

81. Catalytic One-Pot Osmylation of Cyclohexadienes: Stereochemical and Conformational Studies of the Resulting Polyols

by Théophile Tschamber^{a)}*, Frédérique Backenstrass^{a)}, Hans Fritz^{b)}, and Jacques Streith^{a)}

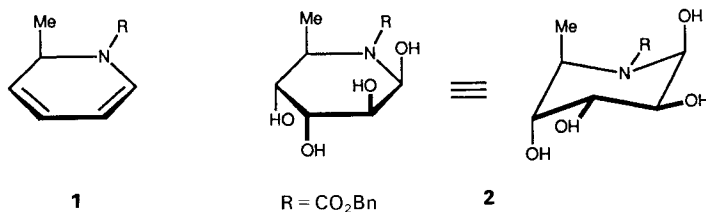
^{a)} Ecole Nationale Supérieure de Chimie, Université de Haute-Alsace, 3, rue Alfred Werner, F-68093 Mulhouse Cédex

^{b)} Physikalische Abteilung, Ciba-Geigy AG, CH-4002 Basel

(8.IV.92)

Catalytic double osmylation is described for a series of cyclohexadienes in acetone/H₂O in the presence of the co-oxidant *N*-methylmorpholine *N*-oxide (NMO). The formation of polyols occurred stereospecifically with cyclohexadienes **3**, **7**, and **11a**, leading thereby to tetrols **5a**, and **9a** and to *allo*-inositol (**14a**), respectively. To the contrary, *trans*-cyclohexadiene-diol **15a** gave a mixture of the stereoisomeric inositols **18a** (*epi*), **19a** (*neo*), and **20a** (*chiro*). High-field NMR let to clearcut conformational analyses of the polyhydroxylated derivatives.

Introduction. – In a preliminary communication, we described the one-pot double catalytic osmylation of dihydropyridine **1** which led stereospecifically to the aminodeoxyaltrose derivative **2** [1]. The stereospecific ‘*cis/trans/cis/trans*’ outcome of this multistep reaction – which led in good yield to 5 asymmetric centres – is most probably due to minimization of steric interactions during each osmylation step. Due to the strong anomeric effect, which is more pronounced in piperidinoses than in pyranoses, compound **2** occurred exclusively in the chair conformation [1].

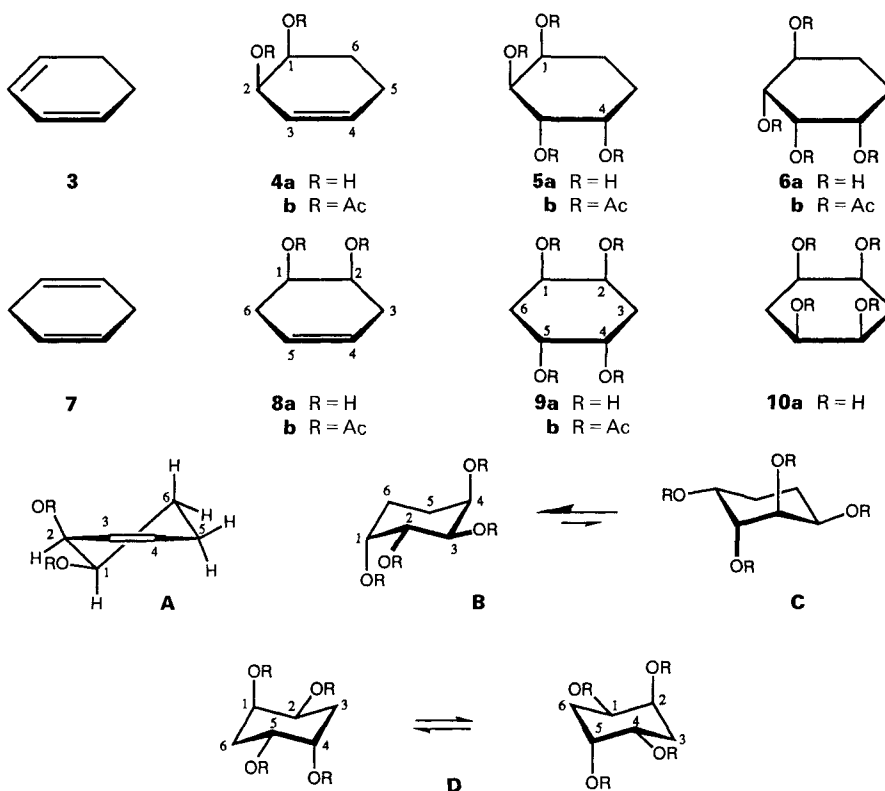


In the meantime, Sharpless, Park, and Moon Kim reported a similar catalytic osmylation methodology with open-chain conjugated (*E,E*)-dienes, and found stereoselectivities which were strongly in favour of the ‘*cis/trans/cis*’ stereoisomeric tetrols [2]. Besides these two preliminary studies [1] [2], osmylation of 1,3-dienes has received but little attention, despite its obvious utility in the synthesis of polyhydroxylated compounds, KMnO₄ being used more frequently than OsO₄, leading usually with poor stereoselectivity to a mixture of tetrols [3–9].

We describe herein catalytic single and double osmylation of cyclohexadienes **3**, **7**, **11a**, and **15a** with *N*-methylmorpholine *N*-oxide (NMO) as the co-oxidant, the experi-

mental conditions being similar to those used for the synthesis of aminodideoxyaltrose **2**, and to those described by *Sharpless* and coworkers [2] (see below).

Catalytic Osmylation of Cyclohexa-1,3-diene (3) and Cyclohexa-1,4-diene (7). – Catalytic osmylation of **3** in the presence of an excess of NMO (more than 2 equiv.) at room temperature in acetone/H₂O led exclusively to tetrol **5a** which was characterised as its tetraacetate **5b** (overall yield 86%). The same procedure applied to cyclohexadiene **7** gave tetrol **9a** and thence the corresponding tetraacetate **9b** (81% overall yield). Since **5a** and **9a** are the less hindered tetrols, it is most likely that they were formed under steric control. Incomplete osmylation of **3** in the presence of 1 equiv. of NMO, followed by acetylation gave **4b** (79%) and **5b** (5%). Diene **7**, treated likewise, led to a mixture of **8b** (32%) and **9b** (27%). These results indicate that diol **8a** is more reactive (less hindered) than diol **4a**.

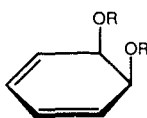


While *Pasternak* and *Friedli* could show that catalytic osmylation of enediol **4a** in the presence of an excess of AgClO₃ led stereospecifically to tetrol **5a** in good yield [3], *Zelinski et al.* found that hydroxylation of diene **3** with KMnO₄ gave **6a** in poor yield only [4]. The stereochemical outcome of this latter reaction was explained by assuming a 'cis-complexation' of both double bonds by KMnO₄ followed by an 'anti'-hydrolysis of the so-formed 'manganic ester' [5]. As to the double hydroxylation of diene **7** with

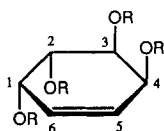
KMnO₄, it gave a mixture **8a/9a**, both in poor yield [6]. Eventually, *McCasland et al.* described the catalytic osmylation of **7** in the presence of AgClO₃, which led to **9a/10a**, the former being the major product [10].

It appears, therefore, that the OsO₄/NMO hydroxylation is the method of choice, since it leads to higher yields; furthermore, it is stereospecific, at least with dienes **3** and **7**.

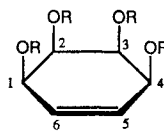
Catalytic Osmylation of Cyclohexa-3,5-diene-1,2-diols **11a (*cis*) and **15a** (*trans*).** – Catalytic mono- and bis-osmylation of cyclohexadiene-diols **11a** and **15a** in the presence of NMO led to conduritols and to cyclitols. Thus, osmylation of **11a** for 15 h in the presence of 2 equiv. of NMO followed by acetylation led to conduritols E tetraacetate (**12b**; 48%) and *allo*-inositol hexaacetate (**14b**; 31%), whereas conduritols D tetraacetate (**13b**) could not be isolated. This indicates that steric crowding in **12a** is such that the second osmylation step occurs with difficulty. According to *Nakajima et al.*, catalytic osmylation of **11a** in the presence of AgClO₃ gave mostly phenol (48%) and small amounts of conduritols E (4% **12b** after acetylation) [7]. Furthermore, when bis-acetate



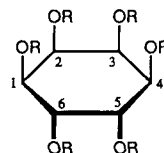
11a R = H
b R = Ac



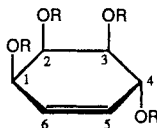
12a R = H,
conduritols E
b R = Ac



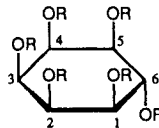
13a R = H,
conduritols D
b R = Ac



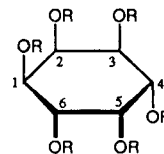
14a R = H,
allo-inositol
b R = Ac



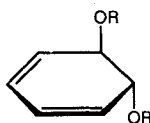
16a R = H,
conduritols C
b R = Ac



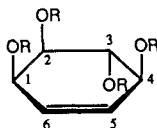
18a R = H,
epi-inositol
b R = Ac



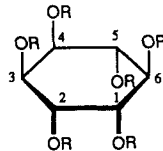
19a R = H,
neo-inositol
b R = Ac



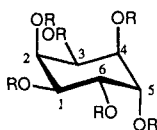
15a R = H
b R = Ac



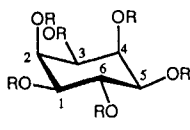
17a R = H,
conduritols F
b R = Ac



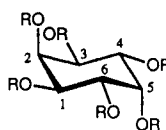
20a R = H,
chiro-inositol
b R = Ac



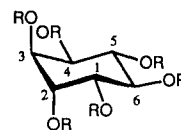
E (*allo*)



F (*epi*)



G (*neo*)



H (*chiro*)

11b was hydroxylated with KMnO_4 in neutral medium, these authors isolated **12b** only (61 %) [8], whereas catalytic osmylation of **11a** in the presence of 1 equiv. of NMO led to a mixture **12b/13b** after acetylation (see below). When **12b** was hydroxylated with KMnO_4 , *allo*-inositol was isolated as its acetate **14b** (45 %) [8].

Catalytic osmylation of **11a** in the presence of 1 equiv. only of NMO followed by acetylation gave **12b/13b** (overall yield 88 %), **12b** being the major product; under these conditions, **11a** was entirely consumed after 2 h and *allo*-inositol (**14a**) did not form.

From these two experiments, *i.e.* catalytic osmylation of **11a** in the presence of 1 or 2 equiv. of NMO, one concludes that conduritol D (**13a**) is transformed into *allo*-inositol (**14a**) at a faster rate than its stereoisomer conduritol E (**12a**). This is clearly due to a steric interaction between OsO_4 and the OH functions which is more pronounced in **12a** than in **13a**, a conclusion which had also reached *Angyal* and *Gilham* [11]. Furthermore, since **13a** and **12a** gave the same inositol **14a**, bis-osmylation of **11a** is stereospecific overall.

Diol **15a**, which was prepared according to *Platt* and *Oesch*'s procedure [12], was osmylated in the presence of a 2-fold excess of NMO (more than 2 equiv.) for 5 d and the reaction mixture acetylated to give **18b** (*epi*)/**19b** (*neo*)/**20b** (*chiro*) in a 42:11:47 ratio (overall yield 60 %). The same experimental conditions applied to diacetate **15b** for 3 d led, after acetylation, to **16b/18b/19b/20b** in a 50:9:13:28 ratio.

Mono-osmylation of **15a** or of **15b** in the presence of 1 equiv. of NMO, followed by peracetylation, led to **16b/17b/19b/20b** in a 66:25:2:7 ratio (as determined by $^1\text{H-NMR}$).

These results indicate that conduritol F (**17a**) is more reactive (towards OsO_4) than its stereoisomer conduritol C (**16a**). This again is best explained by steric crowding (in the transition states) which is more severe in **16** than in **17**. Nevertheless, it is rather surprising that the seemingly more crowded conduritol C (**16a**) was formed at a faster rate than conduritol F (**17a**).

Structural Analyses of the Polyols. – The above described conduritols and cyclitols all are known compounds. Their relative configurations were established decades ago, in most cases by applying low-field NMR techniques. We used 250- and 400-MHz $^1\text{H-NMR}$ spectroscopy to analyze unequivocally their conformations.

Cyclohexene-diols and Diacetates 4 and 8. Compounds **4** are asymmetric; a large coupling constant ($J(1,6_{\text{ax}}) = 10.2 \text{ Hz}$) clearly demonstrates that diol **4a** occurs essentially in its pseudo-chair conformation **A**¹⁾. As a consequence, all CH atoms of **4** appear with well differentiated chemical shifts (8 *m*, see Table 1). Compounds **8** also belong to the C_1 point group; nevertheless a fast equilibrium between the two enantiomeric pseudo-chair conformations leads to a simplification of the $^1\text{H-NMR}$ spectrum at room temperature with only 3 differentiated chemical shifts (see Table 1).

Table 1. Selected $^1\text{H-NMR}$ Data of Diols **4a** and **8a**, and of Their Acetates **4b** and **8b**. 250 MHz, 300 K. δ in ppm, J in Hz, internal standard TMS.

	Solvent	H–C(1)	H–C(2)	H–C(3)	H–C(4)	H–C(5)	H _{ax} –C(6)	H _{eq} –C(6)
4a	CD_3OD	3.73	4.04	5.69	5.81	2.05 ^a , 2.21 ^b	1.82	1.68
b	CDCl_3	5.10	5.42	5.67	5.99	2.25 ^c	1.95	1.78
8a	CD_3OD	3.86	3.86	2.24 ^d	5.53	5.53	2.24 ^d	
b	CDCl_3	5.18	5.18	2.35 ^d	5.62	5.62	2.35 ^d	

¹⁾ The pseudo-chair conformation **A** is also corroborated by the $J(1,2)$, $J(2,3)$, and $J(2,4)$ coupling constants (see Table 1). For similar half-chair conformations, see [13–15].

Tab. 1 (cont.)

	$J(1,2)$	$J(1,6a)$	$J(1,6e)$	$J(2,3)$	$J(2,4)$	$J(2,6e)$	$J(3,4)$	$J(3,5)^d$	$J(4,5)^d$	$J(4,6e)$	$J(5a,5e)$	$J(5,6a)^d$	$J(5,6e)^e$	$J(6a,6e)$
4b CDCl ₃	3.9	10.2	3.3	4.3	1.0	1.0	9.9	2.1	3.6	0.3	18.2	7.0	5.5	12.6

^{a)} H_{ax}–C(5).
^{b)} H_{eq}–C(5).
^{c)} AB pattern for H_{eq}–C(5) and H_{ax}–C(5).
^{d)} Mean values for H_{eq}–C(3)/H_{eq}–C(6) and for H_{ax}–C(3)/H_{ax}–C(6).
^{e)} These data represent mean values of coupling constants with H_{ax}–C(5) and H_{eq}–C(5).

Cyclohexane-tetrols and Tetraacetates 5 and 9. Tetrol **5a** is C₂-symmetric, so that only 3 peaks appear in the ¹³C-NMR spectrum at room temperature (and 4 peaks in the ¹H-NMR), the interconversion between the chair conformations **B** and **C** being fast (see Table 2). According to the coupling constant $J(2,3)$, measured from the ¹³C-satellites of the H–C(2), H–C(3) signal, **B** is the major conformation (60%) at equilibrium. At –60°, both chair conformations **B** and **C** appear as well separated C₂-symmetric entities, the **B/C** ratio being 56:44 for **5a**, and 90:10 for **5b** (see Table 2).

Tetrol **9a** and its tetraacetate **9b** have C_i symmetry, so that 3 peaks should appear in the ¹³C-NMR spectrum for the ring C-atoms, and 4 peaks in the ¹H-NMR spectrum for the cyclohexane H-atoms; this is indeed observed at high field and at low temperature (–30°), with $J(2ax,3ax) = J(5ax,6ax) = 12$ Hz (these are the only J values which can be determined) for **9b**. At room temperature though, the fast conformational inversion between the 2 identical chair conformations **D** leads to 2 peaks in the ¹³C- and to 2 peaks in ¹H-NMR, the fast inverting molecule having statistical C_{2h} symmetry. Such a conformational analysis could not be achieved at 60 MHz [10].

Conduritols E–F and Tetraacetates 12, 13, 16, and 17. Structure and conformation of conduritols C, D, E, and F were already studied by Abraham *et al.* at high-field ¹H-NMR [16]. Vogel and coworkers undertook a higher-resolution ¹H- and ¹³C-NMR study of conduritols C, D, and F and confirmed Abraham's conformational analyses [17][18]. Our own highfield ¹H- and, in part, ¹³C-NMR investigations led to similar conclusions in terms of conformational analysis (see *Exper. Part*). Conduritol **D** (**13a**) undergoes a fast conformational equilibrium between the 2 enantiomeric half-chairs (3 peaks in ¹H- and 3 in ¹³C-NMR at room temperature). Conduritol **E** (**12a**) is C₂-symmetric and its ¹H-NMR shows 3 peaks both at –30° and at room temperature. Conduritol **F** (**17a**) being asymmetric (C₁ point group) appears with 6 peaks in the ¹H-NMR at room temperature.

Inositols and Hexaacetates 14 (allo), 18 (epi), 19 (neo), and 20 (chiro). *allo*-Inositol hexaacetate (**14b**) is asymmetric and leads to 6 peaks both in the ¹H- and in the ¹³C-NMR for the ring H- and C-atoms at low temperature (see Table 3), the absorption bands having been attributed *via* double-irradiation techniques. At –30°, $J(1,6) = 11$ Hz can be determined which demonstrates the *trans*-diaxial configuration of these 2 protons as indicated in **E** (the other J values are smaller than 3 Hz). At 45°, a fast equilibrium occurs between the two enantiomeric chair conformations as demonstrated by the simplified spectrum, the molecule having now a statistical C_s symmetry (3 peaks in the ¹H- and 3 in the ¹³C-NMR).

epi-Inositol hexaacetate (**18b**) having C_s symmetry shows 4 peaks in the ¹H- and in the ¹³C-NMR, as expected (see Table 3). Only one chair conformation is present, *i.e.* **F**, with 4 equatorial AcO groups ($J(1,6) = J(5,6) = 10.4$ Hz).

Table 2. Selected ^1H -NMR (400 MHz) and ^{13}C -NMR (100.6 MHz) Data of Tetrols **5a** and **9a** and of Their Tetraacetates **5b** and **9b**. δ in ppm, internal standard TMS (^1H) and CDCl_3 (δ (CDCl_3) = 77 ppm; ^{13}C)^a.

	Solvent, temp.	H-C(1)	H-C(2)	H-C(3)	H-C(4)	H-C(5)	H-C(6)	C(1)	C(2)	C(3)	C(4)	C(5)	C(6)
5a	CD_3OD , 25°	3.93	3.73	3.73	3.93	1.76, 1.63	1.63, 1.76	70.23	73.29	73.29	70.23	26.59	26.59
	CD_3OD , -60°	3.98	3.64	3.64	3.98	1.82, 1.58	1.58, 1.82	71.22	72.01	72.01	71.22	26.22	26.22
5b	C	3.82	3.84	3.84	3.82	1.70, 1.58	1.58, 1.70	68.30	74.12	74.12	68.30	27.00	27.00
	CDCl_3 , -25°	5.32	5.13	5.13	5.32	1.86, 1.71	1.71, 1.86	68.96	69.21	69.21	68.96	23.29	23.29
9a	B	5.34	5.03	5.03	5.34	ca. 1.75	ca. 1.75	69.16	69.59	69.59	69.16	23.39	23.39
	C	4.97	5.19	5.19	4.97	b)	b)	68.54	68.62	68.62	68.54	23.54	23.54
9b	D_2O , 25°	4.00	4.00	1.87	4.00	4.00	1.87	70.30	70.30	34.88	70.30	70.30	34.89
	CDCl_3 , 25°	5.30	5.30	2.05	5.30	5.30	2.05	67.85	67.85	29.06	67.85	67.85	29.06
	CDCl_3 , -30°	5.44	5.08, 1.95	2.10	5.44	5.08, 1.95	2.10	67.93	67.07	28.58	67.93	67.07	28.58

^a) Assignment of the ^1H - and ^{13}C -NMR signals by selective decoupling experiments.^b) Not determined.Table 3. Selected ^1H - and ^{13}C -NMR Data of Inositol Hexaacetates **14b**, **18b**, **19b**, and **20b**. δ in ppm, internal standard TMS (^1H) and CDCl_3 (δ (CDCl_3) = 77 ppm; ^{13}C).

	Frequency, solvent, temp.	H-C(1)	H-C(2)	H-C(3)	H-C(4)	H-C(5)	H-C(6)	C(1)	C(2)	C(3)	C(4)	C(5)	C(6)
14b	400 MHz, CDCl_3 , -30°	5.28 ^a)	5.63	5.24	5.29 ^a)	5.42	5.46	66.58	68.46	65.22	67.59	67.22	66.09
	400 MHz, CDCl_3 , 45°	5.32	5.45	5.45	5.32	5.47	5.47	67.72	67.50	67.50	67.72	67.34	67.34
18b	250 MHz, C_6D_6 , 27°	5.02	5.82	4.66	5.82	5.02	6.04	68.9	68.6	67.4	68.6	68.9	65.9
19b	250 MHz, CDCl_3 , 27°	5.35	5.66	5.35	5.35	5.66	5.35	67.5	68.1	67.5	67.5	68.1	67.5
20b	250 MHz, CDCl_3 , 27°	5.32	5.40	5.40	5.32	5.43	5.43	68.9	69.8	69.8	68.9	67.3	67.3

^a) Can be inverted.

neo-Inositol hexaacetate (**19b**) is C_{2h} -symmetric so that only 2 peaks appear in the ^1H - and ^{13}C -NMR (see Table 3). The coupling constants ($J(1,6) = J(3,4) = 11.1$ Hz and $J(1,2) = J(2,3) = J(4,5) = J(5,6) = 2.5$ Hz) can be calculated by simulation *via* the LAOCOON-III (PANIC) iteration program [19], showing thereby that conformation **G** is the predominant one for **19b**.

chiro-Inositol hexaacetate **20b** is C_2 -symmetric and leads to 3 peaks in the ^1H - and ^{13}C -NMR (see Table 3), as expected. The coupling constants are calculated as above *via* the LAOCOON-III program [19] and permit to state that **H** is the dominant chair conformation, as expected ($J(2,3) = 2.5$ Hz; $J(1,2) = J(3,4) = 1.4$ Hz; $J(1,6) = J(5,6) = J(4,5) = 10.3$ Hz).

Conclusion. – Double catalytic osmylation in the presence of 2 equiv. of NMO in acetone/ H_2O proved to be '*cis/trans/cis*'-stereospecific with (*Z,Z*)-cyclohexadienes **3**, **7**, and **11a**, leading thereby to tetrol **5a**, to tetrol **9a**, and to *allo*-inositol (**14a**), respectively. The C_2 -symmetric *trans*-cyclohexadiene-diol **15a** gave a mixture of the three stereoisomeric inositols **18a–20a**, *via* conduritol C (**16a**) and conduritol F (**17a**), the overall yield being moderate. The absence of stereoselection in this latter instance is reminiscent of the non-stereoselective osmylation as described recently by Campbell *et al.* for a 1,5-disubstituted cyclohexa-1,3-diene [20].

The support of the *Centre National de la Recherche Scientifique (URA-135)* is gratefully acknowledged. We also wish to thank the *Ministère de la Recherche et de la Technologie* for a research grant to one of us (F.B.) and Prof. C. Le Drian for a sample of (+)-conduritol F tetraacetate (**17b**).

Experimental Part

General. Flash chromatography (FC): silica gel (Merck 60, 230–400 mesh). TLC: Al roll silica gel (Merck 60, F_{254}). M.p.: Kofler hot bench or Büchi-SMP-20 apparatus; corrected. IR Spectra (cm^{-1}): Perkin-Elmer 157-G. ^1H - and ^{13}C -NMR Spectra: Bruker AC-F-250, and AM-400 using double-irradiation techniques; tetramethylsilane (TMS; ^1H) and CDCl_3 or C_6D_6 (δ (CDCl_3) = 77.0 or δ (C_6D_6) = 128.0 ppm rel. to TMS; ^{13}C) as internal references; δ in ppm and J in Hz. High-resolution (HR) MS were measured on a MAT-311 spectrometer at the University of Rennes. Microanalyses were carried out by the 'Service Central de Microanalyses' of the CNRS, at Vernaison.

Reagents. The catalyst was prepared according to [21] from OsO_4 (1 g) and 1 ml of 70% *t*-BuOOH in 200 ml of *t*-BuOH, a soln. which is *ca.* 0.02 mmol per ml. Amberlyst A-26 (OH^- form) was prepared starting from its chloride precursor A-26 (Cl^- form; 10 g) by adding 1N aq. NaOH and letting the ion exchange occur for 1 h. The Amberlyst A-26 (OH^- form) beads were then rinsed several times with H_2O and MeOH and could be kept for several weeks in the cold.

Cyclohexane-*r*-1,*c*-2,*t*-3,*t*-4-tetrayl Tetraacetate (5b**).** To a stirred soln. of cyclohexa-1,3-diene (**3**; 3.12 g, 38.9 mmol) in acetone/ H_2O 9:1 (30 ml) were added NMO (16.02 g, 118 mmol, *ca.* 3 equiv.) and cat. OsO_4 soln. (5 ml). After 2 h at r.t., the soln. was evaporated and the crude residue taken up in Ac_2O (25 ml) and pyridine (50 ml) and left to react at r.t. for 15 h. Some AcOEt was added and the soln. washed sequentially with 10% aq. Na_2SO_3 soln., 10% aq. NaHCO_3 soln., and brine. The combined org. soln. was dried (MgSO_4) and evaporated and the residue separated by column chromatography (AcOEt): **5b** (10.58 g, 86%). Colourless crystals. M.p. 163–164° (AcOEt). IR (KBr): 1740, 1375, 1365, 1240, 1220, 1200. ^1H -NMR: Table 2. ^{13}C -NMR: Table 2. Anal. calc. for $\text{C}_{14}\text{H}_{20}\text{O}_8$ (316.30): C 53.16, H 6.37; found: C 53.2, H 6.3.

Cyclohexane-*r*-1,*c*-2,*t*-3,*t*-4-tetrol (5a**).** To a stirred soln. of **5b** (1.88 g, 5.94 mmol) in MeOH (20 ml) was added Amberlyst A-26 (OH^-). After 1 h at r.t., the suspension was filtered, the resin washed with hot MeOH, and the org. soln. evaporated: **5a** (867 mg, 100%). Colourless crystals. M.p. 214–215° (MeOH; [22]: 216°). IR (KBr): 3350 (br.), 2950, 2935, 2895, 1450, 1250. ^1H -NMR: Table 2. ^{13}C -NMR: Table 2. Anal. calc. for $\text{C}_6\text{H}_{12}\text{O}_4$ (148.16): C 48.64, H 8.16; found: C 48.6, H 8.3.

cis-Cyclohex-3-ene-1,2-diyl Diacetate (**4b**). As described for **5b**, with **3** (1.87 g, 23.4 mmol), acetone/H₂O 9:1 (15 ml), NMO (3.21 g, 23.7 mmol, 1 equiv.), cat. OsO₄ soln. (5 ml), Ac₂O (10 ml), and pyridine (20 ml). FC of the crude (AcOEt/cyclohexane 3:7) gave **4b** as an oil (3.64 g, 79%) and **5b** (360 mg, 5%). **4b**: IR (KBr): 1735, 1365, 1245, 1225. ¹H-NMR: Table 2. ¹³C-NMR: Table 2. HR-MS: 138.0685 (C₈H₁₀O₂ [M – CH₃CO₂H]⁺, calc. 138.06807).

cis-Cyclohex-3-ene-1,2-diol (**4a**). As described for **5a**, with **4b** (742 mg, 3.74 mmol), THF (6 ml), MeOH (15 ml), and Amberlyst A-26 (OH[–]): **4a** (426 mg, 100%). Oil. IR (KBr): 3395 (br.), 2920, 1075. ¹H-NMR: Table 1.

Cyclohexane-*r*-1,*c*-2,*t*-4,*t*-5-tetrayl Tetraacetate (**9b**). As described for **5b**, with cyclohexa-1,4-diene (7; 925 mg, 11.9 mmol), acetone/H₂O 9:1 (14 ml), NMO (4.19 g, 31.0 mmol, 2.6 equiv.), cat. OsO₄ soln. (5 ml), Ac₂O (6 ml)/pyridine (12 ml; 40° for 15 h): **9b** (3.05 g, 81%). Colourless crystals. M.p. 170–171° (CH₂Cl₂/hexane; [23]: 170°). IR (KBr): 1540, 1525, 1380, 1365, 1245, 1235. ¹H-NMR: Table 2. ¹³C-NMR: Table 2. Anal. calc. for C₁₄H₂₀O₈ (316.30): C 53.16, H 6.37; found: C 53.1, H 6.4.

Cyclohexane-*r*-1,*c*-2,*t*-4,*t*-5-tetrol (**9a**). As described for **5a**, with **9b** (727 mg, 2.3 mmol), MeOH (10 ml), and Amberlyst A-26 (OH[–]): **9a** (339 mg, 100%). Colourless crystals. M.p. 246° (MeOH; [23]: 241°). IR (KBr): 3370, 3260, 2940, 1390, 1350, 1225. ¹H-NMR: Table 2. ¹³C-NMR: Table 2. Anal. calc. for C₆H₁₂O₄ (148.16): C 48.64, H 8.16; found: C 48.6, H 8.3.

cis-Cyclohex-4-ene-1,2-diyl Diacetate (**8b**). As described for **5b**, with **7** (905 mg, 11.3 mmol), acetone/H₂O 9:1 (10 ml), NMO (1.54 g, 11.4 mmol, 1 equiv.), cat. OsO₄ soln. (5 ml), Ac₂O (6 ml), and pyridine (12 ml). FC (AcOEt/cyclohexane 3:7) gave **8b** (825 mg, 37%) as an oil and **9b** (973 mg, 27%). **8b**: IR (KBr): 1740, 1365, 1250, 1220. ¹H-NMR: Table 1. HR-MS: 138.0685 (C₈H₁₀O₂ [M – CH₃CO₂H]⁺, calc. 138.06807).

cis-Cyclohex-4-ene-1,2-diol (**8a**). As described for **5a**, with **8b** (685 mg, 3.45 mmol), THF (2 ml), MeOH (5 ml), and Amberlyst A-26 (OH[–]): **8a** (371 mg, 94%), colourless crystals after FC (AcOEt) of the crude. M.p. 79° (crystal washed with Et₂O; [6] [24]: 74–78.5°). IR (KBr): 3260 (br.), 2890. ¹H-NMR: Table 1. Anal. calc. for C₆H₁₀O₂ (114.14): C 63.13, H 8.83; found: C 63.0, H 8.9.

Conduritol E Tetraacetate (**12b**) and allo-Inositol Hexaacetate (**14b**). As described for **5b**, with *cis*-cyclohexa-3,5-diene-1,2-diol (**11a**; 685 mg, 6.10 mmol), acetone/H₂O 9:1 (14 ml), NMO (1.74 g, 12.7 mmol, 2.1 equiv.), cat. OsO₄ soln. (5 ml; 15 h at r.t.), and Ac₂O (5 ml)/pyridine (10 ml; 40° for 15 h). FC (AcOEt/cyclohexane 4:6) of the crude gave **14b** (830 mg, 31%) and **12b** (920 mg, 48%), both as colourless crystals.

12b: M.p. 156° (AcOEt/hexane; [7]: 153°). IR (KBr): 1745 (br.), 1370, 1245, 1215. ¹H-NMR (400 MHz, CDCl₃, 25°): 5.68 (H–C(1), H–C(4)); 5.44 (H–C(2), H–C(3)); 5.91 (H–C(5), H–C(6)); 2.08 (Ac); 2.02 (Ac); |J(1,6) + J(4,5)| ≈ 4, |J(1,2) + J(3,4)| ≈ 3.5, J(2,3) ≈ 10, J(5,6) ≈ 10. ¹H-NMR (400 MHz, CDCl₃, –30°): 5.68; 5.41; 5.92; 2.10; 2.04. ¹³C-NMR (100.6 MHz, CDCl₃, 25°): 170.2 (C=O); 169.9 (C=O); 128.2 (C(5), C(6)); 66.6 (C(2), C(3)); 66.1 (C(1), C(4)); 20.8 (Me); 20.6 (Me). Anal. calc. for C₁₄H₁₈O₈ (314.28): C 53.50, H 5.77; found: C 53.7, H 5.7.

14b: M.p. 141–143° (CH₂Cl₂/hexane; [25]: 144°). IR (KBr): 1745 (br.), 1435, 1370, 1225. ¹H-NMR: Table 3. ¹³C-NMR: Table 3. Anal. calc. for C₁₈H₂₄O₁₂ (432.37): C 50.00, H 5.60; found: C 49.8, H 5.5.

Conduritol E (**12a**). As described for **5a**, with **12b** (290 mg, 0.92 mmol), THF (2 ml), MeOH (5 ml), and Amberlyst A-26 (OH[–]): **12a** (126 mg, 94%). Colourless crystals. M.p. 183° (MeOH; [7]: 180°). IR (KBr): 3365 (br.), 1105, 1095. ¹H-NMR (400 MHz, CD₃OD, 25°): 4.23 (H–C(1), H–C(4)); 3.90 (H–C(2), H–C(3)); 5.75 (H–C(5), H–C(6)). ¹³C-NMR (100.6 MHz, CD₃OD): 130.8 (C(5), C(6)); 71.2 (C(2), C(3)); 67.7 (C(1), C(4)). Anal. calc. for C₆H₁₀O₄ (146.16): C 49.31, H 6.90; found: C 49.4, H 7.1.

allo-Inositol (**14a**). As described for **5a**, with **14b** (334 mg, 0.77 mmol): **14a** (132 mg, 95%). Colourless crystals. M.p. 310° (dec.; [25]: 320° (dec.)). IR (KBr): 3480–3180, 2920, 1440, 1420, 1110. Anal. calc. for C₆H₁₂O₆ (180.16): C 40.00, H 6.71; found: C 39.9, H 6.8.

Conduritol E Tetraacetate (**12b**) and Conduritol D Tetraacetate (**13b**). As described for **5b**, with **11a** (218 mg, 1.94 mmol), acetone/H₂O 9:1 (4 ml), NMO (269 mg, 1.99 mmol, 1 equiv.), cat. OsO₄ (2 ml), Ac₂O (3 ml)/pyridine (6 ml; 5 h at 40°). FC (AcOEt/cyclohexane 4:6) of the crude residue gave **12b** (343 mg, 56%; see above) and **13b** (195 mg, 32%), both as colourless crystals. **13b**: M.p. 107–108° (AcOEt/hexane; [7]: 102–104°). IR (KBr): 1745, 1730, 1365, 1230, 1215. ¹H-NMR (250 MHz, CDCl₃, 25°): 5.58 (*m*, H–C(1), H–C(4)); 5.37 (*m*, H–C(2), H–C(3)); 5.90 (*m*, H–C(5), H–C(6)); 2.01 (Ac).

Conduritol C Tetraacetate (**16b**), epi-Inositol Hexaacetate (**18b**), neo-Inositol Hexaacetate (**19b**), and chiro-Inositol Hexaacetate **20b**. To a stirred soln. of diacetate **15b** (303 mg, 1.54 mmol, 2.15 equiv.) in acetone/H₂O 9:1 (8 ml) were added NMO (446 mg, 3.29 mmol) and cat. OsO₄ soln. (1.5 ml). After 24 h at r.t., the soln. was evaporated, the residue taken up in Ac₂O (3.5 ml) and pyridine (7 ml), and the resulting soln. heated overnight to 45°. Et₃N (5 ml) was added and the soln. kept for 24 h at 45°. After addition of toluene and MeOH, the soln. was

evaporated and the crude residue separated into several fractions by column chromatography. Each fraction was partly resolved by FC (AcOEt/cyclohexane 4:6) whereby **16b**, **18b**, **19b**, and **20b** could be isolated.

16b: M.p. 93° (AcOEt/cyclohexane; [7]: 92°). IR (KBr): 1740, 1360, 1225. ¹H-NMR (C₆D₆, 250 MHz, 25°): 5.48 (H–C(1)); 5.90 (H–C(2)); 5.33 (H–C(3)); 5.96 (H–C(4)); 5.58 (H–C(5)); 5.40 (H–C(6)); 1.68 (Ac); 1.66 (Ac); 1.63 (Ac). ¹³C-NMR (62.9 MHz, C₆D₆, 25°): 170.0 (CO); 169.6 (CO); 169.1 (CO); 128.4 (C(6)); 127.6 (C(5)); 71.2 (C(3)); 70.3 (C(2)); 69.9 (C(4)); 68.2 (C(1)); 20.5 (Me); 20.3 (Me); 20.3 (Me); 20.2 (Me).

18b: M.p. 183–184° (CH₂Cl₂/hexane; [9]: 186°). ¹H-NMR: Table 3. ¹³C-NMR: Table 3.

19b: M.p. 257–259° (MeOH; [9]: 250–252°). ¹H-NMR: Table 3. ¹³C-NMR: Table 3.

20b: M.p. 113–114° (Et₂O/hexane; [26]: 111–112°). ¹H-NMR: Table 3. ¹³C-NMR: Table 3.

Conduritol C Tetraacetate (16b) and Conduritol F Tetraacetate (17b). As described for **5b**, with *trans*-cyclohexa-3,5-diene-1,2-diol (**15a**; 200 mg, 1.8 mmol), acetone/H₂O 9:1 (5 ml), NMO (241 mg, 1.8 mmol, 1 equiv.), cat. OsO₄ soln. (2 ml; 20 h at r.t.), Ac₂O (2.5 ml)/Et₃N (5 ml; 15 h at 40°). ¹H-NMR (250 MHz): **16b/17b** (ca. 70%) 3:1, **18b/19b/20b** (< 20%). Compound **17b** could not be isolated in pure form, but its ¹H-NMR was identical with the one of a pure sample of **17b** as synthesized by *Le Drian et al.* [18]. ¹H-NMR (non-purified **17b**; 250 MHz, C₆D₆): 5.90 (*dd*, *J* = 11.0, 7.5, H–C(3)); 5.65 (*m*, H–C(1)); 5.55 (*ddd*, *J* = 7.5, 1.7, 1.2, H–C(4)); 5.39 (*m*, H–C(5), H–C(6)); 5.23 (*dd*, *J* = 11.0, 3.9, H–C(2)); 1.71, 1.69, 1.63, 1.56 (4s, AcO).

REFERENCES

- [1] F. Backenstrass, J. Streith, Th. Tschamber, *Tetrahedron Lett.* **1990**, 31, 2139.
- [2] Ch. Y. Park, B. Moon Kim, K. B. Sharpless, *Tetrahedron Lett.* **1991**, 32, 1003.
- [3] Th. Posternak, H. Friedli, *Helv. Chim. Acta* **1953**, 36, 251.
- [4] N. D. Zelinski, Ya. I. Denisenko, M. S. Eventova, *C. R. Acad. Sci. URSS* **1935**, 1, 313.
- [5] H. Z. Sable, K. A. Powel, H. Katchian, C. B. Niewoehner, S. B. Kadlec, *Tetrahedron* **1970**, 26, 1509.
- [6] K. A. Powell, A. L. Hughes, H. Katchian, J. F. Jerauld, H. Z. Sable, *Tetrahedron* **1972**, 28, 2019.
- [7] M. Nakajima, I. Tomida, S. Takei, *Chem. Ber.* **1957**, 90, 246.
- [8] M. Nakajima, I. Tomida, S. Takei, *Chem. Ber.* **1959**, 92, 163.
- [9] M. Nakajima, I. Tomida, N. Kurihara, S. Takei, *Chem. Ber.* **1959**, 92, 173.
- [10] G. E. McCasland, S. Furuta, L. F. Johnson, J. N. Shoolery, *J. Org. Chem.* **1963**, 28, 894.
- [11] S. J. Angyal, P. T. Gilham, *J. Chem. Soc.* **1958**, 375.
- [12] K. L. Platt, F. Oesch, *Synthesis* **1977**, 449.
- [13] H. Günther, G. Jikeli, *Chem. Rev.* **1977**, 77, 599.
- [14] M. Barfield, R. J. Spear, S. Sternhell, *Chem. Rev.* **1976**, 76, 593.
- [15] A. Defoin, H. Fritz, G. Geffroy, J. Streith, *Helv. Chim. Acta* **1988**, 71, 1642.
- [16] R. J. Abraham, H. Gottschalck, H. Paulsen, W. A. Thomas, *J. Chem. Soc.* **1965**, 6268.
- [17] C. Le Drian, E. Vieira, P. Vogel, *Helv. Chim. Acta* **1989**, 72, 338.
- [18] C. Le Drian, J.-P. Vionnet, P. Vogel, *Helv. Chim. Acta* **1990**, 73, 161.
- [19] LAOCOON-III program provided by *Bruker-Spectrospin*.
- [20] M. M. Campbell, M. F. Mahon, M. Sanisbury, P. A. Searle, *Tetrahedron Lett.* **1991**, 32, 951.
- [21] K. Akashi, R. E. Palermo, K. B. Sharpless, *J. Org. Chem.* **1978**, 43, 2063.
- [22] G. E. McCasland, *Adv. Carbohydr. Chem.* **1965**, 20, 10.
- [23] E. O. N. Lippmann, *Ber. Dtsch. Chem. Ges.* **1901**, 34, 1159.
- [24] G. R. Krow, R. Carmosin, A. Maucuso, *Org. Prep. Proced. Lut.* **1977**, 9, 285 (*CA*: **1978**, 88, 37305w).
- [25] S. J. Angyal, L. Andersen, *Adv. Carbohydr. Chem.* **1959**, 14, 135.
- [26] H. G. Fletcher, Jr., G. R. Frindlay, *J. Am. Chem. Soc.* **1948**, 70, 4050.