

TWO POLYMETHOXYFLAVONES FROM *AGERATUM HOUSTONIANUM**

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Abstract—Besides agecorynin C, eupalestin and lucidin dimethyl ether, two new highly oxygenated flavones were isolated from *Ageratum houstonianum*. Their structures were established by spectroscopic and degradative evidence as 5, 6, 7, 8, 2', 3', 4', 5'-octamethoxyflavone and 5, 6, 7, 2', 3', 4', 5'-heptamethoxyflavone.

INTRODUCTION

In previous papers we have published the isolation and structure elucidation of several flavonoids from *Ageratum corymbosum* and *A. strictum* [1, 2]. Now we report the results of our study of *Ageratum houstonianum* Mill. which resulted in the isolation of the already known lucidin dimethyl ether (2a) [3] eupalestin (2b) and agecorynin C (3) [1] as well as two new highly oxygenated flavones with a novel substitution pattern in the B-ring, named agehoustin A (1a) and B (1b). To our knowledge this is the first report of flavones with the 2', 3', 4', 5'-oxygenation pattern and the second of flavones with a tetra-O-substituted B-ring [4]. Agehoustin A (1a) is the first octamethoxyflavone from the Asteraceae and the third compound of this type yet reported from nature. Exoticin (3,5,6,7,8,3',4',5'-octamethoxyflavone) [5] and purpurascenin (3,5,6,7,8,2',4',5'-octamethoxyflavone) [6] are the other two octamethoxyflavonoids isolated so far. Some other octasubstituted flavonoids, with the same substitution pattern as exoticin have been also isolated [7].

RESULTS AND DISCUSSION

The petrol extract of the aerial parts of *Ageratum houstonianum* afforded a mixture of several flavones. Three have been isolated before: lucidin dimethyl ether (2a) from *Lindera lucida* [3], eupalestin (2b) and agecorynin C (3) from *Ageratum corymbosum* [1]. In addition two new fully methylated and highly oxygenated flavones, agehoustin A (1a) and B (1b), were isolated.

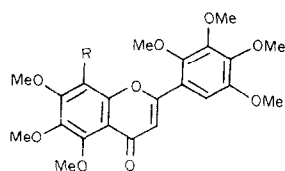
Agehoustin A (1a), $C_{23}H_{26}O_{10}$, was the less polar flavone isolated from the petrol extract, mp 116–118°. Both the UV (268, 318 nm) and the IR (1636, 1580 cm^{-1}) absorptions were typical of non-phenolic flavones [8]. The 1H NMR spectrum of 1a clearly

indicated it to be an octasubstituted flavonoid, since it showed two singlets (1H each) at δ 6.88 and 7.13 indicating the presence of only two flavone-nucleus protons. Six sharp singlets at 3.89 (3H), 3.91 (3H), 3.95 (9H), 3.98 (3H), 3.99 (3H), 4.09 (3H), indicated the presence of eight methoxy groups. The mass spectrum of agehoustin A (1a) confirmed the assumption that it is an octamethoxyflavone, since the molecular ion peak was observed at m/z 462 and a base peak at 447 due to $[M - Me]^+$. Other spectral peaks at m/z 225 $[A_1 - Me]^+$, 197 $[A_1 - Me - CO]^+$ due to A-ring fragmentation [9], suggested that, as in exoticin [5] and purpurascenin [6], agehoustin A (1a) must have all A-ring positions substituted. Therefore the remaining four methoxyl groups can be placed on the B-ring since the 1H NMR displayed two singlet flavone-nucleus protons. In addition, since the 1H NMR spectrum of 1a is different from that of purpurascenin (3, 5, 6, 7, 8, 2', 4', 5'-octamethoxyflavone) [6], it must have the proposed structure 1a. Final confirmation of structure 1a was achieved by alkaline degradation which furnished 2-hydroxy-3, 4, 5, 6-tetramethoxyacetophenone (4a) [1] and 2, 3, 4, 5-tetramethoxybenzoic acid (5a) identified by its mp 82–84° [10], IR, NMR and mass spectra. Hence agehoustin A is 5, 6, 7, 8, 2', 3', 4', 5'-octamethoxyflavone (1a).

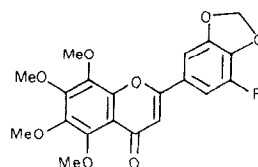
As in the case of agecorynin C (3) [1], additional degradation products were isolated and identified as the keto-enol 6a and 2, 3, 4, 5-tetramethoxyacetophenone (4b) and 2-hydroxy-3, 4, 5, 6-tetramethoxybenzoic acid (5b). 4b and 5b can be explained as further degradation products of 6a (or its corresponding β -diketone).

A second new flavone, agehoustin B (1b), $C_{22}H_{24}O_9$, mp 85–86°, was isolated from the petrol extract. The 1H NMR of 1b was very similar to that of agecorynin C (3) [1]. It showed three singlets (1H each) at δ 6.70, 6.74 and 6.94 due to three flavone nucleus protons and five sharp singlets between 3.8 and 4.0 due to seven methoxy groups. Hence 1b must be a new heptamethoxylated flavone whose structure can be

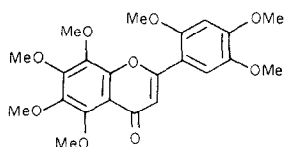
*Part 3 in the series "Flavonoids from *Ageratum* Species". For Part 2 see ref. [2]. This is contribution No. 604 of the Instituto de Química, U.N.A.M.



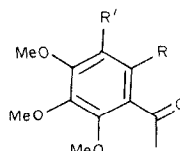
1a R = OMe
1b R = H



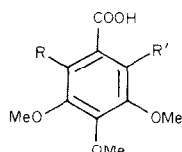
2a R = H
2b R = OMe



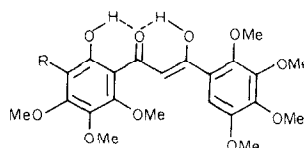
3



4a R = OH, R' = OMe
4b R = H, R' = OMe
4c R = OH, R' = H



5a R = H, R' = OMe
5b R = OH, R' = OMe



6a R = OMe
6b R = H

represented as 5,6,7,2',3',4',5'-heptamethoxyflavone (**1b**). The mass spectrum was in agreement with this assumption, the molecular ion peak being observed at m/z 432 and the base peak at 417 due to $[M - Me]^+$ and other significant fragmentation peaks at m/z 195 $[A_1 - Me]^+$, 167 $[A_1 - Me - CO]^+$ which suggested the tri-substitution of the A-ring [9]. Alkaline degradation of agehoustin B (**1b**) gave the same 2,3,4,5-tetramethoxybenzoic acid (**5a**) as in the case of **1a**. The neutral product was identified as 6-hydroxy-2,3,4-trimethoxyacetophenone (**4c**) by comparison with an authentic sample previously obtained from agecorynin A [1]. Therefore the structure of agehoustin B corresponds to 5,6,7,2',3',4',5'-heptamethoxyflavone (**1b**).

As in the case of **1a** three additional degradation products were also isolated and identified as the same acetophenone **4b** obtained from **1a** and 6-hydroxy-2,3,4-trimethoxybenzoic acid [11], both further degradation products of **6b** (or its corresponding β -diketone) which was also identified.

EXPERIMENTAL

Ageratum houstonianum Mill. was collected in October 1980, in México: Morelos, road to Cuernavaca, ca 60 km south of México City. A voucher, Quijano 40, is on deposit at the Herbarium of Instituto de Biología (UNAM), México.

The air-dried plant material, leaves and flowers (2.9 kg), were extracted with petrol as described before [1], giving 45 g crude syrup which was chromatographed over 300 g Si gel, using petrol- $CHCl_3$ and $CHCl_3$ - Me_2CO mixtures as

eluants. Fractions eluted with $CHCl_3$ - Me_2CO (9 : 1) gave a mixture of several flavonoids. Crystallization from $CHCl_3$ - Et_2O gave a mixture of lucidin dimethyl ether (**2a**) and a new octamethoxyflavone (**1a**). The mother liquor residue (14 g) was rechromatographed over Si gel (200 g) and gave on elution with petrol- $EtOAc$ (4 : 1) further quantities of **1a** and lucidin dimethyl ether (**2a**), mp 166–168° (lit. [3] 163–168°). Further elution with petrol- $EtOAc$ (7 : 3) gave eupalestin (**2b**) and a new heptamethoxyflavone (**1b**). Later fractions eluted with petrol- $EtOAc$ (3 : 7) afforded agecorynin C (**3**). Eupalestin (**2a**) and agecorynin C (**3**) were identified by comparison with authentic samples previously isolated from *Ageratum corymbosum* [1].

Agehoustin A (1a). Earlier fractions eluted with petrol- $EtOAc$ (4 : 1) from the rechromatography gave 600 mg **1a** as needles from $CHCl_3$ - Et_2O , mp 116–117°. UV λ_{max}^{MeOH} nm (ϵ): 268 (20 970), 318 (18 980). IR ν_{max}^{film} cm^{-1} : 1636, 1580, 1560, 1490, 1460, 1410, 1400. 1H NMR (80 MHz, $CDCl_3$): δ 3.89 (3H, s), 3.91 (3H, s), 3.95 (9H, s), 3.98 (3H, s), 3.99 (3H, s), 4.09 (3H, s) (8 \times OMe); 6.88 (1H, s, H-3), 7.13 (1H, s, H-6'). EIMS (probe) 70 eV m/z (rel. int.): 462 $[M]^+$ (19.5), 447 $[M - Me]^+$ (100), 417 $[M - Me - CH_2O]^+$ (19.9), 389 $[M - Me - CH_2O - CO]^+$ (6.09), 225 $[A_1 - Me]^+$ (4.8), 197 $[A_1 - Me - CO]^+$ (9.7), 222 $[B_1]^+$ (2.4), 207 $[B_1 - Me]^+$ (3.6). (Found: C, 59.61; H, 5.71; O, 34.60. $C_{25}H_{26}O_{10}$ requires: C, 59.73; H, 5.67; O, 34.60%.)

Agehoustin B (1b). Rechromatography fractions eluted with petrol- $EtOAc$ (13 : 7) gave 300 mg **1b** as crystals from $CHCl_3$ - Et_2O , mp 85–86°. UV λ_{max}^{MeOH} nm (ϵ): 237 sh (22 500), 265 sh (13 050), 313 (17 500). IR ν_{max}^{film} cm^{-1} : 1732, 1595, 1485, 1410, 1400. 1H NMR (80 MHz, $CDCl_3$): δ 3.86 (3H, s), 3.91

(6H, s), 3.96 (3H, s), 3.97 (6H, s), 3.99 (3H, s) ($7 \times \text{OMe}$), 6.70 (1H, s, H-3 or H-8), 6.75 (1H, s, H-3 or H-8), 6.94 (1H, s, H-6'). EIMS (probe) 70 eV m/z (rel. int.): 432 $[\text{M}]^+$ (18.5), 417 $[\text{M} - \text{Me}]^+$ (100), 401 $[\text{M} - \text{MeO}]^+$ (48.4), 387 $[\text{M} - \text{Me} - \text{CH}_2\text{O}]^+$ (33.8), 195 $[\text{A}_1 - \text{Me}]^+$ (6.5), 167 $[\text{A}_1 - \text{Me} - \text{CO}]^+$ (14.6), 207 $[\text{B}_1 - \text{Me}]^+$ (3.6). (Found: C, 60.64; H, 5.68; O, 33.50. $\text{C}_{22}\text{H}_{20}\text{O}_9$ requires: C, 61.10; H, 5.59; O, 33.30.)

Alkaline degradation of agehoustin A (1a). A 200 mg sample of **1a** was refluxed with 50% KOH (25 ml) in EtOH (10 ml) under N_2 for 72 hr. The reaction mixture was cooled, acidified with dil. HCl and extracted with EtOAc. The EtOAc extract was washed with 10% NaHCO_3 , then H_2O , dried, *concd in vacuo* and the residue purified by prep. TLC to afford the oily 2-hydroxy-3, 4, 5, 6-tetramethoxyacetophenone (**4a**) identified by comparison with a sample previously obtained from eupalestin [1]. The more polar compound was spectroscopically identified as 2, 3, 4, 5-tetramethoxyacetophenone (**4b**). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 215 (15600), 265 (6717), 310 (2934). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 1670, 1585, 1480, 1460, 1400. EIMS (probe) 70 eV m/z (rel. int.): 240 $[\text{M}]^+$ (20.1), 225 $[\text{M} - \text{Me}]^+$ (41.6), 197 $[\text{M} - \text{MeCO}]^+$ (8.5), 43 $[\text{MeCO}]^+$ (100). ^1H NMR (80 MHz, CDCl_3): δ 2.58 (3H, s, COMe), 3.83, 3.84, 3.86, 3.93 ($4 \times 3\text{H}$, s, $4 \times \text{OMe}$), 7.02 (1H, s, H-6).

The NaHCO_3 soln was acidified with HCl, extracted with EtOAc, washed with H_2O , dried, *concd in vacuo* and purified by prep. TLC to give 2, 3, 4, 5-tetramethoxybenzoic acid (**5a**), mp 82–84° (lit. [10], 87–88°). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 206 (35417), 246 sh (4600), 285 (2016). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 2600–3300, 1695, 1675, 1595, 1570. EIMS (probe) 70 eV m/z (rel. int.): 242 $[\text{M}]^+$ (100), 227 $[\text{M} - \text{Me}]^+$ (53.9), 184 $[\text{M} - 58]^+$ (60.1). ^1H NMR (80 MHz, CDCl_3): δ 3.87, 3.90, 3.95, 4.05 ($4 \times 3\text{H}$, s, $4 \times \text{OMe}$), 7.37 (1H, s, H-6), 11.9 (1H, br, COOH).

A second acidic product was isolated and identified as 2-hydroxy-3, 4, 5, 6-tetramethoxybenzoic acid (**5b**). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 2600–3400, 1700, 1450, 1420, 1380. EIMS (probe) 70 eV m/z (rel. int.): 258 $[\text{M}]^+$ (32.4), 240 $[\text{M} - \text{H}_2\text{O}]^+$ (100), 225 $[\text{240} - \text{Me}]^+$ (96.8), 197 $[\text{225} - \text{CO}]^+$ (86.9), 182 $[\text{197} - \text{Me}]^+$ (31.7). ^1H NMR (80 MHz, CDCl_3): δ 3.80, 3.86, 4.04, 4.09 ($4 \times 3\text{H}$, s, $4 \times \text{OMe}$).

Keto-enol 6a. A 100 mg sample of **1a**, refluxed with 50% KOH (20 ml) in EtOH (10 ml) under N_2 for 4 hr gave the keto-enol **6a** as main product. Mp 100–102°. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 205 (37600), 282 (10266), 373 (15600). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 2500–3500, 1600 sh, 1580, 1555. EIMS (probe) 70 eV m/z (rel. int.): 480 $[\text{M}]^+$ (7.3), 449 $[\text{M} - 31]^+$ (20.6), 241 $[\text{C}_{11}\text{H}_{13}\text{O}_6]^+$ (6.3), 240 $[\text{C}_{11}\text{H}_{12}\text{O}_6]^+$ (9.7), 225 $[\text{C}_{11}\text{H}_{13}\text{O}_5]^+$ (100). ^1H NMR (80 MHz, CDCl_3): δ 3.82 (3H, s), 3.88 (9H, s), 3.90 (3H, s), 3.93 (3H, s), 3.95 (3H, s), 4.10 (3H, s, $8 \times \text{OMe}$); 7.18 (1H, s, H-3), 7.66 (1H, s, H-6), 12.35 (1H, s, $\phi_{\text{A}}\text{-OH}$), 15.81 (1H, s, $\text{C}_2\text{-OH}$).

Alkaline degradation of agehoustin B (1b). A 200 mg sample of **1b** was treated and worked-up as described above.

The neutral material after TLC purification afforded 6-hydroxy-3, 4, 5-trimethoxyacetophenone (**4c**), identical with an authentic sample obtained from agecorynin A [1] and the same acetophenone **4b** obtained from **1a**. A third neutral compound was isolated and identified as the keto-enol **6b**, mp 93–94°. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 206 (33750), 281 (12421), 333 (10078), 385 (13664). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 2500–3400, 1600, 1560. EIMS (probe) 70 eV m/z (rel. int.): 450 $[\text{M}]^+$ (17.5), 419 $[\text{M} - \text{MeO}]^+$ (26.1), 225 $[\text{C}_{11}\text{H}_{13}\text{O}_5]^+$ (100), 211 $[\text{C}_{10}\text{H}_{11}\text{O}_5]^+$ (11.0). ^1H NMR (80 MHz, CDCl_3): δ 3.78 (3H, s), 3.87 (12H, s), 3.94 (3H, s), 3.95 (3H, s) ($7 \times \text{OMe}$), 6.25 (1H, s, H-8), 7.17 (1H, s, H-3), 7.65 (1H, s, H-6'), 12.74 (1H, s, $\phi_{\text{A}}\text{-OH}$), 15.75 (1H, s, $\text{C}_2\text{-OH}$).

The acidic material after TLC purification gave the same tetramethoxybenzoic acid **5a** obtained from **1a** and 6-hydroxy-2, 3, 4-trimethoxybenzoic acid, mp 107–110° (lit. [11], 113–115°). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} : 2500–3500, 1630, 1460, 1420, 1370, 1260, 1090. EIMS (probe) 70 eV m/z (rel. int.): 228 $[\text{M}]^+$ (26.8), 210 $[\text{M} - \text{H}_2\text{O}]^+$ (87.8), 195 $[\text{210} - \text{Me}]^+$ (86.9), 167 $[\text{195} - \text{CO}]^+$ (100), 94 (29.2), 69 (40.7). ^1H NMR (80 MHz, CDCl_3): δ 3.78, 3.87, 4.14 ($3 \times 3\text{H}$, s, $3 \times \text{OMe}$), 6.32 (1H, s, H-3), 11.94 (2H, s, COOH and OH).

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