A Novel Mimic of the Sialyl Lewis x Determinant

Nigel M. Allanson*, Alan H. Davidson and Fionna M. Martin

Medicinal Chemistry Department, British Bio-technology, Brook House, Watlington Road, Cowley, Oxford. OX4 5LY U.K.

Abstract: The syntheses of two novel disaccharides are described which block the interaction of the lectin domain of E-selectin with its glycosylated ligands on neutrophils.

The interaction of tumour cells and neutrophils with the activated endothelium of blood vessels is the initial event which allows them to pass from the circulation into the tissue. The process is mediated by several factors, one of which is the interaction of the lectin domain in E-selectin with its glycosylated ligands on the circulating cells¹. Blockade of this interaction might represent a possible therapy for; (i) reperfusion injury, ARDS and Crohn's disease in which excessive accumulations of neutrophils are found in the inflamed tissue, (ii) prevention of metastasis of certain types of tumour.

Figure 1

NeuNAcα (2→3)Galβ(1→4)GlcNAc Fucα (1→3)↓	NeuNAca (2	NcuNAc α(2→3)Galβ (1→4)Glc Fucα (1→3)—

sLex², sLea³ and 3'-sialyl-3-fucosyl lactose⁴ (Figure 1) all have been shown to bind to the E-selectin. It would appear that they possess a common epitope^{4,5}, formed by the fucose, sialic acid and by parts of the galactose residue which interacts with the receptor. In order to interfere with this process using simpler molecules, we decided to link the C-4 position of fucose to the C-2 position of sialic acid by a 6 atom chain, hoping to create a mimic of this common domain. The chain would serve not only to maintain the relative distance and orientation of these residues but would also result in the introduction of functionality with which we could represent the galactose.

Figure 2



a) Silver salicylate/ CH2=CHCH2OH/ 4A sieves 95% b) O3/ MeOH 09C then Me2S 60%.

The linker was constructed by coupling a sialic acid aldehyde 1 to a fucosyl ketophosphonate 2a. The sialic acid aldehyde was prepared in two steps from methyl tetra-O-acetyl- α 2-chloro-N-acetylneuraminidate⁶ (Figure 2). Reaction with allyl alcohol in the presence of silver salicylate⁶ and powdered 4A sieves gave the α -allyl glycoside 3 in 95% yield. Ozonolysis in methanol at 0°C followed by a dimethyl sulphide workup gave the sialic acid aldehyde 1 in 55% yield after chromatography on silica.

The starting material for the fucose ketophosphonate 2a was methyl 2,3-di-O-benzoyl- α -L-fucopyranoside 47 (Figure 3). Oxidation with pyridinium chlorochromate and 4A sieves gave the fucosyl ketone 5 in 80% yield. Elaboration of 5 to the desired ketophosphonate proved difficult because of its sensitivity to base, namely the elimination of the 2-O benzoyl group to the enone 6. It was therefore decided to add a 3-carbon nucleophile to the *Si* face of 5 under Lewis acid conditions and then to develop this into an equatorial ketophosphonate group. Although addition of allyl silane using TiCl₄⁸ or SnCl₄ in CH₂Cl₂ at -78%C gave excellent yields of a homoallylic alcohol 7, NOE studies⁹ showed that attack had occurred exclusively from the opposite face. Although the undesired isomer had been obtained, it was decided to take this through to the ketophosphonate 2b. After much experimentation we found this could be achieved via epoxidation with MCPBA (92%), formation of a pair of bromohydrins with Li₂NiBr₄¹⁰(98%), oxidation with PCC and 4A sieves (78%), and an Abusov reaction with triisopropylphosphite¹¹ at 80%C (39%) which yielded 2b¹². However, attempts to invert the 4-hydroxyl on either 7 or 2b were unsuccessful. This forced us to re-examine basic nucleophilic additions to compound 5.





a. PCC CH₂Cl₂ 4A sieves 12hr 80% b. Ph₃PCHCO₂Et PhH 20% c. (1) CH₃COCH₂PO(OMe)₂, 2nBuLi THF 0^oC (2) CITi(Oi-Pr)₃-78^oC (3) 5 -78^oC 2hr 43% d. CH₂=CHCH₂Si(Mc)₃ TiCl₄ CH₂Cl₂ -78^oC 2hr 95% e. MCPBA CH₂Cl₂ 48hr 92% f. Li₂NiBr₄ THF 2hr 98% g. PCC CH₂Cl₂ 4A sieves 12hr 78% h. P(Oi-Pr)₃ 80^oC 2hr 39%.

Addition of the lithium dianion of dimethyl-2-oxopropyl phosphonate¹³ gave a 15% yield of the desired equatorial fucose ketophosphonate **2a** with no contaminating axial byproduct. The stereochemistry at C-4 was assigned from the NOE enhancements of 3% and 5% observed from the axial H-3 and H-5 protons of the fucose ring to the CH₂ protons between the ketone and C-4. No enhancement was observed between the CH₂ group and H-2. Encouraged by this result, we examined the effects of metal additives to modify the basicity of the nucleophile. Conducting the addition in the presence of CeCl₃¹⁴gave a 29% yield of **2a**, however the best yield was obtained using TiCl(Oi-Pr)₃¹⁵. Thus addition of one equivalent of **5** to two equivalents of the lithium dianion of dimethyl-2-oxopropyl phosphonate with 1.2 equivalents of chlorotriisopropoxy titanium in THF at -78°C gave only the desired equatorial fucose ketophosphonate **2a** in 43% yield.

The Wadsworth-Emmons reaction of sialic acid-aldehyde 1 with the fucosyl ketophosphonate 2a (Figure 4) proved unexpectedly demanding owing to the base sensitivity of the aldehyde component and of the disaccharide enone 8. Caesium carbonate in t-butanol¹⁶ was much superior to any of fifteen other conditions tried for this reaction. The reaction of 1 with 2a gave an optimal 58% yield of 8 after 2hr. The enone was quantitatively converted to the ketone 9 by reduction with hydrogen over a 10% palladium on charcoal catalyst. The alcohol protecting groups were removed with a catalytic amount of sodium in methanol to give the methyl ester 10 in 65% yield. 10 was quantitatively saponified to the sodium carboxylate salt 11 with sodium hydroxide in water.

Figure 4



a. Cs2CO3 t-BuOH 2hr 57% b. H2 10% Pd-C McOH 100% c. NaOMe (cat) MeOH 12hr 65% d. NaOH, H2O 100%.

Both methyl ester 10 and the sodium carboxylate salt 11 blocked the adhesion of an sLex expressing myeloid cell line U937 to activated endothelial cell cultures at twenty-five and thirty-fold higher concentrations than sLex. Full details of the biological results will be published elsewhere.

References

- 1. (a) Springer, T. A. Nature, 1990, 346, 425. (b) Osborn, L. Cell, 1990, 62, 3.
- (a) Lowe, J. B.; Stoolman, L. M.; Nair, R. P.; Larsen, R. D.; Berhend, T. L.; Marks, R. M. Cell 1990, 63, 475. (b) Phillips, M. L.; Nudelman, E.; Gaeta, F. C. A.; Perez, M.; Singhal, A. K.; Hakomori, S.; Paulson, J. C. Science 1990, 250, 1130. (c) Walz, G.; Aruffo, A.; Kolanus, W.; Bevilacqua, M. P.; Seed, B. Science 1990, 250, 1132.
- (a) Berg, E. L.; Robinson, M. K.; Mansson, O.; Butcher, E.C.; Magnani, J. L. J. Biol. Chem., 1991, 266, 14869. (b) Takada, A.; Ohmori, K.; Takahashi N.; Tsuyuoka, K.; Yago, A.; Zenita K.; Hasegawa A.; Kannagi, R. Biochem. Biophys. Res. Comm. 1991, 179, 713.

- 4. Tyrrell, D.; James, P.; Rao, N.; Foxall, C.; Abbas, S.; Dasgupta, F.; Nashed, M.; Hasegawa, A.; Kiso, M.; Asa, D.; Kidd, J.; Brandley B. K. Proc. Natl. Acad. Sci. USA, 1991, 88, 10372.
- (a) Ball G. E.; O'Neill, R. A.; Schultz, J. E.; Lowe, J. B.; Weston, B. W.; Nagy, J. O.; Brown, E. G.; Hobbs, C. J.; Bednarski, M. D. J. Am. Chem. Soc. 1992, 114, 5449. (b) Lin, Y-C.; Hummel, C. W.; Huang, D-H.; Ichikawa, Y.; Nicolaou, K. C.; Wong, C-H. J. Am. Chem. Soc. 1992, 114, 5452.
- 6. Roy, R.; Laferriere, C. A. Can. J. Chem., 1990, 68, 2045.
- 7. Lindhorst, T. K.; Thiem J. Carbohydr. Res., 1991, 209, 119.
- 8. Hosomi, A.; Sakurai, H. Tet. Lett., 1976,1295.
- 9. An NOE enhancement of 11% was observed between H-2 ring proton and the allylic CH₂ group.
- 10. Turner, J. V. Tet. Lett., 1984, 25, 2061.
- 11. Karanewsky, D. S.; Badia, M.C.; Ciosek, C. P.; Robl J. A.; Sofia, M. J.; Simpkins, L. M.; DeLange, B.; Harrity, T.W.; Biller, S. A.; Gordon, E.M. J. Med. Chem., 1990, 33, 2952.
- 12. In the presence of Cs₂CO₃ and t-butanol **2b** also reacted with aldehyde **1** to give a 38% yield of the 4fucosyl hydroxy epimer of disaccharide **8**.
- 13. Grieco P.A.; Pogonowski, C. S. J. Amer. Chem. Soc., 1973, 95, 3072.
- 14. Paquette, L. A.; Combrink, K. D.; Elmore, S. W.; Rogers, R. D. J. Amer. Chem. Soc., 1991, 113, 1335.
- 15. Reetz, M. T.; Steinbach, R.; Westermann, J.; Peter, R. Angew. Chem. Int. Ed. Eng., 1980, 19, 1011.
- Blackwell C. M.; Davidson, A. H.; Launchbury S. B.; Lewis, C. N.; Morrice, E. M.; Reeve, M. M.; Roffey, J. A. R.; Tipping, A. S.; Todd, R. S. J. Org. Chem., 1992, 57, 1935.

Selected analytical data on compounds 2b and 10. The following abbreviations are used: S for sialic acid, F for fucose and L for the linker. The numbering system in the linker runs away from the fucosyl ring.

Compound **2b:** ¹<u>H nmr</u>: (CDCl₃) δ 7.99-7.87 (4H, 2d, 2 x OBz); 7.52-7.26 (6H, m, 2 x OBz); 5.82(H, d, J = 10.2 Hz, H-3); 5.49 (H, dd, J = 3.8 and 10.2 Hz, H-2); 5.14 (H, d, J = 3.8 Hz, H-1); 4.24 (H, q, J = 6.4 Hz, H-5); 4.10 (H, br s, OH); 3.71 (3H, d, J = 11.3 Hz, MeOP); 3.64 (3H, d, J = 11.3 Hz, MeOP); 3.41 (3H, s, F1-OMe); 3.10-2.99 (2H, dd, J = 5.5 and 22.6 Hz, (MeO)₂OP<u>CH₂CO-</u>); 2.86 (2H, s, CO<u>CH₂C-4); 1.30 (3H, d, J = 6.4 Hz).</u> ¹³<u>C nmr</u>: (CDCl₃) δ 200.6 (-<u>C</u>O-), 166.2, 166.0 (2 x O<u>C</u>OPh), 133.3, 133.3, 129.8, 129.7, 129.3, 129.2, 128.4, 128.2 (2 x Ph); 97.0 (C-1), 75.1 (C-4), 72.9, 70.6, 67.8, 60.3 C1-O<u>C</u>H₃), 53.0, 52.9 (P(O<u>C</u>H₃)₂), 46.8 (CO<u>C</u>H₂C-4), 43.58 and 41.6 (P<u>C</u>H₂CO), 13.5 (C-6). High resolution mass spectrum: C₂₆H₃₁O₁₁P [M+H]+ calc.551.16823; found 551.16769.

Compound 10: ^{1}H nmr: (D₂O) δ 4.84 (H, d, F-1); 4.07 (H, q, J = 6.4 Hz, F-5); 3.83 (3H, s, S-OMe); 3.82-3.39 (11H, m, F-2, F-3, S-4, S-5, S-6, S-7, S-8, 2 x S-9, 2 x L-5); 3.36 (3H, s, F-OMe); 2.87 (H, d, J = 16.4 Hz, L-1); 2.66-2.59 (4H, m, L-1', 2 x L-3, S-3e); 1.99 (3H, s, NHAc); 1.87-1.70 (3H, m, 2 x L-4, S-3a); 1.12 (3H, d, J = 6.4 Hz, F-6). ^{13}C nmr: (D₂O) δ 213.5 (L-2), 175.2 (S-1), 178.4 (NHAc), 99.5 (S-2), 99.4 (F-1), 76.0 (F-4), 73.1 (S-8), 71.3 (F-2), 70.9 (F-3), 69.2 (S-7), 68.5 (S-6), 68.1 (F-5), 67.4 (S-4), 64.1 (L-5), 63.3 (S-9), 55.3 (F-OMe), 53.6 (S-OMe), 52.0 (S-5), 45.5 (L-1), 40.6 (L-3), 39.4 (S-3), 23.2 (L-4), 22.3 (NHAc), 13.0 (F-6).

High resolution mass spectrum: $C_{24}H_{41}NO_{15}$ [M+Na]+ calc.606.23739; found 606.23763.

(Received in UK 27 April 1993)