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2,3,6-Trideoxy-5-O-(4-nitrobenzoyl)-3-trifluoroacetamido-L-ribo-hexofuranosyl Bromide — A Suitable Furanoid Ristosamine Glycosylation Reagent

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5-O-Acetyl-2,3,6-trideoxy-3-phthalimido-β-L-ribo-hexofuranose (2) was converted to its corresponding methyl glycoside 3. After deprotection with 33 % methylamine in ethanol and subsequent protection of the amino group with trifluoroacetic anhydride and the hydroxy group with 4-nitrobenzoyl chloride, the resulting compound 5 was transformed into the appropriate glycosyl bromide 7 via the 1-O-acetyl derivative 6. Compound 7 was coupled with daunomycinone in the presence of yellow mercury(II) oxide and mercury(II) bromide.

The 3-amino-2,3,6-trideoxyhexoses are distributed in nature as the glysosidic moiety of several important antibiotics which exhibit anticancer activity. ¹⁻³ In addition, anthracycline antibiotics such as doxorubicin, daunorubicin and their analogues showed strong inhibitory effect on HIV-reverse transcriptase. ^{4,5} Their administration is, however, accompanied by various undesirable side-effects, especially a cumulative, dose-related cardio-

pathogenicity, 6 which seriously impedes their broader utilization in chemotherapy. Daunosamine, the sugar component of these antibiotics, has been replaced by other aminodeoxy sugars, 7,8 as well as by neutral sugars, 9,10 all in their pyranoid form. The first glycosylation with a furanoid analogue at 7-0 in carminomycinone was performed by Medgyes et al. 11 with a 3-amino-3,5-dideoxy-L-lyxo-hexofuranosyl derivative. Shortly after, the synthesis and the cytostatic activity of daunorubicin analogues with furanoid structures was also published.12 The prerequisite of the described routes is the multistep synthesis of the appropriate furanosides with easily removable protecting groups. In our laboratory, we have previously synthesized an L-ristosamine derivative possesing the furanose configuration. 13 In this investigation we synthesize its glycosyl bromide with the same protecting August 1993 SYNTHESIS 791

Scheme

groups that previously have been reported useful¹² for other configurations of 3-amino-2,3,6-trideoxyhexoses when used in glycosylation reactions.

The L-ribo isomer of 5-O-acetyl-2,3,6-trideoxy-3-phthalimidohexofuranose (2) was chosen as the starting material because it could be easily obtained in two steps from di-O-acetyl-L-rhamnal (1). 13 Glycosidation of 2 in 0.1 % hydrogen chloride afforded an anomeric mixture from which the β -anomer 3 crystallized in 46% yield by addition of diethyl ether. Compound 3 was deprotected with 33% methylamine in absolute ethanol to give the corresponding methyl 3-aminohexofuranoside 4 which was treated with trifluoroacetic anhydride and then with 4-nitrobenzoyl chloride in dry pyridine to give methyl 5-O-(4-nitrobenzoyl)-2,3,6-trideoxy-3-trifluoroacetami $do-\beta-L-ribo$ -hexofuranoside (5) in 85% yield. Treatment with acetic anhydride in the presence of acetic acid and sulfuric acid14 resulted in formation of an anomeric mixture ($\alpha/\beta = 1:2$) of the 1-O-acetylated hexofuranose 6 in 57% yield which on bromination with bromotrimethylsilane in dichloromethane afforded the glycosyl bromide under the same conditions as described by Kumar et al.15 An example of using the latter for glycosylation reactions was performed with daunomycinone in dichloromethane in the presence of mercury(II) oxide and mercury(II) bromide. From the anomeric mixture formed, the β -anomer 8 was isolated in 63 % yield by column chromatography.

The structural assignment of the glycoside **8** was confirmed by ${}^{1}\text{H}$ - ${}^{1}\text{H}$ 2D COSY and ${}^{1}\text{H}$ NOE experiments. Due to the *ribo*-configuration, irradiation of 3'-H resulted in 7 % NOE in 2' β -H and 4% NOE in 5'-H. The latter was in accordance with 6% NOE in 3'-H when 5'-H was irradiated. The β -configuration was assigned by the 11 % NOE in 2'- α -H when 1'-H was irradiated. In conformity with this assignment we found $J(1',2'\beta)$ close to zero in the

¹H NMR spectrum. In a similar way ¹H-¹H 2D COSY and ¹H NOE experiments on compound 5 confirmed β-configuration of the compounds 3–5.

NMR spectra were recorded on a Bruker AC 250 FT NMR spectrometer at 250 MHz for ^1H NMR and 62.9 MHz for ^{13}C NMR with TMS as an internal standard. FAB mass spectra were obtained on a Kratos MS 50 TS spectrometer. Analytical silica gel TLC plates 60 F $_{254}$ and silica gel (230–400 mesh) were purchased from Merck. 3,4-Di-O-acetyl-6-deoxy-L-glucal (di-O-acetyl-L-rhamnal) was purchased from Pfanstiehl Laboratories INC, Waukegan. Satisfactory microanalyses obtained for 3, 5, 6: C \pm 0.19, H \pm 0.30, N \pm 0.34.

Methyl 5-*O*-Acetyl-3-phthalimido-2,3,6-trideoxy- β -L-*ribo*-hexofuranoside (3):

Compound 2 (3.6 g, 11.3 mmol) was dissolved in dry MeOH (50 mL) containing 0.1 % HCl (w/w) and stirred at r.t. for 3 h. ${\rm Ag_2CO_3}$ (1 g) was added and the resulting suspension was filtered after 40 min through a pad of Celite. After evaporation in vacuo the resulting oil was treated with dry Et₂O (200 mL) and the precipitate filtered off to give pure 3 (0.75 g). An additional precipitate of 3 was obtained from the mother liquor on standing at 5 °C for one week to give a total yield of 1.75 g (46 %); mp 111–113 °C.

¹H NMR (CDCl₃): δ = 7.88–7.73 (m, 4 H, PhthN), 5.20 (d, 1 H, J = 5.1 Hz, 1-H), 5.01–4.87 (m, 2 H, 3-H, 5-H), 4.34 (t, 1 H, J = 7.0 Hz, 4-H), 3.38 (s, 3 H, MeO), 2.85 (ddd, 1 H, J = 5.1, 8.5, 12.6 Hz, 2α-H), 2.17 (dd, 1 H, J = 8.5, 12.6 Hz, 2β-H), 1.96 (s, 3 H, Ac), 1.30 (d, 3 H, J = 6.3 Hz, 6-H).

 13 C NMR (CDCl₃): $\delta = 170.12$ (Ac), 167.58 (C-2'), 134.07 (C-5'), 131.60 (C-3'), 123.19 (C-4'), 105.10 (C-1), 81.93 (C-4), 72.28 (C-5), 54.82 (MeO), 50.85 (C-3), 35.67 (C-2), 20.98 (Ac), 16.72 (C-6).

Methyl 3-Amino-2,3,6-trideoxy-β-L-ribo-hexofuranoside (4):

Compound 3 (1.30 g, 3.9 mmol) was dissolved in 33% solution of MeNH₂ in abs. EtOH (65 mL) and left at r.t. overnight. The solvent was evaporated in vacuo and the residual sirup purified by silica gel column chromatography (2×20 cm) with MeOH/CH₂Cl₂ (1:9); yield: 0.43 g (68%) which was used for the next step.

Methyl 5-O-(4-Nitrobenzoyl)-2,3,6-trideoxy-3-trifluoroacetamido- β -L-ribo-hexofuranoside (5):

A solution of 4(0.46 g, 2.7 mmol) in dry $\text{Et}_2\text{O}(20 \text{ mL})$ was cooled in an ice bath and treated with cold $(\text{CF}_3\text{CO})_2\text{O}(5.6 \text{ mL})$. The clear

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solution was kept in the ice bath for 20 min and at r. t. for 3 h. After evaporation of the solvent, the resulting sirup was coevaporated twice with toluene (2 × 20 mL). The residual oil was dissolved in ice-cooled MeOH (15 mL). The solution was treated with a few drops of pyridine and stirred at r. t. for 12 h. The solvent was evaporated in vacuo followed by coevaporation twice with pyridine (2 × 10 mL). To the solution was added dry pyridine (8.5 mL) and the solution was cooled to 0 °C. 4-Nitrobenzoyl chloride (0.81 g) was added and the mixture was stirred at 0 °C for 2 h and then at r. t. for 24 h. The mixture was poured into ice-water and extracted with CH₂Cl₂ (3 × 15 mL). The combined extracts were washed with sat. aq NaHCO₃ (10 mL) and H₂O (10 mL), dried (Na₂SO₄) and the solvent evaporated in vacuo. Silica gel column chromatography with petroleum ether/EtOAc (7:3) afforded pure 5; yield: 0.58 (85 %); mp 109-110 °C.

¹H NMR (CDCl₃): $\delta = 8.31-8.19$ (m, 4 H, H_{arom}), 6.52 (d, 1 H, J = 7.0 Hz, NH), 5.29 (quint, 1 H, J = 6.4 Hz, 5-H), 5.10 (dd, 1 H, J = 1.2, 5.2 Hz, 1-H), 4.77 (quint, 1 H, J = 7.8 Hz, 3-H), 4.04 (t, 1 H, J = 6.0 Hz, 4-H), 3.30 (s, 3 H, Ac), 2.45 (ddd, 1 H, J = 1.2, 7.8, 13.4 Hz, 2β-H), 2.07 (ddd, 1 H, J = 5.2, 7.8, 13.4 Hz, 2α-H), 1.43 (d, 3 H, J = 6.4 Hz, 6-H).

¹³C NMR (CDCl₃): δ = 163.92 (ArCO), 156.41 (CF₃CO), 150.54 (C-1'), 135.43 (C-4'), 130.68 (C-3'), 123.42 (C-2'), 117.80 (CF₃), 104.85 (C-1), 85.48 (C-4), 72.47 (C-5), 55.26 (MeO), 51.19 (C-3), 39.19 (C-2), 16.41 (C-6).

1-O-Acetyl-5-O-(4-nitrobenzoyl)-2,3,6-trideoxy-3-trifluoroacetamido-L-ribo-hexofuranose (6):

Compound 5 (0.50 g, 1.2 mmol) was dried by coevaporation with benzene and dissolved in EtOAc (10 mL). The solution was cooled to $-25\,^{\circ}\text{C}$ and mixed with a cold ($-25\,^{\circ}\text{C}$) solution of EtOAc (19.5 mL), Ac₂O (11.0 mL), AcOH (8.3 mL) and conc. H₂SO₄ (0.05 mL). The solution was kept at $-15\,^{\circ}\text{C}$ for 16 h, poured into ice-water and extracted with CH₂Cl₂ (3×25 mL). The combined extracts were washed with H₂O (15 mL) and dried (Na₂SO₄). After evaporation of the solvent, the product was coevaporated with dry toluene; yield: 0.30 g (57%).

Pure β -(80 mg) and α -(40 mg) anomers of **6** were isolated from 0.12 g of the anomeric mixture by silica gel column chromatography with CH₂Cl₂/MeOH (97:3).

Compound 6 (β -anomer): mp 113-114°C.

¹H NMR (CDCl₃): $\delta = 8.32-8.20$ (m, 4 H, H_{arom}), 6.87 (d, 1 H, J = 8.0 Hz, NH), 6.40 (d, 1 H, J = 4.3 Hz, 1-H), 5.31 (quint, 1 H, J = 6.4 Hz, 5-H), 4.82 (quint, 1 H, J = 8.0 Hz, 3-H), 4.13 (t, 1 H, J = 6.0 Hz, 4-H), 2.55 (ddd, 1 H, J = 1.0, 8.0, 13.8 Hz, 2β-H), 2.27 (ddd, 1 H, J = 4.3, 8.0, 13.8 Hz, 2α-H), 1.95 (s, 3 H, Ac), 1.45 (d, 3 H, J = 6.4 Hz, 6-H).

¹³C NMR (CDCl₃): δ = 169.72 (Ac), 163.85 (ArCO), 150.54 (C-1'), 135.19 (C-4'), 130.71 (C-3'), 123.49 (C-2'), 97.29 (C-1), 86.14 (C-4), 72.07 (C-5), 50.58 (C-3), 38.30 (C-2), 20.86 (Ac), 16.04 (C-6).

Compound 6 (α-anomer): mp 146-148°C.

¹H NMR (CDCl₃): δ = 8.31–8.19 (m, 4 H, H_{arom}), 7.08 (d, 1 H, J = 8.3 Hz, NH), 6.47 (d, 1 H, J = 4.7 Hz, 1-H), 5.21 (quint, 1 H, J = 6.4 Hz, 5-H), 4.77 (t, 1 H, J = 8.2 Hz, 3-H), 4.26 (dd, 1 H, J = 1.8, 5.4 Hz, 4-H), 2.52 (ddd, 1 H, J = 4.7, 8.2, 13.7 Hz, 2β-H), 2.13 (d, 1 H, J = 13.7 Hz, 2α-H), 2.11 (s, 3 H, Ac), 1.47 (d, 3 H, J = 6.4 Hz, 6-H).

¹³C NMR (CDCl₃): $\delta = 168.81$ (Ac), 163.73 (ArCO), 156.41 (CF₃CO), 150.59 (C-1'), 135.12 (C-4'), 130.62 (C-3'), 123.46 (C-2'), 98.17 (C-1), 88.68 (C-4), 71.29 (C-5), 49.58 (C-3), 37.53 (C-2), 20.90 (Ac), 16.21 (C-6).

5-O-(4-Nitrobenzoyl)-2,3,6-trideoxy-3-trifluoroacetamido-L-ribo-hexofuranosyl Bromide (7):

Compound 6 (0.20 g, 0.46 mmol) was dissolved in 3 mL of dry alcohol-free CH₂Cl₂ and the solution was cooled to 0 °C. Me₃SiBr (0.2 mL) was added and the mixture was stirred for 10 min and then at r.t. for 3.5 h. When silica gel TLC with CHCl₃/MeOH (19:1) showed complete conversion of 6 into a more polar product, the solvent was evaporated in vacuo at r.t. The product, an odorless solid was immediately used in the following step; yield: 0.20 g (95 %).

7-*O*-[5-*O*-(4-Nitrobenzoyl)-2,3,6-trideoxy-3-trifluoroacetamido-*L-ribo*-hexofuranosyl]daunomycinone 8:

Daunomycinone (0.10 g, 0.25 mmol) was dissolved in dry alcohol-free CH_2Cl_2 (15 mL) and finely ground preignited type 3Å molecular sieves (0.4 g) was added. Dry yellow HgO (0.21 g) and HgBr₂ (0.084 g) was added. To this mixture was added 7 (100 mg, 0.22 mmol) in CH_2Cl_2 (2 mL) with stirring. After 30 min another batch of molecular sieves (0.21 g), yellow HgO (0.13 g) and HgBr₂ (0.032 g) and finally 7 (100 mg, 0.22 mmol) were added. After 6 h, the mixture was diluted with CH_2Cl_2 (50 mL) and filtered. The solid was rinsed with CH_2Cl_2 (20 mL) which was combined with the filtrate and evaporated. The residual red solid was purified by silica gel column chromatography with MeOH/benzene/CHCl₃ (1:5:10) to give 8; yield: 0.12 g (63 %); mp 178–180 °C.

¹H NMR (500 MHz, CDCl₃): $\delta = 14.07$ (s, 1 H, OH), 13.20 (s, 1 H, OH), 8.05, 7.80 and 7.42 (d, t, and d, 3 H, J = 7-8 Hz, 1,2,3-H), 8.04–8.0 (m, 4 H, H_{arom}), 6.49 (d, 1 H, J = 7.4 Hz, NH), 5.75 (d, 1 H, J = 5.8 Hz, 1'-H), 5.50 (d, 1 H, J = 3.0 Hz, 7-H), 5.30 (quint, 1 H, J = 6.4 Hz, 5'-H), 4.68 (quint, 1 H, J = 8.0, 3'-H), 4.11 (s, 3 H, OMe), 4.00 (t, 1 H, J = 6.4 Hz, 4'-H), 3.16 and 3.02 (2 × d, 2 H, J = 19.0 Hz, 10-H), 2.51 (dd, 1 H, J = 8.0, 13.9 Hz, 2'β-H), 2.41 (m, 1 H, partially hidden by Ac, 8e-H), 2.38 (s, 3 H, Ac), 2.25 (ddd, 1 H, J = 5.8, 8.0, 13.9 Hz, 2'-α-H), 2.00 (dd, 1 H, J = 3.0, 15.0 Hz, 8a-H), 1.33 (d, 1 H, J = 6.4 Hz, 6'-H).

FAB (MeOH + 1 % AcOH + 3-O₂NC₆H₄CH₂OH): m/z (%) = 773 (M + H⁺, 1).

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