uncorrected but were taken in a Fisher melting point apparatus with a set of Anschütz thermometers which gave correct melting points with various pure reagents.

DEPARTMENT OF CHEMISTRY HOLY CROSS COLLEGE WORCESTER, MASSACHUSETTS **Received** June 1, 1936

## COMMUNICATIONS TO THE EDITOR

## SYNTHETIC SUBSTRATES FOR PROTEIN-DIGEST-ING ENZYMES

## Sir:

Knowledge regarding the specificity of those enzymes which hydrolyze intact proteins (peptic, tryptic and catheptic proteinases) is meager. In general it is assumed that these enzymes react exclusively on high molecular substrates.

Recently it has been possible to study the specificity of proteinases with the aid of synthetic substrates. Such substrates have been found in this Laboratory for the catheptic enzymes, papain, liver-cathepsin and bromelin. The authors have now observed the splitting of  $\alpha$ -hippuryl-lysine-amide by tryptic proteinase.

 $\alpha$ -Hippuryl- $\epsilon$ -carbobenzoxy-lysine methyl ester was converted into  $\alpha$ -hippuryl- $\epsilon$ -carbobenzoxylysine amide, m. p. 212°, with the aid of methanolic ammonia. *Anal.* Calcd. for C<sub>23</sub>H<sub>28</sub>N<sub>4</sub>O<sub>5</sub>: C, 62.7; H, 6.4; N, 12.7. Found: C, 62.6; H, 6.7; N, 12.8. This amide was hydrogenated catalytically, yielding  $\alpha$ -hippuryl-lysine-amide which was isolated as the strongly hygroscopic hydrochloride, m. p. 248°. *Anal.* Calcd. for C<sub>15</sub>H<sub>23</sub>N<sub>4</sub>O<sub>3</sub>Cl: C, 52.5; H, 6.8; N, 16.3. Found: C, 52.0; H, 7.0; N, 15.9.

The tryptic proteinase was prepared according to E. Waldschmidt-Leitz and A. Purr [*Ber.*, 62, 2217 (1929)]. The preparation contained no dipeptidase, aminopeptidase, and no carboxypeptidase; however, protaminase probably was present (Table I).

In contrast to HCN-papain, which splits only one peptide bond, tryptic proteinase splits two. After a complete splitting, hippuric acid was isolated from the digest (over 70% of the theoretical amount). Therefore, the splitting also must have liberated lysine and ammonia. That the free  $\epsilon$ amino group is an essential condition for the enzymic hydrolysis is shown by the fact that the

ENZYMIC HYDROLYSIS OF $\alpha$ -H AT 40°	IIPPURYL-LY	SINE-AMIDE
(Titration of liberated	carboxyl	groups)
Enzyme	Time, hrs.	Hydrolysis in % of one peptide bond
Tryptic proteinase, pH 8.8	22	12 <b>3</b>
	72	200
Tryptic proteinase, pH 8.8	18	121
_	42	175
HCN-Papain, pH 5.0	5	58

TABLE I

above mentioned  $\alpha$ -hippuryl- $\epsilon$ -carbobenzoxy-lysine amide is not hydrolyzed under the conditions of our experiments. The hydrolysis of our substrate by tryptic proteinase is remarkable since tryptic proteinase is supposed to react exclusively on anionic substrates.

 $\mathbf{24}$ 

49

80

85

It is intended to continue this research by studying the action of pure tryptic proteinases.

The Laboratories of The Rockefeller Institute	Max Bergmann William F. Ro <b>ss</b>
FOR MEDICAL RESEARCH	
New York, N. Y.	
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RECEIVED JULY 10, 1936

## STEROLS. VI. SYNTHETIC PREPARATION OF OESTRONE (THEELIN)

Sir:

The evidence for the accepted structure of oestrone has recently been reviewed [L. F. Fieser, "Chemistry of Natural Products Related to Phenanthrene," A. C. S. Monograph Series, No. 70]. We have been able to prepare a well crystallized compound from ergosterol which by analysis, derivatives and mixed melting points, is identical with oestrone isolated from pregnancy urine. It has been previously shown that ergosterol may be converted into 3-hydroxy-nor-allo-cholanic acid [Chuang, *Ann.*, **500**, 270 (1933); Fernholz and Chakravarty, Ber., 67, 2021 (1934)] which is also obtainable from dihydro-cholesterol. This acid can give rise to isoandrosterone which differs from androsterone only in configuration of the hydroxyl group in the 3-position. Therefore, with the preparation of oestrone from ergosterol, a complete connection between the hormones, male and female, has been established.



In the present work, dehydro-neo-ergosterol, III, was prepared from ergosterol by the method of Windaus [Windaus and Borgeaud, Ann., 520, 235, 460 (1928)], Inhoffen [Inhoffen, ibid., 497, 130 (1932)], and Honigmann [Honigmann, ibid., 511, 292 (1934)]. Taking advantage of the fact that naphthalene derivatives may be reduced to tetrahydro derivatives with sodium and amyl dehydro-neo-ergosterol, III, m. p. alcohol, 147-150°, was reduced to give tetrahydrodehydro-neo-ergosterol, IV, m. p. 170.5-171.5°. This substance is phenolic and forms sodium salts.

Anal. Calcd. for C<sub>27</sub>H<sub>41</sub>O: C, 85.1; H, 10.8. Found: C, 85.7; H, 10.8.

The tetrahydro-dehydro-neo-ergosterol was then acetylated and this product oxidized with chromic acid. The total neutral oxidation product was hydrolyzed with alcoholic sodium hydroxide and then treated with semicarbazide acetate. The crude semicarbazone was hydrolyzed with alcoholic sulfuric acid to the free ketone, which was then distilled at 200° under high vacuum. The sublimate was crystallized from

95% alcohol to give a white crystalline compound m. p. 259-261.5° (uncorr.). This gave no depression in melting point with a sample of natural oestrone of m. p. 255°. It is soluble in alkali. It gave a rotation of  $[\alpha]^{32}D + 159^{\circ}$  in alcohol, c = 514 mg. per 100 cc.

Anal. Calcd. for  $C_{18}H_{22}O_2$ : C, 79.9; H, 8.3. Found: C, 79.4; H, 8.4.

C<sub>9</sub>H<sub>18</sub>

The product gave a benzoate by the Schotten-Baumann reaction, m. p. 205-207° (uncorr.), which gave no depression to a sample of natural oestrone benzoate m. p. 205° (uncorr.).

Anal. Calcd. for C25H26O3: C, 80.2; H, 7.0. Found: C, 80.6; H, 7.2.

It gave a semicarbazone of m. p. 252-253° (uncorr.). From the analysis this semicarbazone calculates to have one-half molecule of water of crystallization. Butenandt [Z. physiol. Chem., 208, 129 (1932); **208**, 149 (1932)] observed the same thing on the preparation of the semi-

carbazone from natural oestrone.

Anal. Calcd. for  $(C_{19}H_{25}O_2N_3)_2 \cdot H_2O$ : C, 67.9; H, 7.9. Found: C, 68.1; H, 8.2.

This synthesis was repeated independently by two of us. We wish to thank Dr. George H. Fleming of this Laboratory for the micro-analyses reported in this paper.

School of Chemistry and Physics The Pennsylvania State College State College, Pa. The Parke, Davis & Co. Research Laboratories Detroit, Mich.			Russell E. Marker Oliver Kamm Thomas S. Oakwood Joseph F. Laucius
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RECEIVED JULY 20, 1936

SYNTHESIS OF VITAMIN B1

Sir:

As foreshadowed in a recent communication [THIS JOURNAL, 58, 1063 (1936)] we have effected a synthesis of the vitamin by the following route.

$$\begin{array}{c} \text{EtoCOCH}_{2}\text{CH}_{2}\text{OEt} \xrightarrow{+\text{HCOOEt}} \\ \xrightarrow{+\text{NH}_{2}-\text{C(CH}_{2})=\text{NH}} \\ \text{EtoCOCHCH}_{2}\text{OEt} \xrightarrow{-\text{C(CH}_{2})=\text{NH}} \\ \xrightarrow{\text{O=CH}} \end{array}$$