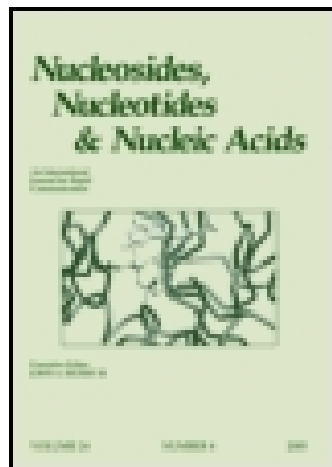


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## Nucleosides, Nucleotides and Nucleic Acids

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### Efficient Synthesis of Benzamide Riboside, a Potential Anticancer Agent

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## EFFICIENT SYNTHESIS OF BENZAMIDE RIBOSIDE, A POTENTIAL ANTICANCER AGENT

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□ *An efficient five step synthesis of benzamide riboside (BR) amenable for a large scale synthesis has been developed. It allows for extensive pre-clinical studies of BR as a potential anticancer agent.*

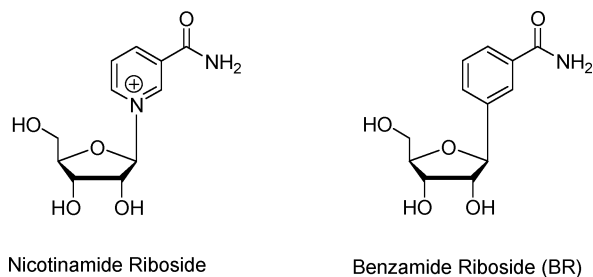
**Keywords** Benzamide riboside; anticancer agent

Benzamide riboside (BR) belongs to the C-nucleoside family of nucleosides, which has a ribofuranosyl moiety linked to the heterocyclic or aromatic base through a stable carbon-carbon bond rather than a carbon-nitrogen bond as in natural nucleosides<sup>[1–4]</sup> (Figure 1). BR exhibits potent antitumor activity in a variety of cultured human tumor cells.<sup>[1,2]</sup>

BR is a prodrug that is phosphorylated by adenosine kinase to the corresponding 5'-mononucleotide, which is converted to the active metabolite, benzamide adenine dinucleotide (BAD), an analog of nicotinamide adenine dinucleotide (NAD), by the action of nicotinamide mononucleotide adenylyltransferase (NMNAT) (Scheme 1).<sup>[1]</sup>

The primary target of BR is IMP-dehydrogenase (IMPDH), the rate-limiting enzyme in the purine nucleotide pathway of guanine nucleotide synthesis. Inhibition of IMPDH causes depletion of GTP and dGTP pools and results in inhibition of cancer cell proliferation.<sup>[1,5]</sup> An ideal anticancer drug is one which produces minimal toxic side effects and exhibits its anticancer properties through selective triggering of apoptosis in tumor cells. BR has been shown to induce apoptosis selectively in human ovarian

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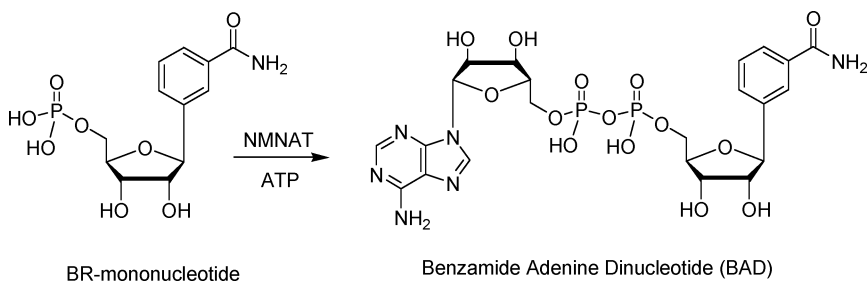


**FIGURE 1** Structure of nicotinamide riboside and benzamide riboside.

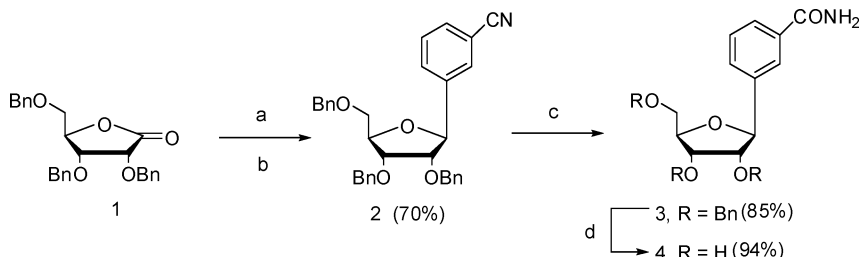
carcinoma-N-1 cells, lung cancer (H520) cells, and in leukemic cells (HL-60) in culture.<sup>[5–7]</sup>

The first synthesis of BR reported by Krohn and coworkers<sup>[8,9]</sup> is a ten-step procedure not amenable for large scale preparation. We report herein a new efficient synthesis of BR in five steps from a commercially available D-(+)-ribonic- $\gamma$ -lactone, protected as the 2,3,5-tri-O-benzyl lactone **1**<sup>[10]</sup> (Scheme 2). The key reaction is a stereo-specific coupling of the Grignard reagent<sup>[11]</sup> prepared from 3-iodobenzonitrile with **1** followed by reduction to give the  $\beta$  anomer **2** exclusively. The nitrile group is stable during coupling reaction and is easy to hydrolyze. Indeed, treatment of **2** with  $\text{Me}_3\text{SiOK}$  afforded the carboxyamido derivative **3**, which was deprotected with  $\text{BBr}_3$  to give the desired BR in 56% overall yield.

In Krohn's synthesis the protected ribonolactone was obtained from ribose by selective protection of the anomeric hydroxyl group followed by the further benzylation of the sugar OH groups. Then removal of the anomeric protective group and oxidation gave the protected ribonolactone (**1**). For coupling Krohn et al. used an oxazoline protected compound obtained in three steps from the benzoic acid. The choice of this protection offered stability during organometallic coupling reaction, but protection and deprotection adds seven steps to the process.



**SCHEME 1** Enzymatic synthesis of benzamide adenine dinucleotide.



Reagents: a) *i*PrMgCl, 3-iodobenzonitrile, THF; b) BF<sub>3</sub>OEt<sub>2</sub>, Et<sub>3</sub>SiH, CH<sub>2</sub>Cl<sub>2</sub>; c) Me<sub>3</sub>SiOK, THF; d) BBr<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>

SCHEME 2 Synthesis of benzamide riboside.

## EXPERIMENTAL

### General Methods

All commercial reagents (Sigma-Aldrich, Acros) were used as provided unless otherwise indicated. An anhydrous solvent dispensing system (J. C. Meyer) using two packed columns of neutral alumina was used for drying THF, Et<sub>2</sub>O, and CH<sub>2</sub>Cl<sub>2</sub>, while two packed columns of molecular sieves were used to dry DMF. Solvents were dispensed under argon. Nuclear magnetic resonance spectra were recorded on a Varian 600 MHz with Me<sub>4</sub>Si or DDS as the internal standard for <sup>1</sup>H. Chemical shifts are reported in ppm, and signals are described as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), bs (broad singlet), and dd (double doublet). Values given for coupling constants are first order. High resolution mass spectra were recorded on an Agilent TOF II TOF/MS instrument equipped with either ESI or APCI interface.

**3-(2,3,5-Tri-*O*-benzyl-1-β-D-ribofuranosyl)benzonitrile (2).** A solution of 3-iodobenzonitrile (3 g, 13.1 mmol) in dry THF (195 mL, 15 mL/mmol) under nitrogen was cooled to  $-78^{\circ}\text{C}$ , a solution of *i*PrMgCl (6.55 mL, 2 M in THF) was added, and the mixture was stirred for 1 hour at  $-78^{\circ}\text{C}$ . The mixture was transferred (via canule) into a solution of 2,3,5-tri-*O*-benzyl-D-ribo-1,4-lactone (**1**)<sup>[10]</sup> (5.47 g, 13.1 mmol) in dry THF (35 mL, 2.5 mL/mmol) under nitrogen cooled at  $-78^{\circ}\text{C}$  and stirred at  $-78^{\circ}\text{C}$  for 1 hour. It allowed to reach room temperature and then stirred for 4 hours to give a pink-red solution. A saturated solution of NaHCO<sub>3</sub> was added and the mixture was extracted with ether. The organic layer was separated, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was diluted with dry CH<sub>2</sub>Cl<sub>2</sub> (26 mL, 2 mL/mmol) under nitrogen, cooled to  $-78^{\circ}\text{C}$  and BF<sub>3</sub>Et<sub>2</sub>O (3.3 mL, 26.2 mmol) was slowly added (5 minutes) to the solution followed by Et<sub>3</sub>SiH (4.2 mL, 26.2 mmol). After 1 hour at  $-78^{\circ}\text{C}$  the solution was allowed to reach room temperature and then stirred overnight.

After addition of sat. NaHCO<sub>3</sub> the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer is separated, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated, and chromatographed on a silica gel column with CH<sub>2</sub>Cl<sub>2</sub> to give compound **2** (4.64 g, 70%) as an oil. <sup>1</sup>H NMR (δ, CDCl<sub>3</sub>, 600 MHz), 7.65 (s, 1H), 7.59 (d, *J* = 7.2 Hz, 1H), 7.51 (d, *J* = 7.8 Hz, 1H), 7.37–7.12 (m, 16H), 4.97 (d, *J* = 8.2 Hz, 1H), 4.61–4.51 (m, 16H), 4.36 (m, 1H), 4.00 (dd, *J* = 4.8, 3.0 Hz, 1H), 3.73 (dd, *J* = 7.8, 4.8 Hz, 1H), 3.65 (dd, *J* = 10.2, 4.2 Hz, 1H), 3.58 (dd, *J* = 10.2, 3.6 Hz, 1H). HRMS calcd for C<sub>33</sub>H<sub>30</sub>NO<sub>4</sub> [M-H]<sup>-</sup> 504.2180 found 504.2166.

**3-(2,3,5-Tri-*O*-benzyl-1-β-D-ribofuranosyl)benzamide (3).** To a solution of compound **2** (0.397 g, 0.78 mmol) in dry THF (2 mL, 2.5 mL/mmol, dry toluene can also be used) placed in a pressure tube Me<sub>3</sub>SiOK (200 mg, 1.56 mmol) was added in one portion and the mixture was refluxed for 24 hours. After reaching room temperature EtOH (20 mL) was added and the mixture was concentrated. The residue was purified on a silica gel column with CH<sub>2</sub>Cl<sub>2</sub>/MeOH(5%) to give compound **3** (350 mg, 85%) as a white solid. <sup>1</sup>H NMR (δ, CDCl<sub>3</sub>, 600 MHz), 7.78 (s, 1H), 7.75 (d, *J* = 7.8 Hz, 1H), 7.50 (d, *J* = 7.8 Hz, 1H), 7.33–7.17 (m, 16H), 5.95 (m, 2H), 5.05 (d, *J* = 6.0 Hz, 1H), 4.62–4.52 (m, 6H), 4.36 (m, 1H), 4.04 (t, *J* = 4.8 Hz, 1H), 3.82 (dd, *J* = 6.0, 5.4 Hz, 1H), 3.73 (dd, *J* = 10.8, 4.2 Hz, 1H), 3.64 (dd, *J* = 10.2, 4.2 Hz, 1H). HRMS calcd for C<sub>33</sub>H<sub>34</sub>NO<sub>5</sub> [M-H]<sup>+</sup> 524.2431 found 524.2483.

**3-(1-β-D-ribofuranosyl)benzamide (4, BR).** Compound **3** (2.15 g, 4.11 mmol) was diluted with dry CH<sub>2</sub>Cl<sub>2</sub> (85 mL, 20 mL/mmol) under nitrogen at -78°C and a 1 N solution of BBr<sub>3</sub> in CH<sub>2</sub>Cl<sub>2</sub> (16.5 mL) was added slowly (15 minutes). The reaction was stirred for 1 hour at -78°C and then overnight at room temperature. The reaction was quenched with a mixture of Et<sub>2</sub>O and MeOH (4/1, 200 mL), stirred for 20 minutes, and concentrated. The residue was chromatographed on a silica gel column with CH<sub>2</sub>Cl<sub>2</sub>-MeOH (25%), the fractions containing **4** were treated with charcoal and filtered through celite pad to give **4** (980 mg, 94%) as a white solid rather than an oil reported previously.<sup>[8]</sup> Spectral and biological properties of **4** were identical to those reported earlier.

## REFERENCES

1. Gharehbaghi, K.; Grunberger, W.; Jayaram, H.N. Studies on the mechanism of action of benzamide riboside: A novel inhibitor of IMP dehydrogenase *Curr. Med. Chem.* **2002**, *9*, 743–748.
2. Khanna, N.; Jayaram, H.N.; Singh, N. Benzamide riboside induced mitochondrial mediated apoptosis in human lung cancer H520 cells. *Life Sciences* **2004**, *75*, 179–190.
3. Kabat, M.; Pankiewicz, K.W.; Watanabe, K.A. Synthesis of 5-β-D-ribofuranosylnicotinamide and its N-methyl derivative. The isosteric and isoelectronic analogs of icotinamide nucleoside. *J. Med. Chem.* **1987**, *30*, 924–927.

4. Pankiewicz, K.W.; Watanabe, K.A.; Lesiak-Watanabe, K.; Goldstein, B.M.; Jayaram, H.N. The chemistry of nicotinamide adenine dinucleotide (NAD) analogues containing C-nucleosides related to nicotinamide riboside. *Curr. Med. Chem.* **2002**, *9*, 733–741.
5. Damaraju, V.L.; Visser, F.; Zhang, J.; Mowles, D.; Ng, A. M. L.; Young, J.D.; Jayaram, H.N.; Cass, C.E. Role of human nucleoside transporters in the cellular uptake of two inhibitors of IMP dehydrogenase, tiazofurin and benzamide riboside. *Mol. Pharmacol.* **2005**, *67*, 273–279.
6. Khanna, N.; Jayaram, H.N.; Singh, N. Benzamide riboside induced mitochondrial mediated apoptosis in human lung cancer H520 cells. *Life Sciences* **2004**, *75*, 179–190.
7. Grusch, M.; Polar, D.; Gfatter, S.; Leuhuber, K.; Huettnerbrenner, S.; Leisser, C.; Fuhrmann, G.; Steinkellner, S.; Smid, K.; Peters, G.J.; Jayaram, H.N.; Klepal, W.; Szekeres, T.; Knasmuller, S.; Krupitza, G. Maintenance of ATP favors apoptosis over necrosis triggered by benzamide riboside. *Cell Death Diff.* **2002**, *9*, 169–178.
8. Krohn, K.; Heins, H.; Wielckens, K. Synthesis and cytotoxic activity of C- glycosidic nicotinamide riboside analogs. *J. Med. Chem.* **1992**, *35*, 511–517.
9. Krohn, K.; Dorner, H.; Zukowski, M. Chemical synthesis of benzamide riboside. *Curr. Med. Chem.* **2002**, *9*, 727–731.
10. Jensen, H.S.; Limberg, Gerrit; W.; Pedersen, C. Benzylolation of aldonolactones with benzyl trichloroacetimidate. *Carbohydrate Res.* **1997**, *302*(1–2).
11. Varchi, G.; Kofink, C.; Lindsay, D.M.; Ricci, A.; Knochel, P. Direct preparation of polyfunctional amino-substituted arylmagnesium reagents via an iodine-magnesium exchange reaction. *Chem. Commun.* **2003**, *3*, 396–397.
12. Kevin, J. Potassium trimethylsilanolate mediated hydrolysis of nitriles to primary amides. *Tetrahedron Lett.* **2000**, *41*, 3747–3749.