

## Stereochemistry of the Decarboxylation of Phenolic Cinnamic Acids by *Saccharomyces cerevisiae*

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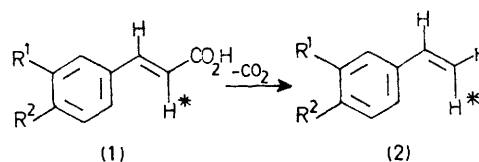
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**Summary** The decarboxylation of (*E*)-3,4-dimethoxycinnamic acid to the corresponding styrene by *S. cerevisiae* proceeds with retention of the hydrogen atom at the  $\alpha$ -position of the acid; the configuration of the double bond is also retained.

ALTHOUGH the non-oxidative decarboxylation of phenolic cinnamic acids [*e.g.* *p*-coumaric (**1a**), ferulic (**1b**), and caffeic (**1c**) acids] to the corresponding styrenes (**2**) occurs widely in bacteria<sup>1,2</sup> and in yeast,<sup>1,3</sup> the enzymic mechanism is unknown. As a preliminary approach to its clarification, the stereochemical course of the reaction has been investigated. We now report that the decarboxylation of 3,4-dimethoxycinnamic acid (**1d**) by a strain of *Saccharomyces cerevisiae*<sup>†</sup> results in retention of configuration at the side-chain double bond (Scheme).

A culture medium<sup>‡</sup> containing a suspension of (*E*)-3,4-dimethoxy[ $\alpha$ -<sup>2</sup>H]cinnamic acid (**1d**, H\* = <sup>2</sup>H) (D atoms per molecule  $0.80 \pm 0.03$  by m.s.)<sup>4</sup> was inoculated with

yeast and shaken at 25° for 24 h. The ether extract of the fermentation medium, when evaporated and chromatographed on silica gel [light petroleum (b.p. 40–70°)–benzene 1:1], gave (*Z*)-3,4-dimethoxy[ $\beta$ -<sup>2</sup>H]styrene (**2d**, H\* = <sup>2</sup>H)<sup>5</sup> (62% yield; D atoms per molecule  $0.75 \pm 0.03$



- (a), R<sup>1</sup> = H, R<sup>2</sup> = OH  
 (b), R<sup>1</sup> = OMe, R<sup>2</sup> = OH  
 (c), R<sup>1</sup> = OH, R<sup>2</sup> = OH  
 (d), R<sup>1</sup> = OMe, R<sup>2</sup> = OMe

SCHEME

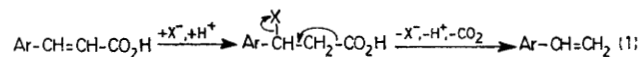
<sup>†</sup> This strain (28 C)<sup>3</sup> is unique in decarboxylating both 3,4-dimethoxycinnamic acid and ferulic acid. Its use made our investigation easier, 3,4-dimethoxystyrene being more stable than 4-hydroxystyrenes.

<sup>‡</sup> Glucose (100 g), yeast nitrogen base (Difco) (7 g), NaH<sub>2</sub>PO<sub>4</sub> (13 g), water (1 l).

by m.s.). The position of the deuterium atom was assigned by comparison of the  $^1\text{H}$  n.m.r. spectrum (vinyl group region) of (2d) with the spectral patterns calculated for each of the three 3,4-dimethoxystyrenes monodeuteriated in their side-chain. The calculations, (using the second-order perturbation method), were based on the chemical shifts and spin-coupling constants of the vinyl protons of 3,4-dimethoxystyrene ( $\delta_A$  5.56,  $\delta_B$  5.12,  $\delta_X$  6.66 p.p.m.;  $J_{AX}$  17.5,  $J_{BX}$  10.6,  $J_{AB}$  1.4 Hz in  $\text{CDCl}_3$ ) and assumed (a) that  $J_{HD} = (\gamma_D/\gamma_H)J_{HH}$  and (b) that the chemical shifts are not affected by deuterium substitution.<sup>6</sup> The correctness of these assumptions was confirmed by comparing the observed and theoretical spectrum of (*E*)-3,4-dimethoxy- $[\beta\text{-}^2\text{H}]$ styrene prepared by unequivocal synthesis *via*  $\text{D}_2\text{O}$  decomposition of the Grignard reagent<sup>6</sup> of *trans*-3,4-dimethoxy- $\beta$ -bromostyrene.<sup>7</sup>

If the hypothesis is made that the *in vivo* decarboxylation of cinnamic acids takes place similarly to the *in vitro* pyridine or thioacetic acid-catalysed decarboxylation of benzylidenemalononic acid derivatives,<sup>8</sup> *i.e.* by a 1,2-addition,

1,2-elimination mechanism (equation 1), then a *cis*-addition followed by a *trans*-decarboxylative elimination (or a *trans*-addition and *cis*-elimination) must be assumed to account for the overall stereochemistry of the process.



( $\text{X}^-$  = nucleophilic group of the enzyme, *e.g.*  $\text{RS}^-$ ,  $\text{RO}^-$ )

It is also remarkable that (*Z*)-3,4-dimethoxycinnamic acid<sup>9</sup> does not undergo decarboxylation by the above strain of *S. cerevisiae*.

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<sup>1</sup> R. D. Steinke and M. C. Paulson, *J. Agric. Food Chem.*, 1964, **12**, 381.

<sup>2</sup> S. R. Indahl and R. R. Scheline, *Appl. Microbiol.*, 1968, **16**, 667; B. J. Finkle, J. C. Lewis, J. W. Corse, and R. E. Lundin, *J. Biol. Chem.*, 1962, **237**, 2926.

<sup>3</sup> G. Albagnac and P. Dubois, Les phénols volatils des milieux fermentés in 'Comptes rendus de l'Assemblée 1972 du Groupe Polyphénols,' Station de Technologie des Produits Végétaux (I.N.R.A.), Narbonne, 1973.

<sup>4</sup> P. Manitto, D. Monti, P. Gramatica, and E. Sabbioni, *J.C.S. Chem. Comm.*, 1973, 563.

<sup>5</sup> G. Redeuilh, P. Rumpf, and C. Viel, *Bull. Soc. chim. France*, 1973, 2665.

<sup>6</sup> T. Yoshino, Y. Manabe, and Y. Kikuchi, *J. Amer. Chem. Soc.*, 1964, **86**, 4670.

<sup>7</sup> E. Adler and K. J. Björkqvist, *Acta Chem. Scand.*, 1951, **5**, 241; E. R. Trumbull, R. T. Finn, K. M. Ibne-Rasa, and C. K. Sauers, *J. Org. Chem.*, 1962, **27**, 2339.

<sup>8</sup> E. J. Corey and G. Fraenkel, *J. Amer. Chem. Soc.*, 1953, **75**, 1168.

<sup>9</sup> E. Adler and B. Gustafsson, *Acta Chem. Scand.*, 1963, **17**, 27.