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Direct Synthesis, Substitution, and Structure of 1-(2'-Deoxy-β-D-*erythro*pentofuranosyl)-4-pentafluorophenylpyrimidin-2*H*-one

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Abstract: Direct methods have been developed to access the title nucleoside 1 from 2'-deoxyuridine (dU). The C-4 pentafluorophenyl group of 1 is readily displaced by amine nucleophiles forming N-4 substituted cytosines in good to excellent yields.

Our interest in the development of antisense and antigene oligonucleotides has prompted an investigation of cytosine nucleosides modified at N-4 necessitating efficient syntheses of these adducts. The presence of N-methyl substituents attached at the amino groups of the bases usually has a destabilising effect on the base pairing of oligonucleotides¹. However, the observation that inclusion of aralkyl substituents² in hexose-DNA³ significantly enhances hybridisation warrants a closer study of pentose oligonucleotides containing these substituents. We chose to substitute the N-4 of cytosine initially since modification of this position for other purposes has already attracted attention. Binding of DNA by proteins has been studied using cytosine with a ¹⁵N label at C-4, for example⁴. Introduction of ¹⁵N and protected amino groups at C-4 can be achieved at the nucleoside level using C-4 azolyl derivatives of uridines⁵. Isotopic labelling and substitution at the C-4 position is also possible at the oligonucleotide level⁶. To date, successful formation of C-4 azolyl derivatives of deoxy nucleosides has required time consuming protection/deprotection strategies for the reactive 5'-OH and 3'-OH positions. Protecting groups have included; acetoyl, benzoyl, di-*t*-butylmethylsilyl, trityl, dimethoxytrityl (DMT) and phosphoramidite⁵⁻⁷. An additional complication is the requirement for protection of the 2'-OH during the formation of C-4 azolyl derivatives of ribose nucleosides which has been met by the use of the tetrahydropyranyl group⁵.

Our aim was to develop an efficient, direct method for the synthesis of N-4 modified cytosines requiring only *in situ* protection of the sugar hydroxyls. Thus, we have established two convenient one-pot syntheses which give direct access to the cytosine precursor 1 in good yield from 2'-deoxyuridine (dU). We chose the pentafluorophenyl (Pfp) group for subsequent displacement instead of the more familiar azolyl⁵. Transient protection of the reactive 5'-OH and 3'-OH groups was achieved through formation of either trifluoroactetate esters⁸ or trimethylsilyl (TMS) ethers⁹. The first one-pot transient protection method we investigated involved treatment of dU in pyridine with TMSCl (2.2 eq.) at room temperature (Scheme). After 30 minutes,



(CH₃CH₂)₃N. 4-chlorophenylphosphodichloridate (2.5 eq.) was added at 0°C and the mixture allowed to warm to room temperature. After 24 h, pentafluorophenol (PfpOH, 5 eq.) was added and the solution left to stir for 72 h. The product mixture was concentrated and the residual oil subjected to flash chromatography (EtOAc-MeOH 9:1) giving 1 in 65% yield¹⁰. A second, higher yielding synthesis of 1 was possible involving addition of

trifluoroacetic anhydride (4 eq.) to a solution of dU in pyridine at 0°C. The solution was stirred for 24 h at room temperature before PfpOH (10 eq.) was added¹¹. After a further 48 h the product solution was concentrated and the residual oil purified as before giving 1 in 78% yield. Recrystallization of 1 from a mixture of methanol and toluene gave crystals suitable for x-ray analysis. An ORTEP¹² plot of 1 is shown in the Figure and the relevant crystallographic data given in the Table.

Figure X-ray Crystallographic Structure of 1.

Table Crystallographic Data¹³ for 1.



Formula	C ₁₅ H ₁₁ N ₂ F ₅ O ₅
System	monoclinic
Space group	<i>P</i> 2 ₁
Cell parameters (Å)	
а	7.739(2)
b	5.502(1)
с	18.546(4)
β (°)	98.54(2)
Volume V (Å ³)	780.9(3)
Ζ	2

The bridging oxygen at O-4 is less conjugated with the Pfp ring than with the pyrimidine, which is oriented *anti* to the sugar ring with its C-2' *endo*, C-3' *exo* pucker and *gauche-gauche* 5'-OH group.¹⁴ The Pfp group in 1 can be readily displaced by amine nucleophiles¹⁵ to give the N-4 substituted 2'-deoxycytosines 2 to 4 (Scheme)¹⁶. Reaction of 1 with N-phenethylamine in dioxane at 80°C overnight gave 2 in 85% yield after purification by flash column chromatography. Under similar conditions 1 reacted with N-methyl-N-phenethylamine to give 3 in excellent yield (98%). The Pfp group of 1 was also displaced by the N-protected polyamine derivative; N1, N8-bis-Boc-N4-propylaminospermidine¹⁷ giving 4 but in more modest yield (58%). In order to make modified cytosine bases after oligomer synthesis, we have made available phosphoramidite 6. Dimethoxytritylation of 1 to give 5 (85%) followed by conversion to the phosphoramidite 6 (75%) was achieved using standard methods^{9,18}. Phosphoramidite 6 is fully compatible with automated solid-supported oligonucleotides containing 2 to 4 and post-synthetic substitution of oligonucleotides containing 1 are currently under investigation.

The one-pot transient protection procedure may be extended to the ribose series without interference from the 2'-OH. Uridine (U) was treated with trifluoroacetic anhydride and PfpOH as described and the ribose nucleoside 7 isolated in 59% yield. We have recently reported the crystal structure¹⁹ of a protected analogue of AZT; 1-(3'-azido-2',3'-dideoxy-5'-O-acetyl- β -D-*erythro*-pentofuranosyl-5-methyl-4-(1,2,4-1*H*triazolyl)-pyrimidin-2*H*-one (ATAZT), which has anti-HIV properties. In order to make unprotected C-4 azolyl analogues directly for comparison with ATAZT, we applied the one-pot transient protection methodology to **dU** and **U**. Thus, **dU** was treated with trifluoroacetic anhydride and with 1,2,4-1*H*-triazole in place of PfpOH giving 8 in good yield (79%). Similarly, the ribose analogue 9 was directly available from U (56%). The antiviral properties of a series of C-4 modified analogues are currently being evaluated and will be reported elsewhere.

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- 10. (a) Analytical data for 1: mp 176°C; TLC (EtOAc-MeOH) R_f 0.50; UV (95% EtOH) λ_{max} 286 nm (ϵ 6730); IR (KBr) v_{max} 3479, 3403, 3083, 2940, 1654, 1556, 1450, 1295 and 1114 cm⁻¹; ¹H NMR [(CD₃)₂SO] δ 2.07 (m, 1H, CH₂-2'), 2.29 (m, 1H, CH₂-2'), 3.62 (s, 2H, CH₂-5'), 3.88 (d, 1H, J = 3.4 Hz, CH-4') 4.23 (s, 1H, CH-3'), 5.14 (s, 1H, OH-5'), 5.28 (s, 1H, OH-3'), 6.06 (t, 1H, J = 6.1 Hz, CH-1'), 6.59 (d, 1H, J = 7.3 Hz, CH-5), 8.59 (d, 1H, J = 7.3 Hz, CH-6); ¹³C NMR [(CD₃)₂SO] δ 41.2 (CH₂-2'), 61.0 (CH₂-5'), 70.0 (CH-4'), 87.2 (CH-3'), 88.5 (CH-5), 93.0 (CH-1'), 148.0 (CH-6), 154.0 (C-4), 168.3 (C-2); ¹⁹F NMR [(CD₃)₂SO] δ -14.72 (t, 2F, J = 20.2 Hz, 2 x meta CF), -19.57 (t, 1F, J = 21.2 Hz, para CF), -21.12 (d, 2F, J = 24.2 Hz, 2 x ortho CF); Mass spectrum (FAB) m/z (I_r) 417 (M + Na⁺, 15%), 395 (M + H⁺, 22%), 305 (14%) and 279 (100%) [Found: m/z 395.067 (M + H⁺). C1₅H1₁F₅N₂O₅ requires 395.068]; Anal. Calcd for C1₅H1₁F₅N₂O₅: C, 45.69%; H, 2.79%; F, 24.11%; N, 7.11%. Found: C, 45.64%; H, 2.56%; F, 24.14%; N, 7.06%. (b) Positive chemical shifts are downfield of the TMS reference for ¹H and ¹³C NMR and negative chemical shifts are upfield of the CF₃CCl₃ reference for ¹⁹F NMR.
- 11. An NMR investigation showed that formation of the O-5' and O-3' trifluoroacetate esters occurred in minutes whereas formation of the intermediate C-4 pyridinium adduct was much slower. The product yield was lowered if addition of PfpOH was not delayed until approximately 24 h after initial reaction of **dU** with triflouroacetic anhydride.
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- 13. Enraf-Nonius CAD4 diffractometer, 2969 reflections measured, final R = 0.0361 for 2609 observed data $[I>2\sigma(I)]$, $wR(F^2) = 0.113$ for 2734 independent data.
- 14. C4-O4 = 1.365(2)Å, O4-O7 = 1.386(3)Å, N3-C4-O4-C7 = $-1.4(3)^{\circ}$, C4-O4-C7-C8 = $104.7(2)^{\circ}$, C2-N1-C1'-O4' = $-124.7(2)^{\circ}$; pseudorotation parameters P = 174.4° , t_m = 35.5° ; O4'-C4'-C5'-O5' = $-63.0(3)^{\circ}$, C3'-C4'-C5'-O5' = $55.9(3)^{\circ}$.
- 15. General method: To a solution of 1 (0.5 mmol) in 1,4-dioxane (10 cm⁻³), the appropriate amine (1 to 1.2 eq.) was added and the mixture stirred at 80°C overnight. The product solution was concentrated and subjected to flash column chromatography to give the pure product.
- 16. All reported compounds gave satisfactory TLC, ¹H NMR, ¹³C NMR, ¹⁹F NMR, mass spectral, and elemental analysis data, in full agreement with their assigned structures.
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