Synthesis of 5-(Alkylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamides, Key Intermediates for the Synthesis of 3-Alkyl-9- β -D-ribofuranosylpurine Derivatives

Taisuke Itaya,* Tohru Saito, Tsunehiro Harada, Seiya Kagatani, and Tozo Fujii

Faculty of Pharmaceutical Sciences, Kanazawa University, Takara-machi, Kanazawa 920, Japan. Received April 13, 1989

An alternative synthesis of 2', 3', 5'-tri-O-benzoyl-N, N, 3-trimethyladenosine iodide (9a) was attained by the reaction of N, N, 3-trimethyladenine (11a) with 1-O-acetyl-2, 3, 5-tri-O-benzoyl- β -D-ribofuranose (10) in the presence of $SnCl_4$ followed by treatment with NaI. Although 3-benzyl-N, N-dimethyladenine (11c) did not react with 10 under similar conditions, the ribosylation of 3-ethyl-N, N-dimethyladenine (11b) followed by alkaline hydrolysis led to the first synthesis of 5-(ethylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamide (15b). A more general procedure for the synthesis of 5-(alkylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamides (15) was developed v a series of reactions: alkylation of N'-benzyloxy-5-formamido-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamidine (12) with alkyl halides in the presence of K_2CO_3 , catalytic hydrogenolysis, and alkaline hydrolysis. By means of this method, 5-(benzylamino)- (15c) and 5-(isopropylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamide (15d) were synthesized for the first time.

Keywords imidazole-4-carboxamide; imidazole nucleoside; ribosylation; formamido *N*-alkylation; *N*-alkoxyamidine hydrogenolysis; imidazolecarboxamidine; carboxamidine hydrolysis; (alkylamino)imidazole

Remarkable lability to acidic hydrolysis at the N-glycosidic bond is a characteristic of the tricyclic compounds 1,1) the putative structures for the fluorescent nucleosides isolated from unfractionated transfer ribonucleic acids (tRN-As) of extremely thermophilic archaebacteria²⁾ and yeast phenylalanine tRNAs.3) The structures 1 are unique in that they contain the 3-methyl-9-β-D-ribofuranosylpurine moiety. In order to establish the relationship between the structure and the rate of hydrolysis at the glycosidic bond, we have synthesized 3-methyladenosine (3), 3-methylguanosine (2a), $^{1b, e, 5)}$ 3-methylinosine (4), 3-methyl xanthosine (5), $^{7)}$ and 3-methylisoguanosine (6), $^{8)}$ as typical examples of 3-methyl-9-β-D-ribofuranosylpurines, and found that all these nucleosides undergo unusually fast cleavage at the glycosidic bond under acidic conditions. One plausible explanation for the high sensitivity is that this is a case of steric assistance of the methyl group situated at the peri position. ^{1d} Taking this as a working hypothesis, we

> > $R = \beta - \mathbf{p}$ -ribofuranosyl

attempted to obtain higher 3-alkyl homologues of these nucleosides, none of which had been synthesized. Since 2a, 4, 5, and 6 have been synthesized from 5-(methylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamide (15a), the title compounds 15 should be good intermediates for the syntheses of our targets. This paper presents a detailed account of a general synthesis of 15.

Compound 15a and its 2',3'-O-isopropylidene derivative have been synthesized by reductive methylation of appropriate imidazole nucleosides. 1a, 10) Alternatively, we have obtained 15a by methylation of 2',3',5'-tri-O-benzoyl-N,Ndimethyladenosine (8) followed by alkaline hydrolysis. 64,11) In the present study, however, ethylation of 8 with EtI or benzylation with PhCH₂Br did not afford the corresponding quaternary salt [9b or 9c (Br for I)]. We have shown that alkylation of either N, N, 9- (type 8, alkyl for the tri-Obenzoylribofuranosyl group) or N,N,3-trialkyladenines (type 11) gives N, N, 3, 9-tetraalkyladeninium salts (type 9, alkyl for the tri-O-benzoylribofuranosyl group) and that the latter reaction proceeds faster than the former. 12) This knowledge encouraged us to try the condensation of N,N,3trimethyladenine (11a)¹³⁾ with 1-O-acetyl-2,3,5-tri-O-ben-zoyl- β -D-ribofuranose (10).¹⁴⁾ Treatment of 11a with 10 in (CH₂Cl)₂ at room temperature in the presence of SnCl₄¹⁵⁾ followed by anion exchange afforded the desired 9a11) in 35% yield. Similar treatment of 3-ethyl-N,N-dimethyladenine (11b)¹³⁾ followed by alkaline hydrolysis¹¹⁾ gave 15b in 12% overall yield. 16) The structure 15b was assignable by comparison of the ultraviolet (UV) and the nuclear magnetic resonance (NMR) spectra with those of 15a. 3-Benzyl-N,N-dimethyladenine (11c), however, hardly reacted with 10.

The third access to **15** was achieved by utilization of N'-benzyloxy-5-formamido-1- β -D-ribofuranosyl-1H-imidaz-ole-4-carboxamidine (**12**),¹⁷⁾ the stable intermediate of the Dimroth rearrangement of 1-benzyloxyadenosine (7). We have already established the synthesis of 1-alkyl-5-(alkyl-amino)-1H-imidazole-4-carboxamides (type **15**, alkyl for the ribofuranosyl group) by alkaline hydrolysis of 1-alkyl-5-(N-alkylformamido)-1H-imidazole-4-carboxamidines (type **14**, alkyl for the ribofuranosyl group), which are obtainable by alkylation of N'-alkoxy-1-alkyl-5-form-

amido-1H-imidazole-4-carboxamidines (type 12, alkyl for the ribofuranosyl group) followed by catalytic hydrogenolysis. 18) We have also reported the synthesis of 14a by methylation of 12 with MeI in the presence of K₂CO₃ followed by catalytic hydrogenolysis using Raney Ni in the presence of 1 molar equivalent of TsOH. 4) Thus, hydrolysis of 14a should lead to a new synthesis of 15a. In fact, when heated in 1 N aqueous NaOH under reflux, 14a gave 15a in 46% overall yield based on 12. Compound 12 was similarly treated with EtI in HCONMe2 in the presence of anhydrous K₂CO₃ at room temperature, furnishing 13b in 79% yield. Removal of the N'-benzyloxy group from 13b and successive hydrolysis of the resulting 14b were conducted in a manner similar to that described above for the preparation of 15a to afford 15b in 62% yield. The reactions of 12 with PhCH₂Br and with Me₂CHI under similar conditions gave the corresponding N-alkylformamido derivatives 13c, d in 66% and 26% yields, respectively. The use of 18-crown-6 in the latter reaction improved the yield of 13d to 61%. Compounds 13c, d were also transformed into 15c, d through 14c, d in 50% and 53% yields, respectively.

In the present study, we also improved the procedure for the preparation of the key intermediate 12, which had been obtained in 79% yield by hydrolysis of 1-benzyloxyadenosine perchlorate (7·HClO₄) in 0.5 m NaHCO₃–Na₂CO₃ buffer of pH 9.5 at 39—41 °C for 4h. ^{17a}) The reaction mixture had been concentrated under reduced pressure at the initial stage of work-up. Since this process is accompanied with a pH shift to the higher region owing to partial decomposition of the buffer component NaHCO₃ to Na₂CO₃, it promotes the ring closure of 12 to N-benzyloxyadenosine, ^{17a}) causing a severe reduction in yield in the case of a large-scale preparation. When a solution of 7·HClO₄ in plain water was adjusted to pH 9.5 with aqueous NaOH and kept at 40 °C for 4h, we found that the pH changed to a much lesser extent throughout the reaction than we had

expected. Such a small change in pH may be interpreted as a beneficial consequence of the buffering action of the $HClO_4$ –7 and 12–NaOH system, since 7 (basic p K_a 7.90¹⁹) and 12 (acidic p K_a 9.93¹⁹) have their p K_a 's near 9.5. Accordingly, the reaction mixture can be easily maintained at pH 9.5, without resort to the carbonate buffer, by occasional supply of a small amount of aqueous NaOH. This procedure improved the yield of 12 to 95%.

In conclusion, among the above three synthetic routes to 15, the third route $12 \rightarrow 13 \rightarrow 14 \rightarrow 15$ has proved to be the most general. Since we have already accomplished the synthesis of 2b - d from 15b - d, $^{1d,20)}$ syntheses of 4-alkyl homologues of 1a - c should be attainable according to our own synthetic routes to 1a - c from 2a. The present synthesis of 15b - d should also permit the syntheses of 3-alkyl homologues of 4 - 6. Quite recently, syntheses of azepinomycin (16), an antitumor antibiotic from Streptomyces species, have been reported by Issiki et al. 11 and Fujii et al. 12 The latter group utilized the third of the methods for the synthesis of the key intermediate 15e.

Experimental

General Notes All melting points were taken on a Yamato MP-1 capillary melting point apparatus and are corrected. Spectra reported herein were recorded on a Hitachi 320 UV spectrophotometer using solutions in 95% aqueous EtOH, 0.1 N aqueous HCl (pH 1), 0.005 M phosphate buffer (pH 7), and 0.1 N aqueous NaOH (pH 13), and a JEOL

JNM-FX-100 NMR spectrometer at 25 °C using Me₄Si as an internal standard. Optical rotations were measured with a JASCO DIP-181 polarimeter. Elemental analyses were performed by Mr. Y. Itatani and his associates at Kanazawa University. The following abbreviations are used: br=broad, d=doublet, dq=doublet-of-quartets, m=multiplet, s=singlet, sh=shoulder, t=triplet.

2',3',5'-Tri-O-benzoyl-N,N,3-trimethyladenosine Iodide (9a) Anhydrous SnCl₄ (0.15 ml, 1.3 mmol) was added to a solution of $11a^{13}$ (177 mg, 1.0 mmol) and 10^{14} (504 mg, 1.0 mmol) in $(CH_2Cl)_2$ (5 ml). The mixture was stirred at room temperature for 4 h. The resulting clear solution was washed successively with H_2O (2×10 ml), saturated aqueous NaHCO₃ (2×10 ml), and H_2O (3×20 ml), dried over Na₂SO₄, and concentrated in vacuo to leave a colorless foam. This was dissolved in EtOH (4.5 ml) and a solution of NaI (300 mg, 2 mmol) in EtOH (0.7 ml) was added. The resulting precipitate was collected by filtration, washed successively with H_2O (0.5 ml) and EtOH (0.5 ml), and dried to give 9a (264 mg, $35\%_o$), mp 186-189 °C (dec.). Recrystallization from MeOH gave colorless prisms, mp 189-190 °C (dec.). This sample was identical [by mixture melting point test and comparison of the infrared (IR) spectrum] with an authentic specimen. 111

N'-Benzyloxy-5-formamido-1-β-D-ribofuranosyl-1H-imidazole-4-carboxamidine (12) A suspension of 1-benzyloxyadenosine perchlorate monohydrate (7·HClO₄·H₂O)²³⁾ (35.98 g, 74.2 mmol) in H₂O (1400 ml) was adjusted to pH 9.5 with 10% aqueous NaOH at 40 °C. The mixture was stirred at that temperature for 4h, being kept at pH 9.5 by occasional addition of 10% aqueous NaOH. The resulting solution was chilled, neutralized with 10% aqueous HCl, and concentrated *in vacuo* to a small volume. The precipitate that separated was collected by filtration, washed with cold H₂O (60 ml), and dried to give 12 (27.26 g, 95%), mp 156—160 °C (dec.) [lit. 17a) mp 158—160 °C (dec.)], identical [by mixture melting point test and comparison of the IR spectrum] with an authentic sample. 17a)

N'-Benzyloxy-5-(N-ethylformamido)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamidine (13b) A mixture of 12 (783 mg, 2.0 mmol), anhydrous K₂CO₃ (415 mg, 3.0 mmol), and HCONMe₂ (12 ml) was stirred at room temperature for 1 h. Ethyl iodide (940 mg, 6 mmol) in HCONMe₂ (1 ml) was added and stirring was continued for a further 24 h. The mixture was concentrated in vacuo and the residue was dissolved in H₂O (10 ml). Saturated aqueous NaCl (10 ml) was added to the solution and the resulting precipitate was filtered off, washed with a little H2O, and dried to give a colorless solid (665 mg, 79%), mp 146—148 °C. Recrystallization from AcOEt afforded an analytical sample as colorless prisms, mp 147-148 °C, $[\alpha]_D^{20}$ – 44.6° (c = 0.999, MeOH); UV $\lambda_{\max}^{95\%}$ EiOH 250 nm (sh) (ε 6500); $\lambda_{\max}^{H_2O}$ (pH 1) 246 (sh) (8400); $\lambda_{\max}^{H_2O}$ (pH 7) 250 (sh) (6100); $\lambda_{\max}^{H_2O}$ (pH 13) 250 (sh) (6000); ${}^1\text{H}\text{-NMR}$ [(CD₃)₂SO] δ : 24) 0.87 (3H, t, J = 7 Hz, Me), 3.54 [4H, m, $MeC\underline{H}_2$, $C(5')-\underline{H}_2$], 3.89 [1H, m, $C(4')-\underline{H}$], 4.05 [1H, m, $C(3')-\underline{H}$], 4.32 [1H, m, C(2')-H], 4.85 with a shoulder at 4.89 (2H, s, $PhC\underline{H}_2$), 5.02 (1H, t, J=5 Hz, 5'-OH), 5.21 (1H, d, J=5 Hz, 3'-OH), 5.24 [1H, d, J=6 Hz, C(1')-H], 5.47 (1H, d, J=6 Hz, 2'-OH), 5.78 with a small peak at 5.70 (2H, br, NH₂), 7.33 (5H, m, Ph), 7.94 with a small peak at 8.22 (1H, s, CHO), 25) 8.09 with two small peaks at 8.03 and 8.06 [1H, s, C(2)-H].²⁵⁾ Anal. Calcd for $C_{19}H_{25}N_5O_6$: C, 54.41; H, 6.01; N, 16.70. Found: C, 54.12; H, 5.96; N, 16.70.

N'-Benzyloxy-5-(N-benzylformamido)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamidine (13c) A mixture of 12 (587 mg, 1.5 mmol), anhydrous K₂CO₃ (311 mg, 2.2 mmol), and HCONMe₂ (9 ml) was stirred at room temperature for 1 h, and then PhCH₂Br (260 mg, 1.5 mmol) in HCONMe₂ (1 ml) was added. After stirring had been continued for 30 h, the reaction mixture was concentrated in vacuo. The residue was mixed with H₂O (1 ml) and saturated aqueous NaCl (4 ml). The mixture was extracted with AcOEt (2×10 ml). The combined extracts were washed with saturated aqueous NaCl (2 ml), dried over MgSO₄, and concentrated in vacuo. The residue was purified on a silica gel (20 g) column [CHCl₃-EtOH (6:1, v/v)] to give **13c** (480 mg, 66%) as a colorless foam, UV $\lambda_{\text{max}}^{95\%}$ ErOH 250 nm (sh) (ϵ 6800); $\lambda_{\text{max}}^{\text{H}_2O}$ (pH 1) 246 (sh) (7900); $\lambda_{\text{max}}^{\text{H}_2O}$ (pH 7) 250 (sh) (6000); $\lambda_{\text{max}}^{\text{H}_2\text{O}}$ (pH 13) 250 (sh) (6000); ¹H-NMR [(CD₃)₂SO] δ :²⁴⁾ 3.53 [2H, m, C(5')-H₂], 3.78 [1H, m, C(4')-H], 3.85—4.25 [2H, m, C(3')-H and C(2')-H], 4.40—4.75 (2H, m, NCH_2Ph), 4.88 (2H, s, OCH_2Ph), 4.90— 5.20 [3H, m, C(1')-H, 3'-OH, and 5'-OH], 5.35 (1H, br, 2'-OH), 5.66 (2H, br, NH₂), 7.15 (5H, m, NCH₂Ph), 7.34 (5H, m, OCH₂Ph), 8.12 with two small peaks at 8.32 and 8.42 (1H, s, CHO), 25) 7.97 with a small peak at 8.00 $[1H,\,s,\,C(2)\text{-}H].^{25)}$

N'-Benzyloxy-5-(N-isopropylformamido)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamidine (13d) A mixture of 12 (5.48 g, 14 mmol), anhydrous K_2CO_3 (1.93 g, 14 mmol), and $HCONMe_2$ (70 ml) was stirred at

room temperature for 1 h, then 18-crown-6 (2.04 g, 7.7 mmol) was added. Stirring was continued for a further 0.5h and then Me₂CHI (2.62 g, 15.4 mmol) was added. The whole was stirred at 30°C for 48 h and concentrated in vacuo. The residue was mixed with ice-water (50 ml) and saturated aqueous NaCl (50 ml). The mixture was extracted with CHCl₃ (3×60 ml) and the combined organic layers were dried over MgSO₄ and then concentrated in vacuo to leave a brown oil. The residue was washed with Et₂O (3×20 ml) and purified on a silica gel (200 g) column [CHCl₃-MeOH (5:1, v/v)] to give 13d (3.70 g, 61%) as a yellowish foam, UV $\frac{256 \text{ nm (sh)}}{3} (\epsilon 6000); \lambda_{\text{max}}^{\text{H}_2\text{O}} (\text{pH 1}) 246 (\text{sh}) (7300); \lambda_{\text{max}}^{\text{H}_2\text{O}} (\text{pH 7}) 250$ (sh) (5600); $\lambda_{\text{max}}^{\text{H}_2\text{O}}$ (pH 13) 247 (sh) (5800); ¹H-NMR [(CD₃)₂SO] δ :²⁴⁾ 0.87 and 0.93 (a total of 3H, d each, J = 6.5 Hz, Me), 1.11 and 1.16 (a total of 3H, d each, J = 6.5 Hz, Me), 3.54 [2H, m, C(5')-H₂], 3.88 [1H, m, C(4')-H], 3.94-4.54 [3H, m, C(3')-H, C(2')-H, and Me₂CH], 4.85 with a shoulder at 4.89 (2H, s, PhC \underline{H}_2), 5.04 (1H, t, J=6 Hz, 5'-OH), 5.18 (1H, d, J=4 Hz, 3'-OH), 5.27 and 5.30 [a total of 1H, d each, J=6.5 Hz, C(1')-H], 5.45 and 5.47 (a total of 1H, d each, J = 6.5 Hz, 2'-OH), 5.78 with a shoulder at 5.64 (2H, br, NH₂), 7.32 (5H, m, Ph), 7.88 and 7.90 with a small peak at 8.37 (a total of 1H, s each, CHO), 25) 8.10 with a small peak at 8.07 [1H, s, C(2)-H]. 25)

5-(Methylamino)-1-β-D-ribofuranosyl-1*H*-imidazole-4-carboxamide (15a) Compound 14a was prepared from 13a (405 mg, 1 mmol) according to the reported procedure^{4b}) and dissolved in 1 N aqueous NaOH (10 ml). The solution was refluxed for 30 min and applied to a column of Dowex 50W-X8 (H⁺) (10 ml). The column was washed with H₂O (50 ml). The product was eluted with 5% aqueous NH₃ and the eluate (550 ml) was concentrated *in vacuo*. The residue was washed with EtOH (3 ml) to give a chromatographically pure sample of 15a (146 mg, 54%), mp 168—170 °C. Recrystallization from EtOH afforded almost colorless prisms, mp 179—181 °C (lit.¹¹¹ mp 182—184 °C). This sample was identical (by comparison of the IR spectrum and paper chromatographic behavior) with an authentic specimen.¹¹²

5-(Ethylamino)-1-β-D-ribofuranosyl-1*H*-imidazole-4-carboxamide (15b) i) From 11b: The reaction of 11b¹³ (382 mg, 2.0 mmol) and 10¹⁴) was carried out in a manner similar to that described above for the ribosylation of 11a to give a crude product as a colorless foam. This was dissolved in EtOH (5 ml) and a solution of NaI (600 mg, 4 mmol) in EtOH (1.6 ml) was added. The resulting mixture was concentrated in vacuo and the residue was washed with H_2O (3 × 20 ml) and dried to give a colorless solid (812 mg), mp 102—129 °C (dec.). A part (500 mg) of this sample was refluxed in a mixture of 2N aqueous NaOH (6 ml) and EtOH (3 ml) for 2h, concentrated in vacuo to half the initial volume, and then cooled. Ion exchange resin [Bio-Rad AG 50W-X8 (H+)] (5 ml) and EtOH (3 ml) was added to the solution. The resulting mixture was put on a column packed with more resin (3 ml). Then, the column was washed successively with H₂O-EtOH (2:1, v/v) (40 ml) and H₂O (20 ml). The column was eluted with cold 5% aqueous NH₃ (6 ml) and then with concentrated aqueous NH₃ (60 ml). The combined ammoniac eluates were concentrated in vacuo to leave a caramel (150 mg). This was purified on a silica gel (5 g) column [CHCl3-MeOH (5:1, v/v) to give 15b $(43 \text{ mg}, 12\%)^{16}$ as a colorless glass. Identity of this sample with that described below was confirmed by 1H-NMR spectroscopy.

ii) From 13b: A solution of 13b (4.20 g, 1.0 mmol) and p-toluenesulfonic acid monohydrate (1.90 g, 1.0 mmol) in H₂O (250 ml) was hydrogenated over Raney Ni (7 ml) at room temperature and atmospheric pressure for 2h. The catalyst was filtered off and washed with H₂O (90 ml). The combined filtrate and washings were concentrated in vacuo to give 14b as an oily residue. This was dissolved in 1 N aqueous NaOH (100 ml) and the solution was refluxed for 10 min and then put on a column of Dowex 50W-X8 (H⁺) (100 ml). The column was washed with H₂O (500 ml) then eluted successively with 5% aqueous NH₃ (350 ml) and concentrated aqueous NH₃ (1200 ml). The combined ammoniac eluates were concentrated in vacuo and the residue was purified on a silica gel (96 g) column [CHCl₃-MeOH (5:1, v/v)] to give 15b (1.79 g, 62%) as a colorless glass, UV $_{\text{max}}^{95\%}$ EtOH 266 nm (ε 9200); $\lambda_{\text{max}}^{\text{H}_2\text{O}}$ (pH 1) 257 (6500); $\lambda_{\text{max}}^{\text{H}_2\text{O}}$ (pH 7) 263 (7500); $\lambda_{\text{max}}^{\text{Ha2O}}$ (pH 13) 265 (7500); ¹H-NMR [(CD₃)₂SO] δ : 1.08 (3H, t, J=7 Hz, $MeCH_2$), 3.15 (2H, dq, J=7 Hz each, $MeC\underline{H}_2$), 3.56 [2H, br, $C(5')-H_2$], 3.88 [1H, m, C(4')-H], 4.05 [1H, m, C(3')-H], 4.31 [1H, m, C(2')-H], 5.03 (1H, t, J=5 Hz, 5'-OH), 5.18 (1H, d, J=5 Hz, 3'-OH), 5.44 [d, J=6 Hz, overlapped with signals due to 2'-OH and NH, C(1')-H], 6.89 and 7.03 (1H each, br, NH₂), 7.60 [1H, s, C(2)-H].

5-(Benzylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamide (15c) A solution of 13c (3.31 g, 6.87 mmol) and p-toluenesulfonic acid monohydrate (1.31 g, 6.89 mmol) in a mixture of H_2O (140 ml) and EtOH (40 ml) was hydrogenated over Raney Ni (5 ml) at room temperature and atmospheric pressure for 6 h. The mixture was worked up in a manner

similar to that described above for the hydrogenolysis of **13b**. The product **14c** was hydrolyzed by refluxing in 1 N aqueous NaOH (80 ml) for 30 min. The resulting solution was neutralized with concentrated hydrochloric acid, concentrated *in vacuo* to *ca*. 60 ml, and extracted with AcOEt (10 × 100 ml). The combined organic layers were dried over MgSO₄ and concentrated *in vacuo* to leave **15c** (1.19 g, 50% yield) as a colorless solid, mp 149—150 °C. Recrystallization from EtOH gave an analytical sample as colorless needles, mp 153—154 °C; [α] $_D^{60}$ –47.7° (c=1.00, MeOH); UV $\lambda_{\max}^{95\%}$ EtOH 266 nm (ϵ 9200); λ_{\max}^{HsO} (pH 1) 258 (7500); λ_{\max}^{HsO} (pH 7) 265 (8000); λ_{\max}^{HsO} (pH 13) 268 (8000); ¹H-NMR [(CD₃)₂SO] δ : 3.58 [2H, br, C(5')-H₂], 3.90 [1H, m, C(4')-H], 4.06 [1H, m, C(3')-H], 4.31 [1H, m, C(2')-H], 4.36 (2H, br, PhCH₂), 5.06, 5.17, and 5.45 (1H each, br, 5'-, 3'-, and 2'-OH), 5.54 [1H, d, J=6 Hz, C(1')-H], 6.12 (1H, br, NH), 6.88 and 7.02 (1H each, br, NH₂), 7.31 (5H, m, Ph), 7.60 [1H, s, C(2)-H]. *Anal.* Calcd for C₁₆H₂₀N₄O₅: C, 55.16; H, 5.79; N, 16.08. Found: C, 55.19; H, 5.67; N, 16.20.

5-(Isopropylamino)-1- β -D-ribofuranosyl-1H-imidazole-4-carboxamide (15d) Compound 13d (1.73 g, 3.99 mmol) was hydrogenated over Raney Ni and worked up in a manner similar to that described above for 15b. The crude 15d thus obtained was crystallized by treating it with EtOH (3 ml) to give colorless pillars (573 mg), mp 162-164°C. An additional crop (67 mg; the total yield was 53%) was obtained from the mother liquor by silica gel (10 g) column chromatography [CHCl₃-MeOH (5:1, v/v)]. Recrystallization from EtOH gave an analytical sample as colorless pillars, mp 163—165 °C; [α]_D²¹ – 50.6° (c =0.497, MeOH); UV $\lambda_{max}^{95\%}$ EiOH 266 nm (ϵ 9300); $\lambda_{max}^{H_{2O}}$ (pH 1) 257 (6800); $\lambda_{max}^{H_{2O}}$ (pH 7) 264 (8700); $\lambda_{max}^{H_{2O}}$ (pH 13) 265 (8800); ¹H-NMR [(CD₃)₂SO] δ : 1.06 and 1.08 (3H each, d, J=6 Hz, Me₂), 3.44 (1H, m, Me₂CH), 3.55 (2H, m, CH₂), 3.86 [1H, m, C(4')-H], 4.04 [1H, m, C(3')-H], 4.30 [1H, m, C(2')-H], 4.98 (1H, t, J = 5 Hz, 5'-OH), 5.17 (1H, d, J=5 Hz, 3'-OH), 5.39 [1H, d, J=6 Hz, overlapped with signals due to 2'-OH and NH, C(1')-H], 6.86 and 7.02 (1H each, br, NH₂), 7.62 [1H, s, C(2)-H]. Anal. Calcd for $C_{12}H_{20}N_4O_5$: C, 47.99; H, 6.71; N, 18.66. Found: C, 47.82; H, 6.99; N, 18.61.

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References and Notes

- a) S. Nakatsuka, T. Ohgi, and T. Goto, Tetrahedron Lett., 1978, 2579; b) T. Itaya, T. Watanabe, and H. Matsumoto, J. Chem. Soc., Chem. Commun., 1980, 1158; c) B. Golankiewicz and W. Folkman, Nucleic Acids Res., 11, 5243 (1983); d) T. Itaya and T. Harada, J. Chem. Soc., Chem. Commun., 1984, 858; e) T. Itaya, H. Matsumoto, T. Watanabe, and T. Harada, Chem. Pharm. Bull., 33, 2339 (1985); f) B. Golankiewicz, E. Zielonacka-Lis, and W. Folkman, Nucleic Acids Res., 13, 2443 (1985); g) H. Bazin, X-X. Zhou, C. Glemarec, and J. Chattopadhyaya, Tetrahedron Lett., 28, 3275 (1987); h) T. Itaya, Chem. Pharm. Bull., 35, 4372 (1987); i) C. Glemarec, J-C. Wu, G. Remaud, H. Bazin, M. Oivanen, H. Lönnberg, and J. Chattopadhyaya, Tetrahedron, 44, 1273 (1988); j) T. Itaya, M. Shimomichi, and M. Ozasa, Tetrahedron Lett., 29, 4129 (1988).
- 2) J. A. McCloskey, P. F. Crain, C. G. Edmonds, R. Gupta, T. Hashizume, D. W. Phillipson, and K. O. Stetter, *Nucleic Acids Res.*,

- 15, 683 (1987).
- a) S. Takemura, H. Kasai, and M. Goto, J. Biochem., 75, 1169 (1974); b) U. L. RajBhandary, R. D. Faulkner, and A. Stuart, J. Biol. Chem., 243, 575 (1968); c) R. Thiebe and H. G. Zachau, Eur. J. Biochem., 5, 546 (1968); d) S. H. Blobstein, R. Gebert, D. Grunberger, K. Nakanishi, and I. B. Weinstein, Arch. Biochem. Biophys., 167, 668 (1975).
- a) T. Saito and T. Fujii, J. Chem. Soc., Chem. Commun., 1979, 135; b)
 T. Fujii, T. Saito, and T. Nakasaka, Chem. Pharm. Bull., 37, 2601 (1989).
- 5) T. Itaya and K. Ogawa, Tetrahedron Lett., 1978, 2907.
- 6) a) T. Itaya and H. Matsumoto, Tetrahedron Lett., 1978, 4047; b) Idem, Chem. Pharm. Bull., 33, 2213 (1985).
- a) T. Itaya and T. Harada, Heterocycles, 19, 687 (1982); b) Idem, Chem. Pharm. Bull., 37, 1235 (1989).
- 8) T. Itaya and T. Harada, Tetrahedron Lett., 23, 2203 (1982).
- A preliminary form of this report has been published: T. Itaya, T. Saito, T. Harada, S. Kagatani, and T. Fujii, *Heterocycles*, 19, 1059 (1982).
- P. K. Bridson and C. B. Reese, Bioorg. Chem., 8, 339 (1979).
- T. Itaya, H. Matsumoto, and T. Watanabe, Chem. Pharm. Bull., 30, 86 (1982).
- T. Itaya, K. Ogawa, H. Matsumoto, and T. Watanabe, Chem. Pharm. Bull., 28, 2522 (1980).
- T. Itaya, H. Matsumoto, and K. Ogawa, Chem. Pharm. Bull., 28, 1920 (1980).
- E. F. Recondo and H. Rinderknecht, Helv. Chim. Acta, 42, 1171 (1959).
- 15) U. Niedballa and H. Vorbrüggen, J. Org. Chem., 39, 3654 (1974).
- 6) We previously reported the yield $(30\%)^{9}$ erroneously.
- 17) a) T. Fujii, C. C. Wu, T. Itaya, S. Moro, and T. Saito, Chem. Pharm. Bull., 21, 1676 (1973); b) J. A. Montgomery and H. J. Thomas, "Nucleic Acid Chemistry," ed. by L. B. Townsend and R. S. Tipson, John Wiley and Sons, New York, 1978, p. 683.
- a) T. Fujii, T. Saito, and M. Kawanishi, Tetrahedron Lett., 1978, 5007;
 b) T. Fujii, T. Saito, and T. Nakasaka, Heterocycles, 15, 195 (1981);
 c) T. Fujii, T. Itaya, T. Saito, K. Mohri, M. Kawanishi, and T. Nakasaka, Chem. Pharm. Bull., 37, 1504 (1989).
- T. Itaya, T. Saito, S. Kawakatsu, and T. Fujii, Chem. Pharm. Bull., 23, 2643 (1975).
- 20) T. Itaya and T. Harada, unpublished work.
- K. Isshiki, Y. Takahashi, H. Iinuma, H. Naganawa, Y. Umezawa, T. Takeuchi, H. Umezawa, S. Nishimura, N. Okada, and K. Tatsuta, J. Antibiot., 40, 1461 (1987).
- 22) T. Fujii, T. Saito, and T. Fujisawa, Heterocycles, 27, 1163 (1988).
- 23) T. Fujii, C. C. Wu, and T. Itaya, Chem. Pharm. Bull., 19, 1368 (1971).
- 24) The observed complexity of the signals is probably due to *cis-trans* isomerism of the *N*-substituted formamido group (ref. 18c and references cited therein) and diastereoisomerism produced by a combination of atropisomerism caused by restricted rotation about the imidazole-to-nitrogen bond and chirality of the ribofuranosyl group (refs. 4b and 7b and references cited therein).
- 25) Selective deuteration at the 2-position (treatment with boiling D_2O for $4\,h)^{18c)}$ permitted the assignment.