

A New *N*-Acetylneuraminic Acid Donor for Highly Stereoselective α -Sialylation

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The new sialyl donor **6** (prepared from *N*-acetylneuraminic acid in 44% yield over six steps) effects clean α -sialylation of 2-(trimethylsilyl)ethyl 2,3,6,2',4',6'-hexabenzyl- β -D-lactoside in 67% yield.

Sialylation in high yield and stereoselectivity is difficult.^{1,2} Hindered glycosyl acceptors are particularly troublesome, causing a substantial fraction of the sialic acid-derived donor to undergo elimination to the corresponding glycal; yields obtained with such acceptors are consequently modest. However, yields may be increased with donors (e.g. **7**³ and **8**⁴) carrying sulfur-containing leaving groups, and especially when the acceptors have several hydroxy groups unprotected (e.g.

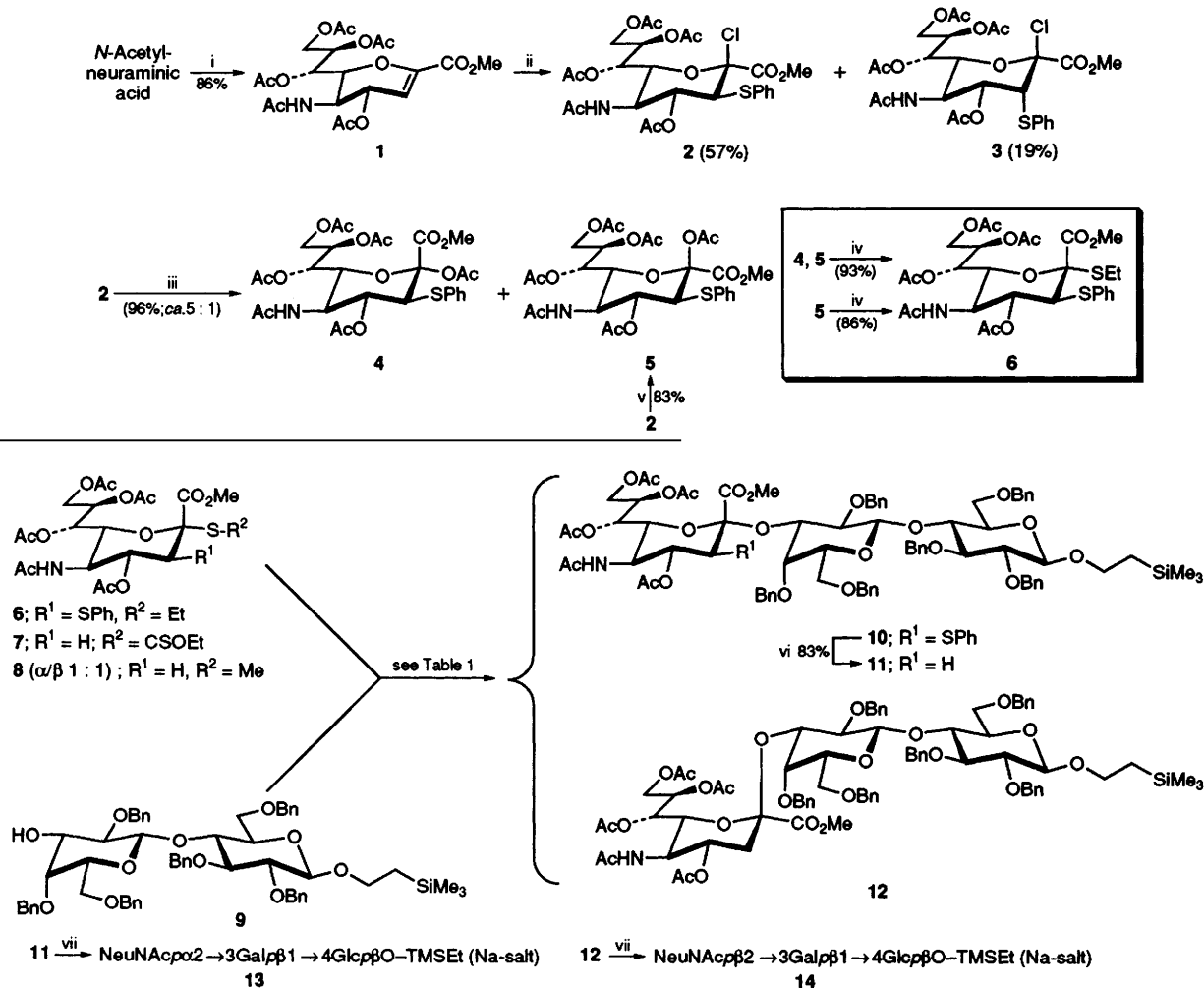
HO-2,3,4 in galactose residues). In order to improve the $\alpha:\beta$ -ratio in the sialylation reaction and also reduce glycal formation, an auxiliary participating 3-PhS-group has been introduced in 2-halogeno-sialic acid donors.⁵ However, the most effective donors carry *O*-benzyl protecting groups and their syntheses require *ca.* six steps, which furnish the desired donor in 20–50% overall yield,^{5,6} starting from the glycal **1**.

We now report the new sialyl donor **6**: (i) its synthesis

Table 1 Sialylation of lactoside acceptor **9** (0.15–0.50 mmol) with the sialic acid donors **6–8**

Donor	Mol. ratio ^a	P1,P2 ^a	Reaction conditions ^b	Product	Yield ^c (%)	α : β
6	1.0 : 1.0 : 1.1 : 1.1	MSB-AgOTf	MeCN, -40 °C	10	54	>99 : 1
6	1.0 : 1.5 : 1.6 : 1.6	MSB-AgOTf	MeCN, -40 °C	10	67	>99 : 1
6	1.0 : 1.5 : 1.7 : 0.5	NIS-TfOH	MeCN, -40 °C	10	57	>99 : 1
7	1.0 : 1.5 : 1.3 : 1.2	MSB-AgOTf	MeCN, CH ₂ Cl ₂ , -60 °C	11 + 12	36 + 4	90 : 10
8	1.0 : 1.5 : 1.5 : 0.7	NIS-TfOH	MeCN, -40 °C	11 + 12	29 + 4	88 : 12

^a Donor/acceptor/promotor 1 (P1)/promotor 2 (P2). ^b The concentration of **9** was $\sim 0.10 \text{ mol dm}^{-3}$. ^c Based on the donor (**6–8**).



Scheme 1 Reagents and conditions: i, MeOH, Dowex-H⁺, then Ac₂O, pyridine, then TMSOTf, MeCN, 0 °C, 6 h; ii, PhSCl, CH₂Cl₂, 20 °C, 7 days, Ar; iii, Hg(OAc)₂, AcOH–Ac₂O 10 : 1, 40 °C, 18 h. iv, EtSH, BF₃Et₂O, CH₂Cl₂, 20 °C, 18 h; v, HgBr₂, Hg(CN)₂, ClCH₂CH₂Cl–H₂O 100 : 1, reflux, 3.5 h, then Ac₂O, pyridine, DMAP, 20 °C, 1 h; vi, Ph₃SnH, AIBN, toluene, reflux, 14 h. vii, Pd/C, AcOH, 20 °C, overnight, then MeONa, MeOH, 20 °C, 2 h, then NaOH, H₂O, 20 °C, 0.5 h

Table 2 Anomeric configuration (NeuNAc residue) based on NMR analysis of compounds **2**, **4–6**, **10–14**

Compound	JC(1)–H(3) _{ax} /Hz ^a	JH _{7,8} /Hz	δ H(3) _{eq} /ppm	Δδ H(9)–H(9')/ppm	Configuration ^b
2	1.7	8.0	—	0.28	β
4	5.9	6.5	—	0.32	α
5	1.4	4.0	—	0.38	β
6	7.5	7.9	—	0.21	α
10	6.3	8.1	—	0.34	α
11	7.4 ^c	8.4	2.45	0.34	α
12	1.1	n.d.	2.71	~1.0 ^d	β
13	5.8	n.d.	2.75	n.d.	α
14	1.0	n.d.	2.45	n.d.	β

^a Identified by long-range HECTOR and measured according to ref. 21. ^b Anomeric (non-carboxyl) substituent. ^c JC(1)–H(3)_{eq} = 1.1 Hz. ^d Estimated from COSY spectrum.

requires only three steps (*ca.* 50% overall yield) from glycal **1**; (ii) it is a stable, pure α-thioglycoside; (iii) it carries a 3-PhS auxiliary group; (iv) unreacted **1** and the potentially useful byproduct **3** can be rescued from the reaction mixture; (v) **6** is an efficient α-sialyl donor, even with sterically congested acceptors such as **9**⁷ (Table 1).

The glycal **18**⁹ was synthesised (Scheme 1) by treatment of fully acetylated neuraminic acid methyl ester^{10,11} (9.3 mmol) with 2 equiv.^{12,6} (not 0.2¹³) of fresh (to reduce 4,5-oxazoline^{14,15} formation) trimethylsilyl trifluoromethanesulfonate.

Addition of fresh phenyl sulfonyl chloride (23 mmol) to **1** (8.55 mmol) in dichloromethane (30 ml) gave the diastereoisomers **2** (57%) and **3** (19%),¹⁶ and unreacted **1** (10%) after chromatography (chloroform–acetone gradient 40:1 → 3:1).

Acetolysis of **2** (0.58 mmol) with Hg(OAc)₂ (0.71 mmol) in acetic acid–acetic anhydride (2.76 ml; 10:1) followed by chromatography (toluene–acetone gradient 4:1 → 3:1) gave pure **4** and a mixture of **4** and **5** in a total yield of 96% (**4**:**5 ca.** 5:1). Hydrolysis of **2** (1.6 mmol) followed by acetylation of the intermediate β-hemiacetal¹⁷ gave pure **5** (83%).

Treatment of the **4–5** mixture (0.53 mmol) with ethanethiol (1.05 mmol) and boron trifluoride etherate (BF₃·Et₂O, 2.7 mmol) in dichloromethane (2.5 ml) gave, after chromatography (toluene–acetone 3:1) pure **6** in 93% yield; no β-anomer was detected. Similar treatment of **5** gave **6** in 86% yield.

A comparative glycosylation of the hexabenzyl lactoside **9**⁷ was performed (Table 1) with donors **6**, **7**³ and **8**⁴ using either methyl sulfonyl bromide–silver trifluoromethanesulfonate¹⁸ or *N*-iodosuccinimide–trifluoromethanesulfonic acid⁴ as promoters.

The new donor **6** gave the GM₃-trisaccharide **10** in good yield and very high stereoselectivity, with both the methods used for anomeric activation. Note also the high yield obtained when **6** and **9** were used in a molar ratio of 1:1.

The donors **7**³ and **8**⁴ have been used extensively for sialylation of the 3-position of galactose residues; good yields (60–80%) have been reported with acceptors having two or three hydroxy groups unprotected.^{4,19,20} However, sialylation of the sterically congested acceptor **9** with **7** and **8** proceeded in only 30–40% yield of GM₃-saccharide **11** (Table 1) and with concomitant formation of the corresponding β-glycoside **12**.

The auxiliary PhS-group was removed by treatment of **10** (0.12 mmol) with triphenyltin hydride (1.2 mmol)–AIBN (0.09 mmol), thus giving **11** (83%) and unreacted **10** (12%) after chromatography (toluene–MeCN gradient 4:1 → 2:1). We found that triphenyltin hydride is superior to tributyltin hydride, which gave **11** in low yield.

De-*O*-benzylation, de-*O*-acetylation, and hydrolysis of the methyl ester of **11** and **12** gave the TMSEt glycosides **13** and **14** (as the sodium salts) in 98 and 96% yields, respectively.

The anomeric configuration of the sialic acid residues of **2**, **4–6**, **10–14** were determined by measuring the long-range JC(1)–H(3)_{ax} coupling constant.²¹ As seen in Table 2, all sialic acid residues having an axial carboxyl (ester) group (as in α-glycosides) show couplings in the range 5.8–7.5 Hz, whereas

the corresponding equatorial carboxyl compounds show couplings in the range 1.0–1.7 Hz.

Values of δ H(3)_{eq} have been suggested to be smaller for β- than for α-glycosides.²² Data in Table 2 serve as a caveat in this respect, since the chemical shift order is reversed in the protected pair **11–12** as compared to the unprotected pair **13–14**.

JH_{7,8} has also been used as an anomeric configuration probe (*ca.* 2 and >7 Hz indicating β and α configuration).²³ In addition, the chemical-shift difference between the two hydrogens at position 9 [Δδ H(9)–H(9')] is reported to depend on the anomeric configuration, Δδ being *ca.* 1 for β glycosides and <0.5 for α glycosides.²³ However, these empirical rules do not apply for the 2-chloro and 2-*O*-acetyl compounds **2** and **5** (Table 2).

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