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Note

Syntheses of spacer-armed carbohydrate components of the *Mycobacterium avium* serocomplex serovar 8⁻¹

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Abstract

p-Nitrophenyl glycosides of 3-*O*-Me- β -D-Glcp-(1 \rightarrow 3)- α -L-Rhap, α -L-Rhap-(1 \rightarrow 2)-6-deoxy- α -L-Talp, and 3-*O*-Me- β -D-Glcp-(1 \rightarrow 3)- α -L-Rhap-(1 \rightarrow 2)-6-deoxy- α -L-Talp have been prepared, related to *Mycobacterium avium*. Various glycosylation methods have been used for the formation of the interglycosidic linkages. The p-nitrophenyl derivatives were converted into p-isothiocyanatophenyl glycosides, capable of forming neoglycoproteins. © 1997 Elsevier Science Ltd.

Keywords: Spacer-armed glycosides; Oligosaccharide synthesis; Mycobacterium

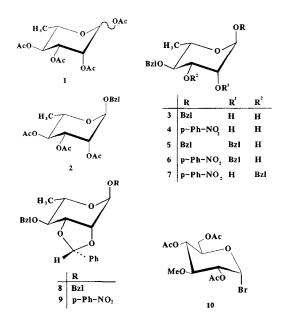
Over the last decade, a number of oligosaccharides related to Mycobacteria have been synthesized in our laboratory [1–5]. It is known that *Mycobacterium tuberculosis* and *M. leprae* are responsible for human tuberculosis and leprosy, respectively. Recently, there has been growing interest in the *Mycobacterium avium–M. intracellulare–M. scrofulaceum* (MAIS) serocomplex because several serovariants, as opportunistic pathogens, cause infections among people with acquired immunodeficiency syndrome (AIDS) [6]. The surface glycolipid of MAIS serovariant 8 has the following oligosaccharide component [7]: 4,6-O-Pyr-3-O-Me- β -D-Glcp-(1 \rightarrow 3)- α -L-Rhap-(1 \rightarrow 2)-6deoxy- α -L-Talp. This pyruvylated trisaccharide has been prepared in our laboratory by a 2 + 1 block synthesis as the p-trifluoroacetamidophenyl glycoside [5]. Now we describe the spacer-armed nonpyruvylated di- and trisaccharides, as p-isothiocyanatophenyl glycosides.

For the preparation of the 3-*O*-Me- β -D-Glcp-(1 \rightarrow 3)-L-Rhap disaccharide glycoside, first suitably protected precursors were synthesized. Benzyl 4-*O*-benzyl- α -L-rhamnopyranoside (3) [8] and p-nitrophenyl 4-*O*-benzyl- α -L-rhamnopyranoside (4) [3] were prepared from the corresponding 2,3-*O*-isopropylidene acetals by acid hydrolysis. One compound in the well-known reaction sequence, namely, benzyl 2,3,4-tri-*O*-acetyl- α -L-rhamnopyranoside (2) [9], was synthesized directly from tetra-*O*-acetyl-L-rhamnose (1) in dry dichloromethane using boron trifluoride etherate as the promoter. It is known that 3 can be

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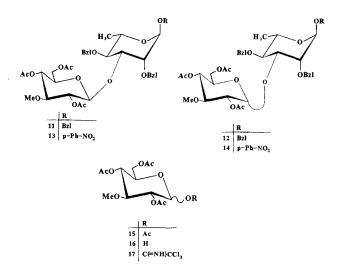
¹ Dedicated to Professor András Lipták on the occasion of his 60th birthday.

converted into the 2,4-di-O-benzyl derivative (5) by employing a phase-transfer-catalysed reaction [10] in a good yield. However, using the same conditions, compound 4 with benzyl bromide gave a mixture of the corresponding tri-O-benzyl, 2,4-di-O-benzyl (6), and 3,4-di-O-benzyl (7) derivatives in a molar ratio of 2:3:3 (TLC). Better selectivities were observed with a kinetically controlled benzylidenation [11] of **3** and 4, yielding benzyl endo-2,3-O-benzylidene (8) *p*-nitrophenyl *endo*-2,3-*O*-benzylidene- α -Land rhamnopyranoside (9), respectively. Reductive ringopening of 8 and 9 with the lithium aluminium hydride-aluminium chloride reagent [8] at ambient temperature then resulted almost exclusively in the 2,4-di-O-benzyl isomers 5 and 6, respectively. It has to be noted that in the case of 9 a prolonged reaction time of 30 min causes the reduction of the nitro group.



Glycosylation of 5 with 10 [12] under *Helferich* conditions gave a mixture of 11 and 12 in a molar ratio of 10:1, demonstrating a non-exclusive stereose-lectivity. Almost the same result was observed in the case of 6, whereby a mercuric bromide-promoted reaction [13] with 10 yielded a 5:1 mixture of 13 and 14.

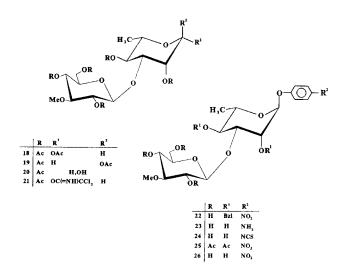
In order to get a better yield and a higher stereoselectivity in the preparation of the disaccharide derivative **11**, the imidate procedure [14] was applied. 1,2,4,6-Tetra-*O*-acetyl-3-*O*-methyl-D-glucopyranose (**15**) [12] was converted into 2,4,6-tri-*O*-acetyl-3-*O*methyl-D-glucopyranose (**16**) using hydrazine acetate [15], and the corresponding imidate **17** was produced using trichloroacetonitrile and 1,8-diazabicyclo-[5.4.0]undec-7-ene [16]. Then, the glycosyl donor 17 was coupled with 5 in the presence of trimethylsilyl triflate (0.2 equiv) and 4 Å molecular sieves (pellets) at -40 °C. It is noteworthy that *powdered* molecular sieves removed the catalyst from the reaction mixture. The reaction was stereospecific, but a little by-product was formed with higher chromatographic mobility.



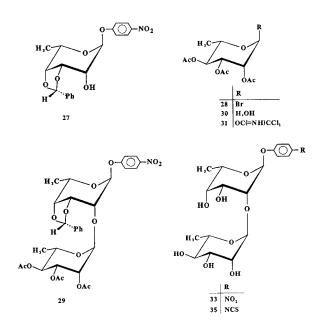
The silver triflate-promoted [17] reaction of 5 with 10 gave exclusively disaccharide derivative 11 in a good yield. Hydrogenolysis (Pd-C, H₂) of 11 and conventional acetylation of the product afforded $(2,4,6-\text{tri-}O-\text{acetyl-}3-O-\text{methyl-}\beta-D-\text{glucopyranosyl}) (1 \rightarrow 3)-1,2,4-\text{tri-}O-\text{acetyl-}L-\text{rhamnopyranose}$ as a mixture of the anomers 18 and 19, and both compounds were isolated and characterised. Compound 18 was converted into 20 by hydrazine acetate, then the corresponding imidate 21 was formed by treatment with trichloroacetonitrile.

In order to prepare 3-O-Me- β -D-Glcp-(1 \rightarrow 3)-L-Rhap with a p-isothiocyanatophenyl aglycone, compound **13** was deacylated with sodium methoxide (2.5 equiv) in methanol (\rightarrow **22**) [18]. Reduction of the nitro and benzyl groups in a one pot reaction (Pd-C, H₂, 80% ethanol) gave **23** with 10% impurity. The crude product was reacted with thiophosgene in the presence of BaCO₃ (\rightarrow **24**) [19], but the impurity could not be removed either by chromatography or by crystallization. Following an other route, pure **24** was prepared from **18** by reaction with *p*-nitrophenol (boron trifluoride etherate; dichloromethane, \rightarrow **25**) and subsequent deacylation, yielding firstly the *p*-nitrophenyl glycoside **26**. Then, reduction of the nitro

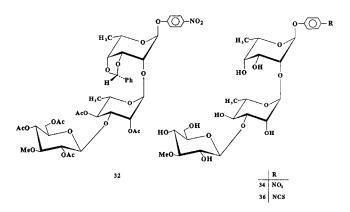
group and reaction with thiophosgene gave the spacer-armed disaccharide 24.



The silver triflate-promoted glycosylation of *p*nitrophenyl *endo*-3,4-*O*-benzylidene-6-deoxy- α -Ltalopyranoside (27) with 2,3,4-tri-*O*-acetyl- α -Lrhamnopyranosyl bromide (28) at -40 °C gave a complex reaction mixture. In order to prepare the disaccharide 29, glycosyl donor 28 was converted into imidate 31 by reaction with water in the presence of silver oxide (\rightarrow 30), followed by treatment with trichloroacetonitrile. Coupling of 27 with the rhamnosyl donor 31 at -40 °C gave 29 in an excellent yield (94%). It is noteworthy that under these conditions cleavage or isomerisation of the dioxolane-type benzylidene group have not been observed.



Reaction of 27 with the disaccharide imidate 21 yielded the fully protected trisaccharide 32 in a good yield (73%). Deprotection of disaccharide 29 and trisaccharide 32 (deacylation, then acid hydrolysis) gave the *p*-nitrophenyl glycosides 33 and 34, respectively. The *p*-nitrophenyl group was converted into the *p*-isothiocyanatophenyl group as described for the preparation of 24, to give the spacer-armed di-(35) and trisaccharides (36).



The structures of the deprotected compounds 24, 26, 33, 34, 35, and 36 were confirmed by 13 C NMR spectroscopy. These compounds have been used for the preparation of the neoglycoproteins, which will be published in a separate paper.

Experimental

General.---Melting points (uncorrected) were determined on a Kofler hot-stage apparatus. Optical rotations were measured with a Perkin-Elmer 241 polarimeter. NMR spectra were recorded with a Bruker WP-200 SY or 400 AM spectrometer for solutions in CDCl₃ (internal Me₄Si) or in D₂O (internal 1,4-dioxane). In the case of compound 34 COSY and HETCOR spectra were acquired at 200 MHz. A HMBC spectrum was recorded with a Bruker DRX 500 instrument to reveal long-range proton-carbon connectivities. Interglycosidic linkages were unequivocally proved by this method [20]. The reactions were monitored by TLC on Kieselgel 60 F₂₅₄ (Merck, Darmstadt) with detection by charring with H_2SO_4 . Kieselgel 60 (Merck) was used for short-column chromatography. The original procedure was used for the regioselective ring-opening reaction of $8 \ (\rightarrow 5)$ with the LiAlH₄-AlCl₃ reagent [8] and for the preparation of compound 27 [4].

Benzyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranoside (2).—To a solution of 1,2,3,4-tetra-O-acetyl-Lrhamnopyranose (1; 39.5 g, 118.9 mmol) in dry CH₂Cl₂ (100 mL) were added 4 Å molecular sieves (10 g pellets) and benzyl alcohol (20.7 mL, 200 mmol), and the mixture was stirred under Ar for 30 min. After the addition of $BF_3 \cdot Et_2O$ (24.6 mL, 200 mmol) at 0 °C, the mixture was stirred for 2 h at 0 °C, then at ambient temperature overnight. Pyridine (20 mL) was added, and the mixture was filtered through Celite, then the cake was washed with CH_2CI_2 (2 × 50 mL). The combined filtrates were washed with aq 5% NaHCO₃ (2×50 mL) and water $(3 \times 50 \text{ mL})$, dried, and concentrated. The crystalline residue was recrystallized from EtOH to give 2 (17.48 g, 38.6%); mp 108–110 °C, lit. 110 °C [9]; $[\alpha]_{D}$ -75° (c 0.52, CHCl₃), lit. -73° (CHCl₃) [9]; ¹H NMR (CDCl₃): δ 7.38–7.32 (m, 5 H, Ph), 5.34 (dd, 1 H, $J_{3,4}$ 10 Hz, H-3), 5.28 (dd, 1 H, $J_{2,3}$ 3.5 Hz, H-2), 5.08 (t, 1 H, $J_{4,5}$ 10 Hz, H-4), 4.80 (d, 1 H, $J_{1,2}$ 1.6 Hz, H-1), 4.63 (dd, 2 H, PhCH₂), 3.91 (m, 1 H, J_{5.6} 6.2 Hz, H-5), 2.13, 2.05, and 1.98 (3 s, each 3 H, 3 Ac), 1.21 (d, 3 H, CMe). The mother liquor was concentrated and subjected to column chromatography (99:1 \rightarrow 95:5 CH₂Cl₂-EtOAc) yielding an additional amount of 2 (13.78 g, 30.5%). Overall yield: 69.1%.

Benzyl 4-O-benzyl-endo-2,3-O-benzylidene- α -Lrhamnopyranoside (8).—To a solution of benzyl 4-*O*-benzyl- α -L-rhamnopyranoside (**3**; 3.44 g, 10 mmol) in α, α -dimethoxytoluene (15 mL) was added ptoluenesulfonic acid (150 mg), and the mixture was stirred for 20 min at room temperature. Then, the mixture was diluted with CH₂Cl₂ (160 mL), washed with aq 5% NaHCO₃ (2 \times 25 mL) and water (3 \times 25 mL), dried, filtered, and concentrated. Column chromatography (hexane \rightarrow 7:3 hexane-EtOAc) of the residue yielded 8 (3.42 g, 79%); mp 52-54 °C (from hexane), lit. 53–54 °C [8]; $[\alpha]_{D}$ – 63° (c 1, CHCl₃), lit. -57° (CHCl₃) [8]; ¹H NMR (CDCl₃): δ 7.50– 7.22 (m, 15 H, 3 Ph), 5.91 (s, 1 H, PhCH), 5.18 (s, 1 H, H-1), 4.84–4.42 (m, 5 H, 2 PhCH₂ and H-3), 4.28 (d, 1 H, H-2), 3.82 (m, 1 H, H-5), 3.27 (dd, 1 H, H-4), 1.26 (d, 3 H, CMe).

p-Nitrophenyl 4-O-benzyl-endo-2,3-O-benzylidene- α -L-rhamnopyranoside (9).—To a solution of *p*nitrophenyl 4-O-benzyl- α -L-rhamnopyranoside [3] (4; 1126 mg, 3 mmol) in α , α -dimethoxytoluene (4 mL) was added *p*-toluenesulfonic acid (100 mg) and the mixture was stirred for 15 min at room temperature, then worked-up as described for **8**. Column chromatography (hexane \rightarrow 7:3 hexane–EtOAc) of the residue yielded **9**, isolated as a syrup (1126 mg, 81%); $[\alpha]_D - 133^\circ$ (*c* 0.11, CHCl₃); ¹H NMR (CDCl₃): δ 8.21 (d, 2 H, aromatic), 7.56–7.10 (m, 12 H, aromatic), 5.99 (s, 1 H, PhC*H*), 5.93 (s, 1 H, H-1), 4.90–4.46 (m, 4 H, PhC*H*₂, H-2,3), 3.78 (m, 1 H, H-5), 3.34 (dd, 1 H, H-4), 1.20 (d, 3 H, CMe). Anal. Calcd for C₂₆H₂₅NO₇: C, 67.37; H, 5.44. Found: C, 67.51; H, 5.40.

p-Nitrophenyl 2,4-di-O-benzyl- (6) and pnitrophenyl 3,4-di-O-benzyl- α -L-rhamnopyranoside (7).—Procedure A.—A mixture of p-nitrophenyl-4-*O*-benzyl- α -L-rhamnopyranoside [3] (4; 751 mg, 2 mmol), CH₂Cl₂ (10 mL), aq 20% NaOH (10 mL), tetrabutylammonium bromide (0.64 g, 2 mmol), and benzyl bromide (0.29 mL, 2.4 mmol) was vigorously stirred overnight at room temperature. Then, the mixture was diluted with CH₂Cl₂ (60 mL), and the organic layer was separated and washed with water $(4 \times 20 \text{ mL})$, dried (MgSO₄), and concentrated. Column chromatography (98:2 CH₂Cl₂-acetone) of the residue gave 6 (257 mg, 27.6%) and 7 (240 mg, 25.8%). Compound **6** had $[\alpha]_D = 80.6^\circ$ (c 1.39, CHCl₃); ¹H NMR (CDCl₃): δ 8.17 and 7.07 (2 d, each 2 H, aromatic), 7.43-7.22 (m, 10 H, aromatic), 5.58 (d, 1 H, J_{1.2} 1 Hz, H-1), 4.95–4.63 (m, 4 H, 2 PhC H_2), 4.13 (m, 1 H, after addition of D₂O dd, $J_{2,3}$ 3.8, J_{3.4} 9 Hz, H-3), 3.95 (dd, 1 H, H-2), 3.68 (m, 1 H, H-5), 3.43 (t, 1 H, H-4), 2.41 (d, 1 H, OH, disappeared after addition of D_2O , 1.29 (d, 3 H, CMe). Anal. Calcd for C₂₆H₂₇NO₇: C, 67.08; H, 5.85. Found: C, 67.19; H, 5.92. Compound 7 had $[\alpha]_{D} = -55.9^{\circ}$ (c 0.86, CHCl₃); Anal. Found: C, 67.26; H, 5.87.

Procedure B.—To a solution of **9** (927 mg, 2 mmol) in 1:1 $\text{Et}_2\text{O}-\text{CH}_2\text{Cl}_2$ (10 mL) was added, with stirring, a solution of LiAlH₄ (114 mg, 3 mmol) and AlCl₃ (400 mg, 3 mmol) in Et_2O (10 mL), and stirring was continued for an additional 10 min. The mixture was diluted with CH_2Cl_2 (60 mL) and the excess reagent was decomposed with EtOAc (5 mL), and Al(OH)₃ was precipitated with water (5 mL). The organic layer was decanted, washed with water (2 × 15 mL), dried (MgSO₄), and concentrated. The residue contained **6** and **7** in a molar ratio of 9:1 (TLC). Compound **6** was isolated by column chromatography yielding 735 mg (78.9%).

Benzyl (2,4,6-tri-O-acetyl-3-O-methyl- β -D-glucopyranosyl)-(1 \rightarrow 3)- (11) and benzyl (2,4,6-tri-Oacetyl-3-O-methyl- α -D-glucopyranosyl)-(1 \rightarrow 3)-2,4di-O-benzyl- α -L-rhamnopyranoside (12).—Procedure A. — A solution of 5 (869 mg, 2 mmol) and Hg(CN)₂ (606 mg, 2.4 mmol) in dry toluene (10 mL)

and nitromethane (10 mL) was concentrated to half its volume at atmospheric pressure. After cooling, 920 mg (2.4 mmol) of 2,4,6-tri-O-acetyl-3-O-methyl- α -D-glucopyranosyl bromide (10) [12] was added, and the mixture was stirred for 2 h at 50 °C, then diluted with CH_2CI_2 (40 mL). After filtration, the filtrate was washed with aq 5% KI (2×10 mL) and water $(2 \times 10 \text{ mL})$, dried (MgSO₄), and concentrated. The products were separated by column chromatography (95:5 CH₂Cl₂-EtOAc) yielding 1017 mg (69%) of 11 and 98 mg (6.6%) of 12. Compound 11 had $[\alpha]_{D} = -57^{\circ} (c \ 1.39, \text{ CHCl}_{3});$ ¹H NMR (CDCl₃): δ 7.43-7.20 (m, 15 H, aromatic), 5.16-4.91 (m, 2 H, H-2',4'), 4.87–4.51 (m, 7 H, 3 PhCH₂ and H-1), 4.40 (d, 1 H, $J_{1'2'}$ 8 Hz, H-1'), 3.42 (s, 3 H, OMe), 2.10, 1.89, and 1.81 (3 s, each 3 H, 3 Ac), 1.27 (d, 3 H, CMe). Anal. Calcd for C₄₀H₄₈O₁₃: C, 65.20; H, 6.57. Found: C, 65.00; H, 6.52. Compound 12 had $[\alpha]_{\rm D}$ +18° (c 0.53, CHCl₃); ¹H NMR (CDCl₃): δ 7.43–7.20 (m, 15 H, aromatic), 5.31 (d, 1 H, $J_{1'2'}$ 3.6 Hz, H-1'), 5.07-4.38 (m, 9 H, 3 PhCH₂, H-1,2',4'), 3.44 (s, 3 H, OMe), 1.99, 1.98, and 1.84 (3 s, each 3 H, 3 Ac), 1.33 (d, 3 H, CMe). Anal. Found: C, 65.39; H, 6.58.

Procedure B. — To a solution of 1,2,4,6-tetra-Oacetyl-3-O-methyl-D-glucopyranose [12] (15; 4.25 g, 11.73 mmol) in dry DMF (12 mL) was added hydrazine acetate [15] (1.30 g, 14.1 mmol) and the mixture was kept for 20 min at 50 °C. After cooling, the mixture was diluted with EtOAc (300 mL) and washed with aq 5% NaCl (3×40 mL). The combined washings were extracted with EtOAc (40 mL). The combined organic phase was washed with water (40 mL), dried, filtered, and concentrated. The obtained crude 2,4,6-tri-O-acetyl-3-O-methyl-D-glucopyranose (16) was dissolved in dry CH_2Cl_2 (40 mL), then trichloroacetonitrile (11.73 mL, 117 mmol) and 1,8-diazabicyclo[5.4.0]undec-7-ene [16] (2.39 mL, 16 mmol) were added. The mixture was stirred for 20 min, then concentrated, and the residue was purified by column chromatography (9:1 CH₂Cl₂-EtOAc containing 0.5% Et₃N) to yield 2,4,6-tri-O-acetyl-3-O-methyl- α -D-glucopyranosyl trichloroacetimidate (17), isolated as a syrup (3.28 g, 60.2%). Only the pure fractions, free of UV active impurities, were collected; $[\alpha]_{\rm D} + 85.7^{\circ} (c \ 0.5, \text{ CHCl}_3)$; ¹H NMR (CDCl₃): δ 8.67 (s, 1 H, NH), 6.52 (d, 1 H, $J_{1,2}$ 3.8 Hz, H-1), 5.12 (t, 1 H, $J_{4.5}$ 10 Hz, H-4), 5.03 (dd, 1 H, J₂, 10 Hz, H-2), 4.27–4.04 (m, 3 H, H-5,6a,6b), 3.82 (t, 1 H, $J_{3,4}$ 10 Hz, H-3), 3.50 (s, 3 H, OMe), 2.12 and 2.08 (2 s, 3,6 H, 3 Ac).

A solution of 5 (435 mg, 1 mmol) and 17 (558 mg,

1.2 mmol) in dry CH₂Cl₂ (8 mL), containing 4 Å molecular sieves (2 g pellets), was cooled to -40° C under Ar. Then, a solution of Me₃SiOTf (19.3 μ L, 0.1 mmol) in dry CH₂Cl₂ (2 mL) was added dropwise and the mixture was stirred for 30 min at -40° C. After the addition of pyridine (1 mL) and CH₂Cl₂ (50 mL), the mixture was filtered through Celite, the filtrate was washed with aq 5% NaHCO₃ (10 mL) and water (2 × 10 mL), dried, and concentrated and co-concentrated with toluene (3 × 20 mL). The residue was purified by column chromatography (95:5 CH₂Cl₂-EtOAc) to give pure **11** (575 mg, 78%); [α]_D - 55.5° (*c* 0.82, CHCl₃). A by-product with higher chromatographic mobility was also isolated (38 mg).

Procedure C. — To a stirred solution of 5 (435) mg, 1 mmol) and 10 (460 mg, 1.2 mmol) in dry CH_2Cl_2 (8 mL) was added 4 Å powdered molecular sieves (1 g), and the mixture was stirred under Ar for 15 min. A solution of silver triflate [17] (308 mg, 1.2 mmol) in 8 mL dry toluene was added to the mixture over a period of 5 min at -50 °C. Stirring was continued for an additional hour while the temperature was raised to -40 °C. The mixture was neutralized with pyridine, diluted with CH₂Cl₂ (100 mL), and filtered through Celite. The filtrate was washed with aq 10% sodium thiosulfate $(2 \times 20 \text{ mL})$ and water $(2 \times 20 \text{ mL})$, dried, and evaporated. The product was purified by column chromatography giving pure 11 (659 mg, 89.4%); $[\alpha]_{D} = -54.4^{\circ}$ (c 0.40, $CHCl_3$).

p-Nitrophenyl (2,4,6-tri-O-acetyl-3-O-methyl-β-Dglucopyranosyl)- $(1 \rightarrow 3)$ - (13) and p-nitrophenyl (2,4,6-tri-O-acetyl-3-O-methyl- α -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzyl- α -L-rhamnopyranoside (14). -A mixture of 6 (400 mg, 0.859 mmol), dry CH₂Cl₂ (10 mL), 2,4,6-tri-O-acetyl-3-O-methyl- α -D-glucopyranosyl bromide (10) [12] (395 mg, 1.03 mmol), and 4 Å powdered molecular sieves (1 g), was stirred under Ar for 15 min. Then, HgBr₂ (371 mg, 1.03 mmol) was added [13] and stirring was continued for 2 days at room temperature. The mixture was diluted with CH_2Cl_2 (100 mL), filtered, and the filtrate was washed with aq 5% KI (2×15 mL), dried (MgSO₄), and concentrated. Column chromatography (95:5 CH_2Cl_2 -EtOAc) of the residue gave amorphous 13 $(292 \text{ mg}, 44.3\%); [\alpha]_{D} - 79^{\circ} (c \ 0.47, \text{CHCl}_{3}); \text{NMR}$ data (CDCl₃): ¹H (400 MHz), δ 8.15 and 7.03 (2 d, each 2 H, aromatic), 7.44–7.25 (m, 10 H, aromatic), 5.43 (d, 1 H, J₁₂ 1.9 Hz, H-1), 5.12 (dd, 1 H, H-2'), 5.03 (t, 1 H, H-4'), 4.86 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 4.90-4.54 (m, 4 H, 2 PhCH₂), 4.19-4.05 (m, 4 H, H-2,3,6a',6b'), 3.65–3.60 (m, 3 H, H-4,5,5'), 3.48 (t, 1 H, H-3'), 3.41 (s, 3 H, OMe), 2.11, 1.95, and 1.91 (3 s, each 3 H, 3 Ac), 1.20 (d, 3 H, CMe); 13 C (100.6 MHz), δ 101.93 (C-1'), 96.62 (C-1), 58.91 (OMe). Anal. Calcd for C₃₉H₄₅NO₁₅: C, 61.01; H, 5.91. Found: C, 60.82; H, 5.97.

Further elution gave **14** (92 mg, 13.9%); mp 176– 177 °C (from cyclohexane–EtOH); $[\alpha]_D - 6^\circ$ (*c* 0.60, CHCl₃); NMR data (CDCl₃): ¹H (400 MHz), δ 8.17 and 7.07 (2 d, each 2 H, aromatic), 7.41–7.27 (m, 10 H, aromatic), 5.49 (d, 1 H, $J_{1.2}$ 2.2 Hz, H-1), 5.38 (d, 1 H, $J_{1'.2'}$ 3.7 Hz, H-1'), 5.02 (dd, 1 H, H-4'), 4.86 (dd, 1 H, H-2'), 4.92–4.58 (m, 4 H, 2 PhCH₂), 4.21 (dd, 1 H, H-3), 4.13 (m, 1 H, H-5'), 4.00–3.92 (m, 3 H, H-2,6a',6b'), 3.77–3.69 (m, 3 H, H-3',4,5), 3.44 (s, 3 H, OMe), 2.02, 1.96, and 1.90 (3 s, each 3 H, 3 Ac), 1.30 (d, 3 H, CMe); ¹³C (100.6 MHz), δ 96.16 (C-1), 92.41 (C-1'), 60.17 (OMe). A mixture of **13** and **14** (125 mg, 18.9%) was also isolated.

(2,4,6-Tri-O-acetyl-3-O-methyl-β-D-glucopyranosyl)- $(1 \rightarrow 3)$ -1,2,4-tri-O-acetyl- α - (18) and β -Lrhamnopyranose (19).—Compound 11 (1 g) was hydrogenated over Pd-C (100 mg) in 10:1 EtOH-HOAc (33 mL) for 2 days. After filtration, the filtrate was concentrated and co-concentrated with toluene (3 \times 15 mL). The residue was conventionally acetylated in 1:1 pyridine-Ac₂O (30 mL), and the product was purified by column chromatography (7:3 CH₂Cl₂-EtOAc) to yield 18 (0.57 g, 71%) and 19 (64 mg, 8%). Compound 18 had mp 185-187 °C (from EtOH); $[\alpha]_{\rm D} - 36.5^{\circ} (c \ 0.72, \text{CHCl}_3); {}^{1}\text{H NMR (CDCl}_3): \delta$ 5.99 (d, 1 H, J_{1.2} 2 Hz, H-1), 5.17 (dd, 1 H, J_{2.3} 3.8 Hz, H-2), 5.11 (t, 1 H, J_{4.5} 10 Hz, H-4), 5.05 (t, 1 H, $J_{4'5'}$ 9.8 Hz, H-4'), 4.93 (dd, 1 H, $J_{2',3'}$ 9.8 Hz, H-2'), 4.55 (d, 1 H, $J_{1',2'}$ 8 Hz, H-1'), 4.15 (d, 2 H, H-6a',6b'), 4.06 (dd, 1 H, J_{3,4} 10 Hz, H-3), 3.81 (m, 1 H, H-5), 3.57 (m, 1 H, H-5'), 3.44 (t, 1 H, J_{3'4'} 9.8 Hz, H-3'), 3.37 (s, 3 H, OMe), 2.13, 2.12, 2.10, and 2.09 (4 s, 6,3,3,6 H, 6 Ac), 1.19 (d, 3 H, CMe). Anal. Calcd for C₂₅H₃₆O₁₆: C, 50.67, H, 6.12. Found: C, 50.50; H, 6.11. Compound 19 had mp 178-180°C (from EtOH); $[\alpha]_{D} + 4.3^{\circ} (c \ 0.28, \text{CHCl}_{3});^{+}\text{H NMR}$ (CDCl₃): δ 5.72 (d, 1 H, $J_{1,2}$ 1.2 Hz, H-1), 5.44 (dd, 1 H, $J_{2,3}$ 3.5 Hz, H-2), 5.08–4.94 (m, 2 H, H-4,4'), 4.88 (dd, 1 H, $J_{2'3'}$ 9.7 Hz, H-2'), 4.48 (d, 1 H, $J_{1'2'}$ 7.8 Hz, H-1'), 4.11 (d, 2 H, H-6a',6b'), 3.79 (dd, 1 H, J_{3.4} 9.8 Hz, H-3), 3.60–3.44 (m, 2 H, H-5,5'), 3.39 (t, 1 H, $J_{3'4'}$ 9.5 Hz, H-3'), 3.33 (s, 3 H, OMe), 2.12, 2.08, 2.06, 2.05, 2.04, and 2.03 (6 s, each 3 H, 6 Ac), 1.22 (d, 3 H, CMe). Anal. Found: C, 50.52; H, 6.15. (2,4,6-Tri-O-acetyl-3-O-methyl-β-D-glucopyrano-

(2,4,6-17)-(2,

trichloroacetimidate (21).—Compound 18 (450 mg, 0.76 mmol) was converted into imidate 21, via 20, as described for the preparation of 17 from 15. Yield: 314 mg (59.5%); mp 149–156°C; $[\alpha]_D - 33°$ (*c* 0.80, CHCl₃); ¹H NMR (CDCl₃): δ 8.74 (bs, 1 H, NH), 6.20 (d, 1 H, $J_{1,2}$ 1.5 Hz, H-1), 5.35 (dd, 1 H, H-2), 5.21–4.91 (m, 3 H, H-4,2',4'), 4.56 (d, 1 H, $J_{1',2'}$ 7.8 Hz, H-1'), 4.19–4.07 (m, 3 H, H-3,6a',6b'), 3.95 (m, 1 H, H-5), 3.58 (m, 1 H, H-5'), 3.44 (t, 1 H, H-3'), 3.37 (s, 3 H, OMe), 2.16, 2.14, 2.12, 2.10, and 2.09 (5 s, each 3 H, 5 Ac), 1.22 (d, 3 H, CMe). This compound was used for the next step without further purification.

p-Nitrophenyl (3-O-methyl-β-D-glucopyranosyl)-(1 \rightarrow 3)-2,4-di-O-benzyl- α -L-rhamnopyranoside (22). To a solution of 13 (307 mg, 0.4 mmol) in dry MeOH (50 mL) was added NaOMe (54 mg, 1 mmol), and the mixture was kept for 3 days at ambient temperature. The solution was neutralized with Amberlite IR 120 (H⁺) resin, filtered, and concentrated. Column chromatography (8:2 CH₂Cl₂-acetone) of the residue gave glassy 22 (160 mg, 62.3%); $[\alpha]_D$ -103° (c 0.43, MeOH); ¹H NMR (CDCl₃): δ 8.19 (d, 2 H, aromatics), 7.42-7.23 (m, 10 H, aromatics), 7.08 (d, 2 H, aromatics), 4.85-4.43 (m, 5 H, H-1 and 2 PhC H_2), 4.37 (d, 1 H, $J_{1',2'}$ 7.6 Hz, H-1'), 3.53 (s, 3 H, OMe), 2.68 (bs, 1 H, OH), 1.70 (bs, 2 H, 2 OH), 1.23 (d, 3 H, CMe). Anal. Calcd for $C_{33}H_{39}NO_{12}$: C, 61.77; H, 6.13. Found: C, 61.92; H, 6.10.

Attempted preparation of p-isothiocyanatophenyl $(3-\text{O-methyl-}\beta-\text{D-glucopyranosyl})-(1 \rightarrow 3)-\alpha-\text{L-}$ rhamnopyranoside (24).—Compound 22 (64 mg, 0.1 mmol) was hydrogenated over Pd-C (120 mg) in aq 80% EtOH (15 mL) and the reaction was monitored by TLC. Conversion of the nitro group into an amino group was rather quick (9:1 CH₂Cl₂–MeOH), but the removal of the benzyl groups was not completed within days (8:2 CH₂Cl₂-MeOH). After 4 days, the catalyst was filtered out, the pH of the filtrate was adjusted to 8 with BaCO₃ and maintained at 8 by adding BaCO₃ whilst thiophosgene (0.2 mL) was added, and the mixture was stirred for 1 h at room temperature. Filtration and concentration yielded a slightly coloured syrup, which was purified by column chromatography (95:5 CH₂Cl₂-MeOH) to give an impure material (37 mg) which could not be purified either by repeated chromatography or by crystallization (from EtOH or EtOAc-MeOH).

p-Nitrophenyl (2,4,6-tri-O-acetyl-3-O-methyl- β -Dglucopyranosyl)-(1 \rightarrow 3)-2,4-di-O-acetyl- α -L-rhamnopyranoside (25).—To a stirred mixture of 18 (148 mg, 250 μ mol), p-nitrophenol (52 mg, 375 μ mol), 4 Å molecular sieves (1 g pellets), and dry CH_2Cl_2 (5 mL) under Ar was added BF₃ · Et₂O (31 μ L, 250 μ mol). After 24 h an additional amount of BF₃ · Et₂O (31 μ L) was added, and stirring was continued for an additional day. Pyridine (1 mL) and CH₂Cl₂ (30 mL) were added, the mixture was filtered through Celite, and the filtrate was washed with aq 5% NaHCO₃ $(2 \times 10 \text{ mL})$ and water $(3 \times 10 \text{ mL})$, dried, and concentrated. The residue was purified by column chromatography to yield pure 25 (69 mg, 41%); $[\alpha]_{D}$ -71.6° (*c* 0.21, CHCl₃); ¹H NMR (CDCl₃): δ 8.22–7.14 (2 d, 4 H, aromatics), 5.58 (d, 1 H, $J_{1,2}$ 1.2 Hz, H-1), 5.34 (dd, 1 H, H-2), 5.22-4.91 (m, 3 H, H-4,2',4'), 4.61 (d, 1 H, $J_{1'2'}$ 8 Hz, H-1'), 4.28– 4.10 (m, 3 H, H-3,6a',6b'), 3.87-3.57 (m, 2 H, H-5,5'), 3.48 (t, 1 H, H-3'), 3.38 (s, 3 H, OMe), 2.18, 2.13, 2.11, 2.09, and 2.08 (5 s, each 3 H, 5 Ac), 1.18 (d, 3 H, CMe). Anal. Calcd for $C_{29}H_{37}NO_{17}$: C, 51.86; H, 5.55. Found: C, 52.04; H, 5.60.

p-Nitrophenyl (3-O-methyl-β-D-glucopyranosyl)-(1 → 3)-α-L-rhamnopyranoside (26).—Compound 25 (67 mg, 0.1 mmol) was deacetylated as described for the preparation of 22 from 13. The product was purified by column chromatography (9:1 CH₂Cl₂– MeOH) yielding 26, isolated as a foam (30 mg, 65%); $[\alpha]_D$ –119° (*c* 0.52, MeOH); ¹³C NMR (D₂O): δ 161.80 and 143.32 (aromatic-q), 127.01 and 117.72 (aromatic), 104.80 (C-1'), 98.81 (C-1), 86.14 (C-3'), 80.93 (C-3), 61.49 (C-6'), 60.63 (OMe), 17.72 (C-6). Anal. Calcd for C₁₉H₂₇NO₁₂: C, 49.45; H, 5.90. Found: C, 49.67; H, 5.93.

p-Nitrophenyl endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (27).—This compound was prepared as described earlier [4]. The product was crystallized from hexane–EtOAc to give pure 27; mp 89–90 °C; $[\alpha]_D = 106.8^\circ$ (*c* 1.1, CHCl₃), lit. = 106° (CHCl₃) [4].

p-Nitrophenyl (2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl)-(1 → 2)-endo-3,4-O-benzylidene-6-deoxyα-L-talopyranoside (29).—A mixture of 2,3,4-tri-Oacetyl-α-L-rhamnopyranosyl bromide (28; 353 mg, 1 mmol), 'old' Ag₂O (463 mg, 2 mmol), CH₂Cl₂ (2 mL) and water (0.1 mL) was stirred for 1 h at room temperature, then diluted with CH₂Cl₂ (20 mL), and filtered through Celite. The filtrate was washed with aq 10% Na₂S₂O₃ (5 mL) and water (3 × 5 mL), dried, and concentrated. The residue was converted into imidate **31** as described for **17**. Column chromatography (9:1 CH₂Cl₂–EtOAc, containing 0.5% Et₃N) gave **31** (276 mg, 63%); [α]_D – 38.3° (c 1.99, CHCl₃); ¹H NMR (CDCl₃): δ 8.74 (bs, 1 H, NH), 6.20 (d, 1 H, J_{1,2} 2 Hz, H-1), 5.46 (dd, 1 H, J_{2,3} 3.5 Hz, H-2), 5.37 (dd, 1 H, $J_{3,4}$ 10 Hz, H-3), 5.17 (t, 1 H, $J_{4,5}$ 10 Hz, H-4), 4.09 (m, 1 H, H-5), 2.19, 2.08, and 2.01 (3 s, each 3 H, 3 Ac), 1.28 (d, 3 H, CMe).

A solution of *p*-nitrophenyl endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (27) (93.3 mg, 250 μ mol) and 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl trichloroacetimidate (31; 163 mg, 375 μ mol) in dry CH_2Cl_2 (4 mL), containing 4 A molecular sieves (0.5 g pellets), was cooled to -40 °C under Ar. A solution of Me₃SiOTf (9.7 μ L, 50 nmol) in dry CH_2Cl_2 (1 mL) was added dropwise and the mixture was stirred for 20 min at -40 °C. After the addition of pyridine (0.5 mL) and CH₂Cl₂ (30 mL), the mixture was filtered through Celite, the filtrate was washed with aq 5% NaHCO₃ (5 mL) and water $(3 \times 5 \text{ mL})$, dried, and concentrated and co-concentrated with toluene $(3 \times 10 \text{ mL})$. The product was purified by column chromatography (95:5 CH₂Cl₂-EtOAc) to give **29** (152 mg, 94.2%); $[\alpha]_{\rm D} = 91.5^{\circ}$ (*c* 1.18, CHCl₃); ¹H NMR (CDCl₃): δ 8.24 and 7.14 (2 d, 4 H, aromatics), 7.67–7.40 (m, 5 H, Ph), 5.82 (s, 1 H, PhCH), 5.73 (d, 1 H, J_{1,2} 6.4 Hz, H-1), 5.30 (dd, 1 H, H-2'), 5.19 (dd, 1 H, $J_{2'3'}$ 3.5, $J_{3'4'}$ 10 Hz, H-3'), 5.06 (t, 1 H, H-4'), 5.06 (bs, 1 H, H-1'), 4.71 (dd, 1 H, J_{2,3} 2.8, J_{3,4} 8.2 Hz, H-3), 2.17, 1.99, and 1.93 (3 s, each 3 H, 3 Ac), 1.29 and 1.21 (2 d, each 3 H, 2 CMe). Anal. Calcd for $C_{31}H_{35}NO_{14}$: C, 57.67; H, 5.46. Found: C, 57.91; H, 5.50.

p-Nitrophenyl (2,4,6-tri-O-acetyl-3-O-methyl-β-Dglucopyranosyl)- $(1 \rightarrow 3)$ -(2,4-di-O-acetyl- α -L-rhamnopyranosyl)- $(1 \rightarrow 2)$ -endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (32).—Compound 27 (101 mg, 0.27 mmol) was reacted with imidate 21 (209 mg, 0.3 mmol) as described for 29. Column chromatography of the crude product yielded amorphous **32** (178 mg, 72.8%); $[\alpha]_{\rm D} = 86.8^{\circ} (c \ 0.99, \text{CHCl}_3);$ ¹H NMR (CDCl₃): δ 8.24 and 7.14 (2 d, 4 H, aromatic), 7.67-7.40 (m, 5 H, aromatic), 5.81 (s, 1 H, PhCH), 5.67 (d, 1 H, J_{1,2} 6 Hz, H-1), 5.22 (dd, 1 H, H-2'), 5.12-4.65 (m, 4 H, H-1',4',2",4"), 4.39 (d, 1 H, $J_{1''2''}$ 7.8 Hz, H-1"), 3.48 (m, 1 H, H-5"), 3.34 (t, 1 H, H-3"), 3.32 (s, 3 H, OMe), 2.10, 2.04, 2.02, and 1.82 (4 s, 3,6,3,3 H, 5 Ac), 1.28 and 1.17 (2 d, each 3 H, 2 CMe). Anal. Calcd for $C_{42}H_{51}NO_{21}$: C, 55.69; H, 5.68. Found: C, 55.81; H, 5.73.

p-Nitrophenyl α -L-rhamnopyranosyl- $(1 \rightarrow 2)$ -6deoxy- α -L-talopyranoside (33).—To a solution of 29 (129 mg, 0.2 mmol) in dry MeOH (4 mL) was added a catalytic amount of NaOMe. The mixture was kept at room temperature overnight, then neutralized with HOAc, and concentrated. The residue was dissolved in aq 60% HOAc (4 mL) and the solution was kept

Table 1 NMR data for trisaccharide **34** in D_2O (δ in ppm, J in Hz)

Sugar unit	Atom No.	δ , ¹³ C	δ , ¹ H	J, ¹ H
6-deoxy-Tal	1	97.86	5.70	$J_{1,2}$ 1.2
	2	76.94	4.05	$J_{2.3}^{-1}$ 3.6
	2 3	66.28	4.16	$J_{3.4}^{-1}$ 3.7
	4	71.87	3.67	$J_{4.5}$ 1.1
	5	69.16	3.99	$J_{5,6}^{n}$ 6.6
	6	16.06	1.10	<i>w</i> , <i>w</i>
Rha	1	103.17	4.98	$J_{1',2'}$ 1.8
	2	70.36	4.27	$J_{2',3'}$ 3.3
	3	80.23	3.92	$J_{3',4'}$ 9.7
	4	71.70	3.55	$J_{4',5'}^{(3)}$ 9.7
	5	70.00	3.78	$J_{5,6}$, 6.1
	6	17.49	1.20	5,0
Gle	1	104.24	4.65	$J_{1'',2''}$ 8.1
	2	73.46	3.34	$J_{2'',3''}$ 9.5
	3	85.86	3.20	$J_{3'',4''}$ 9.5
	4	69.51	3.44	$J_{4'',5''}$ 9.5
	5	76.32	3.38	$J_{5'',6a''}$ 2.3
				$J_{5'',6b''}$ 5.3
	6	61.09	3.81 (a)	5,00
			3.65 (b)	
	Aromatic (q)	161.50		
		143.00		
	Aromatic	126.77	8.08	
		117.29	7.09	

for 20 min at 70 °C. After concentration and co-concentration with toluene (3 × 5 mL), the product was purified by column chromatography (75:25 CH₂Cl₂–MeOH) to give amorphous **33** (58 mg, 67%); $[\alpha]_{\rm D}$ – 64° (*c* 0.73, MeOH); ¹³C NMR (D₂O): δ 103.83 (C-1'), 98.39 (C-1), 77.57 (C-2), 17.75 (C-6'), 16.38 (C-6). Anal. Calcd for C₁₈H₂₅NO₁₁: C, 50.11; H, 5.84. Found: C, 50.50; H, 5.86.

p-Nitrophenyl (3-O-methyl-β-D-glucopyranosyl)-(1 → 3)- α -L-rhamnopyranosyl-(1 → 2)-6-deoxy- α -Ltalopyranoside (34).—To a solution of 32 (90.6 mg, 0.1 mmol) in dry MeOH (5 mL) was added NaOMe (8.1 mg, 0.15 mmol). The mixture was kept for 2 days at room temperature, then neutralized with Amberlite IR 120 (H⁺) resin, filtered, and concentrated. The residue was dissolved in aq 60% HOAc (8 mL), and the solution was stirred for 20 min at 70 °C, then concentrated and co-concentrated with toluene (3 × 10 mL). The product was purified by column chromatography (9:1 CH₂Cl₂-MeOH) to give 34, isolated as a foam (49 mg, 69%); [α]_D - 103° (*c* 0.96, MeOH). For ¹H and ¹³C NMR data, see Table 1. Anal. Calcd for C₂₅H₃₇NO₁₆: C, 49.42; H, 6.14. Found: C, 49.76; H, 6.09.

p-Isothiocyanatophenyl (3-O-methyl-B-D-glucopyranosyl)- $(1 \rightarrow 3)$ - α -L-rhamnopyranoside (24). Compound 26 (23.1 mg, 50 μ mol) was hydrogenated over Pd-C (8 mg) in aq 80% EtOH (4 mL) for 2 h, then the catalyst was filtered out. The pH of the filtrate was adjusted to 8 with BaCO₃ and maintained at 8 by adding BaCO₃ whilst thiophospene (80 μ L) was added, and the mixture was stirred for 1 h at room temperature. Filtration and concentration yielded a slightly coloured syrup. Column chromatography (9:1 CH₂Cl₂-MeOH) of the crude product gave 24 (15 mg, 63%); $[\alpha]_{\rm D} - 72.8^{\circ}$ (c 0.75, MeOH); ¹³C NMR (D₂O): δ 128.07 (aromatic), 118.44 (aromatic), 104.48 (C-1'), 94.64 (C-1), 86.06 (C-3'), 61.54 (C-6'), 60.49 (OMe), 17.97 (C-6). FABMS(+): Calcd for $C_{20}H_{27}NO_{10}S$ (473.49). Found: 473 [M]⁺.

p-Isothiocyanatophenyl α-L-rhamnopyranosyl-(1 → 2)-6-deoxy-α-L-talopyranoside (**35**).—Compound **33** (21.6 mg, 50 µmol) was converted into **35** as described for **24**. Column chromatography (9:1 CH₂Cl₂-MeOH) of the product yielded pure **35** (14.5 mg, 65%); $[\alpha]_D = 90.9^\circ$ (*c* 0.8, MeOH); ¹³C NMR (1:1 D₂O-Me₂SO-d₆): δ 104.33 (C-1'), 99.07 (C-1), 78.34 (C-2), 18.41 (C-6'), 17.10 (C-6). FABMS(+): Calcd for C₁₉H₂₅NO₉S (443.46). Found: 467 [M + 1 + Na]⁺.

p-Isothiocyanatophenyl (3-O-methyl-β-D-glucopyranosyl)-(1 → 3)-α-L-rhamnopyranosyl-(1 → 2)-6deoxy-α-L-talopyranoside (**36**).—Compound **34** (30.4 mg, 50 µmol) was converted into **36** as described for **24**. Column chromatography (85:15 CH₂Cl₂–MeOH) of the product gave pure **36** (18.2 mg, 59%); [α]₁, -85.7° (c 1.89, MeOH); ¹³C NMR (D₂O): δ 103.52 (C-1″), 102.73 (C-1′), 97.48 (C-1), 60.35 (C-6″), 59.79 (OMe), 17.23 (C-6′), 15.84 (C-6). FABMS(+): Calcd for C₂₆H₃₇NO₁₄S (619.63). Found: 644 [M + 1 + Na]⁺.

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References

- Z. Szurmai, J. Kerékgyártó, J. Harangi, and A. Lipták, Carbohydr. Res., 164 (1987) 313–325.
- [2] A. Lipták, J. Kerékgyártó, Z. Szurmai, and H. Duddeck, *Carbohydr. Res.*, 175 (1988) 241–248.

- [3] A. Borbás and A. Lipták, *Carbohydr. Res.*, 241 (1993) 99–116.
- [4] J. Kerékgyártó, Z. Szurmai, and A. Lipták, Carbohydr. Res., 245 (1993) 65–80.
- [5] I. Bajza, J. Kerékgyártó, J. Hajkó, L. Szilágyi, and A. Lipták, *Carbohydr. Res.*, 253 (1994) 111–120.
- [6] A. Lipták, A. Borbás, and I. Bajza, Med. Res. Rev., 14 (1994) 307–352.
- [7] P.J. Brennan, G.O. Aspinall, and J.E. Nam Shin, J. Biol. Chem., 256 (1981) 6817–6822.
- [8] A. Lipták, P. Fügedi, and P. Nánási, *Carbohydr. Res.*, 65 (1978) 209–217.
- [9] J.S. Brimacombe, M.C. Cook, and L.C.N. Tucker, *J. Chem. Soc.*, (1965) 2292–2294.
- [10] V. Pozsgay, Carbohydr. Res., 69 (1979) 284-286.
- [11] J. Kerékgyártó and A. Lipták, Carbohydr. Res., 248 (1993) 361–364.
- [12] T. Fujiwara, S.W. Hunter, and P.J. Brennan, *Carbohydr. Res.*, 148 (1986) 287–298.

- [13] J.-C. Jacquinet, D. Duchet, M.-L. Milat, and P. Sinaÿ, J. Chem. Soc., Perkin Trans. I, (1981) 326–330.
- [14] R.R. Schmidt and J. Michel, Angew. Chem., 92 (1980) 763–764.
- [15] G. Excoffier, D. Gagnaire, and J.-P. Utille, *Carbo-hydr. Res.*, 39 (1975) 368–373.
- [16] S. Sato, Y. Ito, T. Nukada, Y. Nakahara, and T. Ogawa, *Carbohydr. Res.*, 167 (1989) 197–210.
- [17] F.J. Kronzer and C. Schuerch, *Carbohydr. Res.*, 27 (1973) 379–390; S. Hanessian and J. Banoub, *Carbohydr. Res.*, 53 (1977) C13–C16.
- [18] Z. Szurmai, A. Lipták, and G. Snatzke, *Carbohydr. Res.*, 200 (1990) 201–208.
- [19] P.J. Garegg and B. Gotthammar, *Carbohydr. Res.*, 58 (1977) 345–352.
- [20] Gy. Batta and A. Lipták, J. Chem. Soc., Chem. Commun., (1985) 368-370.