Note

Direct fluorination at positions 3' 4', and 6' of β -D-glucopyranosyltheophylline

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Considerable effort has been expended in the synthesis of deoxyfluoro sugars¹⁻⁶ as a result of interest in such analogs for use as hexokinase inhibitors⁷, as carriers of ¹⁸F in positron emission tomography⁸ (PET), and as possible anticancer agents⁹. Also, fluorinated pentofuranose nucleosides have been investigated for the treatment of infections by *Herpes simplex*¹⁰ (HSV) and human immunodeficiency¹¹ (HIV) viruses.

Little has been reported on fluorinated hexopyranose nucleosides¹² and, to our knowledge, the use of diethylaminosulfur trifluoride¹⁴ (DAST) has not been reported hitherto for their synthesis.

We now decribe the synthesis of 3'-, 4'-, and 6'-fluoro derivatives of β -D-glucopyranosyltheophylline.

DAST can effect^{2,13,14} the replacement of OH by F at primary, secondary, and tertiary positions (the last two with inversion of configuration) in high yields.

7-(2,3,4-Tri-O-benzoyl-6-deoxy-6-fluoro- β -D-glucopyranosyl)theophylline (3) was prepared in dry diglyme³ at 60° from the tribenzoate 2. The key precursor of 7-(2,4,6-tri-O-benzoyl-3-deoxy-3-fluoro- β -D-allopyranosyl)theophylline (13) was 7-(4,6-O-isopropylidene- β -D-glucopyranosyl)theophylline (5), which reacted with DAST in CH₂Cl₂ containing 4-dimethylaminopyridine to afford 7-(3-deoxy-3-fluoro-4,6-O-isopropylidene- β -D-allopyranosyl)theophylline (11). 7-(2,3,6-Tri-O-benzoyl-4-deoxy-4-fluoro- β -D-galactopyranosyl)theophylline (9) was obtained by the same procedure from 7-(2,3,6-tri-O-benzoyl- β -D-glucopyranosyl)theophylline (8). If the 4-dimethylaminopyridine was omitted, then 7-(2,3,6-tri-O-benzoyl-4-deoxy- β -D-*erythro*-hex-4-enopyranosyl)theophylline was obtained instead of 9. Attempts to fluorinate 7-(3,4,6-tri-O-benzoyl- β -D-glucopyranosyl)theophylline were unsuccessful even in hot diglyme¹⁵, probably because of steric hindrance in the formation of the reaction intermediate.

Debenzoylation (Zemplén) of 3 and 9 gave 4 and 10, respectively, and deacetalation of 11 gave 12.

All the compounds synthesised were characterised by their proton-coupled ¹⁹F- and proton-decoupled ¹³C-n.m.r. spectra. The n.m.r. data given in Tables I–III

| 20 7 30 | 40 | 6.17 | | H-4' | . <u>с</u> -н | H-6'a | | 9.3 | | J3',4' | | n 0' C | J5',6'b | J _{6'a,6'b} |
|--|----------------|------|------|------|---------------|--|----------|----------------|-----|--------|------|---------------|-----------------------|----------------------|
| ç, 4 % | 0.48 | | 6.04 | 5.74 | 3.80 | 4.10 | 3.91 | | 9.5 | 9.5 | 9.1 | 3.5 | 2.8 | 10.0 |
| 4 ^b 8 <i>a</i> | 6.56 | 6.06 | 6.12 | 5.77 | 4.33 | 4.71 | 4.55 | 8.8 | 9.4 | 9.7 | 9.3 | 2.5 | 2.7 | 7.3 |
| 80 | 5.83 | 4.10 | 3.71 | I | 3.82 | 4.74 | 4.74 | 9.1 | 0.6 | 10.0 | 8.6 | | | |
| | 6.21 | 5.98 | 5.68 | 4.32 | Ţ | ←3.85-4.05→ | * | | 9.5 | 9.4 | 9.1 | ļ | I | ł |
| ъ | 6.55 | 6.18 | 5.65 | 5.29 | 4.63 | 4.52 | 4.43 | | 9.5 | 2.2 | 0 | 4.0 | 1 | 6.3 |
| 10 | 5.85 | 4.44 | 3.99 | 4.99 | 4.10 | +3.9' | 5-3.88→ | | 8.2 | 0 | 0 | 2.3 | 7.4 | ľ |
| 5" | 5.78 | 4.08 | 3.79 | 3.92 | Ĵ, | ←3.85-3.69→ | * | | 7.0 | 5.4 | 10.7 | I | - | |
| 11a | 5.92 | 4.20 | 5.08 | 4.00 | 4.00 | $0 \leftarrow 3.89 - 3.78 \rightarrow$ | 9-3.78→ | | 1.8 | 0.5 | 1 | - | I | |
| 12 ⁶ | 6.05 | 4.49 | 5.20 | 4.00 | Ţ | 4.14-3.78- | * | | 0.5 | 0 | - | I | I | 1 |
| 13" | 5.64 | 4.80 | 5.50 | 4.80 | 4.49 | 3.92 | 3.53 | | 0.5 | 0.5 | 9.2 | 3.8 | | 12.1 |
| R ^C H ₂ X | | | | | H | R | R2 | R ₃ | | R4 | R5 | R | | × |
| | - | | | 1 | | F | OBz | OBz | 2 | Н | Н | OB. | OBz | OTr |
| R ² | | | | 7 | ł | T | OBz | OBi | 2 | Н | Н | OB | Z | НО |
| | R ² | | | e | | Н | OB_{Z} | OB | 2 | Н | Н | OB | 2 | ĹL, |
| 4 4 | | | | 4 | | Н | НО | HO | | Н | Н | HO | L : | ц |
| | | | | ŝ | , - | H | НО | НО | | Н | Н | 0 | CMe ₂ -O | |
| | | | | 9 | | Н | OBz | OB | Z | Н | Н | ò | CMe ₂ -O | |
| | | | | 7 | - | Н | OBz | OB | 2 | Н | Н | HO | | но |
| | | | | œ | H | Н | OBz | OB | 2 | Н | Η | HO | | OBz |
| | | | | 6 | - | Н | OBz | OB | 2 | Н | Ľ. | Η | | OBz |
| | | | | 10 | _ | Н | НО | HO | | Н | ц | Η | | OBz |
| | | | | H | | Н | НО | H | | ц | H | 5 | O-CMe ₂ -O | |
| | | | | 12 | | Н | НО | Η | | Ľ. | H | HO | | HO |
| | | | | 13 | Т | F | OBz | Η | | н | Н | OBz | 2 | OBz |

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TABLE I

NOTE

TABLE II

| Compound | F | J _{F-6',H-6'} | J _{F-6', H-5'} | J _{F-4',H-4'} | $\mathbf{J}_{\textit{F-4'},\textit{H-3'}}$ | J _{F-4',H-5'} | J _{F-3',H-3'} | J _{F-3',H-4'} | J _{F-3',H-2'} |
|------------------------|-------|------------------------|-------------------------|------------------------|--|------------------------|------------------------|------------------------|------------------------|
| 3 ^a | -56.3 | 46.6 | 21.2 | | | | | | |
| 4 ^b | -56.2 | 47.3 | 25.4 | | | | | | |
| 9 ^a | -48.2 | | | 52.1 | 25.6 | 25.6 | | | |
| 10 ⁶ | -46.8 | | | 51.4 | 30.9 | 30.9 | | | |
| 11 ^a | -50.6 | | | | | | 54.0 | 27.0 | 27.0 |
| 12 ^b | -48.2 | | | | | | 53.6 | 28.9 | 28.9 |
| 13ª | -48.1 | | | | | | 53.7 | 28.9 | 28.9 |

¹⁹F-N.M.R. DATA (δ in p.p.m., J in Hz)

^{*a*}In CDCl₃ (internal C_6F_6). ^{*b*}In D₂O (external C_6F_6).

establish the location of the fluorine substituents and the configuration of the products.

The ¹³C-signals (Table III) were assigned by comparing the spectra with those of the parent compounds. Replacement of OH by F causes a downfield shift of the resonance of the attached carbon by 15–20 p.p.m., upfield shifts of the signals of the adjacent carbons by 2–5 p.p.m., and ${}^{1}J_{F,C}$ values of 180–200 Hz (Table III). Thus, the locations of the fluorine substituents were shown to be at position 6 in 3 and 4, at position 4 in 9 and 10, and at position 3 in 11–13.

For 9–13, the ${}^{1}J_{F,C}$ values (Table III) are in accordance with the results of Wray¹⁶ (175.9–183.1 Hz), but those for 3 and 4 (177.5 and 169.5) are higher than the previous values (~167.5)¹⁶. The $J_{F-6,C-5}$ value of 18.5 Hz for 3 and 4 is identical to that for 6-deoxy-6-fluoro- β -D-glucopyranose¹⁶. This author recorded ${}^{2}J_{F,C}$ values of ~17.5 when the oxygen function on the coupled carbon was *gauche* to the coupled fluorine, but smaller values were found for 9 and 11–13 (Table III).

In the ¹H- and ¹⁹F-n.m.r. spectra (Tables I and II, respectively), ² $J_{F,H}$ was 46.6–47.3 for **3** and **4**, 51.4–52.1 for **9** and **10**, and 53.6–54.0 for **11–13**. These values agree with other data^{2,17} on deoxyfluorosugars. The $J_{F.6,H-5}$ values of 21.2–25.4 for **3** and **4** accord with other observations³ and indicate the preponderance of the rotamer with F-6 antiperiplanar to H-5. The values of ³ $J_{F,H}$ for vicinal *trans*-diaxial nuclei in pyranose structures are generally¹⁸ in the range of 21–30 Hz. The observed ³ $J_{F,H}$ values 25.6–30.9 for **9–13** confirm the vicinal *trans*-diaxial H/F relationships. Therefore, the fluorine substituent is axial and the compounds have the ⁴ $C_1(D)$ conformation, which shows that replacement of OH by F does not affect the conformation⁶.

EXPERIMENTAL

General methods. — Melting points are uncorrected. T.l.c. was performed on Silica Gel 60- F_{254} (Merck) and flash chromatography on Silica Gel 60 (240–400

| Compound | C-l' | C-2′ | C-3' | C-4' | C-5' | | J _{F-6', C-6'} | J _{F-6',C-5'} | C-6' JF6, C6' JF6, C5' JF6, C4' JF4, C4' JF4, C3' JF3, C3' JF3, C2' JF3, C4' JF3, C1' | J _{F-4',C-4'} | J _{F-4',C-5'} | J _{F-4',C-3'} | J _{F-3',C-3'} | J _{F-3',C-2'} | J _{F-3',C-4'} | $\mathbf{J}_{F:Y,C:I'}$ |
|--|----------|------------|------------------------|-----------|---------|------------|-------------------------|------------------------|---|------------------------|------------------------|------------------------|------------------------|------------------------|------------------------|-------------------------|
| 2ª | 82.7 | 71.3 | 72.8 | 68.7 | 77.4 | 6.09 | | | | | | | | | | |
| 3ª | 82.5 | 71.9 | 73.8 | 68.3 | 75.8 | 81.0 | 177.5 | 18.5 | 5.4 | | | | | | | |
| 46 | 80.0 | 71.8 | 76.1 | 70.7 | 74.5 | 80.2 | 169.5 | 18.5 | | | | | | | | |
| 8 a | 83.2 | 71.1 | 79.2 | 66.6 | ļ | 61.7 | | | | | | | | | | |
| 9 a | 82.3 | 68.9 | 72.0 | 86.3 | 74.3 | 61.9 | | | | 186.7 | 17.6 | 16.0 | | | | |
| 10 ⁶ | 84.1 | 71.7 | 72.9 | 82.9 | 75.4 | 62.4 | | | | 183.5 | 13.9 | 14.8 | | | | |
| 5ª | 85.7 | 72.9 | 74.6 | 75.0 | 62.0 | 70.8 | | | | | | | | | | |
| 11ª | 84.3 | 6.9 | 90.1 | 70.7 | 61.9 | 66,9 | | | | | | | 181.2 | 14.8 | 14.8 | 3.7 |
| 12 ^b | 85.3 | 68.0 | 95.8 | 71.3 | 78.7 | 63.3 | | | | | | | 175.4 | 14.8 | 14.8 | |
| 13" | 80.7 | 69.7 | 87.8 | 67.2 | 72.2 | 62.4 | | | | | | | 177.6 | 9.1 | 14.8 | 7.4 |
| | | | 2 (| - | | | | | | | | | | | | |
| ^a In CDCl ₃ (internal Me ₄ Si). ^{n} | ternal M | e4Si). "II | וון ט _ג ט ה | nternal 3 | (trimet | hylsilyl)- | -l-propan | esultonic | "In $D_2 O$ [internal 3-(trimethylsilyl)-1-propanesultonic acid, sodium salt]. | ium salt] | | | | | | |

¹³C-N.M.R. DATA (8 IN P.P.M., J IN HZ)

TABLE III

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mesh, Merck). N.m.r. spectra were recorded at room temperature with a Bruker 300 MSL spectrometer.

7-(2,3,4-Tri-O-benzoyl- β -D-glucopyranosyl)theophylline (2). — A solution of β -D-glucopyranosyltheophylline¹⁹ (2 g) and trityl chloride (1.7 g) in anhydrous pyridine (100 mL) was boiled under reflux overnight. Benzoyl chloride (2.2 mL) was then added and, after 2 h, the mixture was cooled, diluted with dichloromethane, washed twice with ice-water, and concentrated. Crystallisation of the residue from ethanol gave 7-(2,3,4-O-benzoyl-6-trityl- β -D-glucopyranosyl)theophylline (1; 4 g, 77%), m.p. 227–228°, $[\alpha]_D^{20} - 22^\circ$ (c 0.1, methanol).

Anal. Calc. for C₅₃H₄₄N₄O₉: C, 72.27; H, 5.00; N, 6.36. Found: C, 72.76; H, 4.96; N, 6.47.

A solution of 1 (5.6 g, 6 mmol) in 3:2 HCOOH–ether²⁰ (20 mL) was kept at room temperature for 30 min, then diluted with ether (150 mL), washed successively with brine and with saturated aq. sodium hydrogencarbonate, dried, and concentrated. Flash chromatography (EtOAc–hexane, 7:3) of the syrupy residue gave 2 (3.3 g, 80%), m.p. 146–147° (from methanol), $[\alpha]_{D^0}^{20}$ -5° (c 0.1, methanol).

Anal. Calc. for $C_{34}H_{30}N_4O_{10} \cdot H_2O$: C, 60.71; H, 4.76; N, 8.33. Found: C, 60.62; H, 4.89; N, 8.17.

7-(2,3,4-Tri-O-benzoyl-6-deoxy-6-fluoro- β -D-glucopyranosyl)theophylline (3). — DAST (0.585 mL, 6 mmol) was added dropwise to a stirred solution of 2 (980 mg, 1.5 mmol) in dry diglyme (6 mL) at -20°. The temperature was raised slowly to 60° and maintained for 1 h. The mixture was poured into ice, the precipitate was collected, and a solution in CH₂Cl₂ was washed once with M NaHCO₃ and twice with water, then concentrated. Flash chromatography (dichloromethanc-ethyl acetate, 4:1) of the resulting oil gave 3 (615 mg, 68%), m.p. 179–180° (from MeOH), [α]_D²⁰ -4° (c 0.1, methanol).

Anal. Calc. for C₃₄H₂₉FN₄O₉·0.5CH₃OH: C, 61.62; H, 4.58; F, 2.83; N, 8.32. Found: C, 61.68; H, 4.47; F, 2.80; N, 8.30.

7-(6-Deoxy-6-fluoro- β -D-glucopyranosyl)theophylline (4). — To a solution of 3 (665 mg, 1 mmol) in anhydrous methanol (10 mL) was added methanolic M sodium methoxide (1 mL). The mixture was stirred for 2 h at room temperature, neutralised with Amberlite IR-120 (H⁺) resin, and concentrated. Recrystallisation of the residue from methanol gave 4 (310 mg, 90%), m.p. 218–220°, $[\alpha]_D^{20} - 12^\circ$ (c 0.12, methanol).

Anal. Calc. for C₁₃H₁₇FN₄O₆·CH₃OH: C, 44.68; H, 5.58; F, 5.05; N, 14.89. Found: C, 44.89; H, 5.53; F. 4.92; N, 15.0.

7-(4,6-O-Isopropylidene- β -D-glucopyranosyl)theophylline (5). — A mixture of 7- β -D-glucoyranosyltheophylline¹⁹ (1.8 g), 2,2-dimethoxypropane (5 mL), and dry *p*-toluenesulfonic acid (15 mg) in dry *N*,*N*-dimethylformamide (15 mL) was stirred at room temperature for 20 h. M NaHCO₃ (1 mL) was added and the mixture concentrated under vacuum. A solution of the residue in dichloromethane was filtered and concentrated, and the residue was crystallised from ethanol to give 5 (1.39 g, 70%), m.p. 208–210°, $[\alpha]_D^{20} - 33^\circ$ (*c* 0.13, methanol).

Anal. Calc. for C₁₆H₂₂N₄O₇: C, 50.26; H, 5.76; N, 14.66. Found: C, 50.06; H, 5.80; N, 14.65.

7-(2,3-Di-O-benzoyl- β -D-glucopyranosyl)theophylline (7). — Conventional treatment of **5** with pyridine and benzoyl chloride and crystallisation of the product from ethanol gave 7-(2,3-di-O-benzoyl-4,6-O-isopropylidene- β -D-glucopyranosyl)-theophylline (**6**, 90%), m.p. 136–139°, $[\alpha]_{D}^{20} - 33°$ (c 0.13, methanol).

Amberlite IR-120 (H⁺) resin was added to a methanolic solution of **6**, which was boiled under reflux for 1 h, then filtered, and concentrated. The syrupy residue was crystallised from ethanol to give **7** (90%), m.p. 225–226°, $[\alpha]_D^{20} - 12^\circ$ (c 0.1, methanol).

Anal. Calc. for $C_{27}H_{26}N_4O_8 \cdot H_2O$: C, 57.04; H, 4.93; N, 9.86. Found: C, 57.23; H, 4.81; N, 10.08.

7-(2,3,6-Tri-O-benzoyl- β -D-glucopyranosyl)theophylline (8). — To a stirred solution of 7 (500 mg, 1 mmol) in pyridine (50 mL) at -30° was added, dropwise, 1 equiv. (0.116 mL) of benzoyl chloride. The mixture was stored at 0° for 72 h and then worked-up in the usual manner. Flash chromatography (ethyl acetate-hexane, 70:30) of the product gave 8 (460 mg, 70%), m.p. 226° (from EtOH), $[\alpha]_{D}^{20} - 12^{\circ}$ (c 0.13, methanol).

Anal. Calc. for $C_{34}H_{30}N_4O_{10}$: C, 62.38; H, 4.59; N, 8.56. Found: C, 62.30; H, 4.63; N, 8.56.

7-(2,3,6-Tri-O-benzoyl-4-deoxy-4-fluoro- β -D-galactopyranosyl)theophylline (9). — To a stirred solution of 8 (654 mg, 1 mmol) and 4-dimethylaminopyridine (244 mg, 2 mmol) in dry dichloromethane (20 mL) at -30° was added DAST (0.195 mL, 2 mmol) during 15 min under nitrogen. The mixture was then allowed to attain room temperature. After 24 h, the mixture was cooled to 0° , methanol was added, solvents were removed under vacuum, and the resulting oil was poured into icewater. The precipitate was collected, and a solution in CH₂Cl₂ was washed with M NaHCO₃ and water to pH 7, dried, and concentrated. Flash chromatography (dichloromethane-ethyl acetate, 9:1) of the residue gave 9 (446 mg, 68%), m.p. 125-126° (from MeOH), $[\alpha]_{D}^{20} + 25^{\circ}$ (c 0.1, ethyl acetate).

Anal. Calc. for C₃₄H₂₉FN₄O₉: C, 62.20; H, 4.42; F, 2.90; N, 8.54. Found: C, 62.05; H, 4.38; F, 2.29; N, 8.29.

7-(4-Deoxy-4-fluoro- β -D-galactoyranosyl)theophylline (10). — Debenzoylation of 9, as for 3, gave 10 (90%), m.p. 215–216° (from MeOH), $[\alpha]_{D}^{20}$ +32° (c 0.1 methanol).

Anal. Calc. for $C_{13}H_{17}FN_4O_6 \cdot 0.5CH_3OH$: C, 45.0; H, 5.27; F, 5.27; N, 15.55. Found: C, 44.84; H, 5.22; F, 5.23; N, 15.52.

7-(3-Deoxy-3-fluoro-4,6-O-isopropylidene- β -D-allopyranosyl)theophylline (11). — A solution of 5 (1.53 g, 4 mmol) and 4-dimethylaminopyridine (1 g, 8 mmol) in dry CH₂Cl₂ at -20° was treated with DAST (0.8 mL, 8 mmol) as described for 9. Column chromatography (ethyl acetate-hexane, 3:1) of the product gave 11 (740 mg, 43%), m.p. 235-236° (from EtOH), $[\alpha]_{D}^{20}$ -13° (c 0.1, ethyl acetate). Anal. Calc. for $C_{16}H_{21}FN_4O_6 \cdot 0.5H_2O$: C, 48.85; H, 5.60; F, 4.83; N, 14.24. Found: C, 49.01; H, 5.59; F, 4.90; N, 14.21.

7-(3-Deoxy-3-fluoro- β -D-allopyranosyl)theophylline (12). — To a solution of 11 in methanol was added Amberlite IR-120 (H⁺) resin, and the mixture was boiled under reflux for 90 min, then filtered, and concentrated to give 12 (90%), m.p. 246° (from EtOH), $[\alpha]_{D}^{20} - 22^{\circ}$ (c 0.1, methanol).

Anal. Calc. for $C_{13}H_{17}FN_4O_6 \cdot 0.25C_2H_5OH$: C, 45.56; H, 5.20; F, 5.34; N, 15.75. Found: C, 45.58; H, 5.13; F, 5.30; N, 15.80.

7-(2,4,6-Tri-O-benzoyl-3-deoxy-3-fluoro-β-D-allopyranosyl)theophylline (13). — Conventional treatment of 12 with benzoyl chloride-pyridine gave 13 (80%), m.p. 126-127° (from MeOH), $[\alpha]_D^{20}$ -35° (c 0.1, methanol).

Anal. Calc. for C₃₄H₂₉FN₄O₉: C, 62.20; H, 4.40; F, 2.90; N, 8.50. Found: C, 62.20; H, 4.34; F, 2.83; N, 8.40.

REFERENCES

- A. B. FOSTER, R. HEMS, AND J. M. WEBBER, Carbohydr. Res., 5 (1967) 292–301; J. S. BRIMACOMBE, A. B. FOSTER, R. HEMS, J. H. WESTWOOD, AND L. D. HALL, Can. J. Chem., 48 (1970) 3946–3952; E. M. BESSELL, A. B. FOSTER, J. H. WESTWOOD, L. D. HALL, AND R. N. JOHNSON, Carbohydr. Res., 19 (1971) 39–48; L. D. HALL, R. N. JOHNSON, J. ADAMSON, AND A. B. FOSTER, Can. J. Chem., 49 (1971) 118–123.
- 2 C. W. SOMAWARDHANA AND E. G. BRUNNGRABER, Carbohydr. Res., 121 (1983) 51-60.
- 3 M. SHARMA AND W. KORYTNYK, Tetrahedron Lett., (1977) 573-576.
- 4 P. J. CARD, J. Org. Chem., 48 (1983) 393-395.
- 5 S. G. WITHERS, D. J. MACLENNAN, AND I. P. STREET, Carbohydr. Res., 154 (1986) 127-144.
- 6 P. KOVÁČ, H. J. C. YEH, AND C. P. J. GLAUDEMANS, Carbohydr. Res., 169 (1987) 23-34.
- 7 W. A. SZAREK, G. W. HAY, AND B. DOBOSZEWSKI, J. Chem. Soc., Chem. Commun., (1985) 663-664.
- 8 P. G. WALTER, in P. N. CAMBELL AND G. D. GREVILLE (Eds.), *Essays in Biochemistry*, Vol. 2, Academic Press, New York, 1966, p. 33.
- 9 J. B. SHATTON, H. P. MORRIS, AND S. WEINHOUSE, Cancer Res., 29 (1969) 1161-1171.
- 10 A. D. BORTHWICK, S. BUTT, K. BIGGADIKE, A. M. AXALL, S. M. ROBERTS, P. M. YOUDS, B. E. KIRK, B. R. BOOTH, J. M. CAMERON, S. W. COX C. L. P. MARR, AND M. D. SHILL, J. Chem. Soc., Chem. Commun., (1988) 656–658.
- 11 P. HERDEWIJN, R. PAUWELS, M. BABA, J. BALZARINI, AND E. DE CLERCQ, J. Med. Chem., 30 (1987) 2131-2137.
- 12 J. KIBURIS, A. B. FOSTER, AND J. H. WESTWOOD, J. Chem. Soc., Chem. Commun., (1975) 44-45, and references therein.
- 13 W. J. MIDDLETON, J. Org. Chem., 40 (1975) 574-578.
- 14 M. HUDLICKY, Org. React., 35 (1988) 513-637.
- 15 P. KOVÁČ, Carbohydr. Res., 153 (1986) 168-170.
- 16 V. WRAY, J. Chem. Soc., Perkin Trans. 2, (1976) 1598-1605.
- 17 L. PHILIPPS AND V. WRAY, J. Chem. Soc., B, (1971) 1618-1624.
- 18 A. A. E. PENGLIS, Adv. Carbohydr. Chem. Biochem., 38 (1981) 195-285.
- 19 F. LECLERCQ, Thèse, Paris, 1971.
- 20 M. BESSODES, D. KOMIOTIS, AND K. ANTONAKIS, Tetrahedron Lett., 27 (1986) 579-580.