A NEW ACCESS TO CHIRAL AZIRIDINES BY ENZYMATIC TRANSESTERIFICATION OF MESO-BIS(ACETOXYMETHYL)AZIRIDINES

Kaoru Fuji*, Takeo Kawabata, Yoshimitsu Kiryu, and Yukio Sugiura Institute for Chemical Research, Kyoto University, Uji, Kyoto 611, Japan Tooru Taga and Yoshinao Miwa

Faculty of Pharmaceutical Science, Kyoto University, Sakyo-ku, Kyoto 606, Japan

Abstract: (2R, 3S)-2-Acetoxymethyl-3-hydroxymethylazirizines of high enantiomeric purity were prepared through enzymatic transesterification of *meso*bis(acetoxymethyl)aziridines.

An antibiotic FR-900482 (1), recently isolated and characterized by Fujisawa research group, has been shown to exhibit exceptionally potent anti-tumor activity.¹ Structually related antibiotic, mytomycin C (2), has already been used for cancer chemotherapy.² These antibiotics are activated in the cells to form interstrand DNA-DNA cross links.^{3,4} This is believed to be the origin of the acute cytotoxity. The aziridine moiety in 1 and 2 is suggested to play a key role in the interaction with DNA. From a synthetic standpoint of view, chiral aziridines possessing cisdisubstituents are expected to be the most promising starting materials for a total synthsis of these molecules. Recently chiral aziridin 3 was prepared in thirteen steps from L-methionine and used for the synthesis of the core nucleus of 1.5 Here we describe a new and convenient method for the synthesis of chiral cisdisubstituted aziridines by means of enzymatic transesterification of meso derivatives. There have been many papers reported concerning enzymatic hydrolysis of *meso* diesters.⁶ To the best of our knowledge, however, enzymatic transformation of meso-disubstitued aziridines to chiral ones has never been reported.



Meso azirizine 5 was prepared starting from commercially available cis-2butene-1,4-diol (4) (Scheme I). Silylation of 4 was followed by epoxidation and Blum's procedure⁷ to give 5. Protection of the aziridine nitrogen and desilylation affoderd diol 6 and 7. The cis-relationship between the two hydroxymethyl groups is ununivocally established by an X-ray crystallographic analysis of 6 (Figure I).⁸ Meso-diacetates 8 and 9 were obtained by acetylation of 6 and 7. The overall yields of 8 and 9 from 4 were 32% and 22%, respectively.

Taking advantage of enzymatic transesterification in organic solvents,⁹ mesodiacetates 8 and 9 were treated with ten equivalents of butanol in diisopropyl ether 10 in the presence of enzyme (Table I). After indicated reaction time at 37 the reaction was terminated by removing the enzyme through filtration. °C.11 Evaporation of the solvent in vacuo gave the residue which was purified by SiO_2 preparative TLC. Enantiomeric excess(ee) of the products was determined by the mesurement of 400 MHz ¹H NMR spectra with Eu(hfc)₃ or with a combination of (S)binaphtol and Eu(hfc)₃,¹² Among a variety of enzyme screened, Amano P has proved to be highly enantioselective in catalyzing the transesterification (runs 1) and 5). Use of larger amount of Amano P or immobilized Amano P on celite¹³ has significantly shorten the reaction time (runs 6 and 7). The absolute configuration of 10 turned out to be (2R, 3S) by an X-ray crystal structure determination of 12.8mp 96-97 °C, $[\alpha]_D^{20}$ +8.1° (c 0.76, CHCl₃), which was obtained by the treatment of 10 with (1S)-(-)-camphanic chloride (Figure II). The absolute configuration of Nbenzyloxycarbonyl aziridine 11 was also determined to be (2R, 3S) by the chemical correlation with 10.14 Interestingly, the major enantiomer obtained in each experiment in runs 1-3 and 5-9 in Table I has the same absolute configuration.



a) TBDMSCI / NEt₃ / DMAP, b) MCPBA, c) NaN₃ / NH₄Cl, d) PPh₃, e) RCl, f) Bu₄NF, g) Ac₂O / Py

Run	Substrate	Enzyme	Time (days)	Product	Yield (%)	ee (%)	Configuration
1	8	Amano Pb)	3	10	76	95h)i)	(2 <i>R</i> , 3 <i>S</i>)
2	8	CCL ^{c)}	4	10	56	72 ^h)	(2 <i>R</i> , 3 <i>S</i>)
3	8	PPLd)	4	10	22	56h)	(2 <i>R</i> , 3 <i>S</i>)
4	8	PLE ^{e)}	4	10	0	-	-
5	9	Amano Pb)	5	11	66	98j)k)	(2 <i>R</i> , 3 <i>S</i>)
6	9	Amano P ^{f)}	6/24	11	68	98j)	(2 <i>R</i> , 3 <i>S</i>)
7	9	Amano Pg)	9/24	11	68	97j)	(2 <i>R</i> , 3 <i>S</i>)
8	9	CCLc)	5	11	14	22j)	(2R, 3S)
9	9	PPL ^{d)}	5	11	30	49j)	(2 <i>R</i> , 3 <i>S</i>)
10	9	PLE ^{e)}	5	11	5	-	•

Table I. Enzymatic Transesterification of 8 and 9 in n-BuOH - i-Pr2O.a)

a) All reactions were run at 37 °C. b) Lipase Amano P: 600 units/mmol substrate c) Lipase from *Candida cylinddracea*: 350000 units/mmol substrate d) Porcine pancreatic lipase: 3000 units/mmol substrate e) Pig liver esterase: 500 units/mmol substrate f) 6000 units/mmol substrate g) Immobilized Amano P on celite; 600 units/mmol substrate h) Determined by 400-MHz ¹H NMR with a combination of Eu(hfc)₃ (0.25 eq.) and (S)-binaphtol (2.0 eq.) i) Mp. 97-99 °C (after recrystallization from ether), $[\alpha]_D^{20}$ -23.8° (c 1.0, CHCl₃) j) Determined by 400-MHz ¹H NMR with Eu(hfc)₃ (0.6 eq.) k) Colorless oil: $[\alpha]_D^{20}$ -17.8° (c 1.3, CHCl₃).

Figure I. X-ray structure of 6.



Figure II. X-ray structure of 12.



In conclusion, an enantioselective method for the preparation of cisdisubstituted aziridine was established through enzymatic transesterification of *meso*-derivatives. The present method may provide promising starting materials for the synthesis of 1 and 2.

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References and Notes

- 1) Uchida, I.; Takase, S.; Kayakiri, H.; Kiyoto, S.; Hashimoto M. J. Am. Chem. Soc., 1987, 109, 4108.
- 2) Carter, S. K.; Crooke, S. T. Mitomycin C.; Current Status and New Developments; Academic Press; New York, 1979.
- 3) Masuda, K.; Nakamura, T.; Shimomura, K.; Shibata, T.; Terano, H.; Kohsaka, M. J. Antibiot., 1988, 41, 1497.
- 4) Tomasz, M.; Lipman, R.; McGuinness, B. F.; Nakanishi, K. J. Am. Chem. Soc., 1988, 110, 5892.
- 5) Jones, R. J.; Rapoport, H. J. Org. Chem., 1990, 55, 1144.
- 6) For leading references, see: a) Huang, Fu-C.; Hsu, Lee, L. F.; Mittal, R. S. D.; Ravikumar, P. R.; Chan, J. A.; Sih, C. J.; Caspi, E.; Eck, C. R. J. Am. Chem. Soc., 1975, 97, 4144. b) Ito, Y.; Shibata, T.; Arita, M.; Sawai, H.; Ohno, M. J. Am. Chem. Soc., 1981, 103, 6739. c) Schneider, M.; Engel, N.; Hönicke, P.; Heinemann, G.; Görisch, H. Angew. Chem. Int. Ed. Engl., 1984, 23, 67.
- 7) Ittah, Y.; Sasson, Y.; Shahak, I.; Tsaroom, S.; Blum J. J. Org. Chem., 1978, 43, 4271.
- 8) Crystal data for $6: C_{11}H_{15}NO_4S$, space group $P2_1/c$ with a = 11.165 (2), b = 13.149 (2), c = 9.024 (2) Å and $D_c = 1.410$ g cm⁻³ for Z = 4. Crystal data for $12: C_{23}H_{29}NO_8S$, space group $P2_1$ with a = 13.090 (2), b = 6.316 (2), c = 14.917 (3) Å and $D_c = 1.305$ g cm⁻³ for Z = 2. Bond lengths, bond angles, and atomic coordinates for 6 and 12 have been deposited with the Cambridge Crystallographic Data Center.
- 9) Klibanov, A. M.; Acc. Chem. Res., 1990, 23, 114.
- 10) Bevinakatti, H. S.; Banerji, A. A.; Newadkar, R. V. J. Org. Chem., 1989, 54, 2453.
- 11) The reactions could be performed also at ambient temperature thougt it requires longer reaction time. For examples, when 8 and 9 were treated with Amano P (600 units/mmol substrate) at ambient temperature, 10 and 11 were obtained in 76% (97% ee) and 68% (97% ee) yield after 7 and 11 days' stirring, respectively.
- 12) Toda, F.; Mori, K.; Okada, J.; Node, M.; Itoh, A.; Oomine, K.; Fuji, K. Chem. Lett., 1988, 131.
- 13) Inagaki, M.; Hiratake, J.; Nishioka, T.; Oda, J. Agric. Biol. Chem., 1989, 53, 1879.
- 14) Tosylate 10 produced through hydrogenolysis (5% Pd-C, H₂) of 11 (93% op) followed by tosylation (TsCl, Py) showed $[\alpha]_D^{25}$ -18.3° (c 1.1, CHCl₃) (77% op). The partial racemization is supposed to occur at the stage of tosylation.

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