

Facile Synthesis of (+)-Brefeldin A Utilizing Two Optically Active Synthons Prepared by Different Enzyme-catalysed Reactions

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The lactone **4a** and the alcohol **6** (both available in optically active form from biocatalytic processes) have been used as synthons in the preparation of (+)-brefeldin A.

Brefeldin A **1** was first isolated in 1958 from *Penicillium decumbens* and was subsequently found as a secondary metabolite in other cultures.¹ The structure and stereochemistry was confirmed by X-ray crystallography in 1971.² Several partial, formal and total syntheses of brefeldin A have been reported^{3,4} while biological testing has shown that the compound exhibits a wide range of biological activities including antibiotic, antiviral, cytostatic and antimitotic effects.⁵

We envisaged that this natural product could be synthesised, in single-enantiomer form, from the *exo*-hydroxylactone **4a**, a compound which we have been able to obtain in an optically pure state by a simple biotransformation.⁶ Fig. 1 shows our retrosynthetic approach. Disconnection of the lactone group and the vinyl side chain of brefeldin A gives a cyclopentenone **2**, bearing a four carbon side chain, that could ostensibly be prepared from the *exo*-hydroxylactone **4a**. The partner in the coupling reaction *i.e.* organometallic reagent **3** is derived from the (*S*)-hept-6-yn-2-ol **6**.

Addition of glyoxylic acid to cyclopentadiene in water produces a mixture of *exo*-**4a** and *endo*-hydroxy bicyclic lactone **5** in a ratio of 1:4.⁶ We have previously reported the enzymatic resolution of both these hydroxylactones by enzyme action at their hydroxyl functions using *Pseudomonas fluorescens* or *Candida cylindracea* lipase, and showed that the *endo*-hydroxylactone **5** and its enantiomer are convenient synthons for the preparation of intermediates for hypocholesteremic agents and the anti-HIV agent (–)-carbovir respectively.⁶

For the synthesis of brefeldin A the minor isomer from the above preparation (*i.e.* the *exo*-hydroxylactone **4a**) is the

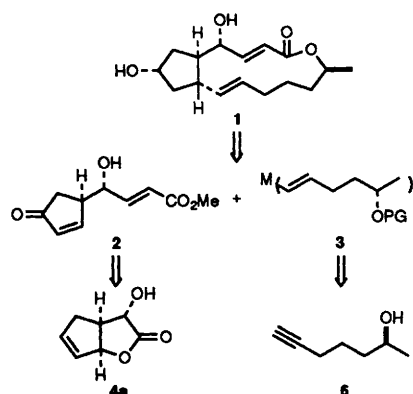


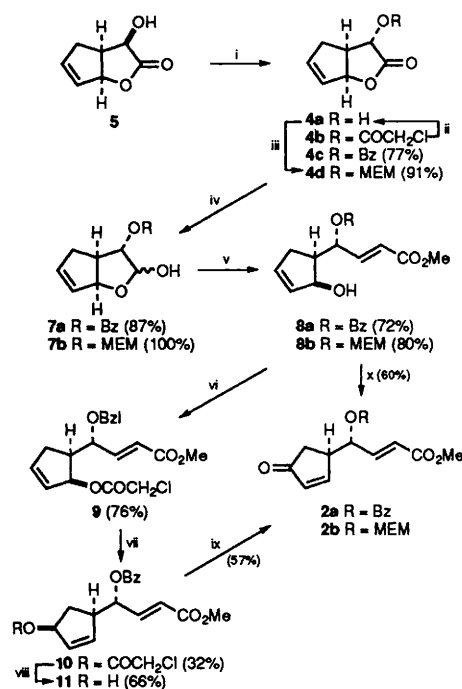
Fig. 1

Table 1 Enzymatic acylation of **6** (using vinyl acetate as solvent and acylating agent)

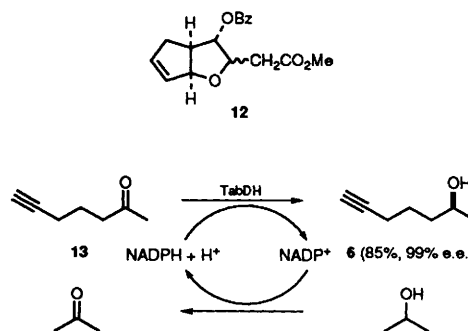
Lipase	t/h	Conversion (%, ±5)	% e.e. Acetate	% e.e. 6
PS	72	50	83	80
P30	41	54	90	90
AK	25	50	93	83
AY30	50	49	0	0

requisite starting material. In order to have suitable amounts of this synthon, the *endo*-isomer **5** was epimerized in two steps (Scheme 1). Thus Mitsunobu inversion of the *endo*-isomer **5** (available as a single enantiomer in kilogram quantities) using chloroacetic acid as the nucleophile provided the ester **4b**, which was treated with thiourea and sodium bicarbonate in refluxing ethanol⁷ (in order to chemoselectively hydrolyse the chloroacetyl group) to yield the *exo*-hydroxylactone **4a** in 86% overall yield.

The protected hydroxylactones **4c** and **4d** were prepared by



Scheme 1 Reagents and conditions: i, Ph_3P , DEAD, THF, benzoic acid (for **4c**) or chloroacetic acid (for **4b**); ii, H_2NCSNH_2 , NaHCO_3 , EtOH, heat [86% from **5**]; iii, MEM-Cl, DIPEA, CH_2Cl_2 ; iv, Diisobutylaluminium hydride (Dibal-H), THF, -78°C ; v, methyl (triphenyl phosphoranyliden) acetate, toluene; vi, chloroacetic acid anhydride, pyridine; vii, $\text{PdCl}_2[\text{MeCN}]_2$, 1,4-benzoquinone; viii, thiourea, NaHCO_3 , EtOH; ix, PCC, CH_2Cl_2 ; x, PCC, *p*-TsOH, CH_2Cl_2 (60%)



Scheme 2

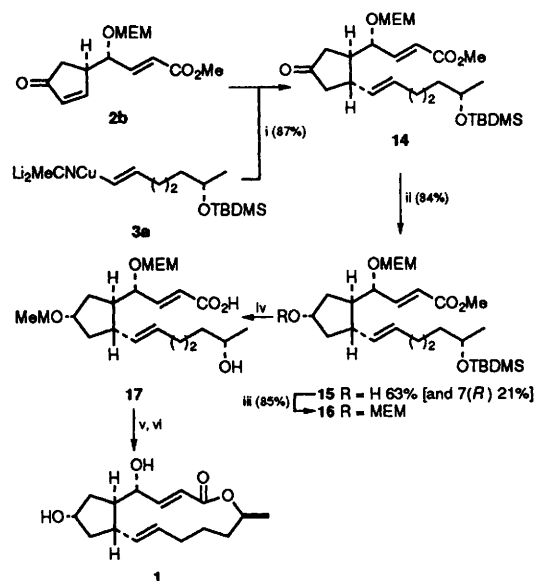
standard methods, and reduced to the corresponding lactols **7a** and **b** using diisobutylaluminium hydride⁸ (Scheme 1). Addition of the Wittig reagent $\text{Ph}_3\text{PCHCO}_2\text{Me}$ to these lactols gave the alcohols **8a** and **b**. As expected, on prolonged reaction an intramolecular conjugate addition of the newly formed hydroxy group to the unsaturated ester took place, forming a bicyclic ether (e.g. **12**).

The cyclopentenol **8a** was acylated to provide the allylic chloroacetate **9**. A (3,3)-sigmatropic rearrangement to a less hindered alcohol was promoted by catalytic amounts of Pd^{II} .⁹ Selective hydrolysis of the chloroacetate with thiourea⁷ and oxidation with PCC gave the desired intermediate **2a**. Notably, the rearrangement and oxidation of cyclopentenols **8a** and **b** to the corresponding cyclopentanones **2a** and **b** was also achieved in one-pot using Baekstrom's conditions¹⁰ in 60% yield.

The lower side chain of brefeldin A was prepared by a second enzyme catalysed process. Thus hept-6-yn-2-one **13** was enantioselectively reduced using the alcohol dehydrogenase from *Thermoanaerobium Brockii* (TabDH) in high optical purity (99% e.e.) and excellent yield in a process that is much cleaner and efficient than the prescribed biotransformation using bakers' yeast (Scheme 2).¹¹ The resolution of (\pm)-hept-6-yn-2-ol using lipases was also not as effective as the dehydrogenase-catalysed reaction for the production of optically active material (Table 1). The alcohol **6** was protected as the *tert*-butyldimethylsilyl ether before being converted into the cuprate reagent **3a**, which was contaminated with 15–20% of the isomeric (*Z*)-alkene.

Conjugate addition¹² of cuprate **3a** to the cyclopentenone **2a** gave complex mixtures. However addition of **3a** to the compound **2b** occurred smoothly at the more reactive cyclopentenone unit from the unhindered α face (Scheme 3).

The disubstituted cyclopentanone **14** so formed was reduced with some selectivity using K. Selectride, and the major product **15** was purified by chromatography before being further protected with a second MEM group. This di-protected diol **16** was identical to Taber's intermediate,⁴ e.g.



Scheme 3 Reagents and conditions: i, Compound **3a**, THF, -78°C ; ii, K. Selectride, THF, -78°C ; iii, MEM-Cl, DIPEA, CH_2Cl_2 ; iv, HCl (1 mol dm^{-3}) then LiOH; v, 2,4,6-trichlorobenzoyl chloride, THF then DMAP, toluene, heat (80%); vi, TiCl_4 , CH_2Cl_2 , 0°C (80%)

$[\alpha]_D = -34^\circ$ (c 1.0, CHCl_3), lit.⁴ $[\alpha]_D = -27.7^\circ$ (c 1.44, CHCl_3), ^{13}C NMR: $\delta(\text{CDCl}_3)$ -4.71 , -4.42 , 18.08 , 23.73 , 25.58 , 25.87 , 32.43 , 32.89 , 39.21 , 40.21 , 42.98 , 48.30 , 51.40 , 58.88 , 58.93 , 66.83 , 67.52 , 68.44 , 71.71 , 71.81 , 75.73 , 76.81 , 94.19 , 94.34 , 121.29 , 131.05 , 133.30 , 148.14 , 166.50 .

Removal of the silyl protecting group and hydrolysis of the methyl ester gave the corresponding hydroxy acid **17**. $[\alpha]_D = -9.33^\circ$ (c 0.4, CHCl_3), ^1H NMR: $\delta(\text{CDCl}_3)$ 6.86 (dd, J 15.8, 6.3 Hz, 1H), 5.95 (dd, J 15.8, 1.2 Hz, 1H), 5.33 (m, 2H), 4.68 (m, 4H), 4.15 (m, 2H), 3.78 (m, 2H), 3.66 (m, 4H), 3.55 (m, 5H), 3.38 (s, 3H), 3.37 (s, 3H), 2.31 (m, 1H), 2.15 (m, 1H), 2.0 – 1.2 (m, 10H), 1.18 (d, J 6 Hz, 3H). Lactone formation and removal of the MEM protecting groups yields brefeldin A **1**.⁴ This new synthesis of (+)-brefeldin A is comprised of only 11 steps starting from the readily available bicyclic lactone **4a**.

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