

SYNTHESIS OF 19-HYDROXY LTB₄, AN ASSUMED METABOLITE OF LEUKOTRIENE B₄.

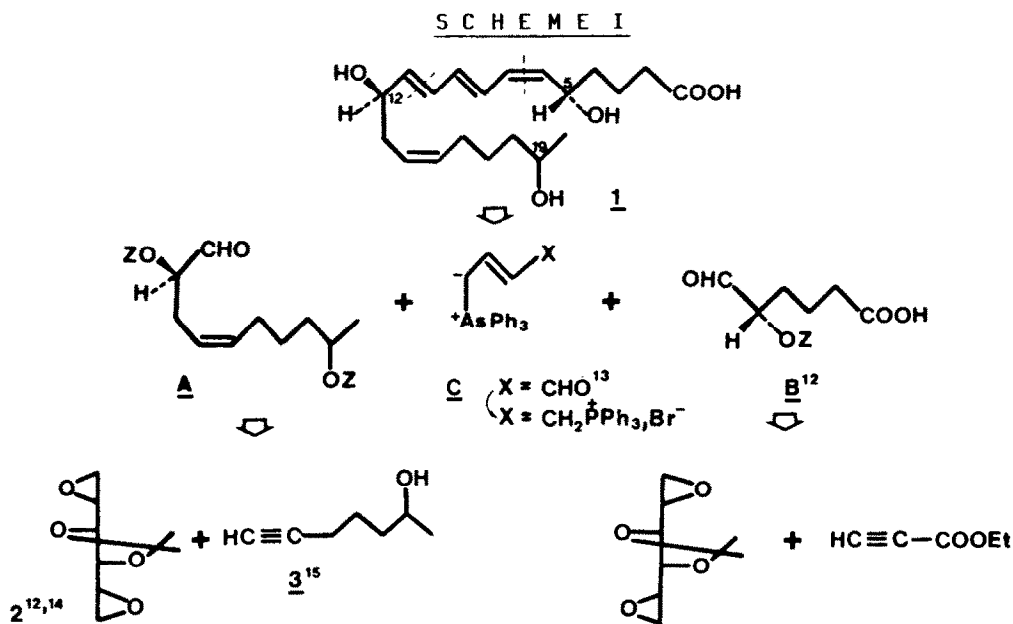
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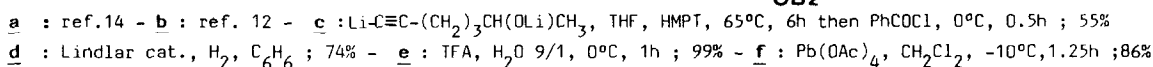
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Summary : From D-mannitol, the total convergent synthesis of 19-hydroxy LTB₄, an assumed metabolite of LTB₄, has been carried out via α -hydroxyaldehydes. Connection at a four carbon atoms interval uses conjugated arsonium ylide.

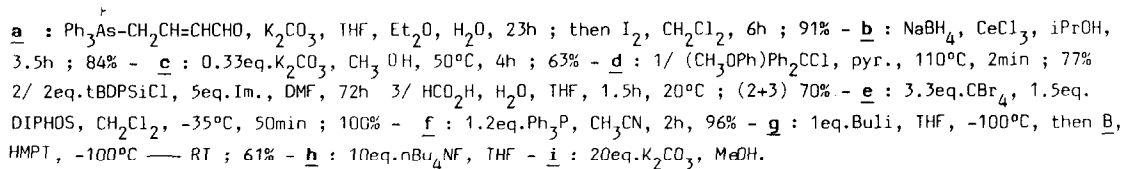
In leucocytes, oxygenation of arachidonic acid in presence of 5-lipoxygenase produces 5-HPETE which is the precursor of leukotrienes (1). Among them, LTB₄ is considered as an important mediator of inflammation (2,3). Biological activity of LTB₄ is reduced by metabolic oxidations. ω -Hydroxy and ω -carboxy LTB₄ have been reported to be the major products of catabolism of LTB₄ in human polymorphonuclear leukocytes (4-7) and have been targets for synthesis (8,9). Recently, preliminary evidence has been provided for the formation of another hydroxylated metabolite of LTB₄ in rat liver microsomes (10) and in rat leucocytes (11). On the basis of mass spectral data, a 19-hydroxy LTB₄ structure was proposed for this metabolite. To confirm the structure we have undertaken the synthesis of this assumed metabolite. Absolute configurations of carbon atoms 5 and 12 are supposed to be respectively S and R as for LTB₄ but the 19-C absolute configuration is unknown, therefore a mixture of the two 19-epimers is required for metabolite identification.

Scheme I outlines the retrosynthetic route to 19-hydroxy LTB₄ **1**.





SCHEME III - Synthesis of 19-hydroxy LTB₄ + $\Delta^{6,7}$ trans isomer (21)



This synthesis is related to the chemistry developed by us for (+)-LTB₄ (12) with regard to the obtention of the two α -hydroxyaldehydes A and B, key intermediates prepared from D-mannitol. To prepare the connection between aldehydes A and B, four atoms are then introduced in one step by a Wittig type condensation using a conjugated arsonium ylide C (13).

From D-mannitol, the synthesis of enantiomerically pure aldehyde B has been previously described (12,14). The synthesis of protected dihydroxy aldehyde A (scheme II) involves diepoxide 2 (12,14) and 1-heptyn-6 ol 3 (15). Nucleophilic opening of enantiomerically pure diepoxide 2 (12,14) by the dilithio-derivative of 3 is followed *in situ* by tetrabenzoylation, controlled hydrogenation of the triple bonds, removal of the acetonide group and oxidative cleavage of the 3,4-diol. Since these transformations preserve the C2 axis of symmetry of 2, two moles of A are obtained from one mole of D-mannitol. Condensation of aldehyde A with arsonium ylide C (13) (scheme III) leads to the E,E-dienal 4 (yield 91% after adjusting experimental conditions). Aldehyde 4 is carefully reduced (16), the primary alcohol 5 obtained is transformed into the phosphonium salt 9a (Z=Bz) from which, even at -100°C, a stable ylide could not be obtained (elimination of the allylic benzoate) (17). To prevent this elimination the secondary allylic alcohol is protected by a silyl group (6→7) (18,19). The ylide prepared from phosphonium salt 9b (Z=tBBPSi) is stable at -100°C and condensation occurs with aldehyde B. Protected 19-hydroxy LTB₄ 10 is obtained with its $\Delta^{6,7}$ trans isomer 11 (ratio 10:11, ca 9/8). The compounds are easily separated by column chromatography and fully characterised (21). Sequential deprotection of 19 leads to the potassium salt of 19-hydroxy LTB₄, which is currently compared with metabolites of LTB₄ produced by different cellular systems (human and rat PMNL) (20).

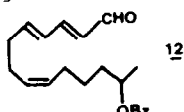
Aknowledgements : We thank Dr. D. Mansuy who drew our attention to 19-hydroxy LTB₄ and Dr. J-L Boucher for performing analytical studies.

Financial aid of this work was partially provided by CNRS in the section ARI-ASP "Lipides Pharmacologiquement actifs". A. Bonnet (on leave from industry) was supported by a FONGECIF grant.

REFERENCES and NOTES

1. B. Samuelsson, *Science*, **220**, 568 (1983).
2. J. Palmblad, C.L. Malmsten, A.M. Uden, O. Radmark, L. Engsted, B. Samuelsson, *Blood*, **58**, 658 (1981).
3. A.W. Ford-Hutchinson, M.A. Bray, M.V. Doig, M.E. Shipley, M.J.H. Smith, *Nature*, **286**, 264 (1980).
4. S. Shak, I.M. Goldstein, *J. Biol. Chem.*, **259**, 10181 (1984).
5. W.S. Powell, *J. Biol. Chem.*, **259**, 3082 (1984).
6. S.J. Feinmark, J.A. Lindgren, M.H. Claesson, C. Malmsten, B. Samuelsson, *FEBS Letters*, **136**, 141 (1981).
7. G. Hansson, J.A. Lindgren, S.E. Dahlen, P. Hedqvist, B. Samuelsson, *FEBS Letters*, **130**, 107 (1981).
8. R. Zamboni, J. Rokach, *Tetrahedron Lett.*, **23**, 4751 (1982).
9. K.C. Nicolaou, Y.S. Chung, P.E. Hernandez, I.M. Taffer, R.E. Zipkin, *Tetrahedron Lett.*, **27**, 1881 (1986).
10. a) J.F. Newton, R.D. Eckardt, P.E. Bender, T.B. Leonard, K.M. Straub, *Biochem. Biophys. Res. Comm.*, **128**, 733 (1985).
b) M.C. Romano, R.D. Eckardt, P.E. Bender, T.B. Leonard, K.M. Straub, J.F. Newton, *J. Biol. Chem.*, **262**, 1590 (1987).
11. W.S. Powell, *Biochem. Biophys. Res. Comm.*, **145**, 991 (1987).

12. Y. Le Merrer, C. Gravier, D. Languin-Micas, J.C. Depezay, *Tetrahedron Lett.*, **27**, 4161 (1986).
13. a) Y. Wang, J. Li, Y. Wu, Y.Z. Huang, L. Shi, J. Yang, *Tetrahedron Lett.*, **27**, 4583 (1986).
b) L. Shi, W. Xia, J. Yang, X. Wen, Y.Z. Huang, *Tetrahedron Lett.*, **28**, 2155 (1987).
14. a) Y. Le Merrer, A. Duréault, C. Gravier, D. Languin, J.C. Depezay, *Tetrahedron Lett.*, **26**, 319 (1985).
b) Y. Le Merrer, A. Duréault, C. Greck, D. Micas-Languin, C. Gravier, J.C. Depezay, *Heterocycles*, **25**, 541 (1987).
15. P.E. Peterson, R.J. Kamat, *J. Amer. Chem. Soc.*, **91**, 4521 (1969).
16. J.L. Luche, *J. Amer. Chem. Soc.*, **100**, 2226 (1978).
17. To prevent benzoate elimination use of phosphonium ylide without protection of secondary allylic alcohol should be a solution, but dienal **12** is obtained instead of primary bromide during transformations of **6** ($-\text{CH}_2\text{OH}-\text{CH}_2\text{Br}$ using CBr_4 -DIPHOS at room temperature)



18. tBDP-silyl group could also be introduced when diepoxyde **2** is opened by lithium acetylide, but removal of the acetonide group is easier in the presence of benzoate (see ref. 12).
19. J. Adams, J. Rokach, *Tetrahedron Lett.*, **25**, 35 (1984).
20. Drs J-L Boucher and M. Delaforge of our department are presently carrying out biological and analytical investigations. Results will be published elsewhere.
21. All new compounds exhibited satisfactory spectra and analytical data.

A : ^1H NMR (250MHz, CDCl_3) : 1.3(2d, 3H, J=7Hz), 1.45(m, 2H), 1.65(m, 2H), 2.1(m, 2H, H_6), 2.7(t, 2H, H_3 , $\text{J}_{2,3}=\text{J}_{3,4}=6.5\text{Hz}$), 5.15(m, 1H, H_9), 5.25(t, 1H, H_2), 5.45(m, 1H, H_4 , $\text{J}_{4,5}=11\text{Hz}$), 5.55(m, 1H, H_5), 8.2-7.3(2m, 10H), 9.6(s, 1H, H_1)— ^{13}C NMR(CDCl_3) : 198.0(C_1), 166.2(PhCO) ; 133.7, 133.5, 132.6, 129.8, 129.5, 128.4, 128.2, 122.5(Ph , C_4 , C_5) ; 78.2(C_2) ; 74.3(C_9) ; 35.7(C_3) ; 32.5, 27.3, 25.2(C_6 , C_7 , C_8) ; 20.1(C_{10}).

4 : ^1H NMR(250MHz, CDCl_3) : 1.3(2d, 3H, J=6Hz), 1.8-1.3(m, 4H), 2.1(m, 2H, H_{10}), 2.6(m, 2H, H_7), 5.15(m, 1H, H_{13}), 5.45, 5.55(2m, 2H, H_8 , H_9), 5.6(m, 1H, H_6 , $\text{J}_{5,6}=6\text{Hz}$), 6.15(dd, 1H, H_2 , $\text{J}_{1,2}=8\text{Hz}$, $\text{J}_{2,3}=15.5\text{Hz}$), 6.25(dd, 1H, H_5 , $\text{J}_{4,5}=15\text{Hz}$), 6.5(dd, 1H, $\text{J}_{3,4}=10.5\text{Hz}$), 7.05(dd, 1H, H_3), 8.1-7.4(2m, 10H), 9.55(d, 1H)— ^{13}C NMR : 193.3(C_1) ; 166.1, 165.5(PhCO) ; 150.3, 141.5, 133.2, 132.7, 129.6, 129.4, 128.4, 128.2, 123.3(Ph , C_2 , C_3 , C_4 , C_5 , C_8 , C_9) ; 73.3(C_6) ; 71.3(C_{13}) ; 35.4(C_7) ; 32.2, 27.2, 25.1(C_{10} , C_{11} , C_{12}) ; 20.0(C_{14}).

10 : ^1H NMR(250MHz, CDCl_3) : 1.05(s, 9H, tBu), 1.2(t, 3H, CO_2Et), 1.3(d, 3H, H_{20} , $\text{J}_{19,20}=6.5\text{Hz}$), 1.35(m, 2H, H_{17}), 1.55(m, 2H, H_{18}), 1.7(m, 4H, H_3 , H_4), 1.85(m, 2H, H_{16}), 2.2(m, 2H, H_2), 2.3(dd, 2H, H_{13}), 4.1(t, 2H, CO_2Et), 4.2(dt, 1H, H_{12}), 5.1(tq, 1H, H_{19}), 5.4-5.2(m, 2H, H_{14} , H_{15}), 5.4(dd, 1H, H_6), 5.66(dd, 1H, H_{11}), 5.9(m, 1H, H_5), 5.98(dd, 1H, H_{10}), 6.1(dd, 1H, H_7), 6.13(dd, 1H, H_9), 6.48(dd, 1H, H_8), 8.1-7.3(4m, 20H) ; $\text{J}_{5,6}=10$, $\text{J}_{6,7}=10.5$, $\text{J}_{7,8}=11$, $\text{J}_{8,9}=14.5$, $\text{J}_{9,10}=10.5$, $\text{J}_{10,11}=15$, $\text{J}_{11,12}=6.5$, $\text{J}_{14,15}=11$, $\text{J}_{18,19}=\text{J}_{19,20}=6.5\text{Hz}$ ^{13}C NMR : 165.8(PhCO) ; 137.6, 135.9, 135.2, 132.8, 131.9, 131.3, 130.0, 129.5, 128.3, 127.5, 126.8, 125.2(Ph , C_6 , C_7 , C_8 , C_9 , C_{10} , C_{11} , C_{14} , C_{15}) ; 73.8(C_{12}) ; 71.6(C_{19}) ; 70.7(C_5) ; 60.3(CO_2Et) ; 36.0, 35.6, 34.4, 34.0, 29.7, 25.3, 20.6(C_2 , C_3 , C_4 , C_{13} , C_{16} , C_{17} , C_{18}) ; 27.1, 19.4(tBu) ; 20.1(C_{20}) ; 14.3(CO_2Et)—SM(NH_3) : 844($\text{M}^+ + 18$)—UV(CH_3OH) : 282, 271, 262nm.

11 : ^1H NMR(250MHz, C_6D_6) : 4.35(q, 1H, H_{12}), 5.2(m, 1H, H_{19}), 5.45-5.3(m, 2H, H_{14} , H_{15}), 5.5(dd, 1H, H_6), 5.65(m, 1H, H_5), 5.7(dd, 1H, H_{11}), 5.95(m, 1H, H_8), 6.0(m, 1H, H_9), 6.1(dd, 1H, H_{10}), 6.3(m, 1H, H_7) ; $\text{J}_{5,6}=7$, $\text{J}_{6,7}=15$, $\text{J}_{7,8}=\text{J}_{9,10}=9$, $\text{J}_{10,11}=14.5$, $\text{J}_{11,12}=6.5$, $\text{J}_{14,15}=11\text{Hz}$ — ^{13}C NMR(CDCl_3) : 173.0(C_1) ; 165.5(PhCO) ; 136.9, 135.9, 133.0, 132.6, 131.2, 130.5, 129.6, 128.2, 127.7, 127.4, 125.1(Ph , C_6 , C_7 , C_8 , C_9 , C_{10} , C_{11} , C_{14} , C_{15}) ; 74.5(C_5) ; 73.6(C_{12}) ; 71.6(C_{19}) ; 60.5(CO_2Et) ; 36.0, 35.6, 34.0, 29.7, 25.3, 20.6(C_2 , C_3 , C_4 , C_{13} , C_{16} , C_{17} , C_{18}) ; 27.1, 19.5(tBu) ; 20.0(C_{20}) ; 14.2(CO_2Et)—SM(NH_3) : 844($\text{M}^+ + 18$)—UV(CH_3OH) : 282, 271, 262nm.

1 (potassium salt) : ^1H NMR(250MHz, D_2O) : 1.15(d, 3H, H_{20}), 1.5-1.35(m, 4H, H_{17} , H_{18}), 1.7, 1.5(m, 4H, H_3 , H_4), 2.05(m, 2H, H_{16}), 2.2(t, 2H, H_2), 2.35(m, 2H, H_{13}), 3.8(m, 1H, H_{19}), 4.25(m, 1H, H_2), 4.65(m, 1H, H_5), 5.45(m, 2H, H_{14} , H_{15}), 5.6(m, 1H, H_6), 5.83(dd, 1H, H_{11}), 6.2(dd, 1H, H_7), 6.45-6.25(2dd, 2H, H_9 , H_{10}), 6.65(dd, 1H, H_8) ; $\text{J}_{2,3}=7.5$, $\text{J}_{6,7}=11$, $\text{J}_{7,8}=11.5$, $\text{J}_{8,9}=15$, $\text{J}_{10,11}=14.5$, $\text{J}_{11,12}=7$, $\text{J}_{19,20}=6.5\text{Hz}$ —UV(CH_3OH) : 281, 270.5, 260nm.