

A PREGNANE ESTER DIGLYCOSIDE FROM *HEMIDESMUS INDICUS*

KANCHAN OBERAI, MAHESHWARI P. KHARE and ANAKSHI KHARE

Department of Chemistry, University of Lucknow, Lucknow, India

(Revised received 20 February 1985)

Key Word Index—*Hemidesmus indicus*; Asclepiadaceae; desinine; steroid; pregnane ester diglycoside.

Abstract—A new pregnane ester diglycoside named desinine has been isolated from the dried twigs of *Hemidesmus indicus*. On the basis of chemical and spectroscopic evidence, its structure has been established as drevogenin B-3-*O*- β -D-oleandropyranosyl-(1 \rightarrow 4)- β -D-oleandropyranoside.

INTRODUCTION

In the roots of *Hemidesmus indicus* R. Br. (family: Asclepiadaceae), the presence has been reported of resin acid, fatty acids, tannins, saponins, an oil containing *p*-methoxysalsaldehyde [1, 2], lupeol, sitosterol and amyryns [3]. In a reinvestigation of the chemical constituents, shade-dried twigs of the plant were extracted which, on repeated CC, afforded a novel diglycoside which we have named desinine. This paper describes the structure elucidation of this glycoside.

RESULTS AND DISCUSSION

Desinine (**1**) mp 115–118°, $[\alpha]_D$ 0°, $C_{37}H_{58}O_{12}$, was obtained from the chloroform-soluble glycoside mixture by CC over silica gel. It gave a positive Liebermann–Burchardt [4] colour reaction and characteristic colour tests for 2-deoxy sugars in the xanthidrol [5, 6] and Keller–Kiliani [7] reactions, indicating it to be a steroidal glycoside of a 2-deoxy sugar.

The 1H NMR spectrum of **1** at 400 MHz showed two methoxy group singlets at δ 3.45 and 3.35, two secondary methyl group doublets ($J = 6$ Hz) at δ 1.23 and 1.20 and the equatorial and axial protons of four C-2 methylene protons of two 2-deoxy sugar units as two sets of multiplets in the regions δ 2.28–2.46 and 1.72–1.88. This suggested it to be a diglycoside of monomethoxy-2,6-dideoxyhexoses for which the anomeric protons could be assigned to the double doublet for two protons at δ 4.88 ($J = 9, 2.5$ Hz) presumably in the axial conformation. This glycoside also contained the signal of one acetyl group at δ 2.10, possibly present in the genin moiety.

The mass spectrum of **1** was also consistent with its diglycoside structure. Although the mass spectrum of **1** did not exhibit the $[M]^+$ expected at m/z 694, there were peaks at m/z 270, 244, 238 and 213 which originated from the disaccharide fragment after the loss of acetaldehyde, methanol and a water molecule(s). The presence of an acetyl group and a keto methyl chain was substantiated by the peaks corresponding to the loss of acetic acid and a keto methyl chain from different fragments.

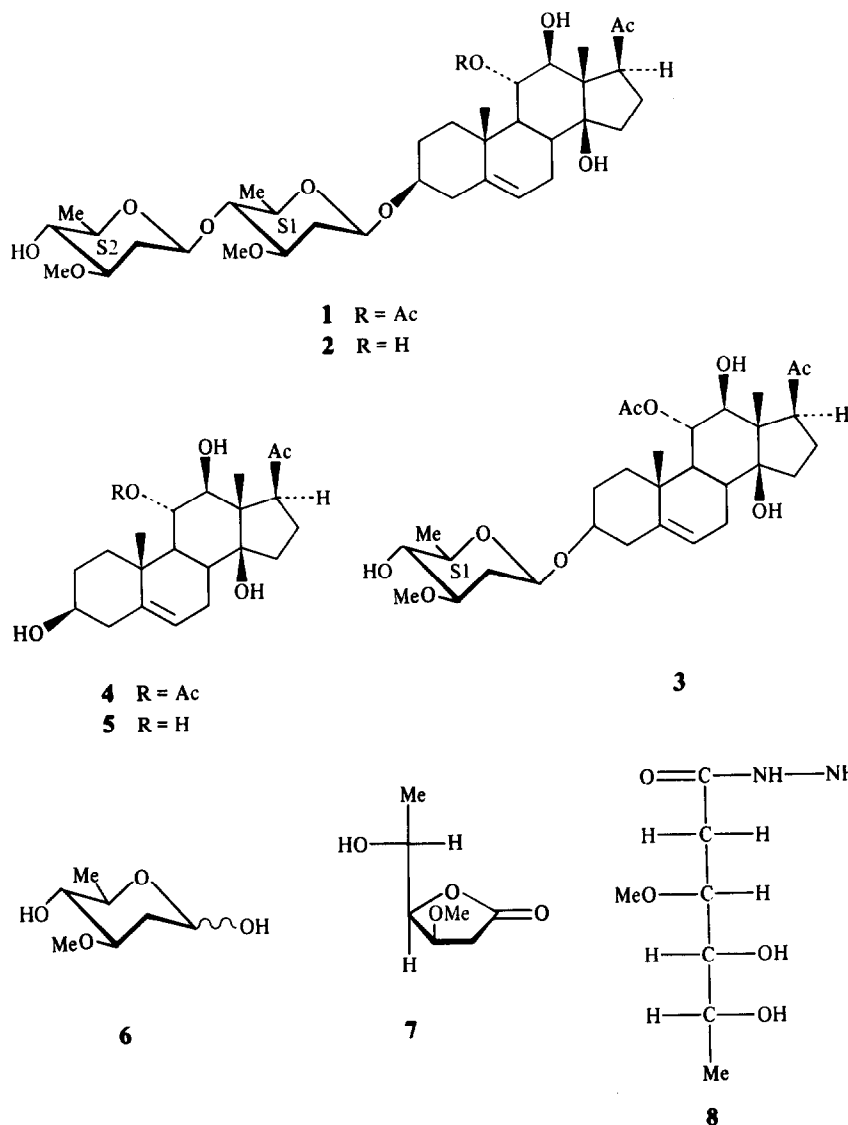
The inertness of **1** to sodium periodate, in contrast to its deacetylated product **2**, suggested that one of the hydroxyl groups, presumably present in the basic genin moiety of **1** in a vicinal diol arrangement, was acetylated. Mild acid

hydrolysis of **1** with 0.025 M sulphuric acid afforded an acetyl genin, **4**, mp 238–240°, $[\alpha]_D + 56^\circ$, $C_{23}H_{34}O_6$, which, on deacetylation with alkali, gave a mixture of two products having very similar mobility. One was obtained from the mixture as a crystalline product, **5**, mp 185–188°, $[\alpha]_D + 32^\circ$, $C_{21}H_{32}O_5$, identified as drevogenin P [8, 9], thus leading to the conclusion that **4** is 11- or 12-mono-*O*-acetyldrevogenin P. A survey of the mps and ORDs of mono-*O*-acetyldrevogenin P indicated **4** to be drevogenin B [10, 11] (11 α -*O*-acetyl, 3 β , 12 β , 14 β -trihydroxy- Δ^5 -pregnen-20-one) which was finally confirmed by its comparison with the authentic material (TLC and undepressed mmp). The 11-*O*-acetyl position in the genin moiety in **1** conforms with its 1H NMR spectrum consisting of a lower field one proton triplet at δ 5.14 ($J = 8$ Hz) attributable to a C-11 methine proton and the higher field one-proton doublet at δ 4.44 ($J = 8$ Hz) assigned to a methine proton at C-12 bearing a free hydroxyl group.

The syrupy sugar **6**, $[\alpha]_D -14^\circ$, was identified as D-oleandrose [12, 13] (2,6-dideoxy-3-*O*-methyl-D-arabinohexose) by comparison of ORD and mobility on PC. Bromine water oxidation of **6** gave a lactone **7** which, with phenylhydrazine, afforded the known D-oleandronic acid phenylhydrazide **8**, mp 132–134° which was characterized by comparison with the authentic material (mmp and IR spectrum) [12, 13].

The difference, $C_{14}H_{24}O_6$, between the molecular formulae of **1** and **4** and the identification of D-oleandrose as the only sugar in the acid hydrolysate clearly indicated that two oleandrose units are present in desinine, suggesting it to be a di-oleandroside. For convenience, the two sugar units in **1** are designated as S1 and S2. A double-resonance experiment was helpful in confirming the assignment of the anomeric protons at δ 4.88. Irradiation of the signal at 1952 Hz resulted in the collapse of the C-2 methylene multiplets, confirming that this signal represented the anomeric protons of 2-deoxy sugar units. The configuration of the anomeric protons in S1 and S2 could be derived easily from the large coupling constant ($J = 9$ Hz) of their signal at δ 4.88, attributed to their axial configuration indicating the sugar units to be present in a 4C_1 (D) conformation and linked through β -D-(1 \rightarrow 4) glycosidic linkage [14].

More direct chemical support that **1** is drevogenin B-dioleandroside was provided by the results of its very mild



acid (0.001 M) hydrolysis [15]. Under these conditions 1 afforded, after 6 days, four spots on TLC. The most polar spot (R_f 0.18) had the same mobility as the starting material 1. The least polar spot (R_f 0.59) was identical in mobility to the genin 4. The spot at R_f 0.31 was found to have identical mobility with oleandrose (6) and the other vanillin perchloric acid positive spot (R_f 0.51), still containing the 2-deoxy sugar, was presumed to be the monoglycoside 3. The hydrolysis was complete in ten days when the hydrolysate exhibited only two spots identical in mobility to sugar 6 and genin 4.

In the light of the foregoing evidence, the structure of desinine (1) was established as drevogenin B-3-*O*- β -D-oleandropyranosyl-(1 \rightarrow 4)- β -D-oleandropyranoside.

EXPERIMENTAL

Mps were determined on a Boetius micro mp apparatus and are uncorr. MS were recorded with an AEI MS-30 instrument. ORs were measured in a 0.1 m long tube with a Jasco-Dip 180 automatic polarimeter. The ^1H NMR spectrum (CDCl_3) was

recorded on a 400 MHz (Brüker) spectrometer with TMS as int. standard. Sugars were made visible with 50% aq. H_2SO_4 for TLC and vanillin- HClO_4 reagent for PC. PC was performed using $\text{C}_6\text{H}_5\text{Me}-\text{BuOH}$ (4:1) satd with H_2O .

Plant extraction. The twigs of *H. indicus* were collected from the National Botanical Garden, Lucknow, India. The identity of the plant was confirmed by Dr. S. L. Kapoor, Systemic Botanist, National Botanical Research Institute, Lucknow, India where a voucher specimen was deposited (voucher No. 67957). Shade-dried powdered twigs (5 kg) of *H. indicus* were extracted by the method employed for pregnane glycosides [16] using 50–95% aq. EtOH. The EtOH extracts were combined and concd under red. pres. and without treatment with $\text{Pb}(\text{OH})_2$, the concentrate was exhaustively extracted successively with petrol, Et_2O , CHCl_3 , $\text{CHCl}_3-\text{EtOH}$ (4:1) and $\text{CHCl}_3-\text{EtOH}$ (3:2). Detailed examination of the CHCl_3 extract was undertaken. Although it did not resolve satisfactorily on TLC, it afforded one pure compound, given the name of desinine (1), by repeated CC over silica gel.

Desinine (1). Mp 115–118°, $[\alpha]_D^{25}$ 0°, $\text{C}_{37}\text{H}_{58}\text{O}_{12}$. (Found: C, 63.75; H, 7.20. $\text{C}_{37}\text{H}_{58}\text{O}_{12}$ requires: C, 63.97; H, 7.35 %.) It gave a

violet colour in the Liebermann–Burchardt test, a pink colour in the xanthidrol and a blue colour in the Keller–Kiliani reactions. $^1\text{H NMR}$ (400 MHz): δ 5.38 (1H, *m*, H-6), 5.14 (1H, *t*, *J* = 8 Hz, H-11), 4.88 (2H, *dd*, *J* = 9, 2.5 Hz, H-1 in S1 and S2), 4.44 (1H, *d*, *J* = 8 Hz, H-12), 3.76–3.88 (2H, *m*, H-5 in S1 and S2), 3.66–3.74 (2H, *m*, H-3 in S1 and S2), 3.45 (3H, *s*, OMe), 3.35 (3H, *s*, OMe), 3.16–3.24 (2H, *m*, H-4 in S1 and S2), 2.28–2.46 (2H, *m*, H-2e in S1 and S2), 2.18 (3H, *s*, COMe), 2.10 (3H, *s*, OAc), 1.72–1.88 (2H, *m*, H-2a in S1 and S2), 1.28 (3H, *s*, Me-18), 1.23 (3H, *d*, *J* = 6 Hz, sec. Me), 1.20 (3H, *d*, *J* = 6 Hz, sec. Me), 1.02 (3H, *s*, Me-19). MS *m/z* (rel. int.) glycoside fragments: 414 [*M* – S2 – HOAc – COMe – Me] $^+$ (16), 397 [*M* – S2 – HOAc – COMe – MeOH] $^+$ (34), 396 [*M* – S2 – HOAc – MeCHO – MeOH] $^+$ (56), 381 [*M* – S2 – HOAc – COMe – MeCHO – Me] $^+$ (12), 357 [*M* – disaccharide fragment (288) – 2OH – Me] $^+$ (5); genin fragments: 406 (genin) not observed, 329 [406 – HOAc – OH] $^+$ (9.8), 314 [406 – HOAc – OH – Me] $^+$ (75), 296 [329 – H₂O – Me] $^+$ (55), 281 [296 – Me] $^+$ (17), 244 [329 – C₃H₅O] $^+$ (92), 238 [281 – COMe] $^+$ (5); sugar fragments: 306 (disaccharide) not observed, 244 [306 – H₂O – MeCHO] $^+$ (92), 270 [306 – 2H₂O] $^+$ (4), 238 [270 – MeOH] $^+$ (5), 213 [306 – MeCHO – MeOH – OH] $^+$ (2).

Mild hydrolysis of 1 with acid. To a soln of crystalline 1 (18 mg) in 80% aq. 1,4-dioxane (1.2 ml) was added 0.05 M H₂SO₄ (1.2 ml) and the soln was warmed for 30 min at 50°. Dioxane was then removed under red. pres. The aq. portion was repeatedly extracted with CHCl₃ and the organic layer was washed in turn with H₂O, 1 M Na₂CO₃, again with H₂O, dried over Na₂SO₄ and evaporated to dryness. Crystallization with MeOH–Me₂CO afforded genin 4 (9 mg), mp 238–240°, $[\alpha]_D^{25} + 56.2^\circ$ (MeOH; *c* 0.18). (Found: C, 67.84; H, 8.20. C₂₃H₃₄O₆ requires: C, 67.98; H, 8.37%). The aq. hydrolysate was neutralized with freshly prepared BaCO₃, filtered and concd under red. pres. to afford the syrupy sugar 6 (6 mg), $[\alpha]_D^{25} - 14.2^\circ$ (MeOH; *c* 0.12). It gave a pink colouration in the xanthidrol and a blue colouration in the Keller–Kiliani reactions. It had identical mobility on PC as oleandrose and its ORD was comparable to D-oleandrose.

Oxidation of compound 6 with bromine water. A soln of 6 (4 mg) in H₂O (0.6 ml) was mixed with Br₂ (10 μ l) and shaken in the dark for 24 hr at room temp. The excess Br₂ was then removed under red. pres. The acidic mixture was neutralized with freshly prepared Ag₂CO₃ and filtered. H₂S was passed through the filtrate to remove Ag $^+$ ions followed by filtration. The filtrate was evaporated to dryness under red. pres. yielding the syrupy lactone 7 (3.2 mg) which gave a violet colour in the spot test with NH₂OH–FeCl₃ reagent [17].

D-Oleandronic acid phenyl hydrazide (8). A soln of lactone 7 (3 mg) in absolute EtOH (0.5 ml) was mixed with freshly distilled phenylhydrazine (0.5 ml) and heated for 30 min at 100°. The viscous mass was cooled and repeatedly triturated with absolute Et₂O (to remove excess of phenylhydrazine) to yield D-oleandronic acid phenylhydrazide which crystallized from MeOH–Et₂O as colourless needles (1.2 mg) mp 132–134°. It had a superimposable IR spectrum and gave no depression in mmp when compared with the authentic material (lit. [12, 13] mp 135°).

Very mild hydrolysis of 1 with acid. To a soln of 1 (2 mg) in aq. 1,4-dioxane (1:1) (0.2 ml) was added 0.001 M H₂SO₄ (0.2 ml) and the soln was kept at room temp. After 6 days the reaction mixture showed four spots on TLC. By co-chromatography the most polar spot (*R_f* 0.18) was identified as the starting material (1). The other spot (*R_f* 0.31) was sugar 6 and the least polar spot (*R_f* 0.59) the genin 4. The last vanillin–HClO₄ positive spot (*R_f* 0.51) which still contained the 2-deoxy sugar was, presumably, the monoglycoside 3.

Alkaline hydrolysis of 4. Compound 4 (7 mg) was dissolved in 5% methanolic KOH (1.5 ml) and heated for 30 min. After adding H₂O (0.8 ml), MeOH was removed under red. pres. The aq. concentrate was extracted with CHCl₃, dried over Na₂SO₄, filtered and evaporated to dryness to yield 5 which crystallized from MeOH–Et₂O as colourless prisms (5 mg), mp 185–188°, $[\alpha]_D^{25} + 32.2^\circ$. (Found: C, 69.15; H, 8.70. C₂₁H₃₂O₅ requires: C, 69.23; H, 8.79%). It was identified as drevogenin P (lit. mp 185–190°, $[\alpha]_D^{25} + 34.4^\circ$) by comparison with the authentic sample on TLC, ORD and undepressed mmp.

REFERENCES

- Sanjiva Rao, B., Subramanian, K. S. and Kelkar, N. C. (1938) *Proc. Soc. Biol. Chemists, India* 3, 35.
- Rao, B. S., Subramanian, K. S. and Kelkar, N. C. (1938) *Chem. Abstr.* 32, 8696.
- Chatterjee, R. C. and Bhattacharya, B. K. (1955) *J. Indian Chem. Soc.* 32, 485.
- Abisch, E. and Reichstein, T. (1960) *Helv. Chim. Acta* 43, 1844.
- Barton, G. M., Evans, R. S. and Gardner, J. A. F. (1952) *Nature* 170, 249.
- Tschesche, R., Grimmer, G. and Seehofer, F. (1953) *Chem. Ber.* 86, 1235.
- Nagata, W., Tamm, C. and Reichstein, T. (1965) *Helv. Chim. Acta* 48, 847.
- Sauer, H. H., Weiss, E. and Reichstein, T. (1965) *Helv. Chim. Acta* 48, 857.
- Sauer, H. H., Weiss, E. and Reichstein, T. (1966) *Helv. Chim. Acta* 49, 1632.
- Winkler, R. E. and Reichstein, T. (1954) *Helv. Chim. Acta* 37, 721.
- Bhatnagar, A. S., Kaufmann, H., Stocklin, W. and Reichstein, T. (1968) *Helv. Chim. Acta* 51, 117.
- Blindenbacher, F. and Reichstein, T. (1948) *Helv. Chim. Acta* 31, 2061.
- Renkonen, O., Schindler, O. and Reichstein, T. (1959) *Helv. Chim. Acta* 42, 182.
- Allgeier, H. (1968) *Helv. Chim. Acta* 51, 311.
- Kapur, B. M., Allgeier, H. and Reichstein, T. (1967) *Helv. Chim. Acta* 50, 2171.
- Schaub, F., Kaufmann, H., Stocklin, W. and Reichstein, T. (1968) *Helv. Chim. Acta* 51, 738.
- Abdel Akher, M. and Smith, F. (1951) *J. Am. Chem. Soc.* 73, 5859.