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Note

# Synthesis of galactosyl and lactosyl derivatives as potential anti-metastasis compounds

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#### Abstract

Based on the known anti-metastasis activities of lactosides and galactosides, a galactosyl and a lactosyl trimannoside were prepared via the conventional Koenigs–Knorr and trichloroacetimidate methods, respectively. Through typical deblocking procedures, a tetrasaccharide  $\alpha$ -D-Galp-(1  $\rightarrow$  2)- $\alpha$ -D-Manp-(1  $\rightarrow$  2)- $\alpha$ -D-Manp-(1  $\rightarrow$  6)- $\alpha$ -D-ManpOCH<sub>3</sub> and a pentasaccharide  $\beta$ -D-Galp-(1  $\rightarrow$  4)- $\beta$ -D-Glcp-(1  $\rightarrow$  2)- $\alpha$ -D-Manp-(1  $\rightarrow$  2)- $\alpha$ -D-Manp-(1  $\rightarrow$  6)- $\alpha$ -D-ManpOCH<sub>3</sub> were obtained. © 2000 Elsevier Science Ltd. All rights reserved.

Keywords: α-Galactosyl trimannoside; β-Lactosyl trimannoside; Glycosylation; Synthesis

# 1. Introduction

Raz and Lotan [1,2] discovered that lactose, D-galactose, D-glucosamine, and N-acetyl-Dgalactosamine can inhibit the adhesion of some cancer cells, such as human melanoma A375, SH4, Hs294, Hs852, human adenoma Hela-S3, murine melanoma B16-F1, and murine fibrosarcoma UV-2237P. Among the aforementioned glycoses, lactose has the strongest inhibition activity, whereas other glycoses such as D-mannose, L-fucose, N-acetyl-D-glucosamine have none. The adhesion of cancer cells to other normal cells is the basis of cancer metastasis and proliferation. Oguchi et al. [3] reported that methyl  $\beta$ -lactoside can significantly inhibit murine melanoma B-16

cells from agglomerating in the lungs. Further study of the lactosides indicated that conjugates having a lactosyl group linked to a lysine-lysine peptide or other polylysine peptides through a carbon chain spacer arm have different activities in regard to inhibiting metastasis. Interestingly some of the glycoconjugates have strong anti-metastasis activity while others show no such activity or even enhance metastasis, a factor apparently related to the difference in length of the spacer arms [4]. In addition, such ramified mannans as hexamannoside Man6 and nonamannoside Man9 are also able to induce cancer cell adhesion, whereas trimannoside Man3 has not [5,6].

We therefore postulated that a combination of a galactosyl or lactosyl group and a trimannoside might lead to new anti-metastasis compounds. For this purpose, a tetrasaccharide  $\alpha$ -D-Galp-(1  $\rightarrow$  2)- $\alpha$ -D-Manp-(1  $\rightarrow$  2)

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- $\alpha$ -D-Manp-(1 $\rightarrow$ 6)- $\alpha$ -D-ManpOCH<sub>3</sub> (8), and a pentasaccharide  $\beta$ -D-Galp-(1 $\rightarrow$ 4)- $\beta$ -D-Glcp-(1 $\rightarrow$ 2)- $\alpha$ -D-Manp-(1 $\rightarrow$ 2)- $\alpha$ -D-Manp-(1 $\rightarrow$ 6)- $\alpha$ -D-ManpOCH<sub>3</sub> (13) have been synthesized, using 1+3 and 2+3 coupling modes (see Scheme 1).

## 2. Results and discussion

The trimannoside **3** was prepared using 2-O-acetyl-3,4,6-tri-O-benzyl- $\alpha$ -D-mannopyranosyl trichloroacetimidate (**1**) as glycosyl donor and the dimannoside **2** [7] as acceptor. The condensation was carried out in a solution of dichloromethane with trimethylsilyl triflate (Me<sub>3</sub>SiOTf) as promoter and the yield was good (72.7%). Zemplén deacetylation gave the trisaccharide acceptor **4** in 83.7% yield. 2,3,4,6-Tetra-O-benzyl- $\alpha$ -D-galactopyranosyl bromide (**6**), obtained readily by in situ con-



Scheme 1.

version of the corresponding ethyl 1-thioglycoside **5** [8] reacting with bromine in dichloromethane [9], was coupled to **4** by the conventional Koenigs–Knorr method with Hg(CN)<sub>2</sub> as promoter to give in 54% yield of an exclusive  $\alpha$ -galacosylated tetrasaccharide **7**, which was then debenzylated by catalytic hydrogenolysis to afford the deblocked tetrasaccharide **8**. Compound **5** could be coupled directly to glycosyl acceptor **4** using methyl triflate as promoter to afford tetrasaccharide **7**, but the yield was relatively low (20–30%). The configuration of **8** was confirmed by the galactosyl anomeric <sup>3</sup>J<sub>H-1,2</sub>, value of 3.68 Hz.

To prepare pentasaccharide 11, hepta-Oacetyllactosyl trichloroacetimidate 10 [9], obtained by reaction of lactoseheptaacetate 9 and trichloroacetonitrile in the presence of DBU as catalyst, was chosen as glycosyl donor to couple with 4 in the presence of Me<sub>3</sub>SiOTf as promoter. The resultant pentasaccharide 11 was obtained in moderate vield (45%). Deacetvlation of 11 in NaOMe-Me<sub>3</sub>OH gave compound 12 in 81.7% yield. Subsequent catalytic hydrogenolysis gave the deblocked pentasaccharide 13 in 82.6% yield. <sup>1</sup>H NMR spectroscopy indicated that the nonreducing terminal galactosyl and glucosyl anomeric  ${}^{3}J_{H-1,2}$  values are 7.97 and 7.99 Hz, respectively, and thus the  $\beta$  configuration of the lactosylation was thus confirmed.

Compounds 8 and 13, along with several other related oligosaccharides synthesized in our laboratory are being tested for their antimetastasis activity, and the results will be reported later.

#### 3. Experimental

General methods.—Spectra were recorded with the following instruments: <sup>1</sup>H and <sup>13</sup>C NMR, Bruker ARX-400; the <sup>1</sup>H NMR spectra were recorded with Me<sub>4</sub>Si as the internal standard and <sup>13</sup>C NMR with CDCl<sub>3</sub> as solvent and internal standard; Mass spectra, VG ZAB-Hs; IR, Perkin–Elmer 983. Elemental analyses were performed on a Perkin–Elmer 240C instrument. Optical rotations were measured at 25 °C with a Optical Activity AA-10R polarimeter. The progress of reactions was monitored by thin-layer chromatography (TLC) on Silica Gel  $GF_{254}$  (Hai Yang Chemical Factory, Qingdao, Shandong, PR China). Detection was effected by examination under UV light and by charring with 5% phosphomolybdic acid hydrate in EtOH or 20% concd  $H_2SO_4$  in EtOH and heating. PTLC was performed on Silica Gel  $GF_{254}$  and column chromatography on silica gel H (Hai Yang Chemical Factory, Qingdao, Shandong, PR China). The solvent systems indicated are volume volume ratios, and the petroleum ether used in the experiment has the boiling range of 60–90 °C.

Methyl 2,3,4-tri-O-benzyl-6-O-[3,4,6-tri-Obenzyl-2-O-(2-O-acetyl-3,4,6-tri-O-benzyl-a-Dmannopyranosyl) -  $\alpha$  - D - mannopyranosyl] -  $\alpha$  - Dmannopvranoside (3). -2-*O*-Acetyl-3.4.6-tri-O-benzyl- $\alpha$ -D-mannopyranosyl trichloroacetimidate (1) (1.04 g, 1.63 mmol) and methyl 2.3.4-tri-O-benzyl-6-O-(3.4.6-tri-O-benzyl-2- $O - \alpha - D - mannopyranosyl) - \alpha - D - mannopy$ ranoside (2) (1.32 g, 1.44 mmol) were dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (25 ml). To the solution was added powdered 4Å molecular sieve (1.0 g) and the mixture was stirred at room temperature (rt) for 1 h and then cooled to -10 °C for another 20 min. Trimethylsilyl triflate (ten drops) was added and the mixture was stirred for 5 h, during which time the mixture attained to rt. The mixture was filtered through a layer of Celite and rinsed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate was washed with water, satd aq NaHCO<sub>3</sub>, and water, and then dried (MgSO<sub>4</sub>). The residue obtained by evaporation of the solvent was subjected to silica gel column chromatography eluting with 6:1 petroleum—acetone to give compound 3 (yield 1.47 g, 72.7%) as a colorless syrup.  $R_f 0.50$  (4:1 cyclohexane-acetone);  $[\alpha]_{\rm D}$  + 10.5° (c 0.39, CHCl<sub>3</sub>). Anal. Calcd. for C<sub>84</sub>H<sub>90</sub>O<sub>17</sub>: C, 73.58; H, 6.57. Found: C, 73.37; H, 6.28. IR:  $1732 \text{ cm}^{-1}$  (C=O).

Methyl 2,3,4-tri-O-benzyl-6-O-[3,4,6-tri-Obenzyl-2-O-(3,4,6-tri-O-benzyl- $\alpha$ -D-mannopyranosyl)- $\alpha$ -D-mannopyranosyl]- $\alpha$ -D-mannopyranoside (4).—To a solution of compound 3 (1.33 g, 0.95 mmol) in abs MeOH (30 ml), metal sodium (30 mg) was added. The solution was stirred at rt overnight, decationized and evaporated. Column chromatography of the residue on silica gel, eluting with 4:1 cyclohexane-acetone gave compound 4 (yield 1.08) g, 83.7%) as a colorless syrup.  $R_f$  0.37 (4:1 cyclohexane–acetone);  $[\alpha]_{\rm D}$  + 19.6° (c, 0.12, CHCl<sub>3</sub>); IR: 3460 cm<sup>-1</sup> (OH); <sup>1</sup>H NMR:  $\delta$ 2.60 (br, 1 H, OH), 3.22 (s, 3 H,  $OCH_3$ ), 3.67-4.70 (m, 49 H, H-1, 2, 3, 4, 5, 6, H-1', 2', 3', 4', 5', 6', H-1", 2", 3", 4", 5", 6", 9× PhC $H_2$ ), 7.20–7.32 (m, 45 H, aromatic H); <sup>13</sup>CNMR:  $\delta$  54.61 (OCH<sub>3</sub>), 65.27–77.36 (C-2, 3, 4, 5, 6, C-2', 3', 4', 5', 6', C-1", 2", 3", 4", 5", 6", 9 × PhCH<sub>2</sub>), 98.82, 99.10, 101.13 (C-1, C-1', C-1"), 127.27-128.41 (aromatic carbon), 138.09, 138.12, 138.25, 138.29, 138.47, 138.55 (2 C), 138.68, 138.72 (aromatic quaternary carbon).

Methyl 2,3,4-tri-O-benzyl-6-O-{3,4,6-tri-Obenzvl-2-O-[3,4,6-tri-O-benzvl-2-O-(2,3,4,6tetra - O - benzvl -  $\alpha$  - D - galactopyranosvl) -  $\alpha$  - Dmannopyranosyl]- $\alpha$ -D-mannopyranosyl}- $\alpha$ -Dmannopyranoside (7).—To a solution of compound 4 (550 mg, 0.40 mmol) in 25 mL of dry  $CH_2Cl_2$ ,  $Hg(CN)_2$  (0.33 g) and powdered 4Å molecular sieve (1.0 g) were added and the mixture was stirred for 1 h at rt. Meanwhile, to a solution of compound 5 (780 mg, 1.33 mmol) in 15 mL of dry CH<sub>2</sub>Cl<sub>2</sub> was added liquid bromine dropwise (0.25 mL) with stirring for 45 min at rt. The solvent was evaporated in vacuo and toluene  $(2 \times 25 \text{ mL})$  was co-distilled from the residue. The resulting galactosyl bromide 6 was dissolved in dry  $CH_2Cl_2$  (10 mL) and the solution added dropwise to the aforementioned mixture. After stirring for 24 h at rt, the mixture was filtered and the filtrate was washed successively with satd brine, satd NaHCO<sub>3</sub> solution, and water, respectively, and dried (MgSO<sub>4</sub>). Evaporation of the solvent in vacuo gave a yellowish residue, which was subjected to column chromatography on silica gel eluting with 5:1 petroleum ether-EtOAc to give tetrasaccharide 7 (yield 420 mg, 54.0%) as a colorless syrup;  $R_f$  0.33 (5:1 petroleum ether-EtOAc);  $[\alpha]_{\rm D}$  + 37.1° (c 0.35, CHCl<sub>3</sub>). <sup>1</sup>H NMR:  $\delta$  3.17 (s, 3 H, OCH<sub>3</sub>), 3.49–5.20 (54 H, sugar ring H and  $13 \times PhCH_2$ ), 7.17–7.45 (65 H, aromatic H). <sup>13</sup>C NMR: δ 53.48 (OCH<sub>3</sub>), 60.42–80.50 (C-2, 3, 4, 5, 6, C-2', 3', 4', 5', 6", C-2", 3", 4", 5", 6", C-2<sup>*'''*</sup>, 3<sup>*'''*</sup>, 4<sup>*'''*</sup>, 5<sup>*'''*</sup>, 6<sup>*'''*</sup>,  $13 \times PhCH_2$ ), 97.96, 98.91, 99.05, 101.12 (C-1, C-1', C-1", C-1"),

127.00–128.53 (aromatic carbon), 138.01, 138.25, 138.36, 138.53, 138.62, 138.70, 138.73, 138.79, 138.85, 138.90, 138.95, 139.17, 139.21 (<sup>13</sup>C, aromatic quaternary carbon).

6-O-[2-O-(2-O-α-D-galactopyran-Methvl  $osyl - \alpha - D - mannopyranosyl) - \alpha - D - mannopyran$ osyl]-α-D-mannopyranoside (8).—Compound 7 (320 mg, 0.17 mmol) was dissolved in 4:1 MeOH-EtOAc (65 mL) and the solution was hydrogenlyzed with palladium on charcoal (200 mg) in a hydrogen stream under a pressure of 330 KPa for 24 h. The mixture was filtered and the resulting solution was evaporated to give compound 8 a colorless syrup, which was then dissolved in distilled water and lyophilized to a colorless foam (98 mg, 83.3%), FABMS: (m/z) 681  $[M + 1]^+$ ;  $[\alpha]_D$  $+ 54.0^{\circ}$  (c 0.63, water). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.43 (s, 3 H, OCH<sub>2</sub>), 3.37–4.25 (m, 37 H, sugar ring H,  $13 \times OH$ ), 4.77 (d, 1 H,  $J_{1,2}$ ) 1.48 Hz, mannosyl H-1), 5.17, 5.42 (2 s, 2 H, mannosyl H-1), 5.18 (d, 1 H, J<sub>1,2</sub> 3.68 Hz, galactosyl H-1); <sup>13</sup>C NMR:  $\delta$  57.33 (OCH<sub>3</sub>), 63.46, 63.60, 63.82 (mannosyl C-6), 68.35 (galactosyl C-6), 69.05, 69.47, 69.68, 71.37, 71.82 (2 C), 72.41, 72.74, 72.88, 73.80 (2 C), 73.93, 75.31, 75.66, 81.48, 82.09 (C-2, 3, 4, 5, C-2', 3', 4', 5', C-2", 3", 4", 5", C-2"', 3"', 4"', 5"), 100.51 (galactosyl C-1), 103.20, 103.57, 103.74 (mannosyl C-1). Anal. Calcd. C<sub>25</sub>H<sub>44</sub>O<sub>21</sub>: C, 44.05; H, 6.47. Found: C, 44.36; H, 6.61.

2.3.6-Tri-O-acetyl-4-O-(2.3.4.6-tetra-O $acetyl - \beta - D - galactopyranosyl) - D - glucopyrano$ svl trichloroacetimidate (10).—To a solution of 2,3,6-tri-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-D-glucopyranose [9] (9, 1.13 g, 1.78 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL), trichloroacetonitrile (1 mL) was added and the mixture was stirred in an ice-water bath for 10 min. DBU (ten drops) was then added and the mixture was stirred for 2 h during which time the bath reached rt. Evaporation of the solvent in vacuo gave a dark-brown oil, which was purified by column chromatography eluting with 1:1 petroleum ether-EtOAc to give compound **10** (1.1 g, 79.7%), as a yellow syrup.  $R_f$  0.31 (1:1 petroleum ether-EtOAc).

Methyl (2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)- (1  $\rightarrow$  4)- (2,3,6-tri-O-acetyl- $\beta$ -Dglucopyranosyl)- (1  $\rightarrow$  2)- (3,4,6-tri-O-benzyl-

 $\alpha$  - D - mannopyranosyl) -  $(1 \rightarrow 2)$  - (3, 4, 6 - tri - O) $benzyl-\alpha$ -D-mannopyranosyl)- $(1 \rightarrow 6)$ -2,3,4-tri-O-benzyl- $\alpha$ -D-mannopyranoside (11).—A mixture of compound 10 (270 mg, 0.35 mmol), compound 4 (290 mg, 0.22 mmol) and powered 4 Å molecular sieve (1 g) in dry  $CH_2Cl_2$ (20 mL) was stirred for 2 h at rt and then cooled to -15 °C. Trimethylsilyl triflate (10 drops) was added and the mixture was stirred for 5 h. The mixture was filtered through a layer of Celite, rinsed with CH<sub>2</sub>Cl<sub>2</sub>, and then dried (MgSO<sub>4</sub>). The residue resulting from evaporation of the solvent was subjected to silica gel column chromatography eluting with 3:1 cyclohexane-acetone to give compound 11 (vield 190 mg, 45%) as a colorless syrup;  $[\alpha]_D$  $+11.5^{\circ}$  (c 0.17, CHCl<sub>3</sub>); IR: 1748 cm<sup>-</sup> (C=O). <sup>1</sup>H NMR: δ 1.97, 2.04, 2.06, 2.08, 2.10, 2.15. 2.17 (7 s. 21 H.  $7 \times CH_2CO$ ). 3.23 (s. 3) H, OCH<sub>2</sub>), 3.68–5.22 (m, 53 H, sugar ring H,  $9 \times PhCH_2$ , 7.20–7.30 (45 H, aromatic H). <sup>13</sup>C NMR:  $\delta$  20.49, 20.62 (2 C), 20.71, 20.76, 20.80, 20.85  $(7 \times CH_3CO)$ , 54.65  $(OCH_3)$ , 60.95-77.40 (39 C, sugar ring carbon,  $9 \times$ PhCH<sub>2</sub>), 98.90, 99.0 (2 C) (C-1, C-1', C-1"), 99.35, 101.14 (C-1<sup>'''</sup>, C-1<sup>''''</sup>), 127.41–128.88 (aromatic carbon), 138.04, 138.30, 138.33, 138.37(2 C), 138.49(2 C), 138.53, 138.62 (aromatic quaternary carbon), 168.99, 169.46, 169.85, 170.03, 170.15, 170.25, 170.36 (7 C, CH<sub>3</sub>CO). Anal. Calcd. for  $C_{108}H_{122}O_{17}$ : C, 66.60; H, 6.27. Found: C, 67.03; H, 6.20.

Methyl  $(\beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ - $(\beta$ -D-glucopyranosyl)- $(1 \rightarrow 2)$ -(3,4,6-tri-O-benzyl- $\alpha$ -D-mannopyranosyl)- $(1 \rightarrow 2)$ -(3,4,6-tri-O-benzyl- $\alpha$ -D-mannopyranosyl)- $(1 \rightarrow 6)$ -(2,3,4-tri-Obenzyl- $\alpha$ -D-mannopyranoside) (12).—To a solution of compound 11 (180 mg, 0.09 mmol) in abs MeOH (20 mL) CH<sub>3</sub>ONa (10 mg) was added and the mixture was stirred for 2 h at rt. TLC showed that the reaction was complete. The solution was decationized, evaporated, and was purified by PTLC (3:1 CHCl<sub>3</sub>-MeOH) to afford compound 12 (yield 125 mg, 81.7%) as a colorless syrup.  $R_f$  0.33 (3:1 CHCl<sub>3</sub>-MeOH).

Methyl  $(\beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ - $(\beta$ -D-glucopyranosyl)- $(1 \rightarrow 2)$ - $(\alpha$ -D-mannopyranosyl)- $(1 \rightarrow 2)$ - $(\alpha$ -D-mannopyranosyl)- $(1 \rightarrow 6)$ - $(\alpha$ -D-mannopyranoside) (13).—To a solution of compound 12 (114 mg, 0.07 mmol) in

MeOH (50 mL) was added Pd-C (10%) and the mixture was hydrogenolyzed in a hydrogen stream under a pressure of 33 KPa for 24 h. The mixture was filtered and the catalyst was rinsed with MeOH and a small amount of distilled water. Evaporation and lyophilization of the filtrate gave compound 13 (yield 48 mg, 82.6%) as a white amorphous solid; FABMS: (m/z) 843  $[M + 1]^+$ , 865  $[M + Na]^+$ ;  $[\alpha]_D$  $+35.6^{\circ}$  (c 0.51, water). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.44 (s, 3 H, OCH<sub>3</sub>), 3.35–3.93 (m, sugar ring H), 4.05 (dd, 1 H, mannosyl H-2), 4.27 (dd, 1 H, mannosyl H-2), 4.48 (d, 1 H,  $J_{1,2}$  7.97 Hz, glucosyl H-1), 4.59 (d, 1 H,  $J_{1,2}$ 7.99 Hz, galactosyl H-1), 4.76 (d, 1 H, J<sub>1.2</sub> 1.48 Hz, mannosyl H-1), 5.14 (d, 1 H, J<sub>1,2</sub> 1.44 Hz, mannosyl H-1), 5.16 (d, 1 H, J<sub>1.2</sub> 1.28 Hz, mannosyl H-1). <sup>13</sup>C NMR:  $\delta$  57.37 (OCH<sub>3</sub>), 62.58, 63.10, 63.48, 63.52, 68.29 (C-6, C-6', C-6", C-6", C-6""), 69.09, 69.46, 71.13 (2 C), 72.08, 72.45, 72.75, 73.21, 73.32, 73.53, 74.98, 75.09, 75.36, 75.73, 76.66, 77.34, 77.95 (2 C), 79. 86, 80.82, 81.38 (C-2, 3, 4, 5, C-2', 3', 4', 5', C-2", 3", 4", 5", C-2", 3", 4", 5", C-2", 3", 4"", 5""), 100.50, 102.87, 103.62, 104.01, 105.54 (C-1, C-1', C-1", C-1"", C-1""). Anal. Calcd.

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## References

- [1] A. Raz, R. Lotan, Cancer Res., 41 (1981) 3642-3647.
- [2] R. Lotan, A. Raz, Cancer Res., 43 (1983) 2088–2093.
- [3] H. Oguchi, T. Toyokuni, B. Dean, Cancer Commun., 2 (1990) 311–314.
- [4] B. Dean, H. Oguchi, S. Cai, E. Otsuji, K. Tashiro, S. Hakomori, T. Toyokuni, *Carbohydr. Res.*, 245 (1993) 175–192.
- [5] S. Chandrasekaran, M.L. Tanzer, M.S. Gingers, J. Biol. Chem., 269 (1994) 3356–3366.
- [6] S. Chandrasekaran, M.L. Tanzer, M.S. Gingers, J. Biol. Chem., 269 (1994) 3367–3373.
- [7] Z.J. Li, H. Li, M.S. Cai, Carbohydr. Res., 320 (1999) 1-7.
- [8] H. Lönn, Carbohydr. Res., 139 (1985) 105-113.
- [9] F.A.W. Koeman, J.W.G. Meissner, H.R.P. Van Ritter, J.P. Kamerling, J.F.G. Vligenthart, J. Carbohydr. Chem., 13 (1994) 1–25.