

A One-Pot Conversion of α -Aminoalkylphosphonous Acids into Monomethyl α -Aminoalkylphosphonates

Katja Štrancar, Stanislav Gobec*

University of Ljubljana, Faculty of Pharmacy, Aškerčeva 7, 1000 Ljubljana, Slovenia
Fax +386(1)4258031; E-mail: gobecs@ffa.uni-lj.si

Received 4 November 2003; revised 8 December 2003

Abstract: An excellent method for a one-pot conversion of N-protected α -aminoalkylphosphonous acids into the corresponding monomethyl phosphonates, using thionyl chloride in methanol, has been devised. N-Protected monomethyl α -aminoalkylphosphonates are valuable intermediates in the preparation of phosphapeptides.

Key words: α -aminoalkylphosphonous acids, α -aminoalkylphosphonate monomethyl esters, thionyl chloride, oxidation, selective esterification

Replacement of peptide bonds by phosphonamide, phosphonate or phosphinate moieties results in peptide analogues (phosphapeptides) which closely resemble the high-energy tetrahedral transition state of enzyme-catalyzed amide bond hydrolysis or formation.¹ These stable transition-state analogues form an important class of enzyme, particularly protease, inhibitors.² Complex phosphapeptides play an important role in the development of catalytic antibodies with protease-like activity and specificity.³

The three phosphorus peptide bond replacements have both similarities and differences in the ways in which they are assembled. The synthesis of phosphonates and phosphonamides commonly begins with N-protected α -aminoalkylphosphonate monoesters. These are usually first converted into the phosphonochlorides, using chlorinating reagents like thionyl or oxalyl chloride,⁴ and then coupled with the appropriate alcohol or amine component, respectively. For the preparation of phosphonate peptide analogues, N-protected α -aminoalkylphosphonate monoesters can be also activated by BOP-Cl,⁵ BOP,⁶ PyBOP⁶ and TPP/DIAD in a modified Mitsunobu reaction⁷, and then coupled to the appropriate alcohol. Syntheses of phosphonamides from the same precursors (N-protected α -amino phosphonate monoesters), activated by DPPA,⁸ DCC⁹ and BOP,¹⁰ have also been reported, but data on the usefulness of these methods are contradictory. An alternative synthesis of the phosphonates and phosphonamides involves the in situ conversion of N-protected H-phosphinate esters into phosphonochlorides, using carbon tetrachloride and triethylamine.¹¹

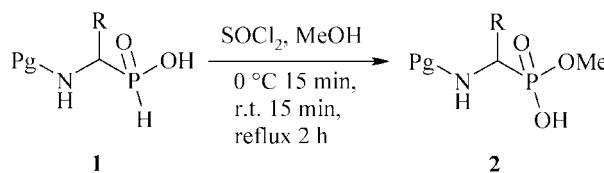
Phosphonic acid monoesters are thus very important synthons for preparing phosphapeptides. They can be pre-

pared by one of three methods: via direct monoesterification of phosphonic acids with alcohols in the presence of condensing reagents (such as DCC,¹² DCC/TEA,¹³ CCl₃CN,¹⁴ SOCl₂/DMF,¹⁵ BroP¹⁶ or TpyClU¹⁶), via hydrolytic (LiOH, NaOH)¹⁷ or nonhydrolytic (NaN₃,¹⁸ LiN₃,¹⁸ Me₃SiBr,^{7a,17b,19} t-BuNH₂,²⁰ DABCO,²¹ NaI,²² PhSH²³) selective ester cleavage of symmetrical or unsymmetrical phosphonate diesters, or via hydrolysis of the corresponding phosphonochlorides,^{24a} prepared by the direct action of PCl₅^{24a} or (COCl)₂^{24b} on a phosphonate diester, or by treatment of a phosphonyl dichloride with one equivalent of the appropriate alcohol.²⁵ In addition, monomethyl esters of alkylphosphonic acids can be successfully converted into other monoesters, since methyl esters are easily removed.^{7a,17b}

In 1986 Karanewsky described an alternative synthesis of phosphonic monoesters from phosphonous acid.²⁶ It is a two step procedure in which phosphonate monoesters are prepared by DCC/DMAP-mediated esterification of phosphonous acid, followed by oxidation of the resulting phosphinate ester in a second reaction step. However, the transformation involves two separate synthetic steps. Fasrez et al. have achieved a one-pot conversion of α -aminoalkyl phosphonous acids into monoethyl α -aminoalkylphosphonates using iodine/ethanol.²⁷ However, there is no indication on the applicability of this reaction for the synthesis of N-protected monomethyl α -aminoalkylphosphonates. The latter are much more common and convenient synthons for preparing phosphonate-,^{4,17b,22b,28} phosphonamidate-^{4,29} and even phosphinate-peptides,³⁰ as well as building blocks for assembling combinatorial libraries³¹ than the ethyl counterparts, since the deprotection of a methyl ester appears to be more facile than that of an ethyl ester.^{4,8a}

To the best of our knowledge, a one-pot transformation of phosphonous acid into phosphonate monomethyl ester has not yet been described. We report here a new, short, one-pot synthesis of N-protected monomethyl α -aminoalkylphosphonates **2** from N-protected α -aminoalkylphosphonous acids **1** (Scheme 1). We succeeded in achieving this single step transformation during the course of our investigations aimed at developing new phosphapeptides with potential biological activity.^{8b,32} The starting compounds **1** were readily synthesized by N-protection of α -aminoalkylphosphonous acids,^{33–35} which were obtained by the method developed by Baylis et al.³⁴

This method, which involves Strecker-type condensation of amines with aldehydes and anhydrous hypophosphorous acid, has been recently improved using microwave irradiation under solvent free conditions.³⁶ α -Aminoalkylphosphonous acids can also be prepared by alkylation of the N-protected aminomethylphosphonous acid at the C_a-position.³⁷



Scheme 1

We have now found that racemic N-protected α -aminoalkylphosphonous acids **1a–h** are rapidly transformed into the corresponding monomethyl phosphonates **2a–h**, using thionyl chloride in methanol (Scheme 1, Table 1). In order to show the generality of the reaction we have prepared several monomethyl phosphonate analogues of α -amino acids, N-protected with the two most commonly used protecting groups, benzyloxycarbonyl (Cbz) and 9-fluorenylmethoxycarbonyl (Fmoc) (Table 1). The overall yields of the reactions are high, especially if we take into consideration the fact that two reactions, oxidation and selective esterification occur in a single step.

Table 1 Monomethyl α -Aminoalkylphosphonates **2a–h** Prepared (Scheme 1)

Product	R	Pg	Yield ^{a,b} (%)	mp (°C)
2a	CH ₃	Cbz	65	105–108 ^c
2b	CH(CH ₃) ₂	Cbz	69	93–95 ^d
2c	CH ₂ CH(CH ₃) ₂	Cbz	77	114–115 ^e
2d	CH(CH ₃)CH ₂ CH ₃	Cbz	79	84–87
2e	CH ₃	Fmoc	71	158–160 ^f
2f	CH(CH ₃) ₂	Fmoc	84	135–137 ^f
2g	CH ₂ CH(CH ₃) ₂	Fmoc	75	136–138
2h	CH(CH ₃)CH ₂ CH ₃	Fmoc	86	147–149

^a Yields of isolated, pure products.

^b The structures of all compounds were confirmed by ¹H NMR, ³¹P NMR, IR and mass spectra. For new compounds satisfactory elemental analyses were obtained.

^c Lit.^{29b} mp 108 °C.

^d Lit.^{14b} mp 93–96 °C.

^e Lit.^{28b} mp 118–119 °C.

^f Melting points not given in the literature.³⁸

To summarize, we have presented here an expeditious one-pot conversion of N-protected α -aminoalkylphosphonous acids into the corresponding monomethyl phosphonates. The reaction is fast, the procedure is simple and it

leads to versatile N-protected monomethyl α -aminoalkylphosphonates, which are valuable intermediates for preparing phosphopeptides. Speed and simplicity make this method attractive and potentially useful in the field of preparative and medicinal chemistry.

All reagents and solvents were of commercial grade and used as such. Melting points were determined using a Reichert hot stage microscope and are uncorrected. Elemental C, H, N analyses were performed at the Faculty of Chemistry and Chemical Engineering, University of Ljubljana, on a Perkin-Elmer elemental analyzer 240 C. IR spectra were measured by a Perkin-Elmer FTIR 1600 instrument on KBr pelleted samples. Mass spectra were obtained by a Micromass AutospecQ mass spectrometer using FAB ionization. NMR spectra were obtained on a Bruker Avance DPX 300 instrument. ¹H NMR spectra were done at 300.13 MHz with tetramethylsilane as an internal standard and ³¹P NMR spectra were done at 121 MHz using H₃PO₄ as an external standard.

1-[(9H-Fluoren-9-ylmethoxy)carbonyl]amino-3-methylbutylphosphinic Acid [1g; Pg = Fmoc, R = CH₂CH(CH₃)₂]

Compound **1g** was prepared from 1-amino-3-methylbutylphosphinic acid according to the literature procedure of Yiotakis.^{35b} Yield: 97%; mp 158–160 °C; R_f 0.59 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3284, 2958, 1724, 1547, 1451, 1267, 1098, 985, 140, 530 cm⁻¹.

¹H NMR (DMSO-*d*₆): δ = 0.83 (d, 3 H, J = 6.4 Hz, CH₃), 0.90 (d, 3 H, J = 6.8 Hz, CH₃), 1.30–1.44 [m, 1 H, CH(CH₃)₂], 1.45–1.72 (m, 2 H, CH₂CH), 3.56–3.71 (m, 1 H, PCH), 4.18–4.39 (m, 3 H, CHCH₂O), 6.725 (d, 1 H, J = 518 Hz, PH), 7.28–7.93 (m, 8 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 29.79.

MS (FAB): *m/z* = 374 (M + H)⁺, 396 (M + Na)⁺.

Anal. Calcd for C₂₀H₂₄NO₄P·0.25H₂O: C, 63.57; H, 6.53; N, 3.71. Found: C, 63.61; H, 6.21; N, 3.40.

1-[(9H-Fluoren-9-ylmethoxy)carbonyl]amino-2-methylbutylphosphinic Acid [1h; Pg = Fmoc, R = CH(CH₃)CH₂CH₃]

Compound **1h** was prepared from 1-amino-2-methylbutylphosphinic acid according to the literature procedure of Yiotakis.^{35b} Yield: 95%; mp 128–130 °C; R_f 0.66 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3290, 2966, 1689, 1534, 1450, 1294, 974, 740 cm⁻¹.

¹H NMR (DMSO-*d*₆): δ = 0.84 [2 t, J = 7.345, 7.345 Hz, 3 H, CH(CH₃)CH₂CH₃], 0.97 [2 d, J = 6.78, 6.78 Hz, 3 H, CH(CH₃)CH₂CH₃], 1.12–1.95 [m, 3 H, CH(CH₃)CH₂CH₃], 3.43–3.78 (m, 1 H, PCH), 4.18–4.38 (m, 3 H, CHCH₂O), 6.79 (2 d, 1 H, ¹J_{PH} = 509.8, 506.7 Hz, PH), 7.27–7.94 (m, 8 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 30.03 (1 P), 29.10 (1 P).

MS (FAB): *m/z* = 374 (M + H)⁺, 396 (M + Na)⁺.

Anal. Calcd for C₂₀H₂₄NO₄P: C, 64.33; H, 6.48; N, 3.75. Found: C, 64.53; H, 6.65; N, 3.60.

N-Protected Monomethyl α -Aminoalkylphosphonates **2a–h** from N-Protected α -Aminoalkylphosphonous Acid **1a–h**; General Procedure

N-Protected α -aminoalkylphosphonous acid **1a–h** (3 mmol) was dissolved in anhyd MeOH (25 mL), cooled to 0 °C, and SOCl₂ (0.714 g, 6 mmol) was then added dropwise. The reaction mixture was stirred at 0 °C for 15 min, then allowed to warm to r.t. and stirred for an additional 15 min. Afterwards the mixture was refluxed for 2 h and concentrated in vacuo. The residue was triturated with CH₂Cl₂ (50 mL), filtered and the filtrate was concentrated in vacuo. The concentrated filtrate was suspended in H₂O (50 mL) (the

pH of the solution was 1–2), triturated, cooled to 0 °C and aq 2 M NaOH added dropwise with constant stirring until pH of the solution became 9–10. The alkaline solution was washed with EtOAc (3 × 30 mL), cooled to 0 °C and acidified to pH 1 by dropwise addition of conc. HCl while stirring. The product **2a–h** was precipitated as a white solid, which was filtered and washed with Et₂O (3 × 10 mL). All products thus obtained were identified by comparison of their NMR, IR, MS data and melting points with the values reported in the literature.^{14b,28b} For all new compounds (**2d**, **2g**, **2h**) satisfactory elemental analyses were also obtained.

Methyl Hydrogen 1-{[(Benzylxy)carbonyl]amino}ethylphosphonate (2a)

Mp 105–108 °C (Lit.^{29b} mp 108 °C); R_f 0.42 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3291, 1693, 1544, 1455, 1305, 1056, 817, 694 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 1.37 (2 d, 3 H, *J* = 7.54, 7.15 Hz, CH₃), 3.73 (d, 3 H, *J* = 10.93 Hz, POCH₃), 4.05–4.30 (m, 1 H, PCH), 5.13 (s, 2 H, ArCH₂), 7.30–7.40 (m, 5 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 29.46.

MS (FAB): *m/z* = 274 (M + H)⁺.

Methyl Hydrogen 1-{[(Benzylxy)carbonyl]amino}-2-methylpropylphosphonate (2b)

Mp 93–95 °C (Lit.^{14b} mp 93–96 °C); R_f 0.51 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3329, 2969, 2284, 1716, 1682, 1531, 1456, 1390, 1294, 1254, 1170, 1049, 1002, 911 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 0.995 (d, 3 H, *J* = 6.78 Hz, CH₃), 1.03 (d, 3 H, *J* = 6.78 Hz, CH₃), 2.14–2.30 [m, 1 H, CH(CH₃)₂], 3.71 (d, 3 H, *J* = 10.93 Hz, POCH₃), 3.99–4.15 (m, 1 H, PCH), 5.15 (s, 2 H, ArCH₂), 7.30–7.40 (m, 5 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 24.00.

MS (FAB): *m/z* = 302 (M + H)⁺, 324 (M + Na)⁺.

Methyl Hydrogen 1-{[(Benzylxy)carbonyl]amino}-3-methylbutylphosphonate (2c)

Mp 114–115 °C; R_f 0.46 (H₂O–MeOH–MeCN, 1:1:3).

IR (KBr): 3284, 2956, 1688, 1544, 1450, 1309, 1269, 1184, 1037, 790, 696, 568 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 0.825 (d, 3 H, *J* = 6.4 Hz, CH₃), 0.88 (d, 3 H, *J* = 6.4 Hz, CH₃), 1.31–1.46 [m, 1 H, CH(CH₃)₂], 1.49–1.70 [m, 2 H, CH₂CH(CH₃)₂], 3.55 (d, 3 H, *J* = 10.55 Hz, POCH₃), 3.75–3.93 (m, 1 H, PCH), 5.05 (s, 2 H, ArCH₂), 7.28–7.38 (m, 5 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 25.43.

MS (FAB): *m/z* = 316 (M + H)⁺, 338 (M + Na)⁺.

Methyl Hydrogen 1-{[(Benzylxy)carbonyl]amino}-2-methylbutylphosphonate (2d)

Mp 84–87 °C; R_f 0.43 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3299, 2963, 1719, 1541, 1457, 1244, 1168, 1059, 990, 742, 568 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 0.83 [2 t, *J* = 7.335, 6.785 Hz, 3 H, CH(CH₃)CH₂CH₃], 0.945 [2 d, *J* = 6.78, 6.78 Hz, 3 H, CH(CH₃)CH₂CH₃], 1.06–1.88 [m, 3 H, CH(CH₃)CH₂CH₃], 3.535 (2 d, 3 H, *J* = 10.565, 10.535 Hz, POCH₃), 3.64–3.99 (m, 1 H, PCH), 5.06 (s, 2 H, ArCH₂), 7.28–7.40 (m, 5 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 24.69 (1 P), 24.54 (0.5 P).

MS (FAB): *m/z* = 316 (M + H)⁺, 338 (M + Na)⁺.

Anal. Calcd for C₁₄H₂₂NO₅P: C, 53.33; H, 7.03; N, 4.44. Found: C, 52.97; H, 7.17; N, 4.50.

Methyl Hydrogen 1-{[(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}ethylphosphonate (2e)

Mp 158–160 °C (Lit.³⁸ mp not reported); R_f 0.42 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3284, 3065, 1690, 1542.5, 1450, 1305, 1048, 739 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 1.24 (2 d, 3 H, *J* = 7.16, 7.16 Hz, CH₃), 3.55 (d, 3 H, *J* = 10.55 Hz, POCH₃), 3.75–3.92 (m, 1 H, PCH), 4.17–4.35 (m, 3 H, CHCH₂O), 7.27–7.94 (m, 8 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 25.02.

MS (FAB): *m/z* = 362 (M + H)⁺.

Methyl Hydrogen 1-{[(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}-2-methylpropylphosphonate (2f)

Mp 135–137 °C (Lit.³⁸ mp not reported); R_f 0.47 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3290, 2960, 1689, 1543, 1450, 1214, 1045, 990, 741 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 0.96 (t, 6 H, *J* = 6.78 Hz, CH₃), 1.98–2.15 [m, 1 H, CH(CH₃)₂], 3.55 (d, 3 H, *J* = 10.55 Hz, POCH₃), 3.62–3.78 (m, 1 H, PCH), 4.16–4.33 (m, 3 H, CHCH₂O), 7.27–7.93 (m, 8 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 24.54.

MS (FAB): *m/z* = 390 (M + H)⁺, 412 (M + Na)⁺.

Methyl Hydrogen 1-{[(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}-3-methylbutylphosphonate (2g)

Mp 136–139 °C; R_f 0.63 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3300, 3066, 2955, 1710, 1536, 1450, 1265, 1044, 988, 739, 563 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 0.82 (d, 3 H, *J* = 6.4 Hz, CH₃), 0.90 (d, 3 H, *J* = 6.4 Hz, CH₃), 1.34–1.51 [m, 1 H, CH(CH₃)₂], 1.53–1.75 (m, 2 H, CH₂CH), 3.57 (d, 3 H, *J* = 10.55 Hz, POCH₃), 3.75–3.95 (m, 1 H, PCH), 4.16–4.35 (m, 3 H, CHCH₂O), 7.27–7.92 (m, 8 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 25.55.

MS (FAB): *m/z* = 404 (M + H)⁺, 426 (M + Na)⁺.

Anal. Calcd for C₂₁H₂₆NO₅P·0.5 H₂O: C, 61.16; H, 6.60; N, 3.40. Found: C, 61.09; H, 6.71; N, 3.67.

Methyl Hydrogen 1-{[(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}-2-methylbutylphosphonate (2h)

Mp 147–149 °C; R_f 0.59 (CHCl₃–MeOH–AcOH, 7:2:1).

IR (KBr): 3313, 2962, 1711, 1530, 1450, 1247, 1048, 740, 566 cm^{−1}.

¹H NMR (DMSO-*d*₆): δ = 0.84 [q, *J* = 7.79 Hz, 3 H, CH(CH₃)CH₂CH₃], 0.99 [t, *J* = 6.97 Hz, 3 H, CH(CH₃)CH₂CH₃], 1.10–1.94 [m, 3 H, CH(CH₃)CH₂CH₃], 3.56 (2 d, 3 H, *J* = 10.55, 10.55 Hz, POCH₃), 3.66–4.03 (m, 1 H, PCH), 4.15–4.33 (m, 3 H, CHCH₂O), 7.27–7.92 (m, 8 H_{arom}).

³¹P NMR (DMSO-*d*₆): δ = 24.95 (1 P), 24.82 (0.9 P).

MS (FAB): *m/z* = 404 (M + H)⁺, 426 (M + Na)⁺.

Anal. Calcd for C₂₁H₂₆NO₅P·0.75 H₂O: C, 60.50; H, 6.65; N, 3.36. Found: C, 60.77; H, 6.58; N, 3.61.

Acknowledgment

The Ministry of Education, Science and Sport of the Republic of Slovenia financially supported this work. The authors thank Professor Dr. Roger Pain, Professor Dr. Danijel Kikelj and Dr. Renata Jakše for critical reading of the manuscript.

References

- (1) (a) Hanson, J. E.; Kaplan, A. P.; Bartlett, P. A. *Biochemistry* **1989**, *28*, 6294. (b) Grobelny, D.; Goli, U. B.; Galardy, R. E. *Biochemistry* **1989**, *28*, 4948.
- (2) (a) Hirschmann, R.; Yager, K. M.; Taylor, C. M.; Witherington, J.; Sprengeler, P. A.; Phillips, B. W.; Moore, W.; Smith, A. B. III *J. Am. Chem. Soc.* **1997**, *119*, 8177. (b) Babine, R. E.; Bender, S. L. *Chem. Rev.* **1997**, *97*, 1359; and references cited therein.
- (3) (a) Lerner, R. A.; Benkovic, S. J.; Schultz, P. G. *Science* **1991**, *252*, 659. (b) Hirschmann, R.; Smith, A. B. III; Taylor, C. M.; Benkovic, P. A.; Taylor, S. D.; Yager, K. M.; Sprengeler, P. A.; Benkovic, S. J. *Science* **1994**, *265*, 234. (c) Smithrud, D. B.; Benkovic, P. A.; Benkovic, S. J.; Taylor, C. M.; Yager, K. M.; Witherington, J.; Philips, B. W.; Sprengeler, P. A.; Smith, A. B. III; Hirschmann, R. *J. Am. Chem. Soc.* **1997**, *119*, 278. (d) Mader, M. M.; Bartlett, P. A. *Chem. Rev.* **1997**, *97*, 1281.
- (4) Malachowski, W. P.; Coward, J. K. *J. Org. Chem.* **1994**, *59*, 7616.
- (5) Morgan, B. P.; Scholtz, J. M.; Ballinger, M. D.; Zipkin, I. D.; Bartlett, P. A. *J. Am. Chem. Soc.* **1991**, *113*, 297.
- (6) Campagne, J.-M.; Coste, J.; Jouin, P. *J. Org. Chem.* **1995**, *60*, 5214.
- (7) (a) Campbell, D. A. *J. Org. Chem.* **1992**, *57*, 6331. (b) Campbell, D. A.; Bermak, J. C. *J. Org. Chem.* **1994**, *59*, 658.
- (8) (a) Yamauchi, K.; Ohtsuki, S.; Kinoshita, M. *J. Org. Chem.* **1984**, *49*, 1158. (b) Gobec, S.; Urleb, U. *Molecules* **2002**, *7*, 394.
- (9) (a) Natchev, I. A. *Bull. Chem. Soc. Jpn.* **1988**, *61*, 3705. (b) Natchev, I. A. *Bull. Chem. Soc. Jpn.* **1988**, *61*, 4491. (c) Yamauchi, K.; Kinoshita, M.; Imoto, M. *Bull. Chem. Soc. Jpn.* **1972**, *45*, 2528.
- (10) Chen, S.; Lin, C.-H.; Kwon, D. S.; Walsh, C. T.; Coward, J. K. *J. Med. Chem.* **1997**, *40*, 3842.
- (11) (a) Atherton, F. R.; Oppenshaw, H. T.; Todd, A. R. *J. Chem. Soc.* **1945**, 660. (b) Sampson, N. A.; Bartlett, P. A. *J. Org. Chem.* **1988**, *53*, 4500.
- (12) Burger, A.; Anderson, J. *J. Am. Chem. Soc.* **1957**, *79*, 3575.
- (13) Gilmore, W. F.; McBride, H. A. *J. Pharm. Sci.* **1974**, *63*, 965.
- (14) (a) Wasielewski, C.; Hoffmann, M.; Witkowska, E. *J. Roczn. Chem.* **1975**, *49*, 1795; *Chem. Abstr.* **1976**, *84*, 90218. (b) Wasielewski, C.; Hoffmann, M.; Witkowska, E.; Rachon, J. *Roczn. Chem.* **1976**, *50*, 1613; *Chem. Abstr.* **1977**, *86*, 89948.
- (15) Hoffmann, M. *Synthesis* **1986**, 557.
- (16) Galéotti, N.; Coste, J.; Bedos, P.; Jouin, P. *Tetrahedron Lett.* **1996**, *37*, 3997.
- (17) (a) Jacobsen, N. E.; Bartlett, P. A. *J. Am. Chem. Soc.* **1981**, *103*, 654. (b) Bartlett, P. A.; Hanson, J. E.; Giannousis, P. P. *J. Org. Chem.* **1990**, *55*, 6268.
- (18) Holý, A. *Synthesis* **1998**, 381.
- (19) Salomon, C. J.; Breuer, E. *Tetrahedron Lett.* **1995**, *36*, 6759.
- (20) (a) Gray, M. D. M.; Smith, D. J. H. *Tetrahedron Lett.* **1980**, *21*, 859. (b) Malachowski, W. P.; Coward, J. K. *J. Org. Chem.* **1994**, *59*, 7625.
- (21) (a) Saady, M.; Lebeau, L.; Mioskowski, C. *J. Org. Chem.* **1995**, *60*, 2946. (b) Hoffman, M. *Phosphorus, Sulfur Silicon Relat. Elem.* **1998**, *134/135*, 109.
- (22) (a) Hoffmann, M. *J. Prakt. Chem.* **1988**, *330*, 820. (b) Brachwitz, H.; Ölke, M.; Bergmann, J.; Langen, P. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1739 SS>. (c) Patel, D. V.; Rielly-Gauvin, K.; Ryono, D. E. *Tetrahedron Lett.* **1990**, *31*, 5591.
- (23) (a) Daub, G. W.; Van Tamelen, E. E. *J. Am. Chem. Soc.* **1977**, *99*, 3526. (b) Norbeck, D. W.; Kramer, J. B.; Lartey, P. A. *J. Org. Chem.* **1987**, *52*, 2174. (c) Campbell, D. A.; Bermauk, J. C. *J. Am. Chem. Soc.* **1994**, *116*, 6039.
- (24) (a) Yamauchi, K.; Kinoshita, M.; Imoto, M. *Bull. Chem. Soc. Jpn.* **1972**, *45*, 2531. (b) Ikeda, K.; Achiwa, K. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 225.
- (25) Green, M.; Hudson, R. F. *J. Chem. Soc.* **1958**, 3129.
- (26) Karanewsky, D. S.; Badia, M. C. *Tetrahedron Lett.* **1986**, *27*, 1751.
- (27) Fastrez, J.; Jespers, L.; Lison, D.; Renard, M.; Sonveaux, E. *Tetrahedron Lett.* **1989**, *30*, 6861.
- (28) (a) Smith, W. W.; Bartlett, P. A. *J. Am. Chem. Soc.* **1998**, *120*, 4622. (b) Giannousis, P. P.; Bartlett, P. A. *J. Med. Chem.* **1987**, *30*, 1603.
- (29) (a) Yang, K.-W.; Brandt, J. J.; Chatwood, L. L.; Crowder, M. W. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 1085. (b) Quang-Vo, Y.; Gravey, A. M.; Simonneau, R.; Quang-Vo, L.; Lacoste, A. M.; Le Goffic, F. *Tetrahedron Lett.* **1987**, *28*, 6167. (c) Musiol, H.-D.; Grams, F.; Rudolph-Böhner, S.; Moroder, L. *J. Org. Chem.* **1994**, *59*, 6144. (d) Camp, N. P.; Perrey, D. A.; Kinchington, D.; Hawkins, P. C. D.; Gani, D. *Bioorg. Med. Chem.* **1995**, *3*, 297. (e) Camp, N. P.; Hawkins, P. C. D.; Hitchcock, P. B.; Gani, D. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 1047. (f) Chen, S.; Lin, C.-H.; Kwon, D. S.; Walsh, C. T.; Coward, J. K. *J. Med. Chem.* **1997**, *40*, 3842.
- (30) Bartlett, P. A.; Kezer, W. B. *J. Am. Chem. Soc.* **1984**, *106*, 4282.
- (31) Campbell, D. A.; Bernak, J. C.; Burkoth, T. S.; Patel, D. V. *J. Am. Chem. Soc.* **1995**, *117*, 5381.
- (32) Gobec, S.; Štrancar, K.; Urleb, U. *Tetrahedron Lett.* **2002**, *43*, 167.
- (33) α -Aminoalkylphosphonous acids were N-protected by Cbz and Fmoc protecting group using standard methods.^{34,35} The structures of new N-protected α -aminoalkylphosphonous acids **1g** [Pg = Fmoc, R = $\text{CH}_2\text{CH}(\text{CH}_3)_2$] and **1h** [Pg = Fmoc, R = $\text{CH}(\text{CH}_3)\text{CH}_2\text{CH}_3$] were confirmed by ¹H NMR, ³¹P NMR, IR, mass spectra, and elemental analysis.
- (34) Baylis, E. K.; Campbell, C. D.; Dingwall, J. G. *J. Chem. Soc., Perkin Trans. I* **1984**, 2845.
- (35) (a) Dumy, P.; Escale, R.; Girard, J. P.; Parello, J.; Vidal, J. P. *Synthesis* **1992**, 1226. (b) Georgiadis, D.; Matziari, M.; Yiotakis, A. *Tetrahedron* **2001**, *57*, 3471.
- (36) Kaboudin, B.; As-habei, N. *Tetrahedron Lett.* **2003**, *44*, 4243.
- (37) McCleery, P. P.; Tuck, B. *J. Chem. Soc., Perkin Trans. I* **1989**, 1319.
- (38) Dumy, P.; Escale, R.; Girard, J. P.; Parello, J.; Vidal, J. P. In *Peptides 1990, Proc. Eur. Pept. Symp.*; Giralt, E.; Andreu, D., Eds.; ESCOM Sci. Publ.: Leiden, **1991**, 440.