Preliminary communication

Synthesis of 1-deoxynojirimycin-containing glycans related to the Lewis X and sialyl-Lewis X epitopes recognized by LEC-CAMs *

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LEC-CAMs (selectins)^{2,3}, a family of cell adhesion receptors, mediate leukocyte trafficking and recruitment to sites of inflamation on vascular endothelium. These receptor proteins contain an *N*-terminal carbohydrate recognition domain homologous to various calcium-dependent mammalian lectins. Very recently, the sialyl-Lewis X epitope, found on neutrophils, monocytes, and tumor cells, has been identified⁴⁻⁸ as a minimal carbohydrate ligand recognized not only by endothelial–leukocyte adhesion molecule 1 (ELAM-1, E-selectin) but also by other LEC-CAMs, such as platelet activation-dependent granule external membrane protein⁹ (PADGEM, P-selectin) and LECAM-1¹⁰ (MEL-14, L-selectin). In the course of synthetic studies on sialoglycoconjugates, we have recently succeeded¹¹ in the first total synthesis of sialyl-Lewis X ganglioside and its analogs. Some of the cell adhesion studies⁷⁻¹⁰ of LEC-CAMs just described have been accomplished by employing those chemically synthesized gangliosides.

1-Deoxynojirimycin (DNJ) and related compounds have been shown¹²⁻¹⁷ not only to be potent inhibitors of α -glucosidases and glycoprotein-processing enzymes, but also to be of potential clinical value as antidiabetic, antineoplastic, and anti-HIV agents. We have synthesized a variety of DNJ derivatives^{18,19}, DNJ-containing disaccharides²⁰, and sialyl oligosaccharides²¹ designed as novel biofunctional glycans. The present paper describes the synthesis of DNJ-containing Lewis X and sialyl-Lewis X epitopes.

4,6-O-Benzylidene-N-benzyloxycarbonyl-1,5-dideoxy-1,5-imino-D-glucitol²⁰ (1) was treated with chloroacetyl chloride (1.2 mol equiv) at -20° in CH₂Cl₂ containing 2,6-lutidine (2 mol equiv), to give a 70% yield of the 3-O-chloroacetyl derivative

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2 having $[\alpha]_D - 5.2^\circ$ (CH₂Cl₂) and showing in its ¹H-NMR spectrum a triplet at δ 5.04 ($J_{2,3} = J_{3,4} = 8.4$ Hz, H-3). Following 2-O-acetylation of 2, dechloroacetylation with aq pyridine gave the glycosyl acceptor 3, $[\alpha]_D - 12.5^\circ$ (CH₂Cl₂), quantitatively. In the ¹H-NMR spectrum of 3 H-2 appeared at $\delta \sim 4.83$ as a one-proton multiplet partially overlapping H-6*eq*. Other significant signals were at δ 2.04 (s, CH₃CO), 3.17 (dd, J_{gem} 13.9, $J_{1ax,2}$ 8.2 Hz, H-1*ax*), 4.11 (dd, $J_{1eq,2}$ 4.4 Hz, H-1*eq*), 3.79 (dd, $J_{2,3}$ 6.6, $J_{3,4}$ 9 Hz, H-3), 3.70 (t, $J_{4,5}$ 9 Hz, H-4), and 3.37 (ddd, $J_{5,6ax}$ 10.8, $J_{5,6eq}$ 4.2 Hz, H-5). Glycosylation of 3 with methyl 2,3,4-tri-O-benzyl-1-thio- β -L-fucopyranoside (4) in the presence of dimethyl(methylthio)sulfonium triflate^{22,23} (DMTST) and molecular sieves 4A in benzene for 2.5 h, with warming from 7° to room temperature, gave the desired disaccharide 5, $[\alpha]_D - 94^\circ$ (CH₂Cl₂), in 92%

yield. The ¹H-NMR spectrum of 5 showed a three-proton doublet at δ 0.83 ($J_{5,6}$ 6.6 Hz, C H_3 CH) and a one-proton doublet at δ 5.21 ($J_{1,2}$ 4 Hz, H-1), characteristic of the α -L-fucopyranosyl unit. Reductive ring-opening²⁴ of the benzylidene group in 5 with sodium cyanoborohydride-hydrogen chloride in dry ether afforded the disaccharide glycosyl acceptor 6, $[\alpha]_D$ -51° (CH₂Cl₂), in 81% yield. Significant signals in the ¹H-NMR spectrum of 6 were at δ 3.38 (dd, J_{gem} 15.4, $J_{1ax,2}$ 2.9 Hz, H-1*ax* of DNJ), 4.12 (~d, J_{gem} 15.4, $J_{1eq,2} < 2$ Hz, H-1*eq* of DNJ), 4.91 (d, $J_{1,2}$ 3.7 Hz, H-1 of Fuc), and 4.35-5.21 (10 d, CH₂Ph).

The glycosylation of 6 with methyl 2,3,4,6-tetra-O-benzoyl-1-thio- β -D-galactopyranoside (7) was performed in the presence of N-iodosuccinimide²⁵⁻²⁷ (NIS, 4 mol equiv) and trifluoromethanesulfonic acid (TfOH, 0.4 mol equiv) in CH₂Cl₂, to give the desired trisaccharide 8, $[\alpha]_D - 10.4^\circ$ (CH₂Cl₂), in 70% yield. Hydrogenolytic removal of the benzyloxycarbonyl and benzyl groups over palladium black in MeOH-formic acid, and subsequent O-deacylation with NaOMe in MeOH, gave the 1-deoxynojirimycin-containing Lewis X epitope 9, $[\alpha]_D - 24^\circ$ (2:1 H₂O-EtOH), in quantitative yield. Significant signals in the ¹H-NMR spectrum of 9 (in D₂O-CD₃OD) were at δ 1.14 (d, J_{5,6} 6.8 Hz, CH₃CH of Fuc), 2.97 (dd, J_{gem} 12, J_{1ax,2} 4.4 Hz, H-1ax of DNJ), 4.51 (d, J_{1,2} 7.7 Hz, H-1 of Gal), and 5.37 (d, J_{1,2} 3.9 Hz, H-1 of Fuc), indicating the desired trisaccharide structure.

Coupling of 6 with methyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$ -2,4,6-tri-O-benzoyl-1thio- β -D-galactopyranoside¹¹ (10) was carried out by using NIS (3 mol equiv) and a catalytic amount of TfOH (0.3 mol equiv) as described for 8, to afford, 11 [α]_D -11° (CH₂Cl₂), in 61% yield. In the ¹H-NMR spectrum of 11, a one-proton doublet at δ 4.64 ($J_{1,2}$ 8 Hz, H-1 of Gal) showed the newly formed glycosidic linkage to be β . Other significant signals were at δ 3.81 (s, CO₂CH₃) and 2.45 (dd, J_{gem} 12.8, $J_{3ea.4}$ 4.6 Hz, H-3eq of sialic acid).

The benzyloxycarbonyl and benzyl groups in 11 were simultaneously removed by catalytic hydrogenolysis over palladium black in 1:1 MeOH-acetic acid to give a quantitative yield of 12 $[\alpha]_D - 20^\circ$ (MeOH), which was converted, by treatment with NaOMe in MeOH and subsequent saponification of its methyl ester group, into the 1-deoxynojirimycin-containing sialyl-Lewis X epitope 13 $[\alpha]_D - 14^\circ$ (3:1 H₂O-EtOH), obtained in quantitative yield after chromatography on a column of Sephadex LH-20. In the ¹H-NMR spectrum of 13 the two anomeric protons of residues bound to the DNJ moiety appeared at δ 4.62 ($J_{1,2}$ 7.9 Hz, H-1 of Gal) and 5.40 ($J_{1,2}$ 4 Hz, H-1 of Fuc), respectively, showing the desired β - and α -glycosidic linkages. Very recently, a stepwise synthesis of the sialyl Lewis X epitope itself was reported²⁸.

In conclusion, by employing methyl thioglycosides 4, 7, and 10 as glycosyl donors, and suitably protected mono- and di-saccharide derivatives of 1-deoxynojirimycin (3 and 6) as glycosyl acceptors, we achieved syntheses of novel 1-deoxynojirimycin-containing glycans structurally related to the Lewis X and sialyl-Lewis X epitopes recognized by LEC-CAMs.

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