Synthesis, structure, and transformations of 1,2,4-phosphites of α -D-xylopyranose

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ABSTRACT

The reaction of D-xylose with tris(3,5-dimethylpyrazolyl) phosphite gave the 1,2,4-phosphite, which was oxidised to the 1,2,4-phosphate and also converted into the 1,2,4-thio- and -seleno-phosphates. These compounds were hydrolysed readily to give, first, the 1,4-phosphates and then the 4-phosphates. The direction of hydrolysis of the 1,2,4-phosphates is discussed on the basis of X-ray data on the starting compounds.

INTRODUCTION

Bicyclophosphites of carbohydrates¹ have potential application in the synthesis of bioregulators² and were obtained first by phosphorylation of partially substituted carbohydrates. We now report on the conversion of D-xylopyranose into a 1,2,4-phosphite.

RESULTS AND DISCUSSION

The reaction of α,β -D-xylopyranose (α,β -ratio 1:1; ¹³C-n.m.r. data) in dry dioxane-pyridine with tris(3,5-dimethylpyrazolyl) phosphite⁴ gave, after chromatography on silica gel, 40% of the syrupy 1,2,4-triphosphite 1 (*cf.* ref. 3). The use of triamidophosphites or phosphorus trichloride gave lower yields of 1.

The structure of 1 was established on the basis of ³¹P-, ¹H-, and ¹³C-n.m.r. data (Tables I and II). Thus, there was a single ³¹P resonance (s) at 111.2 p.p.m., which corresponded to those of other bicyclophosphites¹; H-1,2,4 were involved in ³ $J_{H,P}$ couplings; and the resonances for C-1,2,3,4 were doublets due to coupling with P, whereas that for C-5 was a singlet.

On storage, 1 underwent partial intermolecular alcoholysis due to the free hydroxyl group, but the acetate (2), benzoate (3), and phenylcarbamate (4) were stable derivatives. The structures of 2-4 were confirmed by the n.m.r. data. The 31 P-n.m.r. spectra of 1-4 were similar, but the signals for H-3 of 2-4 were shifted upfield compared to that of H-3 in 1 and indicated the substituents to be at position 3.

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¹H- and ³¹P-n.m.r. data^{*a*} (δ in p.p.m, J in Hz)

Compound	H-1	H-2	H-3	H-4	H-5a	H-5b	³¹ <i>P</i>
	J _{1,2}	J _{2,3}	J _{3.4}	J _{4.5a}	$J_{Sa,5b}$	$J_{4.5b}$	
	J _{<i>I,P</i>}	J _{2,P}		$\mathbf{J}_{\boldsymbol{4},\boldsymbol{P}}$			
1	6.22	4.75	4.47	4.70	4.53	4.26	111.2
	(5.2)	(4.4)	(5.2)	(3.7)	(12.5)	(0)	
	(5.9)	(8.1)		(9.6)			
2	5.88	4.47	5.13	4.52	4.15	4.07	111.5
	(5.2)	(2.9)	(2.2)	(4.4)	(12.5)	(<1)	
	(6.6)	(8.1)		(8.8)			
3	6.21	4.88	5.54	4.84	4.29	4.15	111.5
	(5.1)	(1.9)	(4.4)	(4.8)	(12.0)	(<1)	
	(5.4)	(10.1)		(9.5)			
4	5.92	4.54	5.18	4.62	4.28	4.16	111.3
	(5.1)	(3.0)	(4.2)	(4.5)	(12.2)	(<1)	
	(6.0)	(8.2)		(9.6)			
9	6.01	4.82	5.30	4.78	4.43	4.24	6.6
	(5.5)	(3.1)	(4.5)	(4.3)	(13.0)	(0)	
	(17.1)	(19.5)		(24.7)			
10	6.00	4.75	4.48	4.68	4.40	4.20	6.6
	(5.3)	(3.0)	(4.5)	(4.2)	(12.6)	(<1)	
	(17.0)	(19.1)		(24.0)			
15	6.04	4.72	5.34	4.82	4.43	4.20	74.0
	(5.5)	(4.6)	(2.2)	(4.6)	(13.1)	(0)	$(J_{P.Se} \ 1100)$
	(16.8)	(17.6)		(30.1)			,
16	6.11	4.85	5.62	4.95	4.50	4.30	74.0
	(5.4)	(2.3)	(2.0)	(4.4)	(12.7)	(<1)	$(J_{PSe} \ 1100)$
	(16.6)	(19.5)		(25.9)			
17	6.03	4.75	5.32	4.81	4.44	4.20	70.7
	(5.04)	(1.9)	(4.8)	(4.4)	(12.7)	(0)	
	(17.0)	(19.0)		(23.9)			
18	6.10	4.85	5.60	4.91	4.52	4.31	70.9
	(5.9)	(1.5)	(4.4)	(4.4)	(12.5)	(<1)	
	(16.9)	(18.4)		(21.3)			
19	6.07	4.82	5.40	4.92	4.49	4.26	70.9
	(5.5)	(1.8)	(4.5)	(4.5)	(13.0)	(0)	
	(17.0)	(19.5)		(22.5)			
20	5.46	4.06	6.56	4.45	4.29	4.07	35.8
	(3.6)	(3.7)	(1.5)	(<1)	(11.8)	(<1)	$(J_{P,Se} 800)$
_	(27. 9)			(27.9)			
21	5.48	4.10	6.44	4.45	4.27	4.11	48.6
	(3.4)	(3.9)	(1.4)	(<1)	(12.2)	(<1)	
	(24.9)			(27.3)			
23α	5.2	4.50	6.33	4.65	4.49	4.21	-0.8
	(3.4)	(<1)	(2.2)	(2.9)	(12.7)	(<1)	
•••		(21.5)		(21.9)			
23 β	6.03	4.43	5.95	4.61	4.38	3.90	-0.6
	(4.9)	(<1)	(3.7)	(2.2)	(12.7)	(2.20)	
		(8.8)		(11.2)			

^a The ¹H resonances of the protecting groups are not recorded.



TABLE II

¹³C Chemical shift data^{*a*} (δ in p.p.m., $J_{P,C}$ values in Hz)

Compound	C-1	C-2	С-3	C-4	Ċ-5	
1	98.9	73.1	64.9	77.4	65.9	
	(5.4)	(2.4)	(4.9)	(3.8)		
2	98.5	70.4	65.5	73.7	65.7	
	(5.3)	(2.6)	(6.0)	(3.3)		
3	98.7	70.4	65.8	73.6	66.0	
	(5.6)	(2.7)	(6.1)	(3.5)		
4	98.4	70.7	65.4	74.1	65.7	
	(5.3)	(2.1)	(6.1)	(3.2)		
5	97.2	71.2	64.9	74.4	65.1	
	(8.8)	(5.6)	(4.1)	(10.9)		
6	97.1	71.2	65.0	74.4	64.8	
	(8.8)	(5.7)	(3.2)	(11.1)		
15	97.8	73.2	64.2	75.5	64.9	
	(5.6)	(4.6)	(2.7)	(12.2)		
16	97.8	73.3	64.2	75.5	65.0	
	(5.9)	(4.5)	(2.7)	(12.1)		
17	96.9	72.9	64.2	74.6	64.7	
	(3.5)	(2.7)	(2.3)	(11.1)		
18	96.9	73.1	64.3	74.7	64.9	
	(3.5)	(3.4)	(2.4)	(10.7)		
19	96.8	73.2	64.5	74.8	64.7	
	(3.4)	(2.8)	(2.5)	(10.9)		
20	96.7	73.6	72.4	71.7	64.2	
	(12.1)	(2.0)		(9.8)		
21	96.5	73.6	72.9	71.4	64.4	
	(11.1)			(9.2)		
23α	87.5	71.1	63.0	72.2	60.4	
		(8.6)	(5.3)	(8.5)		
23β	88.1	70.0	60.8	71.5	61.2	
•		(7.6)	(5.0)	(7.9)		

" The ¹³C resonances of the protecting groups are not recorded.

	3	16		3	16
P0-1	1.620(9)	1.594(9)	O-1-P-O-2	92.72(39)	95.47(45)
Р-О-2	1.618(8)	1.619(9)	0-1-P-0-4	101.44(43)	106.66(42)
P0-4	1.636(8)	1.596(7)	O-2-P-O-4	100.18(36)	103.50(46)
O-1-C-1	1.421(10)	1.481(13)	P-O-1-C-1	111.29(66)	107.08(69)
O-2–C-2	1.438(11)	1.459(12)	P-O-2-C-2	106.92(68)	100.94(67)
0-4-C-4	1.459(10)	1.394(11)	P-O-4-C-4	122.11(66)	116.60(63)

Interatomic distances (Å) and bond angles (°) for 3 and 16



Fig. 1. Projection of the 3-O-benzoyl-a-D-xylopyranose 1,2,4-phosphite (3) molecule.

X-Ray analysis of the benzoate 3 (Fig. 1 and Table III) revealed that (a) the carbohydrate moiety had a $B_{3,0}$ conformation; (b) the phosphorinane ring had a ${}^{2}C_{4}$ conformation with C-4 and C-2 deviating from the plane of the other four atoms by 0.570 and 0.845 Å, respectively; and (c) the phospholane ring had an E_{2} conformation with O-2 0.673 Å out of the plane. The constraints on the O-P-O angles result in a significant increase of the *s*-character of the lone pair of electrons on the phosphorus, which should decrease its basicity and nucleophilicity¹.

Reaction of the acetate 2 and the benzoate 3 with rhodium(I) dicarbonyl acetylacetonate gave the chiral rhodium complexes 5 and 6, respectively, which, on treatment with triaminophosphites, regenerated 2 and 3. The n.m.r. spectra of 5 and 6 contained doublets for ³¹P at 135.0 p.p.m. with a ¹J_{P,Rh} value of 290.0 Hz; in the ¹³C-n.m.r. spectra (Table II) signals of C-1,2,3,4 in 5 and 6 have ³J_{C,P}, the value of which differ from those in 2 and 3 and indicated a change in the geometry of the 1,2,4-phosphite upon complexation.

Treatment of 2 and 3 with ozone in dichloromethane at -70° afforded ozonides (7 and 8, respectively) with pentacoordinate phosphorus, detected by ³¹P-n.m.r. spectroscopy (³¹P at 32 p.p.m.), which lost oxygen at 20° to form the 1,2,4-phosphates⁵9 and 10, respectively.

The structures of 9 and 10 were established by the ³¹P- and ¹H-n.m.r. data (Table I). There was a ³¹P resonance at 6.6 p.p.m. and the ¹H resonances, in general, were similar to those of 2 and 3. However, the $J_{H,P}$ values were increased significantly (6 \rightarrow 17 Hz for H-1, 8 \rightarrow 19 Hz for H-2, and 9 \rightarrow 24 Hz for H-4). Compounds 9 and 10 polymerised readily on storage.



Oxidation of 1,2-O-alkylidene-D-glucofuranose 3,5,6-phosphites with aqueous 30% hydrogen peroxide gave the 3,5,6-phosphates that could be hydrolysed selectively to afford the stable 3,5-phosphates⁶. Similar treatment of 2 and 3 with hydrogen peroxide gave, first, the 1,2,4-triphosphates 9 and 10, respectively, which were then hydrolysed to give the 1,4-phosphates (11 and 12) that were converted into the 4-phosphates 13 and 14, respectively.

The ¹³C-n.m.r. spectra of 13 and 14 contained signals for the α and β anomers, and the signals for C-1,2 in each anomer were now singlets.

The 1,2,4-phosphites 2-4 reacted variously with selenium and sulfur in dry dioxane at 100° to give 15-19 with a consequent upfield shift of the ³¹P resonances due to the four-coordinate phosphorus. There were also increases in the $J_{H,P}$ and $J_{C,P}$ values. The structure of the seleno derivative 16 was confirmed by the X-ray analysis data (Fig. 2 and Table III). The phosphorinane ring has a ${}^{4}C_{2}$ conformation with O-4 and O-2 being 0.613 and 0.865 Å, respectively, out of the plane; the phospholane ring has an E_{2} conformation, the deviation of O-2 being 0.722 Å; and the carbohydrate moiety has the $B_{3,0}$ conformation. Compared to those in 2-4, the O-P-O angles are increased by 3-5° and the P-O-C angles are decreased by 4-6°. There are also changes in the P-O and O-C bond lengths. Thus, the P-O-2 bond is the shortest in 3 and the longest in 16. These changes, also observed in previous work⁷, cause an increase in the strain energy.

The seleno derivatives 15 and 16 were reactive compounds. Thus, with hexaethylphosphoric triamide at room temperature, 15 was re-converted into 2.



The strain in the 1,2,4-phosphate molecules can be relieved by conversion into the 1,4-phosphates. Similar to the 1,2,4-phosphates 9 and 10, the 1,2,4-thio- and 1,2,4-seleno-phosphates reacted with aqueous triethylamine to give 20 and 21. Each hydrolysis involved the P-O-2 bond, which is the longest (1.619 Å, cf. 1.594 Å for the P-O-1 bond and 1.596 Å for the P-O-4 bond: see Table III).

The structures of 20 and 21 were established by the ${}^{31}P$ -, ${}^{1}H$ -, and ${}^{13}C$ -n.m.r. data. The ${}^{31}P$ resonance of 20 was shifted to higher field (35.8 p.p.m.) and there was no



Fig. 2. Projection of the 3-O-benzoyl-a-D-xylopyranose 1,2,4-selenophosphate (16) molecule.

coupling between P and H-2, as expected after cleavage of the P-O-2 bond⁶. The signal of C-2 was no longer a doublet and was shifted to higher field. Large $J_{C,P}$ values were observed for C-1 and C-4. Similar results were obtained for the thio derivative 21.

Thus, the regio- and stereo-specific opening of the condensed phospholanephosphorinane system in aqueous triethylamine, to afford the 1,4-phosphates, is similar to that of the furanose analogues⁶.

Reaction of 3 with chlorine under homolytic conditions⁸ gave the glycosyl chloride $22\alpha\beta$, which was highly reactive but could be stabilised by reaction with piperidine to give $23\alpha\beta$. The α and β anomers of 23 were isolated by chromatography on silica gel and their structures were established by the ³¹P-, ¹H-, and ¹³C-n.m.r. data. Each anomer gave two ³¹P signals with similar chemical shifts ($\delta - 0.8$ and -0.6 p.p.m.) indicative⁸ of the absence of geometrical isomerism at phosphorus. The $J_{H-1,H-2}$ values were 3.4 and 4.9 Hz. That the P–O-1 bond in 3 had been cleaved was indicated by the C-1 signals that were now singlets and the absence of coupling between H-1 and P. The upfield shift of the signals for C-1 is characteristic of pyranosyl chlorides⁹.



EXPERIMENTAL

Experiments with trivalent phosphorus derivatives were carried out under dry nitrogen. T.l.c. was performed on Silufol-254 and column chromatography on silica gel L 40–100, using benzene–1,4-dioxane, 3:1 (A) and 1:1 (B); hexane–1,4-dioxane, 3:1 (C) and 1:1 (D); and E, 19:1 chloroform-methanol. ³¹P- (32.4 MHz; external aqueous 85% H_3PO_4), ¹³C- (75.4 MHz), and ¹H-n.m.r. spectra (400.1 MHz; external Me₄Si) were obtained with Bruker WP-80, WM-300, and AM-400 spectrometers, respectively. Optical rotations were determined with a DIP-360 polarimeter. X-Ray analysis was performed with a CAD-4 Enraf–Nonius diffractometer and the detailed results will be published elsewhere.

 α -D-Xylopyranose 1,2,4-phosphite (1) and some derivatives. — To a stirred solution of D-xylose (1.50 g) in 1,4-dioxane (50 mL) and pyridine (20 mL) was added a solution of tris(3,5-dimethylpyrazolyl) phosphite (1.58 g, 0.5 mol) in 1,4-dioxane. The mixture was stirred for 0.5 h at room temperature and more phosphite (0.79, 0.40, and 0.40 g) was added every 0.5 h. After a further 0.5 h, the mixture was filtered, and the solvent was evaporated *in vacuo*. Column chromatography (solvent A) of the residue gave 1 (0.71 g, 40%), isolated as a syrup, $[\alpha]_{D}^{20}$ + 50° (c 0.5, dioxane), R_{F} 0.60 (solvent A), 0.70 (solvent B) (Found: C, 33.51; H, 3.72; P, 17.53. C₅H₇O₅P calc.: C, 33.72; H, 3.96; P, 17.39%). Conventional acetylation of 1 (0.35 g) with acetic anhydride (0.28 mL) and pyridine (15 mL), with column chromatography (solvent A) of the product, gave the 3-acetate 2, isolated as a syrup (0.38 g, 88%), $[\alpha]_{D}^{20}$ +8° (c 1.2, chloroform), R_{F} 0.82 (Found: C, 38.00; H, 3.98; P, 14.22. $C_7H_9O_6P$ calc.: C, 38.20; H, 4.12; P, 14.07%).

Conventional benzoylation of 1 (0.35 g) with triethylamine (0.28 mL), pyridine (5 mL), and benzoyl chloride (0.24 mL), with column chromatography (solvent C) of the product, gave the 3-benzoate 3 (0.40 g, 73%), m.p. 74–75° (from benzene), $[\alpha]_{D}^{20} + 36^{\circ}$ (c 2.7, chloroform), $R_{\rm F}$ 0.49 (Found: C, 50.92; H, 3.80; P, 11.10. C₁₂H₁₁O₆P calc.: C, 51.08; H, 3.93; P, 10.98%).

Conventional reaction of 1 (0.35 g) in pyridine (5 mL) with phenyl isocyanate (0.22 mL), followed by column chromatography (solvent A) of the product, gave the 3-phenylcarbamate 4 (0.41 g, 71%), isolated as a syrup, $[\alpha]_{D}^{20} + 32^{\circ}$ (c 2.0, chloroform), R_{F} 0.74 (Found: C, 48.35; H, 3.95; N, 4.50; P, 10.52. $C_{12}H_{12}NO_{6}P$ calc.: C, 48.50; H, 4.07; N, 4.71; P, 10.42%).

To rhodium(I) dicarbonyl acetylacetonate (0.26 g) was added a solution of 2 (0.22 g) in benzene (5 mL) with stirring at room temperature. Part of the solvent was evaporated and the (acetylacetonato)carbonylrhodium complex 5 (0.34 g, 75%) was precipitated with hexane as a syrup, $[\alpha]_{D}^{20} + 36^{\circ}$ (c 0.8, benzene), R_{F} 0.71 (solvent A) (Found: C, 34.41; H, 3.40; P, 7.02. $C_{13}H_{16}O_{9}PRh$ calc.: C, 34.69; H, 3.58; P, 6.88%).

The (acetylacetonato)carbonylrhodium complex **6** (0.41 g, 80%), prepared from **3** (0.28 g) as described for **5**, had m.p. 75–80° (dec.) (from hexane), $[\alpha]_{D}^{20}$ +34.5° (*c* 2.3, benzene), R_{p} 0.88 (solvent *A*) (Found: C, 42.10; H, 3.36; P, 5.90. C₁₈H₁₈O₉PRh calc.: C, 42.21; H, 3.54; P, 6.05%).

3-O-Acetyl- α -D-xylopyranose 1,2,4-phosphate (9). — Through a solution of 2 (0.44 g) in dichloromethane (10 mL) at -70° was passed a stream of ozone (2% by vol.) at 5 L/h until the solution turned blue. ³¹P-N.m.r. spectroscopy revealed the quantitaive formation of the ozonide 7 (³¹P, δ - 32) which, on raising the temperature to 20–25°, polymerised almost completely. The mixture was filtered and the solvent was evaporated to leave 9 as a syrup, $[\alpha]_{p}^{20}$ + 23° (c 0.7, chloroform), R_{p} 0.43 (solvent A), 0.54 (solvent B) (Found: C, 35.42; H, 3.70; P, 13.24. C₇H₉O₇P calc.: C, 35.61; H, 3.84; P, 13.12%).

3-O-Benzoyl- α -D-xylopyranose 1,2,4-phosphate (10). — Following the procedure for 9, a stream of ozone was bubbled through a solution of 3 (0.28 g) in dichloromethane (10 mL) at -70° . The temperature of the mixture was raised to 20–25°, the mixture was filtered, and the solvent was evaporated to leave 10 as a syrup, $[\alpha]_{D}^{20}$ +18° (c 0.8, chloroform), $R_{\rm F}$ 0.50 (solvent A) (Found: C, 48.12; H, 3.65; P, 10.50. C₁₂H₁₁O₇P calc.: C, 48.34; H, 3.72; P, 10.39%).

3-O-Acetyl-D-xylopyranose 4-phosphate (13). — To a solution of 2 (0.44 g) in 1,4-dioxane (5 mL) was added aqueous 30% hydrogen peroxide (0.34 mL), and the mixture was stored for 0.5 h. ³¹P-N.m.r. spectroscopy revealed the quantitative formation of the 1,4-phosphate 11 (³¹P, δ – 4.5), which was hydrolysed to give 13 $\alpha\beta$ (³¹P, δ – 1.0). The solvent was evaporated *in vacuo* and the residue was washed with hexane to yield 13 $\alpha\beta$ (0.49 g, 90%), as a syrup, R_p 0.20 (solvent *E*) (Found: C, 30.65; H, 4.75; P, 11.58. C₇H₁₃O₉P calc.: C, 30.89; H, 4.82; P, 11.38%).

3-O-Benzoyl-D-xylopyranose 4-phosphate (14). — To a solution of 3 (0.28 g) in 1,4-dioxane (5 mL) was added aqueous 30% hydrogen peroxide (0.17 mL). The mixture was stored for 0.5 h. ³¹P-N.m.r. spectroscopy revealed the quantitative formation of 10, which was hydrolysed to give $14\alpha\beta$ (³¹P, δ – 0.6). The solvent was evaporated *in vacuo* and the residue was washed with hexane to yield $14\alpha\beta$ (0.29 g, 88%), as a syrup, R_r 0.20 (solvent E) (Found: C, 42.95; H, 4.35; P, 9.38. C₁₂H₁₅O₉P calc.: C, 43.12; H, 4.52; P, 9.27%).

3-O-Acetyl- α -D-xylopyranose 1,2,4-selenophosphate (15). — To a solution of 2 (0.44 g) in 1,4-dioxane (5 mL) was added powdered selenium (0.16 g), the mixture was stirred for 8 h at 100°, then filtered, and the solvent was evaporated *in vacuo*. Column chromatography (solvent C, then solvent D) of the residue gave 15 (0.43 g, 72%), isolated as a syrup, $[\alpha]_{p}^{20} + 11^{\circ}$ (c 2.5, chloroform), R_{p} 0.18 (solvent C), 0.73 (solvent D) (Found: C, 28.05; H, 2.91; P, 10.11. $C_{7}H_{9}O_{6}PSe$ calc.: C, 28.11; H, 3.03; P, 10.36%).

3-O-Benzoyl- α -D-xylopyranose 1,2,4-selenophosphate (16). — Following the procedure for 15, 3 (0.56 g) was treated with selenium (0.16 g) in 1,4-dioxane. Column chromatography (solvent C) of the product gave 16 (0.49 g, 68%), m.p. 157–158° (from benzene), $[\alpha]_{D}^{20}$ + 30° (c 1.8, chloroform), R_{F} 0.25 (solvent C), 0.71 (solvent D) (Found: C, 39.72; H, 2.92; P, 8.72. C₁₂H₁₁O₆PSe calc.: 39.11; H, 3.07; P, 8.58%).

3-O-Acetyl- α -D-xylopyranose 1,2,4-thiophosphate (17). — A mixture of 2 (0.44 g) and sulfur (0.10 g) in 1,4-dioxane was stirred for 12 h at 100°, then filtered, and the solvent was evaporated *in vacuo*. Column chromatography (solvent C) of the residue gave 17 (0.30 g, 60%), isolated as a syrup, $[\alpha]_{D}^{20}$ + 18° (c 0.3, chloroform), R_{F} 0.40 (Found: C, 33.25; H, 3.52; P, 12.35; S, 12.60. C₇H₉O₆PS calc.: C, 33.34; H, 3.60; P, 12.28; S, 12.72%.

3-O-Benzoyl- α -D-xylopyranose 1,2,4-thiophosphate (18). — Following the procedure for 17, 3 (0.28 g) was treated with sulfur (0.60 g) in 1,4-dioxane. Column chromatography (solvent C) of the product gave 18 (0.19 g, 60%), m.p. 140–142° (from benzene), $[\alpha]_p^{20} + 14^\circ$ (c 1.3, chloroform), R_p 0.25 (solvent C), 0.72 (solvent D) (Found: C, 45.72; H, 3.39; P, 10.04; S, 9.95. $C_{12}H_{11}O_6PS$ calc.: C, 45.84; H, 3.53; P, 9.86; S, 10.21%).

3-O-Phenylcarbamoyl- α -D-xylopyranose 1,2,4-thiophosphate (19). — Following the procedure for 17, 4 (0.30 g) was treated with sulfur (0.60 g) in 1,4-dioxane. Column chromatography (solvent D) of the product gave 19 (0.21 g, 63%), isolated as a syrup, $[\alpha]_{D}^{20} + 22^{\circ}$ (c 1.6, chloroform), R_{F} 0.64 (Found: C, 43.61; H, 3.49; N, 4.02; P, 9.58; S, 9.60. $C_{12}H_{12}NO_{6}PS$ calc.: C, 43.77; H, 3.67; N, 4.25; P, 9.41; S, 9.74%).

Triethylammonium 3-O-benzoyl- α -D-xylopyranose 1,4-selenophosphate (20). — To a solution of 16 (0.72 g) in 1,4-dioxane (5 mL) were added water (0.06 mL) and triethylamine (0.28 mL). The mixture was stirred for 4 h at 20° and the solvents were then evaporated *in vacuo* to leave 20 as a syrup, $[\alpha]_{D}^{20} - 2.5^{\circ}$ (c 0.8, chloroform), R_{F} 0.33 (solvent E) (Found: C, 44.85; H, 5.70; N, 2.72; P, 6.60. C₁₈H₂₈NO₇PSe calc.: C, 45.01; H, 5.88; N, 2.92; P, 6.45%).

Triethylammonium 3-O-benzoyl- α -D-xylopyranose 1,4-thiophosphate (21). — To a solution of 18 (0.63 g) in 1,4-dioxane (5 mL) were added water (0.06 mL) and triethylamine (0.28 mL). The mixture was stored at 20° for 6 h and the solvents were then

evaporated *in vacuo* to leave **21** as a syrup, $[\alpha]_{D}^{20} + 9^{\circ} (c \ 0.57, \text{pyridine}), R_{F} \ 0.31$ (solvent *E*) (Found: C, 49.62; H, 5.60; P, 3.28; S, 7.28. C₁₈H₂₈NO₇PS calc.: C, 49.88; H, 6.51; N, 3.23; P, 7.15%).

3-O-Benzoyl- α -D-xylopyranosyl chloride 2,4-phosphoropiperididate (23). — Dry chlorine was bubbled through a solution of 3 (0.28 g) in dichloromethane (10 mL) at -10 to -15° until it turned green. The excess of chlorine was removed by evaporating the solvent *in vacuo*. To a stirred solution of the residue in dichloromethane at 0° was added piperidine (0.21 mL). The mixture was stored for 2 h at 20°, then filtered, and the solvent was evaporated. Column chromatography (solvent A) of the residue gave 23 β (0.09 g, 22.5%), isolated as a syrup, $R_{\rm F}$ 0.60, $[\alpha]_{\rm D}^{20} - 33^{\circ}$ (c 0.9, chloroform). Eluted second was 23 α (0.09 g, 22.5%), $R_{\rm F}$ 0.40, m.p. 83–84° (from benzene), $[\alpha]_{\rm D}^{20} - 50^{\circ}$ (c 1.3, chloroform) (Found: C, 50.65; H, 5.05; N, 3.25; P, 7.92. C₁₇H₂₁C1NO₆P calc.: C, 50.82; H, 5.27; N, 3.49; P, 7.71%).

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