# Synthesis and structure elucidation of polyphenols containing the $N^{\prime}$ methyleneformohydrazide scaffold as aurora kinase inhibitors 

Short title: Examination of $N^{\prime}$-methyleneformohydrazide-containing polyphenols

Dongsoo Koh, ${ }^{1, \dagger}$ Yearam Jung, ${ }^{2, \dagger}$ Seunghyun Ahn, ${ }^{2}$ Kenneth Hun Mok, ${ }^{3}$ Soon Young Shin, ${ }^{4, *}$ and Yoongho Lim ${ }^{2, *}$
${ }^{1}$ Department of Applied Chemistry, Dongduk Women's University, Seoul 136-714, Korea
${ }^{2}$ Division of Bioscience and Biotechnology, BMIC, Konkuk University, Seoul 143-701, Korea
${ }^{3}$ School of Biochemistry and Immunology, Trinity College, University of Dublin, Dublin 2, Ireland
${ }^{4}$ Department of Biological Sciences, Konkuk University, Seoul 143-701, Korea
${ }^{\dagger}$ D. Koh and Y. Jung contributed equally to this work.

## Corresponding authors:

Prof. Yoongho Lim, Division of Bioscience and Biotechnology, Konkuk University, 12 0 Neungdong-ro, Gwangjin-gu, Seoul, Korea 05029. Email: yoongho@konkuk.ac.kr, Ph one: 82-2-450-3760, Fax: 82-2-3437-6106

Dr. Soon Young Shin, Department of Biological Sciences, Konkuk University, 120 Neungdong-ro, Gwangjin-gu, Seoul, Korea 05029, Email: shinsy@konkuk.ac.kr , Phone: 82-2-2030-7946

Keywords: NMR, ${ }^{1} \mathrm{H}$ NMR, ${ }^{13} \mathrm{C}$ NMR, polyphenols, $N^{\prime}$-methyleneformohydrazide, HR/MS, aurora kinase inhibitors

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process whi ch may lead to differences between this version and the Version of Record. Please cit e this article as doi: 10.1002/mrc. 4611

## Introduction

Aurora kinases play important roles in mitosis and cell division. ${ }^{[1]}$ Three kinases family has been known as aurora $\mathrm{A}(\operatorname{aurA})$, B (aurB), and C (aurC). They have conserved catalytic domains, but their function and localization are not the same. ${ }^{[2-4]}$ AurA localizes to spindle poles in mitosis and is essential to centrosome maturation and separation. AurB localizes to the microtubules interlinking at the spindle and is required for phosphorylation of histone H3 ${ }^{[5,6]}$ Unlike aurA and aurB, aurC has not been studied well. It was found aurA and aurB were overexpressed in colon and breast tumors. ${ }^{[7,8]}$ Other researches revealed aurA was overexpressed in gastric, ovarian, and pancreatic tumors. ${ }^{[9-12]}$ Similar results were found from aurB. ${ }^{[5]}$ Because their overexpression associates with tumorigenesis, they are targeted for cancer therapy. ${ }^{[2,13,14]}$ Of many possible ways to inhibit the enzymatic activities of aurora kinases, one is to use small compounds as inhibitors. Several effective inhibitors have been known as follows: $N$-[(3Z)-2-oxo-3-[phenyl-[4-(piperidin-1-ylmethyl)anilino]methylidene]1 H -indol-5-yl]ethanesulfonamide (named as hesperadine, Suppl. Fig. 1A), $N$-(2-chlorophenyl)-4-[[2-[[4-[2-(4-ethyl-1-piperazinyl)-2-oxoethyl]phenyl]amino]-5-fluoro-4-pyrimidinyl]amino]-benzamide (named as TC-S7010, Suppl. Fig. 1B), N-[4-[[6-methoxy-7-(3-morpholin-4-ylpropoxy)quinazolin-4-yl]amino]phenyl]benzamide (named as ZM447439, Suppl. Fig. 1C), and $N$-[4-[4-(4-methylpiperazin-1-yl)-6-[(5-methyl-1H-pyrazol-3-yl)aminolpyrimidin-2-yl]sulfanylphenyl]cyclopropanecarboxamide (named as VX-680, Suppl. Fig. 1D). ${ }^{[13-16]}$ Besides, it has been reported flavones [Fig. 1A] inhibit aurB. ${ }^{[17]}$ Benzochalcones bearing pyrazoline moieties [Fig. 1B] show the inhibitory effects on aurora kinases. ${ }^{[18]}$ Chromenylchalcones [Fig. 1C] bind to aurA based on their in silico docking study. ${ }^{[19]}$ They all belong to polyphenols. We designed and synthesized 33 polyphenols containing the $N^{\prime}$-methyleneformohydrazide scaffold based on moieties contained in Figs. 1A, B, and C as listed in Table 1. To confirm whether they show the inhibitory effects on aurA, in
vitro kinase assay was performed. Their half-maximal inhibitory concentration $\left(\mathrm{IC}_{50}\right)$ ranged between 1.14 and $98.93 \mu \mathrm{M}$. We report here the assignments of their ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR data and high-resolution mass spectral data. These data can help us design a series of compounds containing novel aurora kinase inhibitor pharmacophores and identify newly synthesized polyphenols containing the $N^{\prime}$-methyleneformohydrazide scaffold.

## -

## Experimental

## General synthetic route

A synthetic scheme outlining the preparation of the polyphenol derivatives containing the $N^{\prime}$ methyleneformohydrazide scaffold (i.e., derivatives 1-33) is shown in Scheme 1. For the preparation of derivatives $\mathbf{1} \mathbf{- 2 0}$, the intermediate chromone aldehydes II were synthesized from substituted 2-hydroxy acetophenones under Vilsmeier reaction conditions. Substituted 2-hydroxy acetophenone (I) was treated with an excess of phosphorous oxychloride and anhydrous $N, N$-dimethylformamide to afford chromone aldehyde (II), which was subsequently treated with benzohydrazide to give the corresponding derivatives $\mathbf{1 - 2 0}$. Resveratrol derived aldehyde III was also employed under the same reaction conditions to obtain the desired compounds 21-26 and 29-30. ${ }^{[20]}$ Typical synthetic procedures for the preparation of derivatives $\mathbf{9}$ and $\mathbf{2 2}$ are described as follows.


Preparation of $(E)$ - $N^{\prime}$-((6-chloro-4-oxo-4H-chromen-3-yl)methylene)-3-hydroxybenzo hydrazide (9)

A mixture of anhydrous $N, N$-dimethylformamide ( $15 \mathrm{~mL}, 193 \mathrm{mmol}$ ) and phosphoryl chloride ( $1.5 \mathrm{~mL}, 23 \mathrm{mmol}$ ) was heated at $60-65^{\circ} \mathrm{C}$ for 1.5 h , after which time 2-hydroxy-5chloroacetophenone ( $5 \mathrm{mmol}, 850 \mathrm{mg}$ ) was added, and the resulting mixture stirred at the same temperature for a further 2 h . The reaction progress was monitored by TLC. After
reaching completion, the reaction mixture was cooled to room temperature and poured into ice-water ( 400 mL ), then neutralized using a saturated $\mathrm{NaHCO}_{3}$ solution. This resulted in precipitation of the crude 6-chloro-3-formylchromone product, which was subsequently filtered then purified by recrystallization in ethanol (Yield: $80 \%$; m.p: $166-168{ }^{\circ} \mathrm{C}$ ). The purified 6-chloro-3-formylchromone ( $60 \mathrm{mg}, 0.3 \mathrm{mmol}$ ) was then dissolved in ethanol ( 10 mL ) containing $m$-hydroxybenzohydrazide ( $45 \mathrm{mg}, 0.3 \mathrm{mmol}$ ). A catalytic amount of glacial acetic acid was added, and the resulting reaction mixture was heated under reflux at $85^{\circ} \mathrm{C}$ for 5 h . After this time, the reaction mixture was cooled to room temperature, the solvent was removed under vacuum, and the resulting residue was purified by column chromatography to give the analytically pure compound 9 (Yield: $77 \%$; m.p: $196-202{ }^{\circ} \mathrm{C}$ ).

Preparation of $(E)$ - $N^{\prime}$-(2,4-dimethoxy-6-(4-methoxystyryl)benzylidene)-3-methoxybenzo hydrazide (22)
(E)-2-(4-methoxystyryl)-4,6-dimethoxybenzaldehyde (90 mg, 0.3 mmol ) and $m$-anisic hydrazide ( $48 \mathrm{mg}, 0.3 \mathrm{mmol}$ ) were dissolved in ethanol $(10 \mathrm{~mL}){ }^{[20]}$ To this solution was added a catalytic amount of glacial acetic acid, and the reaction mixture was heated under reflux at $85^{\circ} \mathrm{C}$ for 7 h . After this time, the mixture was cooled to room temperature, the crude product was precipitated, and the resulting solid was filtered prior to purification by recrystallization in methanol (Yield: $72 \%$; m.p: $160-164^{\circ} \mathrm{C}$ ).

## NMR spectroscopy

The synthesized polyphenols containing the $N^{\prime}$-methyleneformohydrazide scaffold were dissolved in deuterated dimethyl sulfoxide (DMSO- $d_{6}$ ) to a concentration of $\sim 50 \mathrm{mM}$ and transferred into 2.5 mm NMR tubes. All NMR data were collected using a Bruker AVANCE 400 spectrometer system ( 9.4 T; Bruker, Karlsruhe, Germany) equipped with a 5-mm inverse
broadband probehead at room temperature, and the chemical shifts were referenced to TMS $(0 \mathrm{ppm}) .{ }^{1} \mathrm{H}$ NMR experiments were performed using the following parameters: relaxation delay $=1 \mathrm{~s}, 90^{\circ}$ pulse for $11.8 \mu \mathrm{~s}, 5500 \mathrm{~Hz}$ spectral width, 32 k data points, and $0.34 \mathrm{~Hz} /$ point digital resolution. The equivalent parameters for the ${ }^{13} \mathrm{C}$ NMR and DEPT experiments were $3 \mathrm{~s}, 15.0 \mu \mathrm{~s}, 21000 \mathrm{~Hz}, 64 \mathrm{k}$, and $0.64 \mathrm{~Hz} /$ point, respectively. Twodimensional COSY, NOESY, HMQC, and HMBC experiments were acquired with data points of $2 \mathrm{k} \times 256\left(\mathrm{t}_{2} \times \mathrm{t}_{1}\right)$. The long-range coupling time for HMBC was 70 ms , and the mixing time for NOESY was 1.5 s . Processing and analysis of the NMR data were carried out as previously reported. ${ }^{[21]}$

## General analytical procedures

To confirm the structures of the $N^{\prime}$-methyleneformohydrazide-containing polyphenols, ultraperformance liquid chromatography-hybrid quadrupole-time-of-flight mass spectrometry (UPLC-TOFMS) was carried out using a Waters ACQUITY UPLC system (Waters, Milford, MA) with the help of Prof. Choong Hwan Lee at Konkuk University, Korea. ${ }^{[22]}$ All highresolution mass spectrometry (HR/MS) data were collected as $(\mathrm{M}-\mathrm{H})^{-}$ions.

In vitro aurora A kinase assay
The substrate used in vitro human aurora A kinase assay was $200 \mu \mathrm{M}$ LeuArgArgAlaSerLeuGly and the concentration of ATP was $500 \mathrm{cpm} / \mathrm{pmol}$. The detailed procedure followed to the method provided by EMD Millipore's KinaseProfiler service assay protocol (EMD Millipore Corporation, Billerica, MA). ${ }^{[23]}$ TC-S7010 [Suppl. Fig. 1B] was used as a control. The statistical significance was analyzed using Student's t-test. All experiments were performed in triplicate

## Results and Discussion



All 33 derivatives containing the $N^{\prime}$-methyleneformohydrazide scaffold are listed in Table 1. As mentioned previously, these derivatives can be divided into a number of groups. For example, in the structures of derivatives $\mathbf{1} \mathbf{- 1 7}$, the 4 H -chromenyl group is attached to the N methylene functionality and the carbonyl group bears a substituted phenyl group. Similarly, in derivatives 18-20, the 4 H -chromenyl group is attached to the N -methylene moiety, but the carbonyl group bears a substituted pyridine unit. Furthermore, in derivatives 21-26, both the N -methylene and carbonyl groups bear substituted phenyl groups, while in derivatives 27-30, a phenyl group is attached to the $N$-methylene moiety and the carbonyl group bears a substituted pyridine group. Finally, in derivatives 31-33, methyl and phenyl groups are attached to the $N$-methylene functionality, while a substituted phenyl group is found on the carbonyl group. Of the 33 derivatives, the procedure for assigning the structure of derivative 22, $\quad$ i.e., $\quad(E)-N^{\prime}-((E)-2-((E)$-4-methoxystyryl)-4,6-dimethoxybenzylidene)-3methoxybenzohydrazide [Fig. 2], is described here as an example.

A total of 24 signals were observed in the ${ }^{13} \mathrm{C}$ NMR spectrum of $\mathbf{2 2}$, of which 5 were present at $\sim 160 \mathrm{ppm}$. Four of these peaks (with the exception of the signal at 162.6 ppm ) showed long-range couplings with methoxy protons in the HMBC spectrum, allowing the ${ }^{13} \mathrm{C}$ signal at 162.6 ppm to be assigned to the amide carbon, $\mathrm{C}-1 \mathrm{a}$. In addition, as the two ${ }^{13} \mathrm{C}$ signals at 114.1 and 128.4 ppm exhibited double the intensities of the neighboring signals, they were assigned to the $\mathrm{C}-3^{\prime \prime} / \mathrm{C}-5^{\prime \prime}$ and $\mathrm{C}-2^{\prime \prime} / \mathrm{C}-6^{\prime \prime}$ atoms, respectively. The two proton signals at 6.95 and 7.69 ppm were assigned to the $\mathrm{H}-3^{\prime \prime} / \mathrm{H}-5^{\prime \prime}$ and $\mathrm{H} 2^{\prime \prime} / \mathrm{H}-6^{\prime \prime}$ atoms, respectively. In addition, the ${ }^{13} \mathrm{C}$ peaks at 130.3 and 159.0 ppm exhibiting ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ couplings with $\mathrm{H}-3^{\prime \prime} / \mathrm{H}-5^{\prime \prime}$ and $\mathrm{H} 2^{\prime \prime} / \mathrm{H}-6^{\prime \prime}$, respectively, were assigned as $\mathrm{C}-1^{\prime \prime}$ and $\mathrm{C}-4^{\prime \prime}$, respectively. The ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling of C-4" to a methoxy proton at 3.78 ppm allowed this to be assigned as the
$4^{\prime \prime}-\mathrm{OCH}_{3}$ proton, while the ${ }^{13} \mathrm{C}$ peak at 55.1 ppm was assigned to the 4 " $-\mathrm{OCH}_{3}$ carbon atom based on HMQC results. Furthermore, due to the large coupling constants of the two proton signals at 7.20 and 8.51 ppm (i.e., 16.3 Hz ), these signals were assigned to the double bond protons of the styryl group. Their corresponding carbon atoms were observed at 130.4 and 126.1 ppm , respectively. As a ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling between the former carbon atom and the $\mathrm{H} 2^{\prime \prime} / \mathrm{H}-$ $6^{\prime \prime}$ atoms was observed by HMBC, this carbon atom was assigned as $\mathrm{C}-1$ "a. We also observed three correlating proton signals at $7.15,7.44$, and 7.56 ppm in the COSY spectrum, where the signal at 7.56 ppm exhibited a ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling with $\mathrm{C}-1 \mathrm{a}$, and so it was assigned as $\mathrm{H}-6$. In addition, the carbon atom resonating at 117.4 ppm that was attached directly to the proton resonating at 7.15 ppm exhibited a ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling to $\mathrm{H}-6$, and so the proton signal at 7.15 ppm was assigned to the $\mathrm{H}-4$ atom. The remaining proton signal at 7.44 ppm was assigned to the $\mathrm{H}-5$ atom, while the carbon signal at 159.2 ppm exhibiting a ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling with H-5 was assigned to the C-3 atom. Furthermore, the methoxy proton signal at 3.85 ppm showed a ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling with the $\mathrm{C}-3$ atom, and it was assigned as $3-\mathrm{OCH}_{3}$. Moreover, the two carbon signals at 112.7 and 134.9 ppm that exhibited ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ couplings with H-6 and H-5 were labelled C-2 and C-1, respectively. As expected, the most deshielded proton signal at 11.74 ppm was assigned to the NH group, which also exhibited $\mathrm{a}^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling to the carbon C-1'a signal at 145.1 ppm . The signals corresponding to the $\mathrm{C}-2^{\prime}$ and $\mathrm{C}-6^{\prime}$ atoms (at 139.0 and 160.3 ppm ) were assigned as such due to their ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ couplings with the $\mathrm{H}-1$ 'a signal at 8.86 ppm . We also observed the COSY correlation of the two protons resonating at 7.00 and 6.58 ppm . Since the former was long-range coupled to the $\mathrm{C}-1^{\prime \prime} \mathrm{b}$ signal ( 126.1 ppm ) with ${ }^{3} \mathbf{J}_{\mathrm{CH}}$, it was labeled as $\mathrm{H}-3^{\prime}$. As a result, the proton signal at 6.58 ppm was assigned to the $\mathrm{H}-$ $5^{\prime}$ atom. Furthermore, as the methoxy proton signal at 3.87 ppm exhibited a ${ }^{3} \mathbf{J}_{\mathrm{CH}}$ coupling with the $\mathrm{C}-6^{\prime}$ signal, it was assigned as $6^{\prime}-\mathrm{OCH}_{3}$. Thus, of the four methoxy protons present, the final proton signal at 3.89 ppm was assigned to the $4^{\prime}-\mathrm{OCH}_{3}$ group. The carbon signal at
161.1 ppm exhibiting $\mathrm{a}^{3} \mathrm{~J}_{\mathrm{CH}}$ coupling with this methoxy proton was labeled $\mathrm{C}-4^{\prime}$. Finally, the carbon signal at 112.8 ppm that exhibited ${ }^{3} \mathrm{~J}_{\mathrm{CH}}$ couplings with $\mathrm{H}-3^{\prime}$ and $\mathrm{H}-5^{\prime}$ was assigned to the $\mathrm{C}-1^{\prime}$ atom. The nOe cross peaks among $\mathrm{H}-1^{\prime \prime} \mathrm{b} / \mathrm{H}-6^{\prime \prime}, \mathrm{H}-5^{\prime \prime} / 4^{\prime \prime}-\mathrm{OCH}_{3}, \mathrm{H}-3^{\prime} / \mathrm{H}-1^{\prime \prime} \mathrm{a}, \mathrm{H}-$ $1^{\prime \prime} \mathrm{b} / \mathrm{H}-1^{\prime} \mathrm{a}, \mathrm{NH} / \mathrm{H}-1^{\prime} \mathrm{a}, \mathrm{NH} / \mathrm{H}-2$, and $\mathrm{H}-2 / 3-\mathrm{OCH}_{3}$ were observed in the NOESY spectrum. The key correlations obtained from the COSY, NOESY, and HMBC spectra of derivative 22 are shown in Fig. 3.

Similar procedures were employed to assign the NMR data of the other derivatives, and the complete assignments of the ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR data for all derivatives containing the $N^{\prime}$-methyleneformohydrazide scaffold (see Table 1) are listed in Tables 2 and 3, respectively. To confirm the structures of the various derivatives, HR/MS was carried out, and the calculated molecular masses are listed in Table 1 along with the experimental HR/MS results. All ${ }^{1} \mathrm{H}$ NMR, ${ }^{13} \mathrm{C}$ NMR, and HR/MS spectra are provided in the Supplementary Materials.

The $\mathrm{IC}_{50}$ values of 33 derivatives for aurora A kinase ranged from 1.14 to $98.93 \mu \mathrm{M}$ as listed in Table 1 and they were plotted with error bars [Suppl. Fig. 2]. The $\mathrm{IC}_{50}$ value of TC-S7010 [Suppl. Fig. 1B] used as a control was $0.25 \mu$ M. Derivative 6, ( $E$ )-4-fluoro- $N^{N}$-(( $(4-$ oxo-4H-chromen-3-yl)methylene)benzohydrazide showed the best inhibitory effect and derivative 21, $\quad(E)-N^{\prime}-((E)-2-((E)$-4-methoxystyryl)-4,6dimethoxybenzylidene)benzohydrazide did the worst effect. We could therefore conclude that the analytical data provided herein allows us to identify the novel $N^{\prime}$ -methyleneformohydrazide-containing derivatives, and in vitro kinase assay data on aurora A help us design compounds containing novel aurora kinase inhibitor pharmacophores.

## Acknowledgments

This work was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF-2016R1D1A1A09919045) and the Agri-Bio Industry

Technology Development Program (316028-3, Korea Institute of Planning and Evaluation for Technology in Food, Agriculture, Forestry and Fisheries (IPET)). SY Shin was supported by the KU Research Professor Program of Konkuk University.

## References

[1] R. D. Carvajal, A. Tse, G. K. Schwartz. Clin Cancer Res. 2006, 12, 6869-6875.
[2] S. Hauf, R. W. Cole, S. LaTerra, C. Zimmer, G. Schnapp, R. Walter, A. Heckel, J. van Meel, C. L. Rieder, J. M. Peters. J Cell Biol. 2003, 161, 281-294.
[3] D. Berdnik, J. A. Knoblich. Curr Biol. 2002, 12, 640-647.
[4] T. Hirota, N. Kunitoku, T. Sasayama, T. Marumoto, D. Zhang, M. Nitta, K. Hatakeyama, H. Saya. Cell. 2003, 114, 585-598.
[5] S. M. Lens, E. E. Voest, R. H. Medema. Nat Rev Cancer. 2010, 10, 825-841.
[6] C. Crosio, G. M. Fimia, R. Loury, M. Kimura, Y. Okano, H. Zhou, S. Sen, C. D. Allis, P. Sassone-Corsi. Mol Cell Biol. 2002, 22, 874-885.
[7] J. R. Bischoff, L. Anderson, Y. Zhu, K. Mossie, L. Ng, B. Souza, B. Schryver, P. Flanagan, F. Clairvoyant, C. Ginther, C. S. Chan, M. Novotny, D. J. Slamon, G. D. Plowman. EMBO J. 1998, 17, 3052-3065.
[8] S. Sen, H. Zhou, R. A. White. Oncogene. 1997, 14, 2195-2200.
[9] T. M. Gritsko, D. Coppola, J. E. Paciga, L. Yang, M. Sun, S. A. Shelley, J. V. Fiorica, S. V. Nicosia, J. Q. Cheng. Clin Cancer Res. 2003, 9, 1420-1426.
H. Katayama, T. Ota, F. Jisaki, Y. Ueda, T. Tanaka, S. Odashima, F. Suzuki, Y. Terada, M. Tatsuka. J Natl Cancer Inst. 1999, 91, 1160-1162.
[11] D. Li, J. Zhu, P. F. Firozi, J. L. Abbruzzese, D. B. Evans, K. Cleary, H. Friess, S. Sen. Clin Cancer Res. 2003, 9, 991-997.
[12] Y. Miyoshi, K. Iwao, C. Egawa, S. Noguchi. Int J Cancer. 2001, 92, 370-373.
[13] C. Ditchfield, V. L. Johnson, A. Tighe, R. Ellston, C. Haworth, T. Johnson, A. Mortlock, N. Keen, S. S. Taylor. J Cell Biol. 2003, 161, 267-280.
[14] E. A. Harrington, D. Bebbington, J. Moore, R. K. Rasmussen, A. O. Ajose-Adeogun, T. Nakayama, J. A. Graham, C. Demur, T. Hercend, A. Diu-Hercend, M. Su, J. M. Golec, K. M. Miller. Nat Med. 2004, 10, 262-267.
[15] N. Jetton, K. G. Rothberg, J. G. Hubbard, J. Wise, Y. Li, H. L. Ball, L. Ruben. Mol Microbiol. 2009, 72, 442-458.
[16] I. Aliagas-Martin, D. Burdick, L. Corson, J. Dotson, J. Drummond, C. Fields, O. W. Huang, T. Hunsaker, T. Kleinheinz, E. Krueger, J. Liang, J. Moffat, G. Phillips, R. Pulk, T. E. Rawson, M. Ultsch, L. Walker, C. Wiesmann, B. Zhang, B. Y. Zhu, A. G. Cochran. J Med Chem. 2009, 52, 3300-3307.
[17] Y. Jung, S. Y. Shin, Y. Yong, H. Jung, S. Ahn, Y. H. Lee, Y. Lim. Chem Biol Drug Des. 2015, $85,574-585$.
[18] S. Y. Shin, H. Yoon, D. Hwang, S. Ahn, D. W. Kim, D. Koh, Y. H. Lee, Y. Lim. Bioorg Med Chem. 2013, 21, 7018-7024.
[19] S. Y. Shin, H. Yoon, S. Ahn, D. W. Kim, S. H. Kim, D. Koh, Y. H. Lee, Y. Lim. Bioorg Med Chem. 2013, 21, 4250-4258.
[20] S. Ahn, Y. Lim, D. Koh. Acta Crystallogr E Crystallogr Commun. 2015, 71, o771.
[21] Y. Yong, S. Ahn, D. Hwang, H. Yoon, G. Jo, Y. H. Kim, S. H. Kim, D. Koh, Y. Lim. Magn Reson Chem. 2013, 51, 364-370.
[22] S. Ahn, S. Y. Shin, Y. Jung, H. Jung, B. S. Kim, D. Koh, Y. Lim. Magn Reson Chem. 2016, 54, 382-390.
[23] S. K. Sharma, P. Kumar, B. Narasimhan, K. Ramasamy, V. Mani, R. K. Mishra, A. B. Majeed. Eur J Med Chem. 2012, 48, 16-25.

Table 1. Structures, names, HR/MS data, and $\mathrm{IC}_{50}$ values of polyphenol derivatives (1-33) containing the $N^{\prime}$-methyleneformohydrazide scaffold.


This article is protected by copyright. All rights reserved.



This article is protected by copyright. All rights reserved.


This article is protected by copyright. All rights reserved.

Table 2. ${ }^{1}$ H NMR chemical shifts of the polyphenol derivatives (1-33) containing the $N^{\prime}$-methyleneformohydrazide scaffold. DMSO- $d_{6}$ was used as the NMR solvent. Multiplicity and coupling constants are given in parentheses; d and s represent doublet and singlet peaks, respectively.
The units of the chemical shifts and coupling constants are ppm and Hz , respectively


This article is protected by copyright. All rights reserved.


This article is protected by copyright. All rights reserved.

| Position | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| H2 | $\begin{aligned} & 7.98(\mathrm{dd}, \\ & 7.5,1.7) \end{aligned}$ | 7.52 (s) | 7.97 (d, 7.8) | 7.36 (s) | 8.17 (s) | $\begin{aligned} & 8.06(\mathrm{dd}, \\ & 8.9,5.7) \end{aligned}$ | $\begin{aligned} & 7.81(\mathrm{dd}, \\ & 4.5,1.5) \end{aligned}$ | $\begin{aligned} & 7.82(\mathrm{dd}, \\ & 4.3,1.9) \end{aligned}$ | $\begin{aligned} & 7.89(\mathrm{dd}, \\ & 4.4,1.6) \end{aligned}$ | 9.12 (d, 2.4) | $\begin{aligned} & \hline 7.89(\mathrm{dd}, \\ & 8.2,1.3) \end{aligned}$ | 7.41 (s) | $\begin{aligned} & 7.95(\mathrm{dd}, \\ & 7.8,1.6) \end{aligned}$ |
| H3 | $\begin{gathered} 7.53 \text { (ddd, } \\ 7.5,7.1,1.6 \text { ) } \end{gathered}$ | - | 7.06 (d, 7.8) | - | - | $\begin{aligned} & 7.36(\mathrm{dd}, \\ & 8.9,8.7) \end{aligned}$ | $\begin{aligned} & 8.78(\mathrm{dd}, \\ & 4.5,1.3) \end{aligned}$ | $\begin{aligned} & 8.79 \text { (dd, } \\ & 4.3,1.8) \end{aligned}$ | $\begin{aligned} & 8.79(\mathrm{dd}, \\ & 4.4,1.6) \end{aligned}$ | - | $\begin{gathered} 7.51 \text { (ddd, } \\ 8.2,7.2,1.0) \end{gathered}$ | - | $\begin{gathered} 7.55 \text { (ddd, } \\ 7.8,7.4,1.6) \end{gathered}$ |
| H4 | $\begin{aligned} & 7.59(\mathrm{dd}, \\ & 7.1,1.7) \end{aligned}$ | 7.15 (d, 8.2) | - | 6.98 (d, 7.8) | 7.79 (d, 8.0) | - | - | - | - | $\begin{aligned} & 8.76(\mathrm{dd}, \\ & 4.9,1.7) \end{aligned}$ | $\begin{aligned} & 7.57(\mathrm{dd}, \\ & 7.2,1.3) \end{aligned}$ | 7.14 (d, 7.8) | $\begin{aligned} & 7.63 \text { (dd, } \\ & 7.4,1.6) \end{aligned}$ |
|  | $\begin{aligned} & 7.53 \text { (ddd, } \\ & 7.5,7.1,1.6) \end{aligned}$ | $\begin{aligned} & 7.44(\mathrm{dd}, \\ & 8.2,7.8) \end{aligned}$ | 7.06 (d, 7.8) | $\begin{aligned} & 7.31(\mathrm{dd}, \\ & 7.8,7.4) \end{aligned}$ | $\begin{aligned} & 7.50(\mathrm{dd}, \\ & 8.0,7.9) \end{aligned}$ | $\begin{aligned} & 7.36(\mathrm{dd}, \\ & 8.9,8.7) \end{aligned}$ | $\begin{aligned} & 8.78(\mathrm{dd}, \\ & 4.5,1.3) \end{aligned}$ | $\begin{aligned} & 8.79(\mathrm{dd}, \\ & 4.3,1.8) \end{aligned}$ | $\begin{aligned} & 8.79(\mathrm{dd}, \\ & 4.4,1.6) \end{aligned}$ | $\begin{aligned} & 7.56(\mathrm{dd}, \\ & 8.0,4.9) \end{aligned}$ | $\begin{gathered} 7.51 \text { (ddd, } \\ 8.2,7.2,1.0) \end{gathered}$ | $\begin{aligned} & 7.43 \text { (dd, } \\ & 7.8,7.5) \end{aligned}$ | $\begin{gathered} 7.55 \text { (ddd, } \\ 7.8,7.4,1.6) \end{gathered}$ |
| H6 | $\begin{aligned} & 7.98(\mathrm{dd}, \\ & 7.5,1.7) \end{aligned}$ | 7.56 (d, 7.8) | 7.97 (d, 7.8) | 7.40 (d, 7.4) | 7.98 (d, 7.9) | $\begin{gathered} 8.06(\mathrm{dd}, \\ 8.9,5.7) \end{gathered}$ | $\begin{aligned} & 7.81(\mathrm{dd}, \\ & 4.5,1.5) \end{aligned}$ | $\begin{aligned} & 7.82(\mathrm{dd}, \\ & 4.3,1.9) \end{aligned}$ | $\begin{aligned} & 7.89(\mathrm{dd}, \\ & 4.4,1.6) \end{aligned}$ | $\begin{gathered} 8.31 \text { (ddd, } \\ 8.0,2.4,1.7 \text { ) } \end{gathered}$ | $\begin{aligned} & 7.89(\mathrm{dd}, \\ & 8.2,1.3) \end{aligned}$ | 7.47 (d, 7.5) | $\begin{aligned} & 7.95(\mathrm{dd}, \\ & 7.8,1.6) \end{aligned}$ |
| H1'a | 8.86 (s) | 8.86 (s) | 8.83 (s) | 8.84 (s) | 8.86 (s) | 8.85 (s) | 8.39 (s) | 8.40 (s) | 8.88 (s) | 8.85 (s) | - | - | - |
| H2' | - | - | - | - | - | - | 7.36 (d, 1.8) | 6.90 (d, 2.1) | - | - | 7.82 (d, 7.9) | 7.81 (d, 8.1) | - |
| H3' | 7.00 (d, 1.8) | 7.00 (d, 2.0) | 6.99 (d, 2.2) | 6.99 (d, 2.0) | 7.00 (d, 2.3) | 7.00 (d, 2.0) | - | - | 7.00 (d, 2.2) | 7.00 (d, 2.5) | 6.99 (d, 7.9) | 6.99 (d, 8.1) | 6.93 (d, 7.1) |
| H4' | - | - | - | - | - | - | - | 6.59 (d, 2.1) | - | - | - | - | $\begin{aligned} & 7.31(\mathrm{dd}, \\ & 8.3,7.1) \end{aligned}$ |
| H5, | 6.58 (d, 1.8) | 6.58 (d, 2.0) | 6.58 (d, 2.2) | 6.57 (d, 2.0) | 6.58 (d, 2.3) | 6.58 (d, 2.0) | 7.03 (d, 8.3) | - | 6.58 (d, 2.2) | 6.58 (d, 2.5) | 6.99 (d, 7.9) | 6.99 (d, 8.1) | $\begin{gathered} 6.90(\mathrm{dd}, \\ 8.3,7.6) \end{gathered}$ |
| H6 | - | - | - | - | - | - | $\begin{aligned} & 7.23 \text { (dd, } \\ & 8.3,1.8) \end{aligned}$ | 6.90 (d, 2.1) | - | - | 7.82 (d, 7.9) | 7.81 (d, 8.1) | 7.64 (d, 7.6) |
| H1"a | $\begin{gathered} 7.20(\mathrm{~d}, \\ 16.3) \end{gathered}$ | 7.20 (d, <br> 16.3) | 7.19 (d, 16.4) | $\begin{gathered} 7.20(\mathrm{~d} \\ 16.3) \end{gathered}$ | 7.20 (d, 16.2) | 7.20 (d, 16.2) | - | - | $\begin{gathered} 7.20(\mathrm{~d} \\ 16.3) \end{gathered}$ | 7.20 (d, 16.3) | - | - | - |
| H1"b | $\begin{gathered} 8.51(\mathrm{~d}, \\ 16.3) \end{gathered}$ | 8.51 (d, <br> 16.3) | 8.51 (d, <br> 16.4) | $\begin{gathered} 8.50(\mathrm{~d} \\ 16.3) \end{gathered}$ | 8.51 (d, <br> 16.2) | 8.51 (d, 16.2) | - | - | $\begin{gathered} 8.49(\mathrm{~d}, \\ 16.3) \end{gathered}$ | $\begin{gathered} 8.50(\mathrm{~d}, \\ 16.3) \end{gathered}$ | - | - | - |
| H2" | 7.69 (d, 8.4) | 7.69 (d, 8.5) | 7.68 (d, 7.2) | 7.68 (d, 8.4) | 7.68 (d, 8.6) | 7.69 (d, 8.0) | - | - | 7.68 (d, 8.7) | 7.69 (d, 8.5) | - | - | - |
| H3" | 6.95 (d, 8.4) | 6.95 (d, 8.5) | 6.94 (d, 7.2) | 6.94 (d, 8.4) | 6.95 (d, 8.6) | 6.95 (d, 8.0) | - | - | 6.95 (d, 8.7) | 6.95 (d, 8.5) | - | - | - |
| H5" | 6.95 (d, 8.4) | 6.95 (d, 8.5) | 6.94 (d, 7.2) | 6.94 (d, 8.4) | 6.95 (d, 8.6) | 6.95 (d, 8.0) | - | - | 6.95 (d, 8.7) | 6.95 (d, 8.5) | - | - | - |

This article is protected by copyright. All rights reserved.


This article is protected by copyright. All rights reserved.

Table 3. ${ }^{13} \mathrm{C}$ NMR chemical shifts (DMSO- $d_{6}$ ) of the polyphenol derivatives ( $\mathbf{1} \mathbf{- 3 3}$ ) containing the $N^{\prime}$-methyleneformohydrazide scaffold

| Position 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| C1a 163.0 | 162.7 | 162.3 | 163.1 | 161.4 | 162.0 | 163.1 | 162.7 | 163.1 | 161.4 | 162.1 | 163.2 | 162.8 | 163.2 | 161.3 | 161.5 | 162.0 | 161.6 | 161.5 | 161.6 |
| C1 133.2 | 134.5 | 125.17 | 134.6 | 135.3 | 129.6 | 133.1 | 134.5 | 134.5 | 135.2 | 129.5 | 133.1 | 134.5 | 134.5 | 135.2 | 135.4 | 129.5 | 128.9 | 140.2 | 128.9 |
| C2 127.7 | 112.8 | 129.6 | 114.6 | 130.2 | 130.4 | 127.7 | 112.8 | 114.6 | 130.2 | 130.5 | 127.7 | 112.8 | 114.6 | 130.1 | 114.4 | 130.4 | 148.6 | 121.5 | 148.6 |
| C3 128.5 | 159.2 | 113.7 | 157.5 | 121.8 | 115.5 | 128.5 | 159.2 | 157.5 | 121.8 | 115.6 | 128.6 | 159.3 | 157.5 | 121.7 | 163.1 | 115.5 | - | 150.4 | - |
| C4 131.9 | 117.7 | 162.0 | 118.84 | 134.6 | 163.0 | 131.9 | 117.7 | 118.9 | 134.6 | 163.1 | 132.0 | 117.7 | 118.9 | 134.6 | 118.8 | 162.9 | 152.4 | - | 152.4 |
| C5 128.5 | 129.7 | 113.7 | 129.6 | 130.8 | 115.5 | 128.5 | 129.6 | 129.6 | 130.8 | 115.6 | 128.6 | 129.7 | 129.6 | 130.7 | 130.7 | 115.5 | 123.7 | 150.4 | 123.7 |
| C6 127.7 | 119.9 | 129.6 | 118.2 | 126.9 | 130.4 | 127.7 | 119.9 | 118.2 | 126.9 | 130.5 | 127.7 | 119.9 | 118.2 | 126.8 | 123.8 | 130.4 | 135.5 | 121.5 | 135.5 |
| C1'a 140.4 | 140.5 | 139.6 | 140.3 | 141.0 | 140.5 | 140.0 | 140.0 | 139.8 | 140.6 | 140.2 | 140.1 | 140.1 | 139.9 | 140.5 | 140.5 | 140.1 | 141.1 | 141.7 | 140.7 |
| C2' 154.6 | 154.6 | 154.3 | 154.5 | 154.7 | 154.6 | 154.9 | 154.7 | 154.8 | 154.9 | 155.0 | 154.9 | 154.9 | 154.83 | 154.8 | 154.8 | 154.9 | 154.8 | 154.9 | 155.1 |
| C3, 118.4 | 118.4 | 118.5 | 118.5 | 118.2 | 118.3 | 118.5 | 118.4 | 118.5 | 118.3 | 118.5 | 118.60 | 118.6 | 118.64 | 118.5 | 118.5 | 118.53 | 118.2 | 118.1 | 118.3 |
| C4' $\quad 175.1$ | 175.1 | 175.1 | 175.1 | 175.0 | 175.1 | 174.1 | 174.0 | 174.1 | 174.0 | 174.1 | 174.0 | 174.0 | 174.0 | 173.8 | 173.8 | 173.9 | 175.1 | 175.0 | 174.1 |
| C5, 125.2 | 125.2 | 125.23 | 125.2 | 125.2 | 125.2 | 124.2 | 124.1 | 124.2 | 124.2 | 124.2 | 127.3 | 127.3 | 127.3 | 127.3 | 127.2 | 127.3 | 125.2 | 125.2 | 124.2 |
| C6' 126.1 | 126.1 | 126.0 | 126.1 | 126.1 | 126.1 | 130.6 | 130.5 | 130.5 | 130.6 | 130.6 | 124.9 | 124.9 | 124.9 | 124.8 | 124.8 | 124.9 | 126.2 | 126.2 | 130.6 |
| C7, 134.7 | 134.7 | 134.6 | 134.7 | 134.7 | 134.7 | 134.6 | 134.5 | 134.5 | 134.5 | 134.6 | 137.3 | 137.3 | 137.3 | 137.2 | 137.2 | 137.3 | 134.8 | 134.8 | 134.6 |
| C8, 118.7 | 118.7 | 118.7 | 118.76 | 118.7 | 118.8 | 121.3 | 121.2 | 121.3 | 121.3 | 121.3 | 121.5 | 121.5 | 121.5 | 121.4 | 121.4 | 121.5 | 118.8 | 118.8 | 121.3 |
| C9 ${ }^{\text {¢ }}$, 155.8 | 155.8 | 155.8 | 155.8 | 155.8 | 155.8 | 154.4 | 154.3 | 154.4 | 154.4 | 154.4 | 154.8 | 154.8 | 154.80 | 154.7 | 154.7 | 154.8 | 155.8 | 155.8 | 154.4 |
| C10 $\quad 123.3$ | 123.3 | 123.3 | 123.4 | 123.3 | 123.4 | 124.5 | 124.5 | 124.5 | 124.5 | 124.5 | 118.56 | 118.5 | 118.55 | 118.4 | 118.4 | 118.51 | 123.4 | 123.3 | 124.5 |
| $3-\mathrm{OCH}_{3}$ | 55.4 | - | - | - | - | - | 55.3 | - | - | - | - | 55.4 | - | - | - | - | - | - | - |
| $4-\mathrm{OCH}_{3}$ - | - | 55.4 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |

This article is protected by copyright. All rights reserved.

| Position | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| C1a | 162.9 | 162.6 | 162.4 | 163.0 | 161.3 | 161.8 | 161.5 | 161.7 | 161.5 | 161.4 | 163.6 | 163.3 | 164.4 |
| C1 | 133.6 | 134.9 | 125.6 | 135.0 | 135.7 | 130.0 | 140.6 | 140.4 | 140.6 | 129.3 | 134.2 | 135.6 | 133.0 |
| C2 | 127.7 | 112.7 | 129.6 | 114.6 | 130.18 | 130.4 | 121.5 | 121.5 | 121.6 | 148.7 | 127.8 | 113.0 | 128.1 |
| C3 | 128.4 | 159.2 | 113.6 | 157.4 | 121.7 | 115.3 | 150.3 | 150.3 | 150.2 | - | 128.3 | 159.1 | 128.5 |
| C4 | 131.5 | 117.4 | 161.8 | 118.5 | 134.3 | 162.8 | - | - | - | 152.1 | 131.3 | 117.1 | 131.9 |
| C 5 | 128.4 | 129.5 | 113.6 | 129.4 | 130.6 | 115.3 | 150.3 | 150.3 | 150.2 | 123.5 | 128.3 | 129.5 | 128.5 |
| C6 | 127.7 | 119.9 | 129.6 | 118.2 | 126.9 | 130.4 | 121.5 | 121.5 | 121.6 | 135.4 | 127.8 | 119.9 | 128.1 |
| $\mathrm{Cl}{ }^{\prime} \mathrm{a}$ | 145.1 | 145.1 | 144.4 | 145.0 | 145.6 | 145.1 | 149.3 | 148.9 | 146.3 | 145.7 | 155.8 | 156.1 | 158.2 |
| $\mathrm{Cl}^{\prime}$ | 112.7 | 112.8 | 112.9 | 112.8 | 112.5 | 112.7 | 126.7 | 136.0 | 112.4 | 112.5 | 130.5 | 130.4 | 119.4 |
| C2, | 138.9 | 139.0 | 138.8 | 138.9 | 139.1 | 139.0 | 108.4 | 105.0 | 139.2 | 139.1 | 127.9 | 128.0 | 158.7 |
| C3', | 102.5 | 102.6 | 102.5 | 102.5 | 102.6 | 102.6 | 149.1 | 160.7 | 102.7 | 102.6 | 113.7 | 113.7 | 117.3 |
| C4' | 161.0 | 161.1 | 160.9 | 161.0 | 161.2 | 161.1 | 151.0 | 102.6 | 161.3 | 161.2 | 160.4 | 160.4 | 131.2 |
| C5 | 97.5 | 97.5 | 97.5 | 97.5 | 97.5 | 97.5 | 111.5 | 160.7 | 97.5 | 97.5 | 113.7 | 113.7 | 118.5 |
| C6' | 160.3 | 160.3 | 160.2 | 160.3 | 160.4 | 160.3 | 122.2 | 105.0 | 160.5 | 160.4 | 127.9 | 128.0 | 128.4 |
| $\mathrm{C} 1 " \mathrm{a}$ | 130.33 | 130.4 | 130.31 | 130.30 | 130.4 | 130.31 | - | - | 130.5 | 130.4 | - | - | - |
| C1" ${ }^{\prime}$ | 126.1 | 126.1 | 126.1 | 126.1 | 126.1 | 126.1 | - | - | 126.0 | 126.1 | - | - | - |
| C1" | 130.27 | 130.3 | 130.26 | 130.29 | 130.24 | 130.27 | - | - | 130.2 | 130.2 | - | - | - |
| C2" | 128.4 | 128.4 | 128.4 | 128.4 | 128.4 | 128.4 | - | - | 128.4 | 128.4 | - | - | - |
| C3" | 114.1 | 114.1 | 114.1 | 114.1 | 114.1 | 114.1 | - | - | 114.1 | 114.1 | - | - | - |
| C4" | 159.0 | 159.0 | 159.0 | 159.0 | 159.0 | 159.0 | - | - | 159.0 | 159.0 | - | - | - |
| C5" | 114.1 | 114.1 | 114.1 | 114.1 | 114.1 | 114.1 | - | - | 114.1 | 114.1 | - | - | - |
| C6" | 128.4 | 128.4 | 128.4 | 128.4 | 128.4 | 128.4 | - | - | 128.4 | 128.4 | - | - | - |
| $\mathrm{Cl}^{\prime}{ }^{\prime}-\mathrm{CH}_{3}$ | - | - | - | - | - | - | - | - | - | - | 14.4 | 14.5 | 14.0 |
| $3-\mathrm{OCH}_{3}$ | - | 55.4 | - | - | - | - | - | - | - | - | - | 55.3 | - |
| $4-\mathrm{OCH}_{3}$ | - | - | 55.4 | - | - | - | - | - | - | - | - | - | - |
| 3'-OCH3 | - | - | - | - | - | - | 55.5 | 55.3 | - | - | - | - | - |


| $4^{\prime}-\mathrm{OCH}_{3}$ | 55.4 | 55.3 | 55.3 | 55.4 | 55.4 | 55.4 | 55.6 | - | 55.4 | 55.4 | 55.2 | 55.2 | - |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $5^{\prime}-\mathrm{OCH}_{3}$ | - | - | - | - | - | - | - | 55.3 | - | - | - | - | - |
| $6^{\prime}-\mathrm{OCH}_{3}$ | 56.0 | 56.0 | 56.0 | 56.0 | 56.0 | 56.0 | - | - | 56.0 | 56.0 | - | - | - |
| $4{ }^{\prime}-\mathrm{OCH}_{3}$ | 55.1 | 55.1 | 55.1 | 55.1 | 55.1 | 55.1 | - | - | 55.1 | 55.1 | - | - | - |

This article is protected by copyright. All rights reserved.

## Figure Captions



Figure 1.

(A)

(B)


Figure 1. General structures of (A) flavone, (B) benzochalcone bearing pyrazoline moiety, and (C) chromenylchalcone.

Figure 2.



Figure 2. Structure of derivative 22, $\quad(E)-N^{\prime}-((E)-2-((E)$-4-methoxystyryl)-4,6-dimethoxybenzylidene)-3-methoxybenzohydrazide.


## Figure 3.



Figure 3. Key correlations obtained from the COSY, NOESY, and HMBC spectra of derivative 22.

## Scheme 1.



Scheme 1. Synthetic route to the various polyphenols containing the $N^{\prime}$ methyleneformohydrazide scaffold (1-33).


This article is protected by copyright. All rights reserved.

## Synthesis and structure elucidation of polyphenols containing the $N^{\prime}$ methyleneformohydrazide scaffold as aurora kinase inhibitors

Dongsoo Koh, Yearam Jung, Seunghyun Ahn, Kenneth Hun Mok, Soon Young Shin, Yoongho Lim ${ }^{*}$

We designed polyphenols containing the $N^{\prime}$-methyleneformohydrazide scaffold and synthesized 33 derivatives. To confirm whether they show the inhibitory effects on aurA, in vitro kinase assay was performed, and their half-maximal inhibitory concentration ranged between 1.14 and $98.93 \mu \mathrm{M}$. We report here the assignments of their ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR data and high-resolution mass spectral data. These data can help us design a series of compounds containing novel aurora kinase inhibitor pharmacophores and identify newly synthesized polyphenols containing the $N^{\prime}$-methyleneformohydrazide scaffold.


This article is protected by copyright. All rights reserved.

