Simple access to the nonproteinogenic peptide fragments of lysobactin from azetidin-2-one frameworks

Claudio Palomo,* Jesus M. Aizpurua, Iñaki Ganboa, Beatriz Odriozola, Elena Maneiro, José Ignacio Miranda and Raquel Urchegui

Departamento de Química Organica, Universidad del Pais Vasco, Facultad de Química, Paseo Manuel Lardizabal-3, 20080 San Sebastián, Spain

A convenient route to the β -hydroxy α -aminoacid-derived tripeptides found in the macrocyclic peptide lactone antibiotic lysobactin from azetidin-2-one frameworks is provided for the first time.

Lysobactin, Fig. 1, is a macrocyclic peptide lactone antibiotic isolated from the fermentation of Lysobacter sp.ATCC 53042.1 The mode of antibacterial action of this antibiotic has been shown to be comparable with the observed selectivity and potency of the well known clinically useful antibiotic vancomycin. Since infectious strains of vancomycin-resistant bacteria have been isolated,2 lysobactin might be the alternative antibacterial agent of choice in spite of its relative toxicity. Consequently, the development of new semisynthetic peptides of lysobactin with improved therapeutic index is of considerable interest and, thereby, a concise approach to the key structural elements, the α -amino β -hydroxy acids, present in this antibiotic is essential. Although a number of suitable methods for the stereoselective construction of α -amino β -hydroxy acids^{3,4} exists, it would be desirable and conceptually new to develop a synthesis of non-proteinogenic α-amino acid derivatives ready for direct use in peptide coupling reactions. Here we report that α -hydroxy(alkoxy) β -lactams fulfil this criterion and provide a direct way for the construction of the tripeptides A and **B** of lysobactin.

The synthesis of tripeptide A started from the 4-carboxy β lactam 1⁵ [mp 207–209 °C; $[\alpha]_D^{25} = -115.8^\circ$ (c = 1.0, Me₂CO)] as a β -hydroxy aspartic acid form possessing the β carboxyl group and the α -amino moiety simultaneously protected. The dipeptide unit 2 [mp 197–199 °C, $[\alpha]_D^{25} = -25.0^\circ$ (c = 0.5, CH_2Cl_2)] was obtained in 95% overall yield after activation of the carboxyl group with cyanuric fluoride and subsequent coupling with O-benzyl-L-serine benzyl ester according to Carpino's procedure. To achieve this, 2 was first Ndeprotected in the usual way⁷ with the addition of methylene chloride as cosolvent to give 3 as a pale yellow solid [90%, mp 142-144 °C, $[\alpha]_D^{25} = -10.5$ ° (c = 0.8, CH_2Cl_2)]. Formation of tripeptide 4 was accomplished in 80% yield by ring opening of 3 and subsequent acylation of the resulting free β -amino ester intermediate with BocGlyF and N-methylmorpholine (NMM). Alternatively, to facilitate the β -lactam cleavage by means of other nucleophiles, 8 3 was treated with (Boc)₂O and DMAP to give 5 with complete chemoselectivity [80% yield, mp

Scheme 1 Reagents and conditions: i, cyanuric fluoride, pyridine, CH_2Cl_2 , 6 h, room temp.; ii, $S-NH_2CH(CH_2OBn)CO_2Bn$, CH_2Cl_2 , NMM, 3 h, room temp.; iii, $(NH_4)_2Ce(NO_3)_6$, $MeCN-H_2O-CH_2Cl_2$, 1 h, 0 °C; iv, $CISiMe_3$, MeOH, 1 h, 0 °C, room temp.; v, BocGlyF, NMM, CH_2Cl_2 , 2 h, 0 °C; vi, $(Boc)_2O$ (2 equiv.), DMAP, MeCN, room temp., 16 h; vii, NH_4OH , DMF; viii, CF_3CO_2H , CH_2Cl_2 , room temp.

138–140 °C]. The opening of **5** with aq. NH₄OH (25%) (15–20 equiv.) in DMF proceeded cleanly within about 3 h to furnish **6** [mp 86–88 °C, $[\alpha]_D^{25} = +2.3^\circ$ (c = 0.5, CH₂Cl₂)] in 90% isolated yield. Dipeptide **6** was next *N*-Boc deprotected and transformed into the tripeptide **7** as above [mp 106–108 °C $[\alpha]_D^{25} = +8.4^\circ$ (c = 1.1, CH₂Cl₂)].

The strategy for tripeptide B is based on our recent reported method for the construction of α -amino acid N-carboxy anhydrides (NCAs) via Baeyer-Villiger rearrangement of αketo β -lactams.⁹ As shown in Scheme 2, the [2 + 2]cycloaddition of benzyloxyketene, generated in situ from benzyloxyacetyl chloride and triethylamine, 10 with imines 8a and 8b and subsequent removal of the benzyloxy protective group afforded the α -hydroxy β -lactams **9a** and **9b** in 84 and 92% yield, respectively. Oxidation of both 9a and 9b using P₂O₅ in Me₂SO gave **10a** and **10b** which on Baeyer–Villiger rearrangement provided the respective NCAs 11a [81%, mp 168-170 °C, $[\alpha]_D^{25} = +31.8$ ° $(c = 1.0, CH_2Cl_2)]$ and **11b** [89%, mp 120 °C, $[\alpha]_D^{25} = -70.1$ ° $(c = 1.0, CH_2Cl_2)]$. Nevertheless, we found that the enantiopure NCAs 11a and 11b could directly be obtained in 95 and 96% yield respectively in a single pot operation from the corresponding α -hydroxy β lactams 9a and 9b using a phosphate buffer (pH = 6.9) of

a R = Bu^tPh₂Si; R¹ = Prⁱ **b** R = Bu^tMe₂Si; R¹ = Ph

Scheme 2 Reagents and conditions: i, BnOCH₂COCl (2 equiv.), NEt₃, CH₂Cl₂, $-78\,^{\circ}\text{C}$ → room temp.; 20 h then NH₄HCO₂, Pd/C, PriOH, reflux, 1 h, 70% overall for **9a**; and H₂, Pd/C, EtOH, room temp. 15 h, 78% overall for **9b**; ii, Me₂SO, P₂O₅, 20 h; iii, MCPBA, CH₂Cl₂, $-40\,^{\circ}\text{C}$, 1 h; iv, 1 mol dm⁻³ NaOCl, TEMPO(cat), NaHCO₃, KH₂PO₄-K₂HPO₄ (pH: 6.9), CH₂Cl₂

11a
$$\stackrel{i}{=}$$
 R_2HN . $\stackrel{i}{=}$ R_3HN $\stackrel{i}{=}$ R_3HN

Scheme 3 Reagents and conditions: i, S-leuOMe or S-leuOBn, CH₂Cl₂, room temp., 15 h; ii, H₂, Pd/C, EtOH, room temp., 15 h; iii, **11b**, DMF, NaN₃ (1 equiv.), room temp. 15 h; iv, H₂, Pd/C, (Boc)₂O (2 equiv.), EtOH, room temp.

commercial bleach and a catalytic amount of 2,2,6,6-tetramethylpiperidinyl-1-oxy (TEMPO),11 thus making this method inexpensive and practical for the large-scale production of NCAs.† Completion of the synthesis of the tripeptide B of lysobactin was accomplished as shown in Scheme 3. First, 11a was coupled with S-leucine methyl ester in methylene chloride as solvent to give the dipeptide 12 in 95% yield. Further hydrogenolysis of 12 with H₂ over 10% Pd on charcoal led to 13 in 85% yield. In a similar way the coupling reaction of 11a with S-leucine benzyl ester and subsequent N-debenzylation of 14 gave the dipeptide 15 in 90% yield $\{ [\alpha]_D^{25} = -29.0^{\circ} (c = 1.0,$ CH₂Cl₂). Surprisingly, the NCA 11b proved resistant to ring opening by dipeptide 13 but with the addition of sodium azide in DMF as solvent gave the tripeptide product 16 $\{ [\alpha]_D^{25} =$ -26.9° (c = 1.0, $\widetilde{CH_2Cl_2}$)} which was then converted into 17 under standard conditions.

This work was supported by the Ministry of Education of the Spanish Government (Project: SAF: 95/0749) and by Basque Country University (UPV: 170.215-EA 161-94). A Grant from Ministerio de Educacion y Ciencia (B.O.) is gratefully acknowledged.

Footnote

† To ensure the optical purity of these NCAs both 11a and 11b were opened by methanol under reflux followed by N-debenzylation and subsequent acylation of the resulting α -amino esters with Mosher acid chloride and triethylamine. In each case a single set of signals were obtained in their 1 H, 13 C and 19 F NMR spectra.

References

- J. O'Sullivan, J. E. McCullough, A. A. Tymiak, D. R. Kirsch, W. H. Trejo and P. A. Principe, *J. Antibiot.*, 1988, 41, 1740; T. Kato, H. Hinoo, Y. Tervi, J. Kikuchi and J. Shoji, *J. Antibiot.*, 1988, 41, 719; T. Kato, *J. Antibiot.*, 1989, 42, C-2; A. A. Tymiak, T. J. McCormick and S. E. Unger, *J. Org. Chem.*, 1989, 54, 1149.
- 2 D. M. Shlaes, J. Marino and M. R. Jacobs, Antimicrob. Agents chemoter., 1984, 25, 527; H. W. Horowitz, S. Handwerger, K. G. vanHorn and G. P. Wormser, Lancet, 1987, 2, 1329.
- 3 For some reviews see: R. M. Williams, Synthesis of Optically Active α-Amino Acids, Pergamon, Oxford, 1989; D. Seebach, R. Imwinkelried and T. Weber, in 'Modern Synthetic Methods', ed. R. Scheffold, vol. 4, Springer, Berlin, 1986, p. 165; U. Schöllkopf, Top. Curr. Chem., 1983, 109, 65; R. O. Duthaler, Tetrahedron, 1995, 50, 1539.
- 4 For representative examples, see: D. A. Evans, E. B. Sjogren, A. E. Weber and R. E. Conn, *Tetrahedron Lett.*, 1987, 28, 39; D. Seebach, E. Juaristi, D. D. Miller, C. Schickli and T. Weber, *Helv. Chim. Acta*, 1987, 70, 237; M. E. Jung and Y. H. Jung, *Tetrahedron Lett.*, 1989, 30, 6637; C. G. Caldwell and S. S. Bondy, *Synthesis*, 1990, 34; D. Blaser and D. Seebach, *Liebigs Ann. Chem.*, 1991, 1067; E. J. Corey, D. H. Lee and S. Choi, *Tetrahedron Lett.*, 1992, 33, 6735; T. Sunazuka, T. Nagamitsu, H. Tanaka, S. Omura, P. A. Spregeler and A. B. Smith, *Tetrahedron Lett.*, 1983, 34, 4447; J. S. Yadar, S. Chandrasekhar, Y. R. Reddy and A. V. R. Rao, *Tetrahedron*, 1995, 51, 2749.
- 5 C. Palomo, F. Cabré and J. M. Ontoria, Tetrahedron Lett., 1992, 33, 4819.
- 6 L. A. Carpino, El-Sayed M. E. Mansour and D. Sadat-Aalaee, J. Org. Chem., 1991, 56, 2611.
- 7 D. R. Kronenthal, C. Y. Han and M. K. Taylor, J. Org. Chem., 1982, 47, 2765.
- 8 J. E. Baldwin, R. M. Adlington, D. W. Gollins and C. J. Schofield, Tetrahedron, 1990, 46, 4733; C. Palomo, J. M. Aizpurua and C. Cuevas, J. Chem. Soc., Chem. Commun., 1994, 1957.
- C. Palomo, J. M. Aizpurua, I. Ganboa, F. Carreaux, C. Cuevas, E. Maneiro and J. M. Ontoria, J. Org. Chem., 1994, 59, 3123.
- 10 Y. Kobayashi, Y. Takemoto, T. Kamijo, H. Haraba, Y. Ito and S. Terashima, *Tetrahedron*, 1992, 48, 1853; C. Palomo, F. P. Cossío, J. M. Ontoria and J. M. Odriozola, *Tetrahedron Lett.*, 1991, 32, 3105.
- P. L. Anelli, F. Montarini and S. Amici, Org. Synth. Coll., 1993, 8, 367;
 M. R. Leanna, T. J. Sowin and H. E. Morton, Tetrahedron Lett., 1992, 33, 5029.

Received, 7th August 1995; Com. 5/05291B