July 1997 *SYNLETT* 859

The Synthesis of N,N'-O-Trisubstituted Hydroxylamines via a Mild Reductive Alkylation Procedure: An Improved Synthesis of the MMI Backbone

Eric E. Swayze* and Yogesh S. Sanghvi

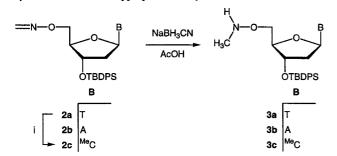
Department of Medicinal Chemistry, Isis Pharmaceuticals, 2292 Faraday Av., Carlsbad, CA 92008

Fax: (760) 603-0036; Bitnet eswayze@isisph.com

Received 19 January 1997

Abstract: A mild procedure for the synthesis of N,N',O-trisubstituted hydroxylamines via the reductive alkylation of N,O-disubstituted hydroxylamines with both aldehydes and ketones has been developed. We have applied this reaction to the synthesis of methylene(methylimino) (MMI) linked dimeric nucleosides, which are of potential use in antisense applications.

Our research toward the development of modified oligonucleotides for use in antisense¹ applications has resulted in a focus on the methylene(methylimino) (MMI) linkage as our preferred backbone modification.² Oligonucleotide analogs which contain alternating phosphodiester and MMI linkages are achiral, have reduced charge, are stable to nucleolytic degradation, and hybridize to their complement RNA with good affinity and base-pair specificity.³ We have therefore concentrated our synthetic efforts to develop a general, convenient, and high yielding synthesis of nucleosidic dimers such as 1a (Scheme 2) which contain the MMI backbone, yet retain the necessary hydroxyl groups for subsequent functionalization and use in automated DNA synthesis. Both of our previous syntheses^{3,4} of MMI dimers required several steps from precursors to the requisite MMI dimers, and are intolerant of common amide base protecting groups, and also the acid labile 4,4'-dimethoxytriphenylmethyl group commonly used for automated solid phase DNA synthesis. We have therefore reduced oxime 2a³ to give the 5'-O-(N-methylamino) thymidine derivative 3a (Scheme 1), and investigated methods for a mild, single-step reductive alkylation of 3a with an appropriate aldehyde.



Scheme 1. (i) 1,2,4-triazole/Et₃N/POCl₃/MeCN then NH₃/1,4-dioxane; T=Thymine, A=adenine, MeC=5-methylcytosine

We initially attempted a coupling of 5'-O-tert-butyldiphenylsilyl-3'-deoxy-3'-C-formylthymidine⁵ with **3a** using standard methods for the reductive alkylation of amines with carbonyl compounds, including NaBH₃CN,⁶ NaBH(OAc)₃⁷ and Zn(BH₄)₂⁸ in a variety of solvents, with and without added acid catalyst (AcOH, HCl, or PPTS). Under all conditions attempted, we observed no formation of the desired dimer **1a** (Scheme 2), and obtained either no reaction, intractable mixtures of products, or preferential reduction of the aldehyde to alcohol. This last difficulty could presumably be overcome by employing excess aldehyde, however, because we intended to use valuable 3'-C-formyl nucleoside derivatives as our precursors, a reaction which required no excess aldehyde was imperative. We also attempted BH₃•pyridine mediated conditions⁹ and observed no reaction, however, upon the addition of 1 eq. of pyridinium para-toluenesulfonate (PPTS), ¹⁰ the solution effervesced, ¹¹ and both starting components were consumed to

form the desired dimer 1a. We subsequently optimized the reaction conditions to 1 eq. aldehyde, 1 eq. 3a, 1 eq. PPTS, and 1 eq. BH₃*pyridine in 3:1 MeOH/THF (0.2 M, THF added for solubility), which provided 1a in 81% isolated yield. We were also able to alkylate 3a with simple aldehydes (benzaldehyde) and ketones (acetone) to prepare 1b and 1c in 88% and 91% isolated yields, respectively, which serves to illustrate in part the generality of this method.

Scheme 2

The versatility of this method for the synthesis of MMI dimers was demonstrated by the synthesis of a series of nucleosidic dimers and the phosphoramidites necessary for oligomerization. In order to test the compatibility of the BH3 • pyridine reaction conditions with the four natural bases necessary for the synthesis of any desired dimer, we prepared the 5'-O-DMT-protected styrenes 4a-c (Scheme 3) in a manner analogous to that previously described for the 5'-O-TBDPS-protected derivatives. 13 Protection of 4c in the standard way14 gave 4d, and the benzoyl-protected 5methylcytosine derivative 4e was prepared from the thymidine derivative 4a by displacement of the corresponding triazolide with sodium benzamide in 75% yield. 15 The aldehydes 5a-e were prepared in acceptable yields by a one-pot cis-dihydroxylation/oxidation sequence, and 5f was prepared by ammonolysis of 5e, then hydrolysis of the resulting imine.

Reductive alkylation of **3a** with **5b** and **5c** proceeded smoothly to give excellent yields of the G^{ibu}*T and A*T dimers **6b** and **6c**, however reaction with the benzoyl-protected aldehyde **5d** resulted in significant depurination, and the A^{Bz}*T dimer **6d** was isolated in only 45% yield. The attempted synthesis of the MeCBz*T dimer **7a** directly from **3a** and **5e** resulted an intractable mixture of products; however, use of the unprotected aldehyde **5f** gave an 84% yield of **6a**, which was then benzoylated to give **7a** in 93% yield (Scheme 4). After desilylation with Et₃N•2HF¹⁶ to provide 95% of **8a**, the appropriately protected phosphoramidite **9a** was prepared in nearly quantitative yield after precipitation of the crude product into hexane.¹⁷ This sequence provides the phosphoramidite necessary for incorporation into oligonucleotides in 70% overall yield from the basic building blocks **5f** and **3a**, and represents a dramatic improvement over previous methods for the synthesis of these dimers.^{3,4}

We also prepared several other mixed base dimers *via* the alkylation of **3b** and **3c**, which were readily prepared from the previously reported oximes **2a** and **3a**. From these hydroxylamines and the aldehydes **5a**, **5c**, and **5f**, the dimers **6e-g** were prepared in 70-85% isolated yields. Benzoylation of **6e**, followed by desilylation and phosphitylation

860 LETTERS SYNLETT

Scheme 3. (i) BzCl/pyridine then 1 N NaOH/EtOH/dioxane (2:5:5 , 0 °C); (ii) 1,2,4-triazole/Et₃N/POCl₃/MeCN then NaH/PhCONH₂/dioxane; (iii) NH₃/MeOH/THF then EtOAc/1 N AcOH; Bz=N-benzoyl, ibu=N-isobutyryl

Scheme 4. (i) Bz_2O/DMF ; (ii) dimethyl-acetamide dimethyl acetal/ MeOH; (iii) BzCVpyridine then 1 N NaOH/EtOH/dioxane (2:5:5, 0 °C); aca=N-[1-(dimethylamino)ethylene]

provided the phosphoramidite **9e** in similar overall yield to that obtained for **9a**. Since 6-*N*-benzoyl-2',3'-dideoxyadenosine derivatives underwent partial depurination under our mildly acidic reaction conditions, we reasoned that they would be unstable to detritylation conditions used for solid phase oligomerization. We therefore opted to protect dimers containing a 2',3'-dideoxyadenosine moiety as the 'upper' unit as their dimethylacetamidine derivatives, ¹⁹ which provided the desired stability toward depurination.

To illustrate this strategy, **6f** was converted to the *bis*-dimethylacetamidine derivative **7f** in quantitative yield, however the amidine protecting group on the 5-methylcytosine unit was lost during desilylation to give **8f**. This material was selectively benzoylated in 80% yield to give **8h**, then converted to the phosphoramidite **9h**. For protection of **6g**, a simpler strategy could be used, as the adenine-containing unit is a 2'-deoxy nucleoside, and therefore the 6-N-benzoyl derivative is stable to detritylation conditions. As Bz₂O/DMF is insufficient to protect an adenine nucleobase, **7g** was prepared in 89% yield by per-benzoylation then partial hydrolysis. Desilylation and phosphitylation gave the phosphoramidite **9g** in excellent yield.

In conclusion, an expedient synthesis of MMI dimers has been developed, which utilizes a new method of reductive alkylation of N,Odisubstituted hydroxylamines as the key step. The mild conditions of this reaction allow use of the highly acid labile DMT group for protection of the 5'-hydroxyl groups throughout the sequence, which is especially convenient since it is required in standard automated synthesis of oligonucleotides. We have demonstrated that all four bases are compatible with this methodology, and elucidated base-protection strategies amenable to the synthesis and oligomerization of all sixteen possible MMI dimer phosphoramidites, ²⁰ which should allow a rigorous evaluation of the value of the MMI backbone in antisense strategies. Additionally, the BH3 • pyridine/PPTS conditions have been shown to effect the reductive alkylation of an N,O-disubstituted hydroxylamine with a representative non-nucleosidic aldehyde and ketone, and the high yields may allow for use in the rapidly expanding field of combinatorial chemistry.²¹ Furthermore, the near neutral conditions of this reaction permit the presence of both acid and base sensitive groups in either substrate, and the 1:1 stoicheometry should facilitate the convergent synthesis of complex molecules which contain substituted hydroxylamines.²²

Acknowledgement We would like to thank Drs. T. Hoshiko, L. Bellon, B. Bhat and S. Dimock for conducting some preliminary experiments toward the development of this procedure.

References and Notes

- Therapeutic Applications of Oligonucleotides Crooke, S. T., Ed.;
 R. G. Landes Co.: Austin, TX, 1995.
- Sanghvi, Y. S. Swayze, E. E.; Peoc'h, D.; Bhat, B.; Dimock, S. Nucleosides Nucleotides 1997, 16, in press.
- Bhat, B.; Swayze, E. E.; Wheeler, P.; Dimock, S.; Perbost, M.; Sanghvi, Y. S. J. Org. Chem. 1996, 61, 8186-8199, and references cited therein.
- Vasseur, J.-J.; Debart, F.; Sanghvi, Y. S.; Cook, P. D. J. Am. Chem. Soc. 1992, 114, 4006-4007.
- Sanghvi, Y. S.; Bharadwaj, R.; Debart, F.; De Mesmaeker, A. Synthesis 1994, 1163-1166.
- a) Lane, C. F. Synthesis 1975, 135-146.
 b) Mattson, R. J.; Pham, K. M.; Leuck, D. J.; Cowen, K. A. J. Org. Chem. 1990, 55, 2552-2554
- Abdel-Magid, A. F.; Carson, K.; Harris, B. D.; Maryanoff, C. A.; Shah, R. D. J. Org. Chem. 1996, 61, 3849-3862.
- 8. Bhattacharyya, S.; Chatterjee, A.; Duttachowdhury, S. K. *J. Chem. Soc. Perkin Trans. I* **1994**, 1-2.
- a) Bomann, M. D.; Guch, I. C.; DiMare, M. J. Org. Chem. 1995, 60, 5995-5996.
 b) Moormann, A. E. Synthetic Comm. 1993, 23, 789-795.

July 1997 SYNLETT 861

10. Commercial BH₃•pyridine complex (Aldrich) contains excess pyridine, and the resulting solution is pH~5.

- 11. As no gas evolution was noted upon addition of BH₃•pyridine to PPTS in MeOH, the gas evolved was presumably produced as the reaction proceeded. It is reasonable to guess that H₂ was produced as the boron-containing species produced upon donation of a hydride decomposed under the reaction conditions.
- 12. General Procedure for Reductive Alkylation. To a solution of aldehyde or ketone (1 eq), 3a (1 eq), and pyridinium para-toluene sulfonate (1 eq) in 3:1 v/v MeOH/THF (5 mL/mmole) was added pyridine-borane complex (1 eq, 8 M in BH₃) dropwise. Stirring was continued until the reaction was complete as judged by TLC. The mixture was partitioned between EtOAc (20 mL/mmole) and 5% aqueous NaHCO₃ (20 mL/mmole), and the organic layer washed with 5% aqueous NaHCO₃ (10 mL/mmole) and brine, then dried (MgSO₄), concentrated, and purified by flash column chromatography. All new compounds were characterized by ¹H NMR, ¹³C NMR, HRMS and/or elemental analysis.
- Sanghvi, Y. S.; Ross, B.; Bharadwaj, R.; Vasseur, J.-J. Tetrahedron Lett. 1994, 35, 4697-4700.
- Jones, R. A. Preparation of Protected Deoxyribonucleosides. In Oligonucleotide Synthesis; Gait, M. J. Ed.; IRL Press: New York, 1984; pp 24-25.
- Perbost, M.; Sanghvi, Y. S. J. Chem. Soc. Perkin Trans. 1 1994, 2051-2052.
- 16. McClinton, M. A. Aldrichimica Acta 1995, 28, 31-35.
- 17. Atkinson, T.; Smith, M. Solid-phase Synthesis of Oligodeoxyribonucleotides by the Phosphite-triester Method. In

- Oligonucleotide Synthesis; Gait, M. J. Ed.; IRL Press: New York, 1984; pp 41-45.
- 18. This was confirmed by treating 6d with 3% trichloroacetic acid in CH₂Cl₂. In addition to DMT removal, the formation of 6-N-benzoyladenine was evident by TLC, with an estimated t_{1/2} of <5 minutes. This is not surprising as sequential removal of hydroxyl groups from adenosine is known to greatly enhance the rate of depurination (Srivastava, P. C.; Robins, R. K.; Meyer, R. B. Jr. Synthesis and Properties of Purine Nucleosides and Nucleotides. In Chemistry of Nucleosides and Nucleotides, Vol. 1; Townsend, L. B. Ed.; Plenum Press: New York, 1988; pp. 266-267).</p>
- McBride, L. J.; Kierzek, R.; Beaucage, S. L.; Caruthers, M. H. J. Am. Chem. Soc. 1986, 108, 2040-2048.
- 20. Studies toward this end are in progress, and will be reported in due course. The phosphoramidites 9a, 9e, 9g, and 9h, have all been incorporated into antisense oligonucleotides on a Millipore Expidite automated DNA synthesizer with coupling efficiencies equal to the natural DNA monomers. The synthesis and biological evaluation of these oligomers will be reported separately.
- 21. Work at Isis has identified several scaffolds which contain N,O-disubstituted hydroxylamines (An, H.; Cook, P. D. Tetrahedron Lett. 1996, 37, 7233-7236 and Cook, P. D.; Sanghvi, Y. S.; Kung, P.-P. PCT Int. Appl. WO 95/18623, July 13, 1995.). It is straightforward to envision the functionalization of scaffolds of this type with aldehydes and ketones via the methods described herein, and studies directed toward this end are ongoing.
- For an example, see: Nicolaou, K. C.; Groneberg, R. D. J. Am. Chem. Soc. 1990, 112, 4085-4086.