 2.49 (m, 1H), 2.30 (m, 1H), 1.52 (s, 9H), 1.31 (m, 1H), 1.14 (ddd, J=5.0, 7.9, 13.7 Hz) 1.00 (ddd, 1H, J=6.0, 8.0, 13.7 Hz), 0.63 (d, 3H, J=6.3 Hz), 0.57 (d, 3H, J=6.3 Hz); ¹³C NMR (75 MHz, CDCl₃) & 148.3, 147.8, 143.7, 135.0, 129.2, 129.1, 127.4, 127.1, 117.8, 105.1, 91.8, 84.0, 82.8, 71.5, 66.5, 59.0, 56.4, 41.1, 40.6, 27.5, 24.8, 23.3, 22.0; LRMS: (FAB, NBA, m/z, %): 652 (1) ($M + Na^+$), 243 (100).

4(*R*)-Formyl-3(*S*)-isobutyl-5(*S*)-(2-methoxy-ethoxy)-tetrahydro-furan-2(*S*)-carboxylic acid, *tert*-butoxycarbonyltrityloxy-amide (25a). O₃ was bubbled through a vigorously stirred solution of **24a** (217 mg, 0.35 mmol) in CH_2Cl_2 (10 mL) at $-78^{\circ}C$ for 50 min. An excess of dimethylsulfide (0.4 mL) was added, the solution allowed to warm up to 0°C then stirred for a further 15 min. The solution was concentrated in vacuo, and the residue was purified by flash chromatography (hexanes/EtOAc, 4:1) to give **25a** (148 mg, 0.23 mmol, 68%, colorless oil); $R_{\rm f}=0.29-0.39$ (hexanes/EtOAc, 4:1); $[\alpha]_{\rm D}=-9.0$ (c 0.7, CHCl₃); IR (neat/NaCl) 1770.8, 1729.1, 1651.5, 1596.2 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) (2 rotamers 4:1) δ 9.63 (d, 0.2H, J=1.7 Hz), 9.58 (d, 0.8H, J=1.7 Hz) 7.39-7.32 (m, 6H), 7.32-7.24 (m, 9H), 5.47 (d, 0.2H, J=1.8 Hz), 5.38 (d, 0.8H, J=1.8 Hz), 5.28 (d, 0.2H, J=7.0 Hz), 4.29 (d, 0.8H, J=7.6 Hz), 3.91-3.78 (m, 1H), 3.70-3.50 (m, 3H), 3.38 (s, 0.6H), 3.35 (s, 2.4H), 2.87 (m, 0.2H), 2.77 (m, 0.8H), 2.67-2.58 (m, 1H), 1.72-1.55 (m, 1H), 1.52 (s, 7.2H), 1.43 (s, 1.8H), 1.38–1.22 (m, 2H), 0.75 (d, 0.6H, J=6.2 Hz), 0.73 (d, 2.4H, J=6.5 Hz), 0.71 (2d, 3H, J=6.3 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 198.3 (1C), 155.1 (0.2C), 149.3 (0.2C), 147.7 (0.8C), 146.05 (0.8C), 143.4 (2.4C), 143.3 (0.6C), 129.0 (1.2C), 128.9 (4.8C), 127.4 (4.8C), 127.3 (1.2C), 127.2 (0.6C), 127.1 (2.4C), 104.3 (0.2C), 103.7 (0.8C), 92.5 (0.2C), 92.0 (0.8C), 84.3 (0.8C), 84.0 (0.2C), 80.8 (0.8C), 76.7 (0.2C), 71.4 (1C), 67.0 (0.2C), 66.9 (0.8C), 64.3 (0.8C), 63.8 (0.2C), 58.78 (0.8C), 58.73 (0.2C), 42.3 (0.2C), 41.8 (0.8C), 41.4 (0.2C), 39.2 (0.8C), 27.3 (2.4C), 27.2 (0.6C), 26.1 (0.2C), 25.9 (0.8C), 22.9 (0.2C), 22.6 (0.8C), 21.8 (0.8C), 21.2 (0.2C); LRMS: (FAB, NBA, m/z, %): 648 (2) (M+OH⁺), 307 (14), 289 (13), 259 (28), 243 (100), 165 (40).

4(R)-Formyl-3(S)-isobutyl-5(R)-(2-methoxy-ethoxy)-tetrahydro-furan-2-carboxylic acid, tert-butoxycarbonyltrityloxy-amide (25b). This compound was prepared by the same procedure as described above. From 24b (239 mg, 0.38 mmol) in CH₂Cl₂ (10 mL) was obtained 25b (167 mg, 0.26 mmol, 70%, colorless oil) isolated by flash chromatography (hexanes/EtOAc, 9:1 then 4:1); $R_{\rm f}=0.75$ (hexanes/EtOAc, 3:2); $[\alpha]_{\rm D}=-70.9$ (c 0.7, CHCl₃); IR (neat/NaCl) 1770.3, 1724.7, 1646.0. 1598.6 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.56 (d, 1H, J=3.6 Hz), 7.40–7.32 (m, 6H), 7.31–7.22 (m, 9H), 5.29 (d, 1H, J=5.0 Hz), 4.25 (d, 1H, J=8.4 Hz), 3.78 (ddd, 1H, J=8.0, 8.0, 11.0 Hz), 3.54 (ddd, 1H, J=4.9, 6.2, 11.0 Hz), 3.47-3.43 (m, 2H), 3.32 (s, 3H), 3.08 (m, 1H), 2.58 (ddd, 1H, J=3.8, 5.1, 10.6 Hz), 1.53 (s, 9H), 1.23 (m, 1H), 1.18-1.05 (m, 2H), 0.68 (d, 3H, J=6.3 Hz), 0.66 (d, 3H, J=6.3 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 199.8, 148.1, 147.0, 143.6, 129.1, 129.0, 127.4, 127.2, 127.2, 104.2, 92.0, 84.3, 83.0, 71.4, 66.8, 62.4, 58.9, 41.7, 37.8, 27.5, 25.5, 22.4; LRMS: (FAB, NBA, m/z, %): 654 (1) $(M+Na^{+}), 632 (1) (M+H^{+}), 243 (100).$

5(S)-(tert-Butoxycarbonyl-trityloxy-carbamoyl)-4(S)-isobutyl-2(S)-(2-methoxy-ethoxy)-tetrahydro-furan-3(S)carboxylic acid (26a). This compound was prepared following the same procedure as for 22a. From 25a (141 mg, 0.22 mmol) in *t*-BuOH (5 mL), NaH₂PO₄ (52 mg, 0.33 mmol) in water (1 mL), 2-methyl-2-butene (0.4 mL) and sodium chlorite (61 mg, 0.67 mmol) was obtained after purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 9:1), 26a (136 mg, 0.21 mmol, 94%, white crystals); $R_{\rm f}$ =0.45 (CH₂Cl₂/CH₃OH, 9:1); $[\alpha]_{D} = -9.8$ (c 0.7, CH₃OH); IR (neat/NaCl) 3443.8, 1770.3, 1738.0, 1713.2, 1651.4 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) (2 rotamers 4:1) δ 7.35–7.23 (m, 15H), 5.36 (d, 0.2H, J=2.8 Hz), 5.27 (d, 0.8H, J=3.0 Hz), 5.25 (d, 0.2H, J=8.7 Hz), 4.11 (d, 0.8H, J=9.0 Hz), 3.85–3.70 (m, 1H), 3.64-3.46 (m, 3H), 3.31 (s, 0.6H), 3.28 (s, 2.4H), 2.80-2.61

(m, 1.2H), 1.51 (s, 7.2H), 1.43 (s, 1.8H), 1.58–1.38 (m, 1H), 1.33–1.12 (m, 2H), 0.75 (d, 3H, J=6.2 Hz), 0.70 (d, 3H, J=6.5 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 175.6 (1C), 157.0 (0.2C), 151.0 (0.2C), 149.5 (0.8C), 148.0 (0.8C), 145.0 (2.4C), 128.56 (4.8C), 128.4 (3C), 108.5 (0.2C), 108.1 (0.8C), 93.8 (0.2C), 93.3 (0.8C), 85.5 (0.8C), 85.3 (0.2C), 82.4 (0.8C), 77.7 (0.2C), 72.7 (1C), 68.3 (0.2C), 68.1 (0.8C), 27.9 (2.4C), 27.7 (0.6C), 27.1 (0.2C), 26.9 (0.8C), 23.4 (1C), 22.7 (0.8C), 22.4 (0.2C), LRMS: (FAB, NBA, m/z, %): 670 (1) (M+Na⁺), 648 (1) (M+H⁺), 548 (71), 243 (100); HRMS calcd for C_{37H46}O₉N (M+H⁺) 648.31726, found 648.31850.

5(S)-(tert-Butoxycarbonyl-trityloxy-carbamoyl)-4(S)-isobutyl-2(R)-(2-methoxy)-tetrahydro-furan-3(S)carboxylic acid (26b). From 25b (79 mg, 0.12 mmol) in t-BuOH (2.5 mL), NaH₂PO₄ (29 mg, 0.19 mmol) in water (0.5 mL), 2-methyl-2-butene (0.2 mL) and sodium chlorite (34 mg, 0.37 mmol) was obtained after purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 9:1) 26b (76 mg, 0.21 mmol, 94%, white foam); $R_{\rm f}=0.48$ (CH₂Cl₂/CH₃OH, 9:1); $[\alpha]_{D} = -79.3$ (c 0.8, CHCl₃); IR (neat/NaCl) 3412.0, 1769.8, 1723.3, 1642.0, 1596.2 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) & 7.35–7.29 (m, 6H), 7.29–7.21 (m, 9H), 5.20 (d, 1H, J=5.0 Hz), 4.12 (d, 1H, J=9.1 Hz), 3.63 (ddd, 1H, J=3.6, 6.0, 10.0 Hz), 3.58–3.45 (m, 3H), 3.30 (s, 3H), 2.97 (m, 1H), 2.75 (dd, 1H, J=5.0, 11.1 Hz), 1.50 (s, 9H), 1.32-1.22 (m, 2H), 1.10 (m, 1H), 0.68 (d, 3H, ¹³C NMR J=5.9 Hz), 0.65 (d, 3H, J=5.9 Hz); (75 MHz, CD₃OD) δ 172.7, 149.6, 149.2, 145.0, 130.3, 128.5, 128.4, 104.6, 93.2, 85.5, 84.2, 72.5, 68.0, 59.3, 57.8, 43.3, 39.1, 28.0, 26.5, 24.2, 22.3; LRMS: (FAB, NBA, m/z, %): 670 (3) (M+Na⁺), 570 (1), 243 (100); HRMS calcd for $C_{37}H_{45}O_9NNa$ (M+Na⁺) 670.29919, found 670.30100.

3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S),4(S)-dicarboxylic acid 2-(*tert*-butoxycarbonyl-trityloxy-amide) 4-[(1-methylcarbamoyl-2(R)-phenyl-ethyl)amide] (27a). To a solution of 26a (64 mg, 0.099 mmol) in THF (2 mL) were added EDC (25 mg, 0.128 mmol), HOBT (17 mg, 0.128 mmol), N-methyl morpholine 0.277 mmol) and *N*-methyl-D-phenylalanine (31 μL, trifluoroacetate salt (46 mg, 0.148 mmol). After stirring for 15 h, the solution was diluted with CH₂Cl₂, washed with 0.1N HCl, water, 0.5N NaHCO₃, brine, dried over Na₂SO₄ and concentrated in vacuo, the residue was purified by flash chromatography (hexanes/EtOAc, 1:1 to 1:4) to provide 27a (54 mg, 0.067 mmol, 68%, colorless oil); $R_{\rm f}$ =0.53 (hexanes/EtOAc, 2:3); $[\alpha]_{\rm D}$ =-5.7 (c 1.1, CHCl₃); IR (neat/NaCl) 3315.8, 1775.1, 1674.7, 1629.4 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) (2 rotamers 4:1) δ 7.31–7.15 (m, 20H), 5.23 (d, 0.2H, J=9.0 Hz), 4.80 (d, 0.2H, J=3.8 Hz), 4.72 (d, 0.8H, J=3.5 Hz), 4.61 (2dd, 1H, J=6.5, 9.0 Hz), 4.08 (d, 0.8H, J=9.6 Hz), 3.73-3.60 (m, 1H), 3.52–3.35 (m, 3H), 3.32 (s, 0.6H), 3.30 (s, 2.4H), 3.06 (dd, 1H, J=6.5, 13.5 Hz), 2.86 (dd, 0.2H, J=8.9, 13.5 Hz), 2.85 (dd, 0.8H, J=9.0, 13.5 Hz), 2.70–2.54 (m, 2H), 2.65 (s, 3H), 1.48 (s, 7.2H), 1.42 (s, 1.8H), 1.40-1.27 (m, 1H), 1.10 (ddd, 1H, J=5.5, 8.5, 13.5 Hz), 1.00 (ddd, 1H, J=5.1, 9.5, 13.5 Hz), 0.70 (d, 2.4H, J=6.5 Hz), 0.68 (d,

0.6H, J=6.0 Hz), 0.60 (d, 2.4H, J=6.5 Hz), 0.58 (d, 0.6H, J=6.0 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 173.5 (0.8C), 172.9 (1C), 172.5 (0.2C), 156.6 (0.2C), 151.0 (0.2C), 149.4 (0.8C), 147.8 (0.8C), 145.0 (2.4C), 144.9 (0.6C), 138.4 (1C), 130.6, 130.5, 130.3, 130.2, 129.92, 129.88, 129.77, 129.65, 129.4, 129.2, 129.0, 128.8, 128.6, 128.4, 127.8, 127.7, 109.0 (0.2C), 108.6 (0.8C), 93.7 (0.2C), 93.3 (0.8C), 85.6 (0.8C), 85.3 (0.2C), 82.3 (0.8C), 77.2 (0.2C), 72.7 (1C), 68.3 (0.2C), 68.1 (0.8C), 59.1 (0.2C), 59.0 (0.8C), 45.0 (0.2C), 43.5 (1C), 42.8 (0.8C), 27.9 (2.4C), 27.7 (0.6C), 27.1 (0.2C), 26.9 (0.8C), 23.4 (1C), 22.7 (0.8C), 22.4 (0.2C); LRMS: (FAB, NBA, m/z, %): 809 (1) (M+H⁺), 243 (100); HRMS calcd for C₄₇H₅₉O₉N₃ (M+H⁺) 809.42511, found 809.42820.

3(S)-Isobutyl-5(R)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S),4(S)-dicarboxylic acid 2-(*tert*-butoxycarbonyl trityloxy-amide) 4-[(1-methylcarbamoyl-2(R)-phenyl-ethyl)amide] (27b). Following the procedure above, 26b (75 mg, 0.116 mmol) in THF (3 mL), EDC (29 mg, 0.151 mmol), HOBT (20 mg, 0.151 mmol), N-methyl morpholine (36 µL, 0.32 mmol) and N-methyl-D-phenylalanine trifluoroacetate salt (54 mg, 0.174 mmol) afforded, after flash chromatography (hexanes/EtOAc, 1:1 to 1:4), **27b** (87 mg, 0.108 mmol, 93%, colorless oil); $R_{\rm f}$ =0.29 (hexanes/EtOAc, 4:1); $[\alpha]_D = -36.8$ (c 0.8, CHCl₃); IR (neat/NaCl) 3295.6, 3061.6, 1771.6, 1648.0 cm⁻ ί: ¹Η NMR (300 MHz, CDCl₃) δ 7.40-7.17 (m, 20H), 6.26 (d, 1H, J=4.6 Hz), 5.13 (d, 1H, J=3.6 Hz), 4.60 (ddd, 1H, J=6.1, 7.8, 13.5 Hz), 4.13 (d, 1H, J=8.8 Hz), 3.80 (ddd, 1H, J=3.6, 3.6, 11.0 Hz), 3.60 (ddd, 1H, J=4.0, 7.1, 11.0 Hz), 3.48-3.40 (m, 2H), 3.29 (s, 3H), 3.12 (dd, 1H, J=6.1, 13.5 Hz), 3.03 (dd, 1H, J=7.8, 13.5 Hz), 2.72 (d, 3H, J=4.5 Hz), 2.58–2.52 (m, 2H), 1.54 (s, 9H), 1.27–1.05 (m, 3H), 0.54 (d, 3H, J=5.9 Hz), 0.48 (d, 3H, J=5.9 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 168.9, 148.1, 146.8, 143.5, 137.0, 129.2, 129.0, 128.6, 127.5, 127.2, 126.8, 101.9, 92.2, 84.4, 82.1, 70.9, 66.4, 58.6, 57.3, 54.7, 41.2, 40.7, 37.5, 27.6, 26.2, 24.9, 23.2, 21.8; LRMS: (FAB, NBA, *m/z*, %): 830 (1) $(M+Na^+)$, 808 (2) $(M+H^+)$, 243 (100); HRMS calcd for $C_{47}H_{57}O_9N_3$ (M+H⁺) 808.41730, found 808.41490.

3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S),4(S)-dicarboxylic acid 4-benzhydryl-amide 2-(tertbutoxycarbonyl-trityloxy-amide) (28). To a solution of 26a (72 mg, 0.111 mmol) in THF (2 mL) were added EDC (28 mg, 0.144 mmol), HOBT (19 mg, 0.144 mmol), N-methyl morpholine (16 µL, 0.144 mmol) and diphenylmethylamine (29 µL, 0.167 mmol). The resulting solution was stirred for 14 h then diluted with CH₂Cl₂ washed with 0.1N HCl, water, 0.5N NaHCO₃, brine, dried over Na₂SO₄ and concentrated in vacuo, the residue was purified by flash chromatography (hexanes/EtOAc, 1:1 to 1:4) to provide 28 $(70 \text{ mg}, 0.086 \text{ mmol}, 78\%, \text{ colorless oil}); R_f = 0.47 \text{ (hexanes/})$ EtOAc, 3:2); $[\alpha]_D = +3.7$ (*c* 1.1, CH₃OH); IR (neat/NaCl) 3318.9, 1770.5, 1686.0 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) (2 rotamers 8:1, major rotamer described) δ 7.38–7.17 (m, 25H), 6.57 (d, 1H, J=8.2 Hz), 6.26 (d, 1H, J=8.2 Hz), 5.14 (d, 1H, J=5.0 Hz), 4.22 (d, 1H, J=9.5 Hz), 3.84 (ddd, 1H, J=3.1, 4.9, 11.0 Hz), 3.58 (ddd, 1H, J=3.6, 6.1, 11.0 Hz), 3.53-3.44 (m, 2H), 3.22 (s, 3H), 2.83 (m, 1H), 2.58 (dd, 1H, J=4.9, 9.5 Hz), 1.49 (s, 9H), 1.30–1.20 (m, 1H), 1.15 (ddd,

1H, J=5.7, 7.4, 13.3 Hz), 1.08 (ddd, 1H, J=6.0, 8.1, 13.3 Hz), 0.69 (d, 3H, J=6.1 Hz), 0.63 (d, 3H, J=6.1 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 72.4, 149.5, 147.8, 145.0, 142.8, 142.6, 130.3, 130.2, 129.5, 128.9, 128.6, 128.4, 128.3, 109.0, 93.3, 85.5, 82.5, 72.7, 68.8, 60.1, 59.1, 58.1, 44.2, 41.0, 27.9, 27.0, 23.9, 22.0; LRMS: (FAB, NBA, m/z, %): 813 (1) (M+H⁺), 307 (28), 289 (20), 243 (100), 167 (35); HRMS calcd for C₅₀H₆₀O₈N₃ (M+NH₄⁺) 830.43805, found 830.43760.

3(S)-Isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S),4(S)-dicarboxylic acid 2-hydroxyamide 4-[(1methylcarbamoyl-2(R)-phenyl-ethyl)-amide] (11a). To a solution of 27a (50 mg, 0.062 mmol) in CH₂Cl₂ (2 mL) was added a solution of 10% TFA in CH₂Cl₂ (1 mL) as to maintain a deep yellow color. The solution was stirred for 1 h then diluted by CH₂Cl₂ washed with 0.5 N NaHCO₃, then brine, dried over Na₂SO₄ and concentrated in vacuo. Purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 9:1) afforded **11a** (24 mg, 0.052 mmol, 83%, colorless oil); $R_{\rm f}=0.19$ (EtOAc); $[\alpha]_{\rm D}=+32.2$ (c 0.8, CH₃OH); IR (neat/ NaCl) 3290.1, 1652.2, 1644.2 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.30–7.15 (m, 5H), 4.80 (d, 1H, J=2.9 Hz), 4.61 (dd, 1H, J=6.1, 9.1 Hz), 4.02 (d, 1H, J=8.7 Hz), 3.70 (m, 1H), 3.52-3.42 (m, 3H), 3.36 (s, 3H), 3.09 (dd, 1H, J=6.1, 13.5 Hz), 2.87 (dd, 1H, J=9.1, 13.5 Hz), 2.69 (dd, 1H, J=2.6, 7.0 Hz), 2.67 (s, 3H), 1.62-1.35 (m, 3H), 0.86 (d, 3H, *J*=6.2 Hz), 0.76 (d, 3H, *J*=6.2 Hz); ¹³C NMR (75 MHz, CD₃OD) & 173.6, 173.5, 169.9, 138.5, 130.3, 129.4, 127.7, 108.7, 83.2, 72.7, 68.2, 60.0, 59.2, 56.0, 46.5, 43.2, 39.1, 30.7, 27.3, 26.3, 23.8, 22.2; LRMS: (FAB, NBA, *m/z*, %): $488 (20) (M+Na^{+}), 466 (85) (M+H^{+}), 307 (66), 289 (41),$ 205 (52), 107 (67); HRMS calcd for $C_{23}H_{36}O_7N_3$ (M+H⁺) 466.25534, found 466.25400.

3(S)-Isobutyl-5(R)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S),4(S)-dicarboxylic acid 2-hydroxyamide 4-[(1-methylcarbamoyl-2(R)-phenyl-ethyl)-amide] (11b). Following the same procedure as described above, 27b (85 mg, 0.105 mmol) provided **11b** (28 mg, 0.060 mmol, 57%, colorless oil); $R_{\rm f}=0.21$ (EtOAc); $[\alpha]_{\rm D}=-56.7$ (c 0.8, CH₃OH); ¹H NMR (300 MHz, CD₃OD) δ 7.30–7.15 (m, 5H), 5.09 (d, 1H, J=5.1 Hz), 4.53 (dd, 1H, J=5.9, 9.2 Hz), 4.02 (d, 1H, J=8.1 Hz), 3.60-3.37 (m, 4H), 3.37 (s, 3H), 3.08 (dd, 1H, J=5.9, 13.6 Hz), 2.92–2.81 (m, 2H), 2.74 (dd, 1H, J=5.2, 11.0 Hz), 2.68 (s, 3H), 1.62 (m,1H), 1.45 (ddd, 1H, J=6.8, 6.8, 6.8 Hz), 1.31 (ddd, 1H, J=6.8, 6.8, 6.8 Hz), 0.82 (d, 6H, J=6.3 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 174.0, 171.7, 170.0, 138.6, 132.4, 130.2, 129.9, 129.5, 127.8, 105.8, 85.1, 72.2, 69.6, 59.1, 58.1, 56.3, 45.0, 43.0, 39.0, 27.0, 26.3, 23.3; LRMS: (FAB, NBA, *m/z*, %): 488 (7) (M+Na⁺), 466 (21) (M+H⁺), 419 (35), 390 (15), 132 (100); HRMS calcd for $C_{23}H_{36}O_7N_3$ (M+H⁺) 466.25534, found 466.25790.

3(*S*)-**I**sobutyI-5(*S*)-(2-methoxy-ethoxy)-tetrahydro-furan-2(*S*),4(*S*)-dicarboxylic acid 4-benzhydryI-amide 2-hydroxyamide (29). Compound 28 (67 mg, 0.082 mmol) in CH₂Cl₂ (2 mL) and 10% TFA in CH₂Cl₂ (1 mL) afforded, after purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 19:1), **29** (32 mg, 0.068 mmol, 83%, colorless oil); $R_{\rm f}$ =0.58 (EtOAc); [α]_D=+32.6 (*c* 0.9, CH₃OH); IR (neat/ NaCl) 3289.8, 3063.2, 1644.9, 1538.5 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.38–7.20 (m, 10H), 6.21 (s, 1H), 5.27 (d, 1H, *J*=2.5 Hz), 4.13 (d, 1H, *J*=7.9 Hz), 3.83 (ddd, 1H, *J*=3.0, 3.3, 8.1 Hz), 3.65 (ddd, 1H, *J*=3.8, 4.1, 8.5 Hz), 3.56 (m, 2H), 3.35 (s, 3H), 2.88 (dd, 1H, *J*=2.5, 6.0 Hz), 2.65 (m, 1H), 1.56 (m, 2H), 1.45 (m, 1H), 0.81 (d, 3H, *J*=6.2 Hz), 0.71 (d, 3H, *J*=6.2 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 173.0, 170.1, 142.8, 142.7, 129.5, 128.8, 128.5, 128.3, 128.0, 109.0, 83.3, 72.8, 68.5, 60.4, 59.1, 58.3, 47.2, 43.0, 27.3, 23.9, 22.0; LRMS: (FAB, NBA, *m/z*, %): 493 (9) (M+Na⁺), 471 (88) (M+H⁺), 307 (58), 289 (35), 182 (35), 167 (100); HRMS calcd for C₂₆H₃₅O₆N₂ (M+H⁺) 471.25220, found 471.25040.

[4(S)-Azido-3(S)-isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)-ylmethoxy]-tert-butyl-diphenyl-silane (30a) and [4(S)-azido-3(S)-isobutyl-5(R)-(2-methoxyethoxy)-tetrahydro-furan-2(S)-ylmethoxy]-tert-butyl**diphenyl-silane** (30b). To a solution of 18 (1.05 g, 2.32 mmol) were added di(S-2-pyridyl) thiocarbonate (3.03 g, 13.9 mmol), 4 Å molecular sieves in CH₂Cl₂ (60 mL) followed by triethylamine (1.9 mL, 13.9 mmol). After stirring for 36 h, the mixture was quickly purified on silica gel and the filtrate processed as usual. Treatment of the residue with methoxyethanol (0.55 mL, 6.9 mmol) and dried silver triflate (1.79 g, 6.9 mmol) following the same procedure as for 19a and 19b gave the desired products. Purification by flash chromatography (hexanes/EtOAc, 50:1) gave 30a (624 mg, 1.22 mmol, 53%, colorless oil) and 30b (296 mg, 0.58 mmol, 25%, colorless oil) separated from the unreacted starting material (97 mg, 0.21 mmol, 9%). The relative configuration of both anomers was determined by NOE spectroscopy; for **30a** (S): $R_f=0.64$ (hexanes/EtOAc, 4:1); $[\alpha]_D = +52.1$ (*c* 0.9, CHCl₃); IR (neat/NaCl) 2098.1 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.75-7.68 (m, 4H), 7.48-7.38 (m, 6H), 5.06 (s, 1H), 3.86-3.75 (m, 4H), 3.68-3.54 (m, 4H), 3.40 (s, 3H), 2.20 (m, 1H), 1.60 (m, 1H), 1.42 (m, 1H), 1.27 (m, 1H), 1.09 (s, 9H), 0.92 (d, 3H, J=5.8 Hz), 0.89 (d, 3H, J=5.8 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 135.7, 135.6, 133.3, 129.7, 127.6, 106.9, 84.4, 71.7, 71.5, 66.5, 64.8, 59.0, 43.7, 41.4, 26.8, 26.4, 23.1, 22.1, 19.2; LRMS: (FAB, NBA, m/z, %): 408 (5), 257 (38), 213 (10), 199 (49), 197 (48), 135 (90). For 30b (*R*): $R_{\rm f}$ =0.53 (hexanes/EtOAc, 4:1); $[\alpha]_{\rm D}$ =-44.7 (c 1.1, CHCl₃); IR (neat/NaCl) 2107.2 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) & 7.72–7.68 (m, 4H), 7.48–7.35 (m, 6H), 5.04 (d, 1H, J=4.6 Hz), 3.86–3.73 (m, 4H), 3.60 (ddd, 1H, J=4.6, 6.0, 10.8 Hz), 3.48 (m, 2H), 3.32 (s, 3H), 3.12 (dd, 1H, J=4.6, 10.8 Hz), 2.35 (m, 1H), 1.68 (m, 1H), 1.40 (m, 2H), 1.08 (s, 9H), 0.92 (d, 3H, J=5.8 Hz), 0.85 (d, 3H, J=5.8 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 136.1, 133.9, 130.2, 128.1, 103.4, 85.4, 72.2, 67.6, 67.3, 59.4, 43.0, 39.8, 27.3, 26.5, 23.7, 23.1, 19.7; LRMS: (FAB, NBA, m/z, %): 512 (19) (M+H⁺), 510 (18), 408 (49), 378 (20), 351 (40), 322 (38), 257 (100), 135 (92).

5(S)-(*tert*-Butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-2(S)-(2-methoxy-ethoxy)-tetrahydro-furan-3(S)-ylamine (**31a**). A solution of **30a** (220 mg, 0.43 mmol) in absolute ethanol (20 mL) was stirred for 20 h in presence of 10% palladium-on-charcoal (150 mg) under hydrogen atmosphere. The suspension was then filtered on a celite pad and concentrated in vacuo. Purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:19) afforded **31a** (189 mg, 0.39 mmol, 96%, pale yellow oil); $R_{\rm f}$ =0.36 (EtOAc); [α]_D=+41.9 (*c* 0.9, CHCl₃); IR (neat/NaCl) 3375.1 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.75–7.67 (m, 4H), 7.45– 7.35 (m, 6H), 4.84 (s, 1H), 3.87–3.77 (m, 3H), 3.68 (dd, 1H, J=3.5, 10.4 Hz), 3.64–3.53 (m, 3H), 3.40 (s, 3H), 3.10 (m, 1H), 1.93 (m, 1H), 1.70 (m, 2H), 1.55 (m, 1H), 1.46 (m, 1H), 1.30 (m, 1H), 1.09 (s, 9H), 0.89 (d, 3H, J=6.1 Hz), 0.87 (d, 3H, J=6.1 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 135.65, 135.60, 133.3, 129.65, 129.61, 127.63, 127.60, 110.6, 84.8, 71.8, 66.1, 65.2, 63.5, 58.9, 45.8, 42.4, 26.8, 26.2, 22.8, 22.6, 19.2; HRMS calcd for C₂₈H₄₄O₄NSi (M+H⁺) 486.30396, found 486.30280.

5(S)-(tert-Butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-2(R)-(2-methoxy-ethoxy)-tetrahydro-furan-3(S)-ylamine (31b). From 30b (429 mg, 0.84 mmol) in absolute ethanol (20 mL) and 10% palladium-on-charcoal (150 mg) was obtained, after purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:19), **31b** (329 mg, 0.68 mmol, 82%, colorless oil); $R_f=0.21$ (CH₂Cl₂/CH₃OH, 9:1); $[\alpha]_D=$ -35.5 (c 0.8, CHCl₃); IR (neat/NaCl) 3389.0 cm⁻¹; ¹H NMR (400 MHz, CD₃OD) δ 7.72-7.67 (m, 4H), 7.48-7.36 (m, 6H), 4.80 (d, 1H, J=4.5 Hz), 3.81 (ddd, 1H, J=3.5, 5.8, 11.0 Hz), 3.78–3.67 (m, 3H), 3.56–3.44 (m, 3H), 3.28 (s, 3H), 2.83 (dd, 1H, J=4.5, 10.0 Hz), 3.10 (m, 1H), 1.75-1.63 (m, 1H), 1.49-1.33 (m, 2H), 1.25-1.17 (m, 1H), 1.05 (s, 9H), 0.83 (d, 3H, J=6.2 Hz), 0.73 (d, 3H, J=6.2 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 136.8, 130.9, 128.83, 128.81, 104.3, 87.4, 72.9, 69.5, 67.4, 62.4, 59.1, 44.3, 43.3, 27.5, 27.1, 24.3, 22.3, 20.1; LRMS: (FAB, NBA, m/z, %): 486 (24) (M+H⁺), 428 (10), 410 (18), 241 (13), 199 (37), 197 (41), 154 (66), 135 (100); HRMS calcd for $C_{28}H_{44}O_4NSi$ (M+H⁺) 486.30396, found 486.30190.

2(S)-Acetylamino-N-[5(S)-(tert-butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-2(S)-(2-methoxy-ethoxy)tetrahydro-furan-3(S)-yl]-3-phenyl-propionamide (32a). To a solution of N-acetyl-L-phenylalanine (164 mg, 0.79 mmol) in THF (5 mL) were successively added EDC (139 mg, 0.72 mmol), HOBT (98 mg, 0.72 mmol) and N-methyl morpholine (79 µL, 0.72 mmol). After stirring for 30 min, 31a (319 mg, 0.66 mmol) in THF (1 mL) was added dropwise. The resulting mixture processed as described for 27a to provide 32a (291 mg, 0.43 mmol, 66%, colorless oil); $R_f=0.56$ (hexanes/EtOAc, 1:4); $[\alpha]_{D} = +20.3$ (c 0.9, CHCl₃); IR (neat/NaCl) 3285.5, 1633.8 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.70–7.65 (m, 4H), 7.48-7.37 (m, 6H), 7.28-7.15 (m, 5H), 4.49 (t, 1H, J=7.8 Hz), 4.43 (d, 1H, J=1.3 Hz), 3.97 (dd, 1H, J=1.3, 5.0 Hz), 3.77-3.65 (m, 4H), 3.55-3.45 (m, 3H), 3.36 (s, 3H), 2.96 (dd, 1H, J=7.8, 13.2 Hz), 2.87 (dd, 1H, J=7.8, 13.2 Hz), 1.89 (s, 3H), 1.84 (m, 1H), 1.50 (m, 1H), 1.37 (ddd, 1H, J=6.0, 9.1, 14.0 Hz), 1.21 (ddd, 1H, J=5.9, 8.0, 14.0 Hz), 1.03 (s, 9H), 0.80 (d, 3H, J=6.5 Hz), 0.78 (d, 3H, J=6.5 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 172.8, 172.6, 138.2, 136.7, 134.9, 134.7, 130.9, 130.4, 129.4, 128.8, 127.8, 109.0, 85.6, 72.8, 67.3, 66.9, 62.6, 59.1, 56.0, 46.0, 42.9, 39.2, 27.4, 27.2, 23.5, 22.8, 22.4, 20.0; LRMS: (FAB, NBA, m/z, %): 697 (10) (M+Na⁺), 675 (15) $(M+H^+)$, 617 (21), 599 (100), 154 (84), 135 (57), 120 (94); HRMS calcd for $C_{39}H_{55}O_6N_2Si$ (M+H⁺) 675.38293, found 675.38020.

2(S)-Acetylamino-N-[5(S)-(tert-butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-2(R)-(2-methoxy-ethoxy)-tetrahydrofuran-3(S)-yl]-3-phenyl-propionamide (32b). From 31b (321 mg, 0.662 mmol), N-acetyl-L-phenylalanine (206 mg, 0.99 mmol) in THF (5 mL), EDC (165 mg, 0.86 mmol), HOBt (116 mg, 0.86 mmol) and N-methyl morpholine (95 µL, 0.86 mmol) was obtained after flash chromatography (hexanes/EtOAc, 2:3 then 1:4), 32b (315 mg, 0.47 mmol, 70%, colorless oil); R_f =0.64 (hexanes/EtOAc, 1:4); $[\alpha]_{D} = -13.9$ (c 0.9, CHCl₃); IR (neat/NaCl) 3445.2, 3310.4, 1634.0 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.70-7.63 (m, 4H), 7.48-7.35 (m, 6H), 7.29-7.15 (m, 5H), 4.65 (d, 1H, J=4.4 Hz), 4.63 (dd, 1H, J=6.4, 8.4 Hz), 4.06 (dd, 1H, J=4.3, 10.4 Hz), 3.80-3.70 (m, 1H), 3.70-3.63 (m, 3H), 3.38-3.31 (m, 3H), 3.22 (s, 3H), 3.05 (dd, 1H, J=6.3, 13.5 Hz), 2.83 (dd, 1H, J=8.5, 13.5 Hz), 2.05 (m, 1H), 1.89 (s, 3H), 1.42 (m, 1H), 1.37-1.23 (m, 2H), 1.04 (s, 9H), 0.82 (d, 3H, J=6.3 Hz), 0.75 (d, 3H, J=6.3 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 173.3, 172.8, 138.4, 136.7, 134.5, 134.4, 130.9, 130.3, 129.4, 128.8, 127.7, 102.2, 86.4, 72.7, 68.8, 66.9, 59.1, 59.0, 56.0, 43.2, 41.8, 39.1, 27.4, 26.7, 23.4, 23.2, 22.4, 20.0; LRMS: (FAB, NBA, *m/z*, %): 697 (10) (M+Na⁺), 675 (72) (M+H⁺), 617 (21), 599 (95), 307 (9), 241 (10), 199 (25), 197 (26), 154 (100), 135 (64), 120 (99); HRMS calcd for $C_{39}H_{55}O_6N_2Si (M+H^+)$ 675.38293, found 675.38090.

N-[5(S)-(tert-Butyl-diphenyl-silanyloxymethyl)-4(S)-isobutyl-2(S)-(2-methoxy)-tetrahydro-furan-3(S)yl]-4-methoxy-benzenesulfonamide (33). To p-methoxybenzenesulfonyl chloride (117 mg, 0.57 mmol) in CH₂Cl₂ (15 mL) was added triethylamine (78 µL, 0.57 mmol) at 0°C. After stirring for 30 min, **31a** (183 mg, 0.38 mmol) in CH₂Cl₂ (5 mL) was added. The resulting mixture was stirred at 0°C for 1 h then at room temperature for 15 h. The solution was diluted with CH₂Cl₂, washed with diluted HCl, water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash chromatography (hexanes/EtOAc, 9:1 then 4:1) to provide 33 (218 mg, 0.33 mmol, 88%, colorless oil); $R_f=0.75$ (hexanes/EtOAc, 3:2); $[\alpha]_{D} = +23.5$ (c 0.7, CHCl₃); IR (neat/NaCl) 3273.1, 3072.0, 3049.1, 1597.4, cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, 2H, J=8.9 Hz), 7.72-7.63 (m, 4H), 7.47-7.34 (m, 6H), 6.95 (d, 2H, J=8.9 Hz), 4.97 (d, 1H, J=10 Hz), 4.80 (s, 1H), 3.88 (s, 3H), 3.82 (dd, 1H, J=2.5, 11.2 Hz), 3.72 (m, 1H), 3.68–3.42 (m, 6H), 3.36 (s, 3H), 1.98 (m, 1H), 1.55 (m, 1H), 1.43-1.25 (m, 3H), 1.11 (s, 9H), 0.75 (d, 3H, J=6.0 Hz), 0.72 (d, 3H, J=6.0 Hz); ¹³C NMR (75 MHz, CDCl₃) & 162.8, 135.6, 135.5, 132.9, 132.7, 132.6, 129.84, 129.81, 129.2, 127.80, 127.78, 114.2, 107.2, 84.1, 71.5, 65.8, 64.6, 63.7, 58.9, 55.6, 44.3, 42.4, 27.0, 25.9, 22.5, 22.3, 19.2; LRMS: (FAB, NBA, m/z, %): 678 (24) (M+Na⁺), 654 (21), 486 (74), 324 (46), 135 (88), 59 (100).

2(S)-Acetylamino-N-[5(S)-hydroxymethyl-4(S)-isobutyl-2(S)-(2-methoxy)-tetrahydro-furan-3(S)-yl]-3phenyl-propionamide (34a). The deprotection was performed following the same procedure as for 19a and 19b. From 32a (241 mg, 0.36 mmol) in THF (5 mL), AcOH (61 μ L, 1.07 mmol) and a solution of TBAF (1.07 mL, 1.07 mmol, 1 M solution in THF) was obtained, after flash chromatography (CH₂Cl₂/CH₃OH, 97:3), 34a

(135 mg, 0.31 mmol, 87%, colorless oil); $R_f=0.40$ $(CH_2Cl_2/CH_3OH, 9:1); [\alpha]_D = +19.4 (c 0.9, CHCl_3); IR$ (neat/NaCl) 3417.6, 3300.0, 1654.8 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.30–7.17 (m, 5H), 4.49 (dd, 1H, J=7.0, 8.1 Hz, 4.48 (s, 1H), 4.02 (dd, 1H, J=0.8, 3.7 Hz), 3.78 (ddd, 1H, J=2.5, 4.3, 7.0 Hz), 3.57-3.42 (m, 2H), 3.56–3.44 (m, 4H), 3.36 (s, 3H), 3.05 (dd, 1H, J=7.0, 13.5 Hz), 2.87 (dd, 1H, J=8.1, 13.5 Hz), 1.92 (m, 1H), 1.90 (s, 3H), 1.62 (m, 1H), 1.49 (ddd, 1H, J=6.0, 8.8, 13.5 Hz), 1.32 (ddd, 1H, J=6.3, 8.0, 13.5 Hz), 0.88 (d, 3H, J=6.5 Hz), 0.86 (d, 3H, J=6.5 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 172.9, 172.4, 138.3, 130.3, 129.4, 127.7, 109.1, 86.0, 72.8, 67.1, 63.6, 62.0, 59.1, 56.2, 44.8, 43.2, 39.0, 27.3, 23.4, 22.8, 22.4; LRMS: (FAB, NBA, m/z, %): 437 (10) $(M+H^+)$, 361 (100), 242 (35), 154 (45), 136 (33), 120 (61); HRMS calcd for $C_{23}H_{37}O_6N_2$ (M+H⁺) 437.26517, found 437.26630.

2(S)-Acetylamino-N-[5(S)-hydroxymethyl-4(S)-isobutyl-2(R)-(2-methoxy-ethoxy)-tetrahydro-furan-3(S)-yl]-3phenyl-propionamide (34b). The deprotection was performed following the same procedure as for 19a and 19b. From 32b (305 mg, 0.45 mmol), in THF (5 mL), AcOH (75 µL, 1.31 mmol), solution of TBAF (1.36 mL, 1.36 mmol, 1 M solution in THF) was obtained 34b (160 mg, 0.37 mmol, white crystals, 81%); $R_{\rm f}$ =0.40 $(CH_2Cl_2/CH_3OH, 9:1); [\alpha]_D = -54.0 (c 0.7, CHCl_3); mp$ 134°C; IR (neat/NaCl) 3422.6, 3305.9, 1644.4 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.30-7.15 (m, 5H), 4.65 (d, 1H, J=4.5 Hz), 4.63 (dd, 1H, J=6.4, 8.4 Hz), 4.08 (dd, 1H, J=4.5, 10.4 Hz), 3.80-3.72 (m, 2H), 3.63 (dd, 1H, J=2.9, 11.7 Hz), 3.57-3.40 (m, 4H), 3.33 (s, 3H), 3.08 (dd, 1H, J=6.4, 13.5 Hz), 2.85 (dd, 1H, J=8.4, 13.5 Hz), 2.12 (m, 1H), 1.90 (s, 3H), 1.60 (m, 1H), 1.47–1.25 (m, 2H), 0.89 (d, 3H, J=6.5 Hz), 0.85 (d, 3H, J=6.5 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 173.5, 172.9, 138.4, 130.3, 129.4, 127.7, 102.1, 86.8, 72. 8, 67.4, 66.2, 59.3, 58.9, 56.1, 43.0, 40.5, 39.0, 27.0, 23.6, 22.8, 22.4; LRMS: (FAB, NBA, m/z, %): 437 (21) (M+H⁺), 361 (100), 307 (11), 242 (76), 154 (56), 136 (40), 120 (57); HRMS calcd for $C_{23}H_{36}O_6N_2Na$ (M+Na⁺) 459.24710, found 459.24903.

N-[5(S)-Hydroxymethyl-4(S)-isobutyl-2(S)-(2-methoxyethoxy)-tetrahydro-furan-3(S)-yl]-4-methoxy-benzenesulfonamide (35). The deprotection was performed following the same procedure as described above. From 33 (212 mg, 0.32 mmol), in THF (5 mL), AcOH (80 µL, 1.40 mmol) and TBAF (254 mg, 0.98 mmol) was obtained, after flash chromatography (hexanes/EtOAc, 1:1 then 2:3), **35** (123 mg, 0.29 mmol, colorless oil, 91%); $R_{\rm f}$ =0.64 (EtOAc); $[\alpha]_D = +23.2$ (c 1.1, CHCl₃); IR (neat/NaCl) 3483.8, 3272.0, 1597.1, 1579.3 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.81 (d, 2H, J=9.0 Hz), 6.98 (d, 2H, J=9.0 Hz), 5.70 (m, 1H), 4.88 (s, 1H), 3.86 (s, 3H), 3.85 (dd, 1H, J=2.1,11.9 Hz), 3.79 (ddd, 1H, J=2.2, 2.2, 5.2 Hz), 3.66 (m, 1H), 3.50-3.42 (m, 5H), 3.35 (s, 3H), 2.47 (m, 1H), 1.85 (m, 1H), 1.37-1.22 (m, 3H), 0.72 (d, 3H, J=6.1 Hz), 0.88 (d, 3H, J=6.1 Hz); ¹³C NMR (300 MHz, CDCl₃) δ 162.8, 132.1, 129.2, 114.2, 107.7, 84.1, 71.5, 65.8, 63.8, 62.5, 58.8, 55.5, 43.8, 42.3, 25.8, 22.5, 22.1; LRMS: (FAB, NBA, m/z, %): 440 (80) (M+Na⁺), 342 (100), 171 (42); HRMS calcd for $C_{19}H_{31}O_7NSNa$ (M+Na⁺) 440.17191, found 440.17070.

4(S)-(2(S)-Acetylamino-3-phenyl-propionylamino)-3(S)isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)-carboxylic acid (36a). Oxidation of 34a (155 mg, 0.36 mmol) in dry CH₂Cl₂ (5 mL), with N-methyl morpholine N-oxide (61 mg, 0.53 mmol), 4 Å molecular sieves and a catalytic amount of tetrapropylammonium perruthenate gave the corresponding aldehyde. This was dissolved in t-BuOH (5 mL) and water (1 mL) containing NaH₂PO₄ (83 mg, 0.53 mmol) then 2-methyl-2-butene (0.4 mL) and sodium chlorite (96 mg, 1.07 mmol) were added. After stirring for 3 h, the residue was purified by flash chromatography (CH₂Cl₂/CH₃OH, 19:1 to 9:1) to provide 36a (90 mg, 0.13 mmol, 56%, colorless oil); $R_{\rm f}$ =0.20 (CH₂Cl₂/ CH₃OH, 9:1); $[\alpha]_D = +42.3$ (*c* 1.1, CHCl₃); IR (neat/NaCl) 3290.8, massif over 1700 cm^{-1} ; ¹H NMR (400 MHz, CD₃OD) & 7.30–7.14 (m, 5H), 4.71 (s, 1H), 4.52 (dd, 1H, J=6.3, 9.0 Hz), 4.11 (m, 2H), 3.71 (m, 1H), 3.55–3.49 (m, 3H), 3.35 (s, 3H), 3.06 (dd, 1H, J=6.3, 13.5 Hz), 2.86 (dd, 1H, J=9.0, 13.5 Hz), 2.15 (m, 1H), 1.90 (s, 3H), 1.74 (m, 1H), 1.60–1.53 (m, 2H), 0.89 (d, 3H, J=6.5 Hz), 0.85 (d, 3H, J=6.5 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 177.2, 173.2, 172.9, 138.6, 130.3, 129.5, 127.8, 109.6, 83.8, 72.8, 67.5, 62.0, 59.1, 56.7, 43.4, 39.0, 27.1, 23.7, 22.5, 22.4; LRMS: (FAB, NBA, m/z, %): 473 (38) (M+Na⁺), 451 (7) $(M+H^+)$, 176 (32), 154 (100), 137 (65), 136 (66), 120 (26); HRMS calcd for $C_{23}H_{34}O_7N_2Na(M+H^+)$ 473.22638, found 473.22504.

4(S)-(2(S)-Acetylamino-3-phenyl-propionylamino)-3(S)isobutyl-5(R)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)carboxylic acid (36b). Compound 34b was oxidized following the same procedure as described above. From 34b (155 mg, 0.36 mmol) in dry CH₂Cl₂ (5 mL), N-methyl morpholine N-oxide (122 mg, 1.06 mmol), 4 Å molecular sieves, catalytic amount of tetrapropylammonium perruthenate then tBuOH (5 mL), NaH₂PO₄ (83 mg, 0.53 mmol) in water (1 mL), 2-methyl-2-butene (0.4 mL) and sodium chlorite (96 mg, 1.07 mmol) was obtained **36b** (112 mg, 0.24 mmol, 69%, colorless oil); $R_{\rm f}$ =0.27 (CH₂Cl₂/CH₃OH, 9:1); $[\alpha]_{D} = -30.4$ (c 0.6, CHCl₂); IR (neat/NaCl) 3302.0, 1731.2, 1653.3 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.30-7.15 (m, 5H), 4.73 (d, 1H, J=4.5 Hz), 4.64 (dd, 1H, J=6.4, 8.6 Hz), 4.12 (d, 1H, J=7.9 Hz), 4.09 (m, 1H), 3.82 (m, 1H), 3.59-3.42 (m, 3H), 3.33 (s, 3H), 3.07 (dd, 1H, J=6.4, 13.5 Hz), 2.85 (dd, 1H, J=8.6, 13.5 Hz), 2.51 (m, 1H), 1.90 (s, 3H), 1.70 (m, 1H), 1.50 (m, 2H), 0.89 (d, 3H, J=6.5 Hz), 0.87 (d, 3H, J=6.5 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 175.4, 173.6, 172.9, 138.4, 130.3, 129.4, 127.7, 102.6, 82.4, 72.7, 67.0, 59.0, 58.9, 56.0, 48.2, 43.9, 43.6, 39.1, 26.8, 23.2, 22.3; LRMS: (FAB, NBA, m/z, %): 473 (5) (M+Na⁺), 451 (12) (M+H⁺), 154 (100), 137 (70), 136 (69), 120 (40); HRMS calcd for C₂₃H₃₄O₇N₂Na (M+Na⁺) 473.22638, found 473.22504.

3(*S*)-**I**sobutyl-4(*S*)-(4-methoxy-benzenesulfonylamino)-**5**(*S*)-(2-methoxy)-tetrahydro-furan-2(*S*)-car**boxylic acid (37).** A solution of **35** (123 mg, 0.29 mmol) in dry CH₂Cl₂ (5 mL), *N*-methyl morpholine (102 mg, 0.88 mmol), 4 Å molecular sieves, catalytic amount of tetrapropylammonium perruthenate afforded the aldehyde as described above. Oxidation in *t*-BuOH (5 mL) containing NaH₂PO₄ (69 mg, 0.44 mmol) in water (1 mL), 2-methyl-2-butene (0.2 mL) and sodium chlorite (80 mg, 0.88 mmol), followed by flash chromatography (CH₂Cl₂/CH₃OH, 19:1 to 9:1) gave **37** (94 mg, 0.22 mmol, 74%, colorless oil); $R_{\rm f}$ =0.15 (CH₂Cl₂/CH₃OH, 9:1); [α]_D=+30.4 (*c* 0.9, CH₃OH); IR (neat/NaCl) 3486.3, 3264.6, 1714.6, 1597.5 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.80 (d, 2H, *J*=8.9 Hz), 7.08 (d, 2H, *J*=8.9 Hz), 4.70 (s, 1H), 3.96 (d, 1H, *J*=7.5 Hz), 3.88 (s, 3H), 3.61 (m, 1H), 3.46–3.38 (m, 3H), 3.33 (s, 3H), 3.29 (m, 1H), 2.01 (m, 1H), 1.52–1.25 (m, 3H), 0.76 (d, 3H, *J*=6.0 Hz), 0.74 (d, 3H, *J*=6.0 Hz); ¹³C NMR (75 MHz, CD₃OD) δ 177.4, 164.5, 133.9, 130.4, 115.4, 109.2, 83.6, 72.6, 67.4, 65.9, 59.1, 56.2, 49.2, 43.5, 26.8, 23.4, 22.5; LRMS: (FAB, NBA, *m/z*, %): 454 (9%) (M+Na⁺), 432 (8) (M+H⁺), 154 (100), 136 (84); HRMS calcd for C₁₉H₂₉O₈NSNa (M+Na⁺) 454.15115, found 454.15260.

4(S)-(2(S)-Acetylamino-3-phenyl-propionylamino)-3(S)isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)carboxylic acid trityloxy-amide (38a). The coupling was performed according to the procedure used for 23a. Thus 36a (61 mg, 0.135 mmol) in THF (3 mL), EDC (31 mg, 0.162 mmol), HOBT (22 mg, 0.162 mmol), N-methyl morpholine (18 µL, 0.162 mmol) was treated with N-acetyl-L-phenylalanine (42 mg, 0.20 mmol). The residue was chromatographed (CH₂Cl₂/CH₃OH, 1:0 to 19:1) to provide **38a** (45 mg, 0.063 mmol, 47%, colorless oil); $R_{\rm f}$ =0.32 (hexanes/EtOAc, 4:1); $[\alpha]_D = +17.3$ (c 0.9, CH₃OH); IR (neat/NaCl) 3281.8, 1651.0 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) & 7.45-7.38 (m, 6H), 7.33-7.14 (m, 14H), 4.49 (dd, 1H, J=6.8, 8.2 Hz), 4.45 (s, 1H), 3.97 (dd, 1H, J=3.8, 9.0 Hz), 3.88 (d, 1H, J=6.8 Hz), 3.60 (m, 1H), 3.49-3.38 (m, 3H), 3.30 (s, 3H), 3.02 (dd, 1H, J=6.8, 13.5 Hz), 2.86 (dd, 1H, J=8.2, 13.5 Hz), 1.91 (s, 3H), 1.86 (m, 1H), 1.52 (m, 1H), 1.37 (ddd, 1H, J=6.0, 9.1, 13.5 Hz), 1.18 (ddd, 1H, J=5.5, 7.9, 13.5 Hz), 0.88 (d, 3H, J=6.5 Hz), 0.85 (d, 3H, J=6.5 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 173.1, 172.5, 170.2, 143.3, 138.4, 130.3, 130.1, 130.0, 129.9, 129.5, 129.1, 128.9, 128.8, 128.5, 127.9, 127.8, 109.9, 94.8, 82.4, 72.7, 67.6, 61.3, 59.1, 56.4, 47.7, 42.8, 39.0, 27.2, 23.5, 22.54, 22.45; LRMS: (FAB, NBA, m/z, %): 730 (12) $(M+Na^{+})$, 708 (12) $(M+H^{+})$, 307 (52), 243 (100), 154 (27), 137 (19), 136 (20), 120 (12); HRMS calcd for $C_{42}H_{49}O_7N_3Na (M+Na^+)$ 730.34680, found 730.34820.

4(S)-(2(S)-Acetylamino-3-phenyl-propionylamino)-3(S)isobutyl-5(R)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)carboxylic acid trityloxy-amide (38b). The coupling was performed according to the procedure described above. From 36b (109 mg, 0.24 mmol) in THF (4 mL), EDC (56 mg, 0.29 mmol), HOBT (39 mg, 0.29 mmol), N-methyl morpholine (32 µL, 0.29 mmol) and N-acetyl-L-phenylalanine (75 mg, 0.36 mmol) was obtained, after flash chromatography (hexanes/EtOAc, 1:1 to 1:4), 38b (98 mg, 0.138 mmol, 57%, colorless oil); $R_f=0.23$ (hexanes/EtOAc, 2:3); $[\alpha]_{D}$ =-20.1 (*c* 1.1, CH₃OH); IR (neat/NaCl) 3401.5, 1651.7 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.45-7.38 (m, 6H), 7.35–7.13 (m, 14H), 4.65 (dd, 1H, J=6.7, 8.4 Hz), 4.62 (d, 1H, J=4.5 Hz), 4.05 (dd, 1H, J=4.5, 11.0 Hz), 3.87 (d, 1H, J=8.9 Hz), 3.27 (s, 3H), 3.17-3.10 (m, 3H), 3.07 (dd, 1H, J=6.7, 13.6 Hz), 2.82 (dd, 1H, J=8.4, 13.6 Hz), 2.14 (m, 1H), 1.90 (s, 3H), 1.60-1.48 (m, 1H), 1.34–1.28 (m, 2H), 0.75 (d, 3H, J=6.5 Hz), 0.70 (d, 3H, J=6.5 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 173.4,

172.8, 170.5, 143.1, 138.5, 130.3, 130.0, 129.4, 129.0, 128.5, 127.7, 103.1, 94.6, 83.2, 72.5, 68.1, 59.1, 58.5, 56.0, 55.9, 43.4, 43.2, 38.8, 26.4, 23.6, 23.2, 22.4; LRMS: (FAB, NBA, m/z, %): 730 (12) (M+Na⁺), 708 (8) (M+H⁺), 307 (70), 289 (50), 259 (46), 243 (100), 154 (25), 137 (19), 136 (20), 120 (13); HRMS calcd for C₄₂H₄₉O₇N₃Na (M+Na⁺) 730.34680, found 730.34550.

3(S)-Isobutyl-4(S)-(4-methoxy-benzenesulfonylamino)-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)-carboxylic acid trityloxy-amide (39). The coupling was performed according to the procedure described above. From 37 (73 mg, 0.17 mmol) in THF (3 mL), EDC (49 mg, 0.25 mmol), HOBT (34 mg, 0.25 mmol), N-methyl morpholine (28 µL, 0.25 mmol) and N-acetyl-L-phenylalanine (207 mg, 0.25 mmol) was obtained 39 (62 mg, 0.090 mmol, 54%, colorless oil) after flash chromatography (hexanes/EtOAc, 3:2); $R_f=0.53$ (hexanes/EtOAc, 1:1); $[\alpha]_{D} = +9.4$ (c 0.7, CHCl₃); IR (neat/NaCl) 3253.7, 1694.0, 1597.0, 1578.1 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.33 (s, 1H), 7.70 (d, 2H, J=8.9 Hz), 7.51 (m, 5H), 7.90-7.79 (m, 10H), 6.97 (d, 2H, J=8.9 Hz), 4.85 (s, 1H), 4.31 (d, 1H, J=7.5 Hz), 3.88 (s, 3H), 3.86 (d, 1H, J=6.0 Hz), 3.58 (m, 1H), 3.46–3.39 (m, 3H), 3.31 (s, 3H), 3.27 (dd, 1H, J=1.8, 7.5 Hz), 1.50 (m, 1H), 1.41-1.10 (m, 3H), 0.63 (d, 3H, J=6.3 Hz), 0.61 (d, 3H, J=6.3 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 167.4, 163.0, 141.5, 131.3, 129.2, 128.8, 128.2, 128.0, 127.8, 114.2, 108.3, 93.5, 81.5, 71.3, 66.3, 62.8, 55.6, 48.5, 42.6, 25.7, 22.4, 21.9; LRMS: (FAB, NBA, *m/z*, %): 711 (20) (M+Na⁺), 578 (29), 310 (38), 286 (21), 243 (10); HRMS calcd for $C_{38}H_{44}O_8N_2SNa (M+Na^+)$ 711.27161, found 711.27590.

4(S)-(2(S)-Acetylamino-3-phenyl-propionylamino)-3(S)isobutyl-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)carboxylic acid hydroxyamide (12a). The deprotection was performed using the procedure described for 11a, from 38a (39 mg, 0.055 mmol) in CH_2Cl_2 (2 mL) and 10% TFA in CH₂Cl₂ (1 mL) was obtained after purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 19:1), **12a** (20 mg, 0.042 mmol, 78%, colorless oil): $R_{\rm f}$ =0.19 (EtOAc); $[\alpha]_{D} = +47.7 (c \, 0.8, CH_{3}OH); {}^{1}H NMR (300 MHz, CD_{3}OD)$ δ 7.31–7.17 (m, 5H), 4.60 (d, 1H, J=0.8 Hz), 4.51 (dd, 1H, J=7.1, 8.1 Hz), 4.03 (m, 1H), 4.02 (d, 1H, J=6.8 Hz), 3.68 (m, 1H), 3.53-3.48 (m, 3H), 3.35 (s, 3H), 3.05 (dd, 1H, J=7.1, 13.5 Hz), 2.90 (dd, 1H, J=8.1, 13.5 Hz), 2.18 (m, 1H), 1.91 (s, 3H), 1.67 (m, 1H), 1.55-1.42 (m, 2H), 0.88 (d, 3H, J=6.5 Hz), 0.85 (d, 3H, J=6.5 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 173.2, 172.7, 170.1, 138.4, 130.3, 129.5, 127.8, 109.7, 82.6, 72.7, 67.7, 61.7, 59.1, 56.4, 48.4, 43.2, 38.9, 27.2, 23.2, 22.7, 22.3; LRMS: (FAB, NBA, *m*/*z*, %): 466 (13) (M+H⁺), 390 (18), 154 (100), 137 (83), 136 (76), 120 (36); HRMS calcd for $C_{23}H_{35}O_7N_3Na~(M+Na^+)$ 488.23727, found 488.23919.

4(*S*)-(2(*S*)-Acetylamino-3-phenyl-propionylamino)-3(*S*)isobutyl-5(*R*)-(2-methoxy-ethoxy)-tetrahydro-furan-2(*S*)carboxylic acid hydroxyamide (12b). The deprotection was performed using the procedure described above. Compound **38b** (71 mg, 0.100 mmol) in CH₂Cl₂ (2 mL) was treated with 10% TFA in CH₂Cl₂ (1 mL). A purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 19:1) afforded **12b** (42 mg, 0.090 mmol, 90%, colorless oil); *R*_f=0.35 (EtOAc); [α]_D=−76.9 (*c* 0.7, CH₃OH); mp 152–154°C; IR (neat/NaCl) 3282.0, 1651.0 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.30–7.15 (m, 5H), 4.76 (d, 1H, *J*=4.8 Hz), 4.63 (dd, 1H, *J*=6.8, 8.4 Hz), 4.11 (dd, 1H, *J*=4.8, 10.9 Hz), 4.07 (d, 1H, *J*=8.9 Hz), 3.1 (m, 1H), 3.63–3.45 (m, 3H), 3.39 (s, 3H), 3.07 (dd, 1H, *J*=6.8, 13.6 Hz), 2.85 (dd, 1H, *J*=8.4, 13.6 Hz), 2.41 (m, 1H), 1.90 (s, 3H), 1.73 (m, 1H), 1.55–1.38 (m, 2H), 0.87 (d, 3H, *J*=6.4 Hz), 0.84 (d, 3H, *J*=6.4 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 173.5, 173.0, 171.3, 138.4, 130.3, 129.4, 127.8, 103.2, 83.4, 72.5, 69.1, 59.1, 58.6, 56.1, 44.1, 43.2, 39.0, 26.6, 23.3, 23.2, 22.4; LRMS: (FAB, NBA, *m/z*, %): 466 (21) (M+H⁺), 390 (24), 154 (100), 137 (69), 136 (72), 120 (46); HRMS calcd for C₂₃H₃₅O₇N₃Na (M+Na⁺) 488.23727, found 488.23820.

3(S)-Isobutyl-4(S)-(4-methoxy-benzenesulfonylamino)-5(S)-(2-methoxy-ethoxy)-tetrahydro-furan-2(S)-carboxylic acid hydroxyamide (40). The deprotection was performed using the procedure described above. Compound **39** (89 mg, 0.129 mmol) in CH_2Cl_2 (2 mL) saturated with water was treated with 10% TFA in CH₂Cl₂ (1 mL). Purification by flash chromatography (CH₂Cl₂/CH₃OH, 1:0 to 9:1) afforded **40** (42 mg, 0.117 mmol, 90%, colorless oil): $R_{\rm f}=0.28$ (CH₂Cl₂/CH₃OH, 9:1); [α]_D=+40.7 (*c* 0.9, CHCl₃); IR (neat/NaCl) 3271.7, 1668.1 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{ CD}_3\text{OD}) \delta 7.80 \text{ (d, 2H, } J=6.3 \text{ Hz}), 7.09 \text{ (d,}$ 2H, J=6.3 Hz), 4.90 (m, 3H), 4.79 (s, 1H), 3.95 (d, 1H, J=5.1 Hz), 3.88 (s, 3H), 3.62 (m, 1H), 3.47 (m, 2H), 3.40 (m, 1H), 3.34 (s, 3H), 3.33 (m, 1H), 2.08 (m, 1H), 1.45-1.29 (m, 3H), 0.78 (d, 3H, *J*=3.9 Hz), 0.75 (d, 3H, *J*=3.9 Hz); ¹³C NMR (300 MHz, CD₃OD) δ 169.8, 164.5, 133.6, 130.4, 115.4, 109.6, 82.0, 72.6, 67.6, 65.8, 59.1, 56.2, 48.7, 43.2, 26.8, 23.1, 22.7; LRMS: (FAB, NBA, m/z, %): 469 (9) (M+Na⁺), 447 (15) (M+H⁺), 371 (100), 310 (66), 171 (73), 154 (66), 136 (54), 107 (46); HRMS calcd for $C_{19}H_{30}O_8N_2SNa (M+Na^+)$ 469.16205, found 469.16410.

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References

1. For recent reviews, see (a) Beckett, R. P.; Davidson, A. H.; Drummond, A. H.; Huxley, P.; Whittaker, M. *Drug Discov. Today* **1996**, *1*, 16. (b) Pendas, A. M.; Knauper, V.; Puente, X. S.; Llano, E.; Mattei, M. G.; Apote, S.; Murphy, G.; Lopez-Otin, C. *J. Biol. Chem.* **1997**, 272, 4281. (c) Birkedal-Hansen, H.; Moore, W. G. I.; Bodden, M. K.; Windsor, L. J.; Birkedal-Hansen, B.; DeCarlo, A.; Engler, J. A. *Crit. Rev. Oral Biol. Med.* **1993**, *4*, 197. 2. Powell, W. C.; Matrisian, L. M. *Curr. Top. Microbiol. Immunol.* **1996**, 23, 1.

 (a) Walakovits, L. A.; Bhardwaj, N.; Gallick, G. S.; Lark, M. W. Arthritis Rheum. 1992, 35, 35. (b) Lohmander, L. S.; Hoerrher, L. A.; Lark, M. W. Arthritis Rheum. 1993, 36, 181. (c) Flannery, C.; Lark, M. W.; Sandt, J. D. J. Biol. Chem. 1992, 267, 1008. 4. Gijbels, K.; Masure, S.; Carton, H.; Opdenakker, G. J. J. Neuroimmunol. **1992**, 41, 29.

5. For recent reviews, see (a) Whittaker, M.; Floyd, C. D.; Brown, P.; Gearing, A. J. H. *Chem. Rev.* **1999**, *99*, 2735. (b) Michaelides, M. R.; Curtin, M. L. *Curr. Pharm. Des.* **1999**, *5*, 787. (c) Rasmusen,

H. S.; McCann, P. P. *Pharmacol. Ther.* **1997**, *75*, 69. (d) White, A. D.; Bocan, M. A.; Boxer, P. A.; Peterson, J. T.; Schrier, D. Curr. *Pharm. Des.* **1997**, *3*, 45. (e) Browner, M. F. *Perspect. Drug Discov. Des.* **1994**, *2*, 343. (f) Greenwald, R.; Golub, L. M., Eds. *Inhibition of Matrix Metalloproteinases: Therapeutic Potential*; N. Y. Academy of Sciences, 1994; Vol. 372.

6. (a) Campion, C.; Davidson, A. H.; Dickens, J. P.; Crimmin,
M. J. PCT Int. Appl. WO 90 05,719, 1990. (b) *Chem. Abstr.* **1990**, *113*, 212677c. (c) Grams, F.; Crimmin, M.; Hinnes, L.;
Huxley, P.; Pieper, M.; Tschesche, H.; Bode, W. *Biochemistry* **1995**, *34*, 14012.

7. Nemunaitis, J.; Poole, C.; Primrose, J.; Rosemurgy, A.; Malfetano, J.; Brown, P.; Berrington, A.; Cornish, A.; Lynch, K.; Rasmussen, H.; Kerr, D.; Cox, D.; Millar, A. *Clin. Cancer Res.* **1998**, *4*, 1101.

 Broadhurst, M. J.; Brown, P. A.; Lawton, G.; Ballantyne, N.; Bottomley, K. M. K.; Cooper, M. I.; Eatherton, A. J.; Kilford, I. R.; Malsher, P. J.; Nixon, J. S.; Lewis, E. J.; Sutton, B. M.; Johnson, W. H. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 2299.

9. Rockwell, A.; Melden, M.; Copeland, E. A.; Hardman, K.; Decicco, C. P.; DeGrado, W. F. *J. Am. Chem. Soc.* **1996**, *118*, 10337.

10. (a) Bird, J.; Montana, J. G.; Wills, R. E.; Baxter, A. D.; Owen,
D. A. PCT Int. Appl. WO 98 39,024, 1998. (b) *Chem. Abstr.* 1998, 129, 225751c.

11. (a) MacPherson, L. J.; Bayburt, E. K.; Capparelli, M. P.; Carroll, B. J.; Goldstein, R.; Justice, M. R.; Zhu, L.; Hu, S.; Melton, R. A.; Fryer, L.; Goldberg, R. L.; Doughty, J. R.; Spirito, S.; Blancuzzi, V.; Wilson, D.; O'Byrne, E. M.; Ganu, V.; Parker, D. T. *J. Med. Chem.* **1997**, *40*, 2525. (b) Gonnella, N. C.; Li, Y.-C.; Zhang, X.; Paris, C. G. *Bioorg. Med. Chem.* **1997**, *5*, 2193.

12. (a) Zook, S. E.; Dagnino, Jr., R.; Deason, M. E.; Bender, S. L.; Melnick, M. J. PCT Int Appl. WO 97 20,824, 1997. (b) *Chem. Abstr.* **1997**, *127*, 108945s.

13. Campbell, J. A.; Van Hart, H.; Caufield, J. P.; Eugie, E.; Womble, S.; Martin, R.; Browner, M.; Brandl, M.; Fischer, L.; Masjedizadeh, M.; Castelhano, A.; Broka, C.; Hendricks, R. T.; Walker, R. 216th A.C.S. Meeting, Boston, MA, 1998, MEDI 4.

14. Hanessian, S.; Bouzbouz, S.; Boudon, A.; Tucker, G. C.; Peyroulan, D. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1691.

15. (a) Gomis-Rüth, F.-X.; Maskos, K.; Betz, A.; Huber, R.; Suzuki, K.; Yoshida, N.; Nagase, H.; Brew, K.; Bourenkov, G. P.; Bartunik, H.; Bode, W. Nature 1997, 369, 77. (b) Becker, J. W.; Marcy, A. I.; Rokosz, L. L.; Axel, M. G.; Burbaum, J. J.; Fitzgerald, P. M.; Cameron, P. M.; Esser, C. K.; Hagmann, W. K.; Hermes, J. D.; Springer, J. P. Protein Sci. 1995, 4, 1966. (c) Esser, C. K; Bugianesi, R. L.; Caldwell, C. G.; Chapman, K. T.; Durette, P. L.; Girotra, N. N.; Kopka, I. E.; Lanza, T. J.; Levorse, D. A.; MacCoss, M.; Owens, K. A.; Ponpipom, M. M.; Simeone, J. P.; Harrison, R. K.; Niedzwiecki, L.; Becker, J. W.; Marcy, A. I.; Axel, M. G.; Christen, A. J.; McDonnell, J.; Moore, V. L.; Olszewski, J. M.; Saphos, C.; Visco, D. M.; Shen, F.; Colleti, A.; Krieter, P. A.; Hagmann, W. K. J. Med. Chem. 1997, 40, 1026. (d) Cherney, R. J.; Wang, L.; Meyer, D. T.; Xue, C.-B.; Arner, E. C.; Copeland, R. A.; Covington, M. B.; Hardman, K. D.; Wasserman, Z. R.; Jaffee, B. D.; Decicco, C. P. Bioorg. Med. Chem. Lett. 1999, 9, 1279. (e) Pikul, S.; McDow Dunham, K. L.; Almstead, N. G.; De, B.; Natchus, M. G.; Anastasio, M. V.; McPhail, S. J.; Snider, C. E.; Taiwo, Y. O.; Chen, L.; Dunaway, C. M.; Gu, F.; Mieling, G. E. J. Med. Chem. 1999, 42, 87. (f) Pikul, S.; McDow Dunham, K. L.; Almstead, N. G.; De, B.; Natchus, M. G.; Anastasio, M. V.; McPhail, S. J.; Snider C. E.; Taiwo, Y. O.; Rydel, T.; Dunaway, C. M.; Gu, F.; Mieling, G. E. J. Med. Chem. 1998, 41, 3568. (g) For comparative studies see: Dhanaraj, V.; Ye, Q.-Z; Johnson, L. L.; Hupe, D. J.; Ortwine, D. F.; Dunbar, Jr., J. B.; Rubin, J. R.; Pavlovsky, A.; Humblet, C.; Blundell, T. L. Drug Des. Discov. 1996, 13, 3.

16. (a) Hajduk, P. J.; Sheppard, G.; Nettesheim, D. G.; Olejniczak, E. T.; Shuker, S. B.; Meadows, R. P.; Steinman, D. H.; Carrera, Jr., G. M.; Marcotte, P. A.; Severin, J.; Walter, K.; Smith, H.; Gubbins, E.; Simmer, R.; Holzman, T. F.; Morgan, D. W.; Davidsen, S. K.; Summers, J. B.; Fesik, S. W. *J. Am. Chem. Soc.* **1997**, *119*, 5818.
(b) Gooley, P. R.; O'Connell, J. F.; Marcy, A. I.; Cuca, G. C.; Salowe, S. P.; Bush, B. L.; Hermes, J. D.; Esser, C. K.; Hagmann, W. K.; Springer, J. P.; Johnson, B. A. *Nature Struct. Biol.* **1994**, *1*, 111.

For recent reviews, see (a) Hanessian, S.; McNaughton-Smith,
 G.; Lombart, H.-G.; Lubell, W. D. *Tetrahedron* **1997**, *53*, 12789.
 (b) Gante, J. *Angew. Chem., Int. Ed. Engl.* **1994**, *33*, 1699.
 (c) Olson, L. G.; Bolin, D. R.; Bonner, M. P.; Bös, M.; Cook,
 C. M.; Fry, D. C.; Graves, B. J.; Hatada, M.; Hill, D. E.; Kahn,
 M.; Madison, V. S.; Rusiecki, V. K.; Sarabu, R.; Sepinwall, J.;
 Vincent, G. P.; Voss, M. E. J. Med. Chem. **1993**, *36*, 3039.

 See, for example, (a) Robinson, R. P.; Laird, E. R.; Blake, J. F.; Bordner, J.; Donahue, K. M.; Lopresti-Morrow, L. L.; Mitchell, P. G.; Reese, M. R.; Reeves, L. M.; Stam, E. J.; Yocum, S. A. *J. Med. Chem.* 2000, 43, 2293. (b) Natchus, M. G.; Cheng, M.; Wahl, C. T.; Pikul, S.; Almstead, N. G.; Bradley, R. S.; Taiwo, Y. O.; Mieling, G. E.; Dunaway, C. M.; Snider, C. E.; McIver, J. M.; Barnett, B. L.; McPhail, S. J.; Anastasio, M. B.; De, B. *Bioorg. Med. Chem. Lett.* 1998, 8, 2077. See also: (c) Jacobsen, E. J. PCT Int. Appl. WO 97/32,846. (b) Duan, J.; Decicco, C. P.; Wasserman, Z. R.; Maduskuie, Jr., T. P. PCT Int. Apll. WO 99/ 18,074.

 (a) Almstead, N. G.; Bradley, R. S.; Pikul, S.; De, B.; Natchus, M. G.; Taiwo, Y. O.; Gu, F.; Williams, L. E.; Hynd, B. A.; Janusz, M. J.; Dunaway, M.; Mieling, G. E. J. Med. Chem. 1999, 42, 4547.
 (b) Levin, J. I.; DiJoseph, J. F.; Killar, L. M.; Sung, A.; Walter, T.; Sharr, M. A.; Roth, C. E.; Skotnicki, J. S.; Albright, J. D. Bioorg. Med. Chem. Lett. 1998, 8, 2657. (c) Cheng, M.; De, B.; Almstead, N. G.; Pikul, S.; Dowty, M. E.; Dietsch, C. R.; Dunaway, C. M.; Gu, F.; Hsieh, L. C.; Janusz, M. J.; Taiwo, Y. O.; Natchus, M. G. J. Med. Chem. 1999, 42, 5426. (d) Cheng, M.; De, B.; Pikul, S.; Almstead, N. G.; Natchus, M. G.; Anastasio, M. V.; McPhail, S. J.; Snider, C. E.; Taiwo, Y. O.; Chen, L.; Dunaway, C. M.; Gu, F.; Dowty, M. E.; Mieling, G. E.; Janusz, M. J.; Wang-Weigand, S. J. Med. Chem. 2000, 43, 369.

20. (a) Hanessian, S.; Griffin, A.; Devasthale, P. V. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 3119. (b) Martin, S. F.; Oalmann, C. J.; Liras, S. *Tetrahedron* **1993**, *49*, 3521.

21. (a) Sheppard, G.; Florjancic, A. S.; Giesler, J. R.; Xu, L.; Guo, Y.; Davidsen, S. K.; Marcotte, P. A.; Elmore, I.; Elbert, D. H.; Magoc, T. J.; Bouska, J. J.; Goodfellow, C. L.; Morgan, D. W.; Summers, J. B. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 3251. (b) Xue, C.-B.; He, X.; Roderick, J.; DeGrado, W. F.; Cherney, R. J.; Hardman, K. D.; Nelson, D. J.; Copeland, R. A.; Jaffee, B. D.; Decicco, C. P. *J. Med. Chem.* **1998**, *41*, 1745. (c) Duan, J.-J. W.; Chen, L.; Xue, C.-B.; Wasserman, Z. R.; Hardman, K. D.; Covington M. B.; Copeland, R. R.; Arner, E. C.; Decicco, C. P. *Bioorg. Med. Chem.* **1999**, *9*, 1453. (d) Ksander, G. M.; De

Jesus, R.; Yuan, A.; Ghai, R. D.; Trapani, A.; McMartin, C.; Bohacek, R. J. Med. Chem. **1997**, 40, 495.

22. Kiyama, R.; Tamura, Y.; Watanabe, F.; Tsuzuki, H.; Ohtani, M.; Yodo, M. J. Med. Chem. **1999**, 42, 1723.

23. Takano, S.; Kurotaki, A.; Takahashi, M.; Ogawara, K. Synthesis **1986**, 403.

24. (a) Hanessian, S.; Murray, P. J. Tetrahedron 1987, 43, 5055.

(b) Hanessian, S. *Aldrichim. Acta* **1989**, *22*, 3 and references cited therein.

25. Kowalski, C. J.; Dung, J.-S. J. Am. Chem. Soc. 1980, 102, 7950.

26. For the synthesis of 2-azido lactones from the corresponding enolates, see Hanessian, S.; Vanasse, B.; Yang, H.; Alpegiani, M. *Can. J. Chem.* **1993**, *71*, 1407 and references cited therein. See also Evans, D. A.; Britton, T. C.; Ellman, J. A.; Dorow, R. L. *J. Am. Chem. Soc.* **1990**, *112*, 4011.

27. (a) Lou, B.; Huynh, H. K.; Hanessian, S. In *Preparative Carbohydrate Chemistry*; Hanessian S., Ed.; Dekker: New York, 1996; Chapter 19, 431pp. See also (b) Hanessian, S.; Huynh, H. K.; Reddy, G. V.; McNaughton-Smith, G.; Ernst, B.; Kolb, H. C.; Magnani, J.; Sweeley, C. *Bioorg. Med. Chem. Lett.* 1998, *8*, 2803. (c) Hanessian, S.; Reddy, G. V.; Huynh, H. K.; Pan, J.; Pedatella, S. *Bioorg. Med. Chem. Lett.* 1997, *7*, 2729. (d) Hanessian, S.; Qiu, D.; Prabhanjan, H.; Reddy, G. V.; Lou, B. *Can. J. Chem.* 1996, *24*, 1738. (e) Hanessian, S.; Conde, J. J.; Huynh, H. K.; Lou, B. *Tetrahedron* 1996, *52*, 10827.

28. (a) Fischer, E. Ber. Dtsch. Chem. Ges. 1893, 26, 2400.
(b) Methods Carbohydr. Chem. 1962, 2, 328.

29. (a) Schmidt, R. R.; Michel, J. Angew. Chem., Int. Ed. Engl.
1980, 19, 731. (b) Schmidt, R. R. Angew. Chem., Int. Ed. Engl.
1986, 25, 212. (c) Schmidt, R. R. Pure Appl. Chem. 1989, 61, 1257.
30. (a) Griffith, W. P.; Ley, S. V.; Whitcombe, G. P.; White, A. D. J. Chem. Soc., Chem. Commum. 1987, 1625. For a review on TPAP oxidation, see Ley, S. V.; Norman, J.; Griffith, W. P.; Marsden, S. P. Synthesis 1994. 639.

31. Bal, B. S.; Childers, W. E.; Pinnick, H. W. *Tetrahedron* **1981**, 37, 2091. See also (b) Lindgren, B. O.; Nilsson, T. *Acta Chem.*

Scand. 1973, 27, 888. (c) Kraus, G. A.; Tashner, M. J. J. Org. Chem. 1980, 45, 1175.

32. Lutz, W. B. J. Org. Chem. 1971, 36, 3836.

33. For a recent discussion of the hydrolysis of O-substituted hydroxamates, see: Hanessian, S.; Johnstone, S. J. Org. Chem. **1999**, *64*, 5896.

34. Human purified MMPs were purchased or acquired. MMP-2 gelatinase A and MMP-9 gelatinase B from Boehringer Mannheim (Meylan, France), MMP-3 stromelysin 1 from Valbiotech (Paris, France). All enzymes were activated by APMA (4-aminophenylmercuric acetate). Inhibition of MMP-3 was quantified by using the peptidomimetic substrate (7-methoxycoumarine-4-yl)-Arg-Pro-Lys-Pro-Tyr-Ala-Nva-Trp-Met-Lys(Dnp)-NH₂ (Bachem, Bubbendorf, Switzerland) which is cleaved between Ala and Nva. For inhibition studies of the other enzymes, the substrate Dnp-Pro-Cha-Gly-Cys(Me)-His-Ala-Lys(Nma)-NH₂ (Bachem), which is cleaved between amino acids Gly and Cys, was used. The fluorescent cleavage products were measured with a cytofluorometer (Cytofluor 2350, Millipore, PerSeptive Systems, Voisins le Bretonneux, France) equipped with a combination of 340 and 440 nm filters for excitation and emission, respectively.

35. Steinman, D. H.; Curtin, M. L.; Garland, R. B.; Davidsen, S. K.; Heyman, H. R.; Holms, J. H.; Albert, D. H.; Magoc, T. J.; Nagy, I. B.; Marcotte, P. A.; Li, J.; Morgan, D. W.; Hutchins, C.; Summers, J. B. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2087.

36. (a) Weiner, S. J.; Kollman, P. A.; Case, D. A.; Singh, U. C.; Ghio, C.; Alagona, G.; Profeta, Jr., S.; Weiner P. K. *J. Am. Chem. Soc.* **1984**, *106*, 765. (b) Weiner, S. J.; Kollman, P. A. *J. Comput. Chem.* **1986**, *6*, 718.

37. InsightII, Molecular Simulations, 1995, San Diego, CA.

38. Guida, W. C.; Bohacek, R. S.; Erion, M. D. J. Comput. Chem. 1992, 13, 214.

39. Hoops, S. C.; Anderson, K. W.; Merz, K. M. J. Am. Chem. Soc. **1991**, *113*, 8262.

40. Watson, S. C.; Eastham, J. F. J. Organometal. Chem. 1967, 9, 165.

41. Baudat, R.; Petrzilka, M. Helv. Chim. Acta 1979, 62, 1406.