

# SELECTIVE HYDROGENOLYSIS OF THE BENZYLOXYCARBONYL PROTECTING GROUP OF N<sup>ε</sup>-LYSINE IN CYCLOPEPTIDES CONTAINING A BENZYLIC PHENYL ETHER FUNCTION. EVIDENCE FOR N<sup>ε</sup>-METHYLATED LYSINE SIDE PRODUCTS.

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**Abstract:** Selective hydrogenolytic cleavage of the N<sup>ε</sup>-Z protecting group of lysine in cyclopeptides c[-(Glycyl)<sub>n</sub>-A-B-N<sup>ε</sup>-Z-Lysyl-2-Phenoxymethyl-5-Aminobenzoate-] (A = B = Glycyl, n = 2 or A = Phenylalanyl, B = Alanine, n = 2 and 3) occurred in both acidic (MeOH/aq.AcOH) and neutral (MeOH/DMF) solvents, with Pd/C catalyst. In the latter case, a N<sup>ε</sup>-(bis)-methylated lysine side product was isolated.

The recent paper of Rocchi *et al*<sup>1</sup> concerning N-alkylation of aminoacids during hydrogenolytic deprotection prompts us to publish our results concerning hydrogenolytic cleavage of the Z protecting group of N<sup>ε</sup>-lysine in cyclopeptides of type 1 (Fig. 1) incorporating a phenoxymethyl-substituted aminobenzoic residue. In the course of elaboration of new substrates/inhibitors of trypsin-like proteases,<sup>2,3</sup> we examined the hydrogenolytic deprotection conditions of the cyclopeptides 1a-c and others, with the purpose of obtaining on one hand compounds of type 2 resulting from the selective deprotection of the N<sup>ε</sup>-Lys Z group, and on the other hand compounds of type 3 resulting from hydrogenolysis of both the Z group and the benzylic phenoxy group

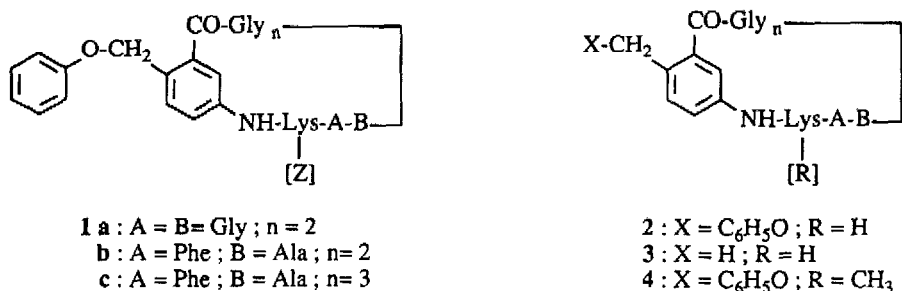


Figure 1

Hydrogenations were conducted in a Parr apparatus, at pressures of 2-3 atmospheres and at room temperature, with always a large excess of palladium on charcoal as catalyst. Using the solvent mixture DMF/MeOH for solubility reasons, long reaction times had to be applied for reaction completion. A selective cleavage of the benzyloxycarbonyl (Z) protecting group of N<sup>ε</sup>-lysine was always observed: compounds of type 2 highly predominated over compounds of type 3. However, analytical t.l.c. (SiO<sub>2</sub>; solvent system EtOAc/n-BuOH/AcOH/H<sub>2</sub>O 1:1:1:1) presented spots (UV light), sometimes of quite high intensity, corresponding to side products of structure 4. Typically, catalytic hydrogenation of 1b<sup>4</sup> in DMF/MeOH 1:9 for

16 h gave **2b** (51 %) <sup>4</sup> with traces amounts of **3b** and **4b** (not isolated). From **1c**, <sup>4</sup> in DMF/MeOH 1:1 for 22 h, the cyclopeptides **2c**, **3c** and **4c** were isolated with 21 %, 7 % and 23 % yield, <sup>4</sup> respectively.

The structure of **4c** was evidenced by <sup>1</sup>H NMR analysis and FAB mass spectroscopy. In <sup>1</sup>H NMR (CD<sub>3</sub>OD) (Fig.2), the main differences between **2c** and **4c** was for the latter a 0.1 ppm downfield shifting of the CH<sub>2</sub><sup>ε</sup> multiplet and the presence of a singlet at 2.82 ppm (integrating for 6 protons), corresponding to N<sup>ε</sup>-[CH<sub>3</sub>]<sub>2</sub> (Lys). The signals for the aromatic protons of the phenoxy group and the AB quartet corresponding to C<sub>6</sub>H<sub>5</sub>O-CH<sub>2</sub>-Ar ( $\delta$  = 5.19 ppm ; J = 12.1 Hz) were present in **4c**, showing that N-(bis)-methylation had occurred faster than reductive cleavage of the benzyl ether.

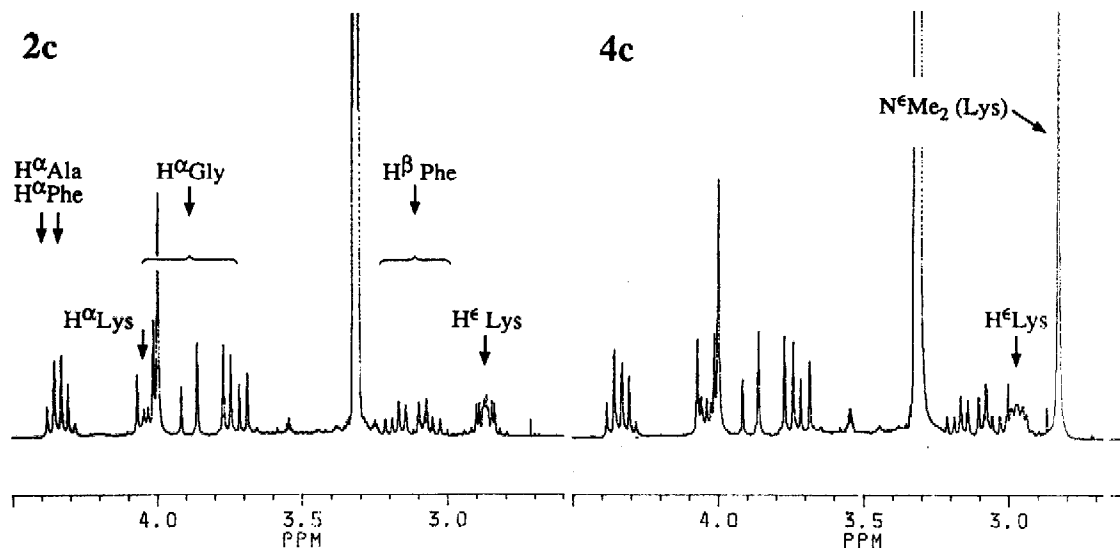


Figure 2

Cleaner reactions and still selective cleavage of the Z group were observed when hydrogenolysis was performed in the presence of water and acetic acid. The cyclopeptide **1a** <sup>2,4</sup> in MeOH/H<sub>2</sub>O/AcOH 7:2:1 gave **2a** (70 %) <sup>4</sup> and traces of **3a** (not isolated) for 0.5 h reaction time, and **3a** (65 %) <sup>4</sup> after 24 h.

Thus, a selective cleavage of the N<sup>ε</sup>-Z protecting group of lysine over a benzyl phenyl ether substituent in cyclopeptides **1** can be performed in both neutral (DMF/MeOH) and acidic solvent systems. In the former case however, longer reaction time results in the formation of N<sup>ε</sup>-methylated lysine side products, resulting from reaction of N<sup>ε</sup>H<sub>2</sub> or N<sup>ε</sup>HCH<sub>3</sub> with formaldehyde, followed by *in situ* reduction.<sup>1</sup>

## References and Notes

1. Filira, F.; Biondi, L.; Gobbo, M. and Rocchi, R., *Tetrahedron Letters*, **1991**, *32*, 7463-7464.
2. Wakselman, M.; Mazaleyrat, J.P.; Xie, J.; Boggetto, N.; Montagne, J.J.; Vilain A.C. and Reboud-Ravaux, M., *Peptides 1990*, Giralt, E. and Andreu, D., Eds., ESCOM Science Pub., Leiden, 1991; pp 794-796.
3. Wakselman, M.; Mazaleyrat J.P.; Xie, J.; Montagne, J.J.; Vilain A.C. and Reboud-Ravaux, M., *Eur. J. Med. Chem.*, **1991**, *26*, 699-707.
4. All cyclopeptides **1-4** gave satisfactory analytical data (<sup>1</sup>H NMR at 300 MHz, C,H,N analysis and/or FAB spectroscopy). Details of synthesis will be published elsewhere. The given yields in **2-4,a-c** correspond to isolated pure compounds.

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