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Identification of novel allosteric nonpeptidergic inhibitors of the human cytomegalovirus-encoded chemokine receptor US28

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1. Introduction

Human cytomegalovirus (HCMV) is an infectious β-herpesvirus that establishes a lifelong latent infection in the majority of the human population and (ab)uses human cells for survival and propagation. Occasional reactivation and shedding of new infectious virions allow viral dissemination to other individuals. In order to establish viral latency and widespread propagation, HCMV has to hijack infected host cells and evade anti-viral immune responses. To this end, HCMV encodes important regulatory proteins that are expressed by infected host cells. One of these virally-encoded genes is the G protein-coupled receptor US28. US28 binds a broad variety of inflammatory chemokines allowing cell migration along chemokine gradients. In addition, recruitment of leukocytes to the microenvironment of HCMV-infected cells is impaired as inflammatory chemokines are depleted by constitutive and rapid US28mediated internalization.¹⁻³ Although latent infection is generally asymptomatic, HCMV has been associated with several chronic inflammatory diseases and cancer.⁴ Importantly, US28 activates in a ligand-independent manner proliferative and inflammatory signal transduction pathways.^{5–7} This US28-mediated constitutive

[†] Both authors contributed equally to this work.

ABSTRACT

Human cytomegalovirus (HCMV) is a widespread human pathogen, possessing onco-modulatory properties. Constitutive signaling of the HCMV-encoded chemokine receptor US28 and its ability to bind a broad spectrum of chemokines might facilitate HCMV-associated tumor progression. Novel nonpeptidergic chemotypes were identified as neutral antagonists or inverse agonists on US28, that allosterically inhibit chemokine binding to US28.

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signaling induces a transformed and pro-angiogenic phenotype in NIH-3T3 cells, and promotes tumorigenesis in a nude mice model.⁸

Identification of small nonpeptidergic ligands is essential to investigate the role of US28 in evasion of the immune system and HCMV-associated cancers. Moreover, such molecules might be considered as promising therapeutics for anti-viral intervention. Previously, we identified VUF2274 (1) [5-(4-(4-chlorophenyl)-4hydroxy-piperidin-1-yl)-2,2-diphenylpentanenitrile] as a nonpeptidergic allosteric inverse agonist for US28 (Fig. 1a).⁹ Compound 1 inhibited US28-mediated constitutive signaling, chemokine binding, and US28-mediated human immunodeficiency virus (HIV) co-receptor activity with a low μ M potency (EC₅₀ and IC₅₀, respectively).⁹ In subsequent lead optimization programs, approximately 100 new analogues were synthesized and pharmacologically characterized.^{10,11} These structure-activity relationship (SAR) studies revealed the importance of the 4-phenylpiperidine moiety. However, the potency of this class of compounds could not be improved beyond the low µM range. In addition to these 4-substituted piperidine derivates (e.g., 1 and 2), series of piperazinyldibenzothiepine (e.g., 3, 4), cinchonidine derivates (e.g., 5) and benzamides like 6 have been described in the patent literature as inhibitors of chemokine binding to US28 with IC₅₀ values in the 0.3–1 μ M range (Fig. 1a).^{12–14}

In order to identify novel chemotypes that can inhibit constitutive US28-mediated signaling and/or chemokine binding that might be used as new starting point for lead optimization, we screened a selection of compounds from our in-house collection.

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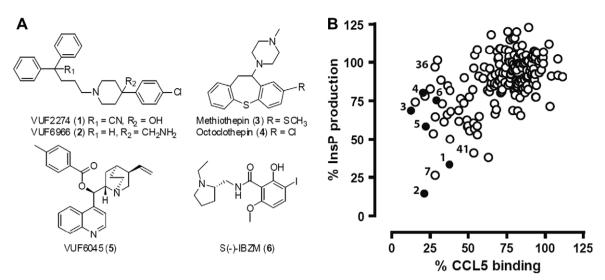
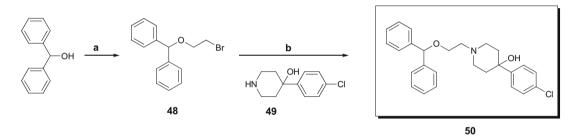


Figure 1. (A) Chemical structures of previously described nonpeptidergic ligands for US28. (B) Screening results using 10 µM nonpeptidergic ligand. Data are represented as percentage CCL5 binding (*X*-axis) and constitutive US28-mediated InsP production (*Y*-axis) in the absence of small ligands. The average of at least three independent experiments are shown. For clarity no error bars (<5%) are shown. Previously described compounds **1–6** are indicated and depicted in black, whereas most significant hits of this screen are indicated **7**, **36**, and **41**.



Scheme 1. Synthetic pathway for the synthesis of compound 50. Reagents and conditions: (a) *p*-TSA, 2-bromoethanol, toluene, reflux; (b) Nal, Na₂CO₃, CH₃CN, microwave (15 min, 160 °C).

Compounds (**7–47**) were selected based on (sub)structural similarity to known ligands acting on US28 (Fig. 1a). Subsequently, the structure–activity relationships (SAR) around three promising hits (i.e., (**7**), (**36**), (**41**)) were explored by synthesizing structural analogues and subsequent pharmacological characterization.

2. Compound selection criteria

For this screening campaign, compounds **7–47** were selected from our in-house compound collection that share (sub)structural similarity to reported US28 ligands. To this end, substructure similarity searches were performed using ChemFinder Std. 7.0 (CambridgeSoft Corporation). In addition, compounds were selected based on subjective structural similarity criteria.

3. Chemistry

3.1. Synthesis of analogues of hit 7

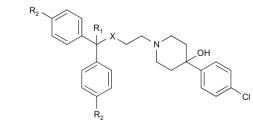
Bromide intermediate **48**, which was synthesized in a quantitative yield by an acid-catalysed condensation of diphenylmethanol with 2-bromoethanol with azeotropic removal of water,¹⁵ was reacted with 4-(4-chlorophenyl)piperidin-4-ol **49** in the microwave in the presence of Nal, Na₂CO₃ and CH₃CN to give compound **50** (Scheme 1). Compound **51** (Table 1) was synthesized in an analogous manner by the alkylation of the commercially available 4,4'-(4-chlorobutane-1,1-diyl)bis(fluorobenzene) with piperidine **49**.

3.2. Synthesis of analogues of hit 36

Reduction of 11,12-dihydrodibenzo[b_f]azocin-6(5H)-one **52** with LiAlH₄ resulted in 5,6,11,12-tetrahydrodibenzo[b_f]azocine **53**,¹⁶ which was alkylated with 1-bromo-3-chloropropane in the microwave (15 min, 200 °C) to yield intermediate **54**. N-alkylation of piperidine moieties **55**, **49** and **56** with chloride **54** afforded inter-

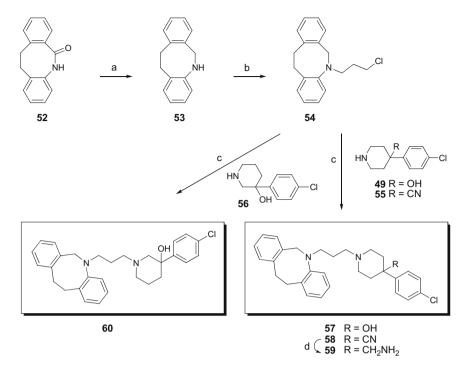
Table 1

Chemical structures and pharmacological properties on US28 of compounds $\mathbf{1},\mathbf{7}$ and $\mathbf{50}\text{-}\mathbf{51}$



No.	R ₁	R ₂	Х	IC_{50} (μM)	EC ₅₀ (μM)
1	CN	Н	CH_2	4.9 (4.4-5.5)	6.3 (5.9-6.8)
7	Н	F	0	2.3 (1.5-3.2)	4.6 (2.5-4.0)
50	Н	Н	0	2.1 (1.8-2.4)	3.5 (1.7-5.4)
51	Н	F	CH_2	3.0 (2.5-3.5)	5.2 (3.2-7.2)

The IC_{50} and EC_{50} values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.



Scheme 2. Synthetic pathway for the synthesis of 57, 59 and 60. Reagents and conditions: (a) LiAlH₄, THF, reflux; (b) 1-bromo-3-chloropropane, Na₂CO₃, microwave (15 min, 200 °C); (c) Nal, Na₂CO₃, CH₃CN, microwave (15 min, 160 °C) or Nal, Na₂CO₃, CH₃CN, reflux; (d) AlCl₃, LiAlH₄, THF.

mediate **58**, and target compounds **57** and **60**, respectively (Scheme 2). Piperidine moiety **55** was synthesized starting from bis(2-chloroethyl)amine hydrochloride as previously described in the literature.^{11,17} The 3-substituted piperidine moiety **56** was synthesized by a Grignard reaction of BOC-protected piperidin-3-one with 4chlorophenyl magnesium bromide followed by a deprotection under acidic conditions.¹⁸ The nitrile group of intermediate **58** was reduced in the presence of AlCl₃ and LiAlH₄ to yield target compound **59**.

3.3. Synthesis of analogues of compound 57

Unsubstituted amine **65** was commercially available, and amines **66** and **67** were synthesized in a quantitative yield via a reductive amination reaction with the corresponding anilines **61–62** and benzaldehydes **63–64** in the presence of sodium triacetoxyborohydride and acetic acid in DCE. The amines **65–67** were deprotonated with NaNH₂ in toluene at reflux temperature followed by an alkylation with 1-bromo-3-chloropropane to give intermediates **68–70**, which were reacted with piperidine **49** in the presence of NaI, Na₂CO₃, and CH₃CN (Scheme 3) to give compounds **71**, **72** and **73**.

3.4. Synthesis of compound 80

Rigid tricyclic analogue **80** was synthesized as depicted in Scheme 4. The reaction of 2-(2-iodophenyl)acetic acid **74** with 4-(methylthio)benzenethiol **75** in the presence of KOH, Cu and water gave intermediate **76**.¹⁹ A subsequent ring closure to the tricyclic system of **77** was performed by a Friedel–Crafts acylation with PPA in toluene.²⁰ Reduction of the carbonyl group of **77** with NaBH₄ in MeOH resulted in alcohol **78**, which was reacted with BF₃·(Et)₂O and 2-bromoethanol in toluene to give bromide **79**.²¹ N-Alkylation of piperidine **49** with **79** yielded target compound **80**.

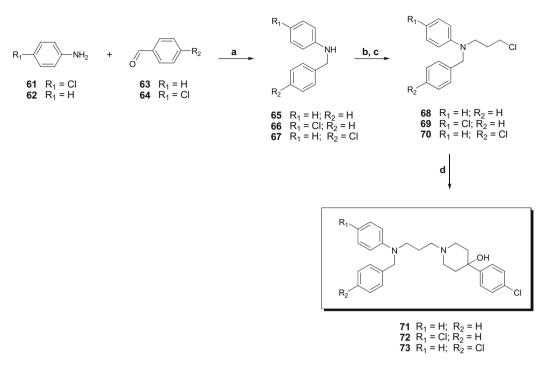
3.5. Synthesis of analogues of hit 41

Indanones **81** were treated with the appropriate substituted benzaldehydes in a saturated solution of K_2CO_3 in EtOH (Scheme

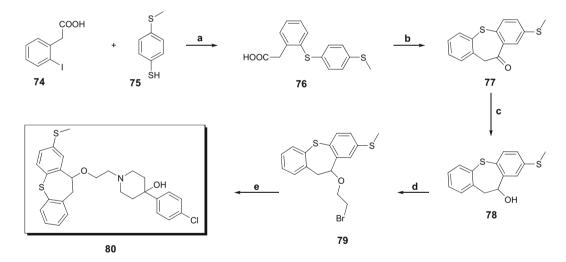
5) to give the substituted 2-(phenylmethylene)-2,3-dihydro-1Hinden-1-ones **82**.²² Reduction of both the carbonyl group as well as the double bond in one pot with lithium aluminium hydride was not successful. Therefore, hydrogenation of the double bond and subsequent reduction of the carbonyl group with sodium borohydride afforded the substituted 2-benzyl-2,3-dihydro-1H-inden-1-ols 83 as 1:1 mixture of both cis/trans isomers in moderate to high yields. Intermediates 83 were converted to the corresponding 1-azido-2-benzyl-2,3-dihydro-1H-indenes 84 in the presence of diphenyl phosphorazidate (DPPA) and 2,3,4,6,7,8,9,10-octahydropy-rimidol[1,2-*a*]azepine (DBU) in THF.^{23,24} For some of the azide intermediates it was possible to separate the two diastereomers via column purification. Hydrogenation of the azide group with palladium on activated carbon afforded target compounds 41 and 85-91. The separation of the two diastereomers could not be accomplished after the last step of the synthesis route. Treatment of 2-benzyl-2,3-dihydro-1H-inden-1-ol derivatives 83 with thionyl chloride gave a 1:1 mixture of the two diastereomers of 1-chloro-2-(2,4-dichlorobenzyl)-2,3-dihydro-1H-indenes 92 in a quantitative yield. Alkylation of chloro derivatives 92 with commercially available amines in the presence of NaI, Na₂CO₃, and CH₃CN at reflux temperature or in the microwave, resulted in mainly elimination of HCl.¹¹ Therefore, the synthesis was performed in a different manner by heating the corresponding amine in CH₃CN to reflux temperature in the presence of Na₂CO₃ and NaI and subsequent addition of the appropriate chloride **92** when reflux temperature was reached. This resulted in the successful synthesis of target compounds **93–98** in moderate to low yields. This synthetic route resulted in the formation of only one diastereomer of the desired compounds.

4. Results and discussion

To identify new chemotypes that inhibit chemokine binding to US28 and/or display inverse agonism, 198 compounds were selected on the merit of (sub)structural similarities to known US28 ligands. Efficacy of these compounds ($10 \mu M$) was deter-



Scheme 3. Synthetic pathway for the synthesis of 71–73. Reagents and conditions: (a) NaBH(OAc)₃, CH₃COOH, DCE; (b) NaNH₂, toluene, reflux; (c) 1-bromo-3-chloropropane, reflux; (d) 49, Nal, Na₂CO₃, CH₃CN, microwave (15 min, 160 °C).

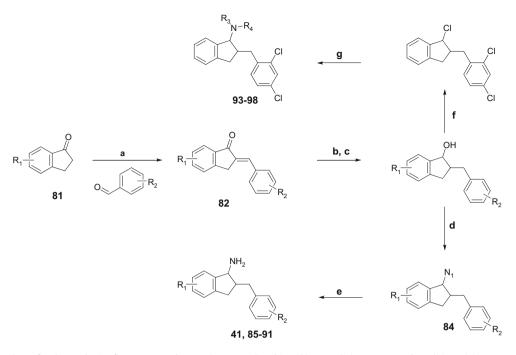


Scheme 4. Synthetic pathway for the synthesis of 80. Reagents and conditions: (a) Cu, KOH, water, reflux; (b) PPA, toluene, reflux; (c) NaBH₄, MeOH; (d) BF₃-(Et)₂O, 2-bromoethanol, toluene; (e) 49, NaI, Na₂CO₃, CH₃CN, reflux.

mined by measuring their effect on the US28-mediated constitutive production of inositol phosphates. Their potency to allosterically inhibit chemokine binding to US28 was analyzed by means of ¹²⁵I-CCL5 radioligand binding assays in the presence of 10 μ M of these compounds (Fig. 1b). Compounds that were identified as allosteric inhibitors of ¹²⁵I-CCL5 binding acted either as antagonists or inverse agonists when considering US28-mediated constitutive signaling. On the other hand, inverse agonistic compounds inhibited ¹²⁵I-CCL5 binding as well. Typically, no inverse agonists were identified that did not affect ¹²⁵I-CCL5 binding (i.e., lower left half of Fig. 1b). Several compounds were identified as novel allosteric inhibitors of US28, and those that inhibited CCL5 binding more than 50% were analyzed more extensively by determining their IC₅₀ and EC₅₀ values from radioligand competition curves and dose–response curves, respectively.

4.1. Structural analogues of VUF2274 (1)

Screening our in-house compound collection yielded the VUF2274 (1)-like compound (7) that displays a slightly higher potency to inhibit constitutive US28 signaling and CCL5 binding as compared with compound 1 (Fig. 1b and Table 1). To evaluate the importance of the fluoro atoms in the phenyl rings and the ether linkage in the alkyl chain between the diphenyl group and the piperidine moiety of 7, compounds 50 and 51 were synthesized (Scheme 1). Removal of both fluoro atoms (i.e., 50) or substitution of the oxygen atom with CH_2 (i.e., 51) did not significantly affect



Scheme 5. Synthetic pathway for the synthesis of 41, 85–91 and 93–98 (R_1 – R_4 are listed in Tables 8 and 9). Reagents and conditions: (a) K_2CO_3 , EtOH; (b) 10% Pd/C, H_2 , MeOH; (c) NaBH₄, MeOH; (d) DPPA, DBU, THF; (e) 10% Pd/C, H_2 , MeOH; (f) SOCfe, DCM; (g) amine, Nal, CH₃CN, 90 °C.

the potency to inhibit chemokine binding and US28-mediated signaling as compared to **7** (Table 1).

4.2. Structural analogues of methiothepin (3) and octoclothepin (4)

The piperazinyldibenzothiepine derivates methiothepin (3) and octoclothepin (4) inhibit CCL5 binding to US28 with a comparable potency as previously described for the binding of CX3CL1 to US28 (Fig. 1b and Table 2).¹³ Compound **3** has a higher potency than the previously described 4-phenylpiperidine scaffold-based compounds (e.g., 1).9-11 Replacement of the thiomethylene group of compound **3** by a chloro atom in **4** resulted in a reduced potency to inhibit CCL5 binding which is comparable to that of 1. On the other hand, both 3 and 4 act as partial inverse agonists on US28 (Fig. 1b) with slightly reduced potency as compared to 1 (Tables 1 and 2). The piperazinyldibenzodiazepine derivate clozapine (8), an atypical antipsychotic drug that binds to a variety of bioaminergic receptors, also diminished (\sim 50%) CCL5 binding to US28 at a concentration of 10 μ M (Table 2). Elongation of the R₂ group of **8** from methyl into ethyl ($\mathbf{9}$), or butyl ($\mathbf{10}$) resulted in IC₅₀ values in the low micromolar range (Table 2). However, compounds 8-10 displayed only weak inverse agonistic activities on US28-mediated constitutive signaling. Substitution of the diazepine with an oxazepine moiety decreases the potency to block CCL5 binding to US28, whereas inverse agonistic activities were similar (11) or abolished (12) (Table 2). Changing the chloro substituent from the 8- (11) to the 2-position (13) of the dibenzo [b, f] [1,4] oxazepine moiety attenuates both potency to block CCL5 binding and inverse agonistic activity (Table 2). Substitution of the oxazepine moiety of compounds 13 and 11 with a thiazepine moiety (compounds 14 and 15, respectively) resulted in neutral antagonists with an increased potency to inhibit CCL5 binding (Table 2). Compounds 16-18 have a high flexibility due to their open structure as compared with the tricyclic moieties of compounds 3-4 and 8-15. Compound 16 displays a lower potency to block CCL5 binding compared to the distinct tricyclic compounds (i.e., **3**, **4**, **8**, **11**, **13**, **14**, **15**). Interestingly, substitution of the methyl group at R_2 position of **16** with a ω - phenylbutyl group (i.e., compound **17**) restores the potency to inhibit CCL5 binding to US28 (Table 2). Subsequent replacement of the phenyl group of **17** with an alcohol moiety (i.e., **18**) diminishes potency again. Compounds **16** and **17**, but not **18**, display weak inverse agonistic efficacy.

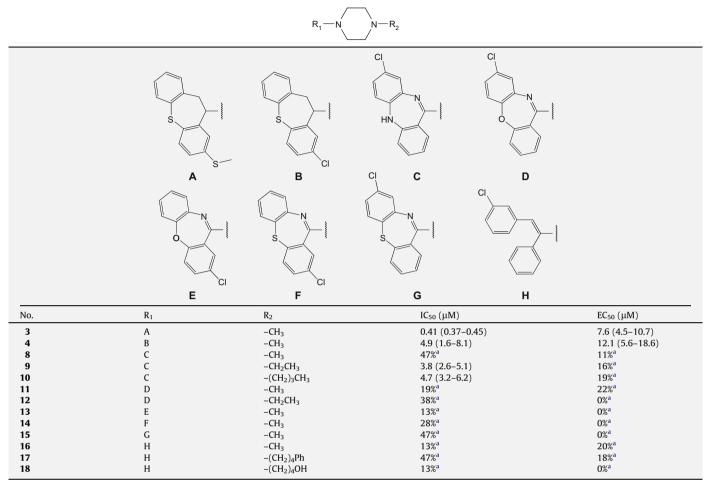
4.3. Tropine derivates

Our screening campaign also identified tropine derivatives as a new hits for US28. Compound B83 (19) inhibited CCL5 binding and constitutive US28 activity with IC₅₀ and EC₅₀ values, respectively, in the low µM range (Table 3). Replacement of the octyl group at the R position of **19** by a phenetyl group (i.e., compound **20**), and subsequent shortening of the alkyl linker between the basic nitrogen atom of the tropine ring and the phenyl ring (i.e., compound **21**) did not significantly affect the inhibitory properties on CCL5 binding or constitutive signaling (Table 3). On the other hand, small methyl and ethyl chains (i.e., compound 22 and 23, respectively), N-hydroxypropionimidamide or ethyl-guanidine groups (i.e., compound 24 and 25, respectively) are detrimental for their interaction with US28. In compounds 26-28 an unsaturated bridge between the two phenyl rings in the tricyclic moiety is introduced, which in combination with short branched alkyl chains at the R position (i.e., compounds 26 and 27) possess no inhibitory properties on US28. However, placing a cyclohexyl group at this R position yielded an inhibitor (i.e., compound 28) with IC₅₀ and EC₅₀ values in the low µM range (Table 3).

4.4. 1-Phenylpropan-2-amine derivates

Various N-substituted analogues of 1-phenylpropan-2-amine (i.e., compounds **29–35**) display low μ M potencies to inhibit CCL5 binding and constitutive activity, even though their tricyclic moieties are quite different with respect to rigidity and the conformation of the two aromatic phenyl rings (Table 4).The two aromatic rings of the tricyclic moiety of compound **29** are tilted out of plane due to the presence of three carbon atoms between the two phenyl rings, whereas the tricyclic ring system of **30** is nearly planar, as deter-

Chemical structures and pharmacological properties on US28 of compounds 3-4 and 8-18



The IC₅₀ and EC₅₀ values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.

^a Inhibition of CCL5 binding and constitutive US28 signaling at a 10 μ M compound concentration.

mined by conformational analysis using the molecular modeling program MOE (molecular operating environment, version 2004.03). Saturation of the double bond of compound **30** (i.e., compound **31**), or introduction of an unsaturated bridge between the two phenyl rings in compound **32** did not significantly affect their potency to inhibit CCL5 binding or constitutive signaling (Table 4). Interestingly, a thioxanthene tricyclic moiety in combination with a *p*-chloro substituent on the phenethyl group confers low μ M potency to block CCL5 binding, but weak inverse agonistic efficacy to compound **33** (Table 4). Similar inhibitory potencies were observed for compound **34** which consists of an acridine tricyclic ring system in combination with a *m*-trifluoromethyl group on the phenethyl group (Table 4). Replacing the tricyclic ring system in **34** with a 4,4'-difluorophenyl group (i.e., compound **35**) reduces the potency to block CCL5 binding to US28 (Table 4).

4.5. Identification and conversion of US28 antagonists into inverse agonists

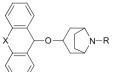
The tricyclic imipramine analogue A194 (**36**) was identified in this screen as the first allosteric neutral antagonist for US28 as it blocks CCL5 binding to US28 with an IC₅₀ value of 4.9 μ M, but displayed no inverse efficacy (Fig. 1b and Table 5). Replacing the *N*-methyl group in **36** with a 2-ethanol group in compound **37** resulted in a slightly lower IC₅₀ value (Table 5). In contrast, convert-

ing the amine nitrogen atom in the tricyclic moiety to an amide nitrogen atom (i.e., compounds **38** and **39**) resulted in a significant reduced inhibition of CCL5 binding to US28 (Table 5). Hence, a basic nitrogen in the tricyclic moiety seemed to be essential for interaction of these antagonists with US28.

Previous SAR studies revealed that a 4-phenylpiperidine moiety of lead compound 1 is essential for inverse efficacy at US28.10 Anticipating that this moiety would confer inverse efficacy to neutral antagonist 36, we synthesized an hybrid compound 57 in which the N-methylpiperazine group of 36 was substituted with 4-(4-chlorophenyl)piperidin-4-ol (Fig. 2 and Scheme 2). Indeed, this hybrid compound 57 displayed inverse efficacy on US28-mediated constitutive signaling with an EC₅₀ of 4.7 µM (Table 5). Moreover, compound **57** was sevenfold more potent in inhibiting CCL5 to binding than the two compounds (i.e., 1 and 36), from which it originated (Table 5). In contrast to the previously described VUF2274-like compound $\mathbf{2}$,¹¹ substitution of the 4-hydroxy group of compound **57** with a methylamine group (i.e., compound **59**), did not yield the anticipated increase in inhibitory potencies (Table 5). Introduction of a chiral center by moving the 4-hydroxy group and the 4-chlorophenyl substituent to the 3-position of the piperidine ring (i.e., compound **60**) did not change the pharmacological properties as compared to compound 57 (Table 5).

Next, flexible analogues of compound **57** were synthesized to explore the significance of rigidity in the tricyclic moiety for the

Chemical structures and pharmacological properties on US28 of compounds 19-28



No.	Х	R	IC ₅₀ (µM)	EC ₅₀ (µM)
19	CH ₂ -CH ₂	$(CH_2)_7CH_3$	5.6 (3.4-7.8)	4.9 (3.7-6.2)
20	CH ₂ -CH ₂	$(CH_2)_2Ph$	11.9 (8.7-15.1)	4.3 (3.5-5.1)
21	CH ₂ -CH ₂	CH ₂ Ph	10.5 (8.3-12.6)	8.8 (5.9-11.7)
22	CH ₂ -CH ₂	CH_3	15% ^a	0% ^a
23	CH ₂ -CH ₂	CH ₂ CH ₃	7% ^a	0% ^a
24	CH ₂ -CH ₂	N-OH √ NH₂	0% ^a	0% ^a
25	CH ₂ -CH ₂	MH H₂N	34% ^a	0% ^a
26	CH=CH	$CH(CH_3)_2$	13% ^a	3% ^a
27	CH=CH	$(CH_2)_2CH_3$	18% ^a	0% ^a
28	CH=CH	Cyclohexyl	8.3 (6.8–9.8)	5.8 (3.3-8.3)

The IC_{50} and EC_{50} values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.

 a Inhibition of CCL5 binding and constitutive US28 signaling at a 10 μM compound concentration.

interaction with US28 (Scheme 3). Exclusion of the ethylene bridge between the two phenyl rings as in open analogue **71** decreased the potency to block CCL5 binding (Table 6). Subsequent introduction of a chloro substituent at the 4-position of one of the phenyl rings revealed an interesting regioselectivity with compound **72** having a slightly higher potency to inhibit CCL5 binding than compound **73**. Not surprisingly, these modifications in the tricyclic moiety did not significantly affect the inverse agonistic potencies of compounds **71**, **72**, and **73** as compared to **57** (Table 6).

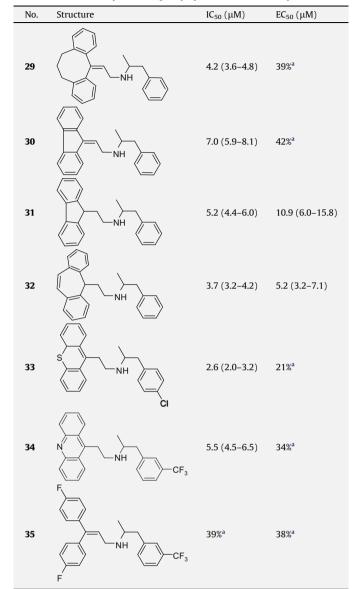
Challenged by the observed increase in potency of compound **57** to block CCL5 binding to US28 as compared to compounds **1** and **36**, we decided to design and synthesize another hybrid compound by fusing the 4-(4-chlorophenyl)piperidin-4-ol scaffold to the tricyclic moiety of compound **3** (Scheme 4). Compound **3** had the lowest IC₅₀ value so far observed and we anticipated to generate a full inverse agonist with an increased potency to inhibit CCL5 binding. Unfortunately, this hybrid compound (**80**) displayed decreased potency to block CCL5 binding to US28 (Table 6).

4.6. Structural analogues of S(-)-IBZM (6)

Automated similarity searches of our in-house compound collection using ChemFinder Std. 7.0 software did not yield any analogues. However, inspired by the substructure of S(-)-IBZM (6) several analogues were selected from our library by subjective visual inspection. Pharmacological analysis of these S(-)-IBZM-like analogues revealed some interesting preliminary SAR. Compound 41 [2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-amine] was identified as a novel scaffold for inverse agonism on US28 (Fig. 1b and Table 7). Conversion of the cyclopentyl group of 41 into cyclohexyl (i.e., compound 42) or cycloheptyl (i.e., compound **43**) did not affect the potency to inhibit CCL5 binding (Table 7). However, compound 43 displayed significantly less inverse agonistic potency as compared to compounds **41** and **42**. Interestingly, increasing the conformational freedom by replacing the rigid ring structure with a more flexible propyl chain as in compound 44 did not significantly affect the pharmacological properties as compared to compounds 41 and 42 (Table 7). However, the presence of 3,4-dichloro substituents on (at least one of) the phenyl rings

Table 4

Chemical structures and pharmacological properties on US28 of compounds 29-35



The IC_{50} and EC_{50} values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.

 a Inhibition of CCL5 binding and constitutive US28 signaling at a 10 μM compound concentration.

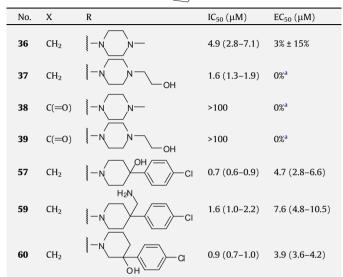
seems to be essential as indicated by compound **45**. Interestingly, compounds **46** and **47**, in which the amine substituent is replaced with a bulky 2-methoxy-2-phenylethanamine group, displayed similar IC₅₀ and EC₅₀ values as the smaller analogues **41** and **42** (Table 7). Replacement of the dichloro substituents (**46**) in the phenyl ring with a *t*-butyl group (**47**) at position 4 did not alter the pharmacological properties.

The structure of compound **41** provided some interesting options for further lead optimization. Firstly, compounds **85–91** with various modifications in the 2,4-dichlorobenzyl moiety were designed and synthesized to optimize the substitution pattern in the benzyl ring (Scheme 5 and Table 8). Substitution of the electron withdrawing chloro groups with electron donating methyl groups in compound **85** resulted in a small decrease in the potency to inhibit CCL5 binding to US28, without affecting inverse efficacy. Subsequent introduction of two methoxy groups at the *ortho* and *para*

pound concentration.

Chemical structures and pharmacological properties on US28 of compounds **36–39** and **57**, **59** and **60**

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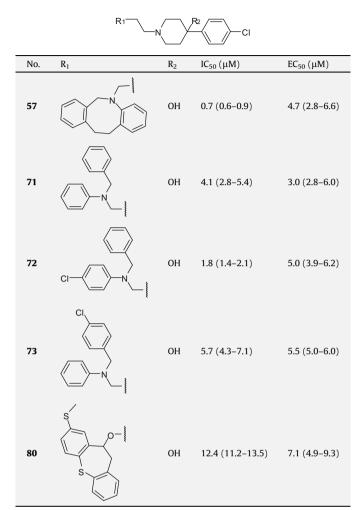


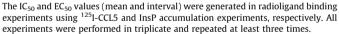
The IC₅₀ and EC₅₀ values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times. ^a Inhibition of CCL5 binding and constitutive US28 signaling at a 10 μ M com-

position of the benzyl ring, significantly attenuated the potency of compound **86** to inhibit CCL5 binding as well as constitutive US28 signaling. Although a 2-methoxy substitution is allowed at the benzyl moiety (compound **87**), the presence of a 4-methoxy group in compounds **86** and **88** seemed to be detrimental for interaction with US28. Interestingly, the potency to inhibit CCL5 binding and inverse agonism was restored in compound **89** by introducing an additional 3-methyl group in 4-methoxyphenyl ring (Table 8).

Table 6

Chemical structures and pharmacological properties on US28 of compounds 57, 71–73 and 80





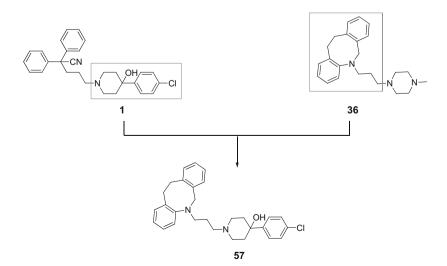
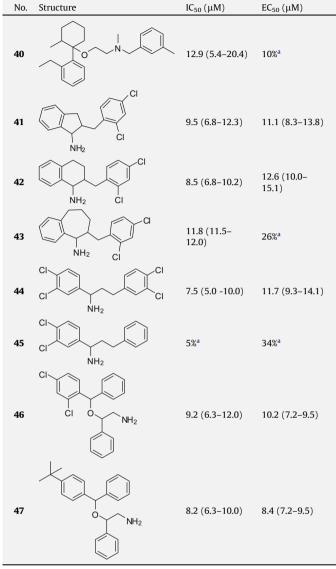


Figure 2. Chemical structures of inverse agonist 1, neutral antagonist 36 and hybrid compound 57.

Chemical structures and pharmacological properties on US28 of compounds 40-47



The IC_{50} and EC_{50} values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.

 $\overset{a}{}$ Inhibition of CCL5 binding and constitutive US28 signaling at a 10 μM compound concentration.

Introduction of an acetamide substituent at the *para* position of the phenyl ring (i.e., compound **90**) or removal of all substituents (i.e., compound **91**) impaired all inhibitory activities at US28 (Table 8).

To evaluate the importance of the amine group of lead compound **41**, several analogues were designed and synthesized in which the nitrogen atom was incorporated into different ring systems (Scheme 5 and Table 9). Substitution of the amine group with a 4-(4-chlorophenyl)piperidin-4-ol group (i.e., compound **93**), a piperidine ring (i.e., compound **94**), or an *N*-methylpiperazine moiety (i.e., compound **95**) are tolerable and did not have a major impact on the pharmacological properties as compared to lead compound **41** (Table 9). On the other hand, introduction of an *N*-phenylpiperazine group (i.e., compound **96**) or unsaturated tetrahydropyridine moiety (i.e., compound **97**), or attachment of a benzyl group to the nitrogen atom (i.e., compound **98**) are not allowed and impaired the potency to inhibit CCL5 binding to US28, as well as US28-mediated constitutive signaling (Table 9).

5. Conclusions

To identify novel nonpeptidergic ligands acting on the constitutively active HCMV-encoded receptor US28, a selection of chemotypes extracted from our in-house compound collection was screened on their potency to inhibit CCL5 binding to US28 and constitutive receptor signaling. Compounds 7 and 41 were identified as novel inverse agonists acting on US28, whereas compound 36 was identified as a neutral antagonist. Combining the tricyclic group of this neutral antagonist with the 4-(4-chlorophenyl)piperidin-4-ol moiety of lead inverse agonist 1, resulted in a hybrid compound **57** displaying inverse efficacy and an increased potency to block CCL5 binding. Hence, inverse efficacy can indeed be attributed to the 4-(4-chlorophenyl)piperidin-4-ol moiety as previously indicated.^{10,11} SAR of compounds 7, 57 and 41 were further explored by generating series of structural analogues, however none of the modifications resulted in improved inhibitory activities on US28. The identification of potent nonpeptidergic inverse agonists and antagonists will provide us with a tool to validate the significance of US28 in HCMV-associated pathologies.

6. Experimental

6.1. General procedures

THF and DCM were freshly distilled from lithium aluminium hydride. All reactions were performed under an atmosphere of dry nitrogen. Microwave reactions were performed in a CEM Explorer single mode MW reactor equipped with auto sampler. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker AC-200 (200 MHz) spectrometer unless otherwise stated. J.T. Baker silica gel was used for flash chromatography. Melting points were measured on a MPA100 OptiMelt automated melting point system apparatus and were uncorrected. HRMS mass spectra were recorded on a Finnigan MAT 900 mass spectrometer. Analytical HPLC-MS analyses were conducted using a Shimadzu LC-8A preparative liquid chromatograph pump system with a Shimadzu SPD-10AV UV-vis detector set at 254 nm, with the MS detection performed with a Shimadzu LCMS-2010 liquid chromatograph mass spectrometer. The analyses were performed using the following conditions; condition I: a Xbridge(C18)5um column (100 mm \times 4.9 mm) with 70% MeOH-30% H₂O-0.1% formic acid (Method Ia); 60% MeOH-40% H₂O-0.1% formic acid (Method Ib); 50% MeOH-50% H₂O-0.1% formic acid (Method Ic); 40% MeOH-60% H₂O-0.1% formic acid (Method Id) or 30% MeOH-70% H₂O-0.1% formic acid (Method Ie). Flow rate = 1.0 mL/min. Total run time 15 min unless otherwise stated. Condition II: a Xbridge(C18)5um column (100 mm \times 4.9 mm) with 50% CH₃CN-50% H₂O-0.1% formic acid (Method IIa); 40% CH₃CN-60% H₂O-0.1% formic acid (Method IIb), 35% CH₃CN-65% H₂O-0.1% formic acid (Method IIc), 30% CH₃CN-70% H₂O-0.1% formic acid (Method IId); 25% CH₃CN-75% H₂O-0.1% formic acid (Method IIe) or 20% CH₃CN-80% H₂O-0.1% formic acid (Method IIf). Flow rate = 1.0 mL/ min. Total run time 20 min. Condition III: a Xbridge(C18)5um column (100 mm \times 4.9 mm) with 60% CH₃CN-40% H₂O with 10% NH₄HCO₃/NH₄OH buffer pH 8 (Method IIIa); 50% CH₃CN-50% H₂O with 10% NH₄HCO₃/NH₄OH buffer pH 8 (Method IIIb). Flow rate = 1.0 mL/min. Total run time 20 min. Compounds that were isolated as fumaric acid salts all showed an extra peak around 2 min. Fumaric acid blancs were used to determine the tR of fumaric acid. Purities calculated are based on RP.

Compounds **1**, **2**, **11** and **12** were previously described.^{9,11,25} Compounds **3**, **4**, **6**, **8**, and **13** were purchased from Sigma–Aldrich

97

98

-NH

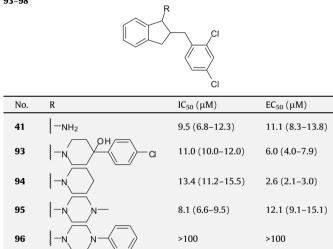
Chemical structures and pharmacological properties on US28 of compounds 41, and 85-91



		\^R	
No.	R	IC ₅₀ (μM)	EC ₅₀ (μM)
41 85 86 87 88 89 90 91	2,4-Cl ₂ 2,4-Me ₂ 2,4-(OMe) ₂ 2-OMe 4-OMe 3-Me-4-OMe 4-NH(C=O)CH ₃ H	9.5 (6.8-12.3) 17.7 (14.5-20.9) >100 37.3 (28.8-45.7) >100 59.7 (50.1-69.2) >100 80.6 (32.4-128.8)	11.1 (8.3-13.8) 13.7 (9.1-18.2) >100 42.7 (41.7-43.7) >100 16.4 (14.1-18.6) >100 >100

The IC_{50} and EC_{50} values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.

Table 9 Chemical structures and pharmacological properties on US28 of compounds 41, and 93–98



³ The IC₅₀ and EC₅₀ values (mean and interval) were generated in radioligand binding experiments using ¹²⁵I-CCL5 and InsP accumulation experiments, respectively. All experiments were performed in triplicate and repeated at least three times.

>100

>100

>100

>100

Co. (USA). Compounds **14** and **15** were kindly donated by Dr. Aebischer from the Sandoz Research Institute in Bern. Compounds **5**, **7**, **9**, **10** and **16–47** were selected from our in-house screening library. Purity of all compounds was $\ge 95\%$ as determined by LC–MS measurements.

6.1.1. General method A: 1-(2-(benzhydryloxy)ethyl)-4-(4-chlorophenyl)piperidin-4-ol fumarate (50)

Bromide intermediate **48** (0.58 g, 1.99 mmol), which was synthesized following a method previously described,¹⁵ 4-(4-chlorophenyl)piperidin-4-ol **49** (0.51 g, 2.41 mmol), NaI (0.30 g, 2.00 mmol), Na₂CO₃ (0.42 g, 3.96 mmol) and 3 mL CH₃CN were added in a 10 mL microwave vessel and this was reacted during 15 min in the microwave at a temperature of 160 °C (settings: ramp time 5 min, hold time 15 min, power 200 watt, pressure 17.2 bar). The solvent was removed in vacuo and the residue was diluted with water (20 mL), followed by an extraction with DCM (3×15 mL). The combined organic layers were washed with water $(3 \times 40 \text{ mL})$ and brine (40 mL), dried over anhydrous MgSO₄, filtered, and evaporated in vacuo. Purification by flash chromatography (0-50% EtOAc in DCM) gave 480 mg (57%) of the free base as an oil. This was dissolved in EtOAc and acidified by the addition of a saturated solution of fumaric acid in Et₂O. The fumaric salt was isolated by filtration and recrystallized from MeOH/Et₂O to give 281 mg(52%) of **50** as a white solid. ¹H NMR (CDCl₃): δ 1.60–1.81 (m, 2H), 2.08–2.43 (m, 2H), 2.84– 3.25 (m, 6H), 3.65-3.78 (m, 2H), 5.28 (s, 1H), 6.63 (s, 2H), 7.11-7.36 (m, 14H). ¹³C (CDCl₃): δ 36.29, 49.42, 56.91, 64.68, 69.54, 84.64, 126.57, 127.30, 128.09, 128.89, 134.06, 135.30, 141.78, 148.74, 169.53. Anal. RP-HPLC *Ib*: $t_{\rm R}$ = 8.60 min (purity 100%), *IId*: $t_{\rm R}$ = 12.13 min (purity 100%). HRMS (ESI) m/z calcd for C₂₆H₂₈ClNO₂: 421.1808; found: 421.1803.

6.1.2. General method B: 1-(4,4-bis(4-fluorophenyl)butyl)-4-(4-chlorophenyl)piperidin-4-ol (51)

A solution of 4,4'-(4-chlorobutane-1,1-diyl)bis(fluorobenzene) (0.56 g, 1.99 mmol), 4-(4-chlorophenyl)piperidin-4-ol **49** (0.51 g, 2.41 mmol), NaI (0.30 g, 2.00 mmol) and Na₂CO₃ (0.43 g, 4.06 mmol) in CH₃CN (20 mL) was refluxed overnight. The solvent was removed in vacuo, the residue was diluted with water (50 mL) and extracted with DCM (3×30 mL). The combined organic layers were washed with water $(3 \times 50 \text{ mL})$ and brine (50 mL), dried over anhydrous MgSO₄, filtered and evaporated in vacuo. Purification by flash chromatography (0-100% EtOAc in DCM) and recrystallization from EtOAc gave 853 mg (94%) of **51** as a white solid. ¹H NMR (CDCl₃): δ 1.48–1.72 (m, 5H), 1.92–2.24 (m, 4H), 2.30–2.55 (m, 4H), 2.69–2.85 (m, 2H), 3.86 (t, J = 7.8 Hz, 1H), 6.90–7.43 (m, 12H). ¹³C (CDCl₃): δ 25.76, 34.27, 38.71, 49.83, 50.14, 58.92, 71.41, 115.57, 115.78, 126.47, 128.82, 129.43, 129.51, 133.22, 140.86, 140.89, 147.16, 160.54, 162.97. Anal. RP-HPLC Ib: *t*_R = 8.78 min (purity 100%), *IIb*: *t*_R = 13.99 min (purity 100%). HRMS (EI) *m*/*z* calcd for C₂₇H₂₈ClF₂NO: 455.1827; found: 455.1826.

6.1.3. General method C: 1-(3-((4-chlorobenzyl)(phenyl)amino)propyl)-4-(4-chlorophenyl)-piperidin-4-ol fumarate (73)

(i) A solution of aniline **62** (1.82 mL, 20.0 mmol), 4-chlorobenzaldehyde **64** (2.35 mL, 20.0 mmol) and CH₃COOH (1.14 mL, 19.9 mmol) in DCE (100 mL) was stirred for 24 h at room temperature and NaBH(OAc)₃ (6.36 g, 30.0 mmol) was added. The reaction mixture was stirred for another 18 h, quenched with 5% Na₂CO₃ (100 mL) and the water layer was extracted with DCM (3 × 100 mL). The combined organic layers were washed with brine (150 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give 4.42 g (100%) of **67** as a brown solid. ¹H NMR (CDCl₃): δ 4.06 (br s, 1H), 4.29 (s. 2H), 6.51–6.58 (m, 2H), 7.07–7.14 (m, 2H), 7.24–7.36 (m, 5H).

(ii) NaNH₂ (0.39 g, 10.0 mmol) was added to a solution of **67** (1.09 g, 5.01 mmol) in toluene (15 mL) and this was refluxed for 18 h. 1-Bromo-3-chloropropane (2.5 mL, 25.3 mmol) was added and the reaction mixture was refluxed for another 18 h. Water (25 mL) was added and the water layer was extracted with EtOAc (3 × 20 mL). The combined organic layers were washed with water (3 × 30 mL) and brine (40 mL), dried over anhydrous Na₂SO and filtered. After evaporation under reduced pressure, the residue was purified by flash chromatography (5% DCM in hexane) to give 142 mg (10%) of **70** as a light yellow oil. ¹H NMR (CDCl₃): δ 1.98–2.18 (m, 2H), 3.45–3.68 (m, 4H), 4.50 (s, 2H), 6.51–6.75 (m, 3H), 7.03–7.28 (m, 6H).

(iii) Following method B using **70** gave 142 mg of the free base as an oil. This was converted to the fumaric salt as described for **50**

to give 178 mg (63%) of **73** as a white solid. ¹H NMR (MeOH- d_4): δ 1.81–1.93 (m, 2H), 1.97–2.17 (m, 2H), 2.19–2.39 (m, 2H), 2.98–3.39 (m, 6H), 3.50 (t, *J* = 7.2 Hz, 2H), 4.54 (s, 2H), 6.59–6.30 (m, 4H), 7.02–7.51 (m, 11H).¹³C (MeOH- d_4): δ 23.81, 36.88, 49.57, 50.09, 55.21, 55.84, 69.75, 114.39, 118.29, 127.48, 129.44, 129.58, 129.61, 130.30, 133.52, 134.07, 137.00, 139.26, 147.47, 149.37, 173.62. Anal. RP-HPLC *Ib*: $t_{\rm R}$ = 4.87 min (purity 100%). *IIb*: $t_{\rm R}$ = 3.89 min (purity 100%). HRMS (EI) *m*/*z* calcd for C₂₇H₃₀Cl₂N₂O: 468.1735; found: 468.1721.

6.1.4. 5-(3-Chloropropyl)-5,6,11,12-tetrahydrodibenzo[*b*,*f*]azo-cine (54)

5,6,11,12-Tetrahydrodibenzo[*b*,*f*]azocine **53** (0.42 g, 2.01 mmol), which was synthesized as previously described in the literature,¹⁶ and Na₂CO₃ (0.42 g, 3.96 mmol) were added in a 10 mL microwave vessel and 1-bromo-3-chloropropane (3 mL) was added. This solution was reacted in the microwave for 15 min at a temperature of 200 °C (settings: ramp time 5 min, hold time 15 min, power 200 watt, pressure 17.2 bar) and filtered. The solvent was evaporated in vacuo and the residue was purified by flash chromatography (15% DCM in hexane) to give 228 mg (40%) of **54** as a colorless oil. ¹H NMR (CDCl₃): δ 1.85–1.98 (m, 2H), 3.04–3.48 (m, 8H), 4.15 (s, 2H), 6.80–7.24 (m, 8H).

6.1.5. 4-(4-Chlorophenyl)-1-(3-(11,12-dihydrodibenzo[*b*,*f*]azocin-5(6*H*)-yl)propyl)piperidine-4-ol fumarate (57)

Following method A using **54** (0.29 g, 1.01 mmol) gave 289 mg of the free base as an oil. This was converted to the fumaric salt as described for **50** and recrystallized from MeOH/Et₂O to give 199 mg (38%) of **57** as white crystals. ¹H NMR (CDCl₃): δ 1.50–1.92 (m, 5H), 2.09–2.36 (m, 2H), 2.41–2.70 (m, 4H), 2.78–3.34 (m, 8H), 4.11 (s, 2H), 6.69 (s, 2H), 6.72–7.35 (m, 12H). ¹³C (CDCl₃): δ 23.15, 33.28, 34.22, 35.92, 48.11, 51.14, 55.08, 60.96, 69.57, 119.32, 122.17, 125.91, 126.82, 127.15, 128.39, 128.79, 129.73, 131.08, 132.94, 135.45, 136.06, 137.65, 141.83, 145.37, 150.02, 171.33. Anal. RP-HPLC *Ib* (total run time 20 min): $t_{\rm R}$ = 14.43 min (purity 100%), *IIc*: $t_{\rm R}$ = 11.35 min (purity 97%). HRMS (EI) *m/z* calcd for C₂₉H₃₃ClN₂O: 460.2281; found: 460.2284.

6.1.6. (4-(4-Chlorophenyl)-1-(3-(11,12-dihydrodibenzo[*b*,*f*]azo cin-5(6*H*)-yl)propyl)piperidin-4-yl)methanamine fumarate (59)

(i) Following method B using **54** and 4-(4-chlorophenyl)-piperidine-4-carbonitrile hydrochloride **55**, which was synthesized as previously described in the literature,¹¹ afforded 675 mg of **58** as a light yellow oil. The crude product was used without further purification.

(ii) AlCl₃ (0.37 g, 2.78 mmol) was added portion wise to a suspension of LiAlH₄ (0.11 g, 2.90 mmol) in THF (10 mL) at 0 °C and this was stirred for 5 min followed by the drop wise addition of a solution of 58 (675 mg, 1.44 mmol) in THF (5 mL). The reaction mixture was allowed to warm to room temperature, stirred overnight, cooled with an ice bath and quenched with a saturated solution of Na₂CO₃ in water until the foaming stopped. Subsequently, the suspension was filtered; the filtrate was dried over anhydrous MgSO₄, filtered again and evaporated in vacuo. Purification by flash chromatography (5% Et₃N in EtOAc) afforded 183 mg (39%) of the free base as an oil. This was converted to the fumaric salt as described for 50 to give 199 mg (24%) of **59** as a light brown solid. ¹H NMR (CDCl₃/DMSO d_6): δ 1.54–1.75 (m, 2H), 1.82–1.93 (m, 2H), 2.05–2.18 (m, 4H), 2.27-2.39 (m, 2H), 2.59-2.68 (m, 2H), 2.86 (s, 2H), 2.92-2.99 (m, 2H), 3.03-3.19 (m, 6H), 4.11 (s, 2H), 6.60 (s, 2H), 6.73 (t, J = 7.3 Hz, 1H), 6.89-7.04 (m, 6H), 7.11-7.39 (m, 5H). ¹³C NMR (CDCl₃/DMSO*d*₆): δ 24.06, 31.36, 32.79, 33.75, 38.67, 48.42, 50.68, 54.91, 60.22, 119.50, 121.68, 125.47, 126.33, 126.68, 128.51, 128.59, 128.79, 129.20, 130.57, 131.95, 134.13, 135.82, 137.06, 141.06, 146.64, 149.77. Anal. RP-HPLC *Ib*: t_{R} = 4.43 min (purity 99%), *Ic* (total run time 20 min): $t_{\rm R}$ = 14.35 min (purity 100%), *Ile* (total runtime 30 min): $t_{\rm R}$ = 21.45 min (purity 96%). HRMS (EI) *m/z* calcd for C₃₀H₃₆ClN₃: 473.2598; found: 473.2579.

6.1.7. 3-(4-Chlorophenyl)-1-(3-(11,12-dihydrodibenzo[*b*,*f*]azocin-5(6*H*)-yl)propyl)piperidin-3-ol fumarate (60)

Following method A using **54** (0.18 g, 0.63 mmol) and 3-(4-chlorophenyl)piperidin-3-ol **56** (0.11 g, 0.52 mmol), which was synthesized as described in the literature,¹⁸ gave 77 mg of the free base as an oil. This was converted to the fumaric salt as described for **50** to give 50 mg (17%) of **60** as a white solid. ¹H NMR (MeOH- d_4): δ 1.76–2.29 (m, 6H), 2.79–3.10 (m, 5H), 2.91 (s, 2H), 3.13–3.39 (m, 6H), 4.13 (s, 2H), 6.69 (s, 2H), 6.79–7.12 (m, 8H), 7.37–7.49 (m, 4H). ¹³C NMR (MeOH- d_4): δ 20.44, 23.57, 34.56, 34.75, 35.80, 52.18, 53.72, 56.81, 61.36, 62.93, 71.47, 121.23, 124.28, 127.22, 127.77, 128.10, 128.49, 129.67, 129.94, 130.90, 132.24, 134.91, 136.16, 138.21, 139.02, 143.20, 144.30, 151.23, 171.08. Anal. RP-HPLC *lb* (total run time 20 min): t_R = 6.78 min (purity 99%), *lla* (total run time 30 min): t_R = 14.64 min (purity 98%). HRMS (EI) *m/z* calcd for C₂₉H₃₃ClN₂O: 460.2281; found: 460.2283.

6.1.8. 1-(3-(Benzyl(phenyl)amino)propyl)-4-(4-chlorophenyl)piperidin-4-ol hemifumarate (71)

Following method C starting with commercially available *N*-benzylaniline gave 331 mg of the free base as an oil. The final product was converted to the fumaric salt as described for **50** and recrystallized from MeOH/Et₂O to give 261 mg (24% over two steps) of **71** as white crystals. ¹H NMR (CDCl₃/DMSO-*d*₆): δ 1.53–1.72 (m, 2H), 1.74–1.97 (m, 2H), 2.00–2.21 (m, 2H), 2.41–2.70 (m, 4H), 2.82–3.01 (m, 2H), 3.26–3.48 (t, *J* = 7.4 Hz, 2H), 4.42 (s, 2H), 6.49–6.88 (m, 5H), 6.98–7.42 (m, 11H). ¹³C NMR (CDCl₃/DMSO-*d*₆): δ 22.90, 36,45, 48.43, 54.32, 54.76, 69.23, 112.30, 116.32, 126.13, 126.43, 126.61, 128.02, 128.34, 129.04, 135.15, 138.49, 146.62, 148.14, 170.21. Anal. RP-HPLC *lb*: *t*_R = 10.01 min (purity 100%), *llb* (total run time 30 min): *t*_R = 8.05 min (purity 100%). *HRMS* (EI) *m/z* calcd for C₂₇H₃₁ClN₂O: 434.2125; found: 434.2113.

6.1.9. 1-(3-(Benzyl(4-chlorophenyl)amino)propyl)-4-(4-chlorophenyl)piperidin-4-ol hemifumarate (72)

Following the analogous route (including the reductive amination and alkylation of the piperidine nitrogen atom) as described in method C starting with benzaldehyde and 4-chloroaniline. Conversion of the final product to the fumaric salt as described for **50** and recrystallized from Et₂O/MeOH resulted in the isolation of 112 mg of **72** as white crystals (6% over three steps) ¹H NMR (CDCl₃/DMSO-*d*₆): δ 1.61–2.05 (m, 4H), 2.13–2.37 (m, 2H), 2.53–2.86 (m, 2H), 2.99–3.12 (m, 2H), 3.37 (t, *J* = 7.1 Hz, 2H), 4.44 (s, 2H), 6.46–6.68 (m, 2H), 6.72 (s, 2H), 7.02–7.39 (m, 11H). ¹³C NMR (CDCl₃/DMSO-*d*₆): δ 22.64, 36.16, 48.38, 48.70, 54.54, 69.37, 113.62, 121.22, 126.02, 126.40, 126.86, 128.26, 128.49, 128.88, 132.73, 135.35, 137.92, 145.84, 146.72, 170.93. Anal. RP-HPLC *Ib* : *t*_R = 5.03 min (purity 100%), *IIa* (total run time 30 min): *t*_R = 14.46 min (purity 98%). HRMS (EI) *m/z* calcd for C₂₇H₃₀Cl₂N₂O: 468.1735; found: 468.1718.

6.1.10. 4-(4-Chlorophenyl)-1-(2-(8-(methylthio)-10,11-dihydrodibenzo[b,f]thiepin-10-yloxy)-ethyl)piperidin-4-ol fumarate (80)

(i) 4-(Methylthio)benzenethiol **75** (5.24 g, 20.0 mmol) was added to a solution of 2-(2-iodophenyl)acetic acid **74** (3.12 g, 20.0 mmol) and KOH (4.49 g, 80.0 mmol) in water (60 mL). After the addition of Cu (205 mg), the reaction mixture was refluxed for 48 h, cooled to room temperature and filtered. The filtrate was acidified with 1.0 M HCl, filtered and the residue was dried under vacuum to yield 4.83 g (83%) of 2-(2-(4-(methylthio)phenyl-

thio)phenyl)acetic acid **76** as a pink solid. ¹H NMR (CDCl₃): δ 2.43 (s, 3H), 7.12–7.35 (m, 8H).

(ii) PPA (50.0 g) was added to a solution of **76** (4.83 g, 16.6 mmol) in toluene (30 mL) and the reaction mixture was heated and refluxed for 18 h. Water (50 mL) was added, the organic layer was separated and the water layer was extracted with EtOAc (3×30 mL). The combined organic extracts were washed with 5% NaOH (50 mL) and brine (50 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give 3.50 g (77%) of 8-(methyl-thio)dibenzo[*b*,*f*]thiepin-10(11*H*)-one **77** as a brown oil. ¹H NMR (CDCl₃): δ 2.46 (s, 3H), 4.36 (s, 2H), 7.13–7.64 (m, 6H), 8.02 (d, *J* = 2.3 Hz, 1H).

(iii) A solution of **77** (3.50 g, 12.8 mmol) and NaBH₄ (1.01 g, 26.7 mmol) in MeOH (100 mL) was stirred overnight at room temperature. The reaction mixture was evaporated in vacuo, water was added (50 mL) and the water layer was extracted with Et₂O (3 × 25 mL). The combined organic extracts were washed with water (3 × 50 mL) and brine (50 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give 3.14 g (89%) of 8-(methylthio)-10,11-dihydrodibenzo[*b*,*f*]thiepin-10-ol **78** as a brown solid. ¹H NMR (CDCl₃): δ 2.44 (s, 3H), 3.31–3.49 (m, 1H), 3.69–3.78 (m, 1H), 5.49 (br s, 1H), 6.95–7.49 (m, 7H).

(iv) BF₃·(Et)₂O (1.60 mL, 12.6 mmol) was added drop wise to a solution of **78** (3.14 g, 11.4 mmol) and 2-bromoethanol (1.22 mL, 17.2 mmol) in toluene (30 mL) and the reaction mixture was stirred for 1 h. Water (25 mL) was added and the water layer was extracted with EtOAc (3×20 mL). The combined organic extracts were washed with water (25 mL), 5% NaOH (2×25 mL) and brine (25 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give 3.37 g of 11-(2-bromoethoxy)-2-(methylthio)-10,11-dihydrodibenzo[*b*,*f*]thiepine **79** as a brown oil. The crude product was used without further purification.

(v) Following method B using crude **79** (0.57 g, 1.49 mmol) and 4-(4-chlorophenyl)piperidin-4-ol **49** (0.21 g, 1.00 mmol) gave 88 mg of a light yellow oil. This was converted to the fumaric salt as described for **50** to give 101 mg (16%) of **80** as a white solid. ¹H NMR (CDCl₃/DMSO-*d*₆): δ 1.67–1.80 (m, 2H), 2.21–2.57 (m, 5H), 2.78–3.31 (m, 6H), 3.42–3.62 (m, 2H), 3.78–3.99 (m, 2H), 5.22–5.33 (m, 1H), 6.73 (s, 2H), 7.00–7.41 (m, 12H). ¹³C NMR (CDCl₃/DMSO-*d*₆): δ 15.53, 35.91, 38.84, 48.92, 49.19, 56.61, 64.57, 68.79, 77.22, 125.02, 125.61, 126.28, 127.43, 128.16, 129.64, 130.97, 131.39, 131.82, 132.58, 133.66, 134.94, 137.86, 138.80, 142.22, 146.10, 169.42. Anal. RP-HPLC *Ia*: *t*_R = 7.76 min (purity 9%), *IIa* (total run time 30 min): *t*_R = 19.48 min (purity 97%). HRMS (EI) *m/z* calcd for C₂₈H₃₀ClNO₂S₂: 511.1406; found: 511.1405.

6.1.11. General method for the synthesis of target compounds 41, 85–91

(i) 2,4-Dimethylbenzaldehyde (1.67 g, 10.0 mmol) and 1-indanone (**77**, R1 = H);1.33 g, 10.0 mmol) were added to a saturated solution of anhydrous K₂CO₃ in EtOH (25 mL).The resulting reaction mixture was stirred vigorously for 24 h, 37% HCl (2 mL) was added and the solvent was evaporated in vacuo. Water (50 mL) was added and the water layer was extracted with DCM (3×25 mL). The combined organic extracts were washed with brine (50 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo. The residue was recrystallized from EtOH to give 1.95 g (79%) of 2-(2,4-dimeth-ylbenzylidene)-2,3-dihydro-1*H*-inden-1-one as white crystals. ¹H NMR (CDCl₃): δ 2.33 (s, 3H), 2.43 (s, 3H), 3.95 (s, 2H), 6.98–7.11 (m, 2H), 7.31–7.60 (m, 3H), 7.80–7.95 (m, 2H).

(ii) A suspension of 2-(2,4-dimethylbenzylidene)-2,3-dihydro-1*H*-inden-1-one (1.14 g, 5.49 mmol) and 10% Pd/C (5 mg) in MeOH (10 mL) was hydrogenated with H₂ (atmospheric pressure) for 18 h. The reaction mixture was filtrated over Hyflo and the residue was washed with EtOH. The filtrate was evaporated in vacuo to give 2-(2,4-dimethylbenzyl)-2,3-dihydro-1*H*-inden-1-one in a quantitative yield. ¹H NMR (CDCl₃): δ 2.28 (s, 3H), 2.30 (s, 3H), 2.41–2.59 (m, 1H), 2.72–3.05 (m, 2H), 3.10–3.22 (m, 1H), 3.37 (dd, *J* = 10.4 Hz and 3.9 Hz, 1H), 6.81–7.01 (m, 3H), 7.25–7.41 (m, 2H), 7.54 (d, *J* = 7.5 Hz, 1H), 7.77 (d, *J* = 7.6 Hz, 1H).

(iii) A solution of 2-(2,4-dimethylbenzyl)-2,3-dihydro-1*H*-inden-1-one (1.15 g, 4.59 mmol) and NaBH₄ (0.17 g, 4.59 mmol) in MeOH (10 mL) was stirred overnight at room temperature. The reaction mixture was evaporated in vacuo, water was added (25 mL) and the water layer was extracted with EtOAc (3 × 20 mL). The combined organic extracts were washed with water (3 × 25 mL) and brine (25 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give a quantitative yield of 2-(2,4-dimethylbenzyl)-2,3-dihydro-1*H*-inden-1-ol as a 1:1 mixture of two diastereomers. ¹H NMR (CDCl₃): δ 1.70 (br s, 1H), 2.30 (s, 6H), 2.35–2.88 (m, 3H), 2.98–3.12 (m, 2H), 4.83–5.01 (m, 1H), 6.89–7.01 (m, 2H), 7.04–7.41 (m, 6H).

(iv) A solution of 2-(2,4-dimethylbenzyl)-2,3-dihydro-1*H*-inden-1-ol (1.15 g, 4.59 mmol) and DPPA (1.39 mL, 6.41 mmol) in THF (10 mL) was stirred for 10 min, cooled to 0 °C and DBU (0.98 mL, 6.42 mmol) was added in a drop wise manner. The reaction mixture was allowed to warm to room temperature and stirred for 18 h. The solvent was evaporated in vacuo, water (25 mL) was added and the water layer was extracted with Et₂O (3 × 15 mL). The combined organic extracts were washed with brine (25 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo. Purification by column chromatography (hexane) afforded 763 mg (60%) of 1-azido-2-(2,4-dimethylbenzyl)-2,3-dihydro-1*H*-indene as a 1:1 mixture of two diastereomers. ¹H NMR (CDCl₃): δ 2.29 (s, 3H), 2.30 (s, 3H), 2.51–2.84 (m, 4H), 2.90–3.11 (m, 1H), 4.52 and 4.70 (d, *J* = 4.7 and 5.0 Hz, 1H), 6.87–7.03 (m, 3H), 7.11–7.39 (m, 4H).

(v) A solution of 1-azido-2-(2,4-dimethylbenzyl)-2,3-dihydro-1*H*-indene (0.76 g, 2.74 mmol) and 10% Pd/C (3 mg) in MeOH (10 mL) was hydrogenated with H₂ (atmospheric pressure) for 3 days. The reaction mixture was filtrated over Hyflo, the residue was washed with EtOH and the filtrate was evaporated in vacuo. Purification by flash chromatography (2% Et₃N in hexane) gave 646 mg of the free base. This was dissolved in a solution of HCl gas in EtOH and the solvent was evaporated in vacuo. The hydrochloride salt was isolated by filtration and recrystallized from Et₂O/MeOH.

6.1.12. 2-(2,4-Dimethylbenzyl)-2,3-dihydro-1*H*-inden-1-amine hydrochloride (85)

Yield: 665 mg (84%) of **85** as a 1:1 mixture of two diastereomers. ¹H NMR (MeOH- d_4): δ 2.30 (s, 3H), 2.33 (s, 3H), 2.61–3.18 (m, 5H), 4.51 and 4.83 (d, *J* = 3.5 and 6.0 Hz, 1H), 6.92–7.15 (m, 3H), 7.21– 7.40 (m, 3H), 7.49–7.63 (m, 1H). ¹³C NMR (MeOH- d_4): δ 18.64, 20.02, 31.51, 35.25, 35.87, 43.08, 58.19, 125.44, 125.73, 126.78, 127.33, 129.29, 129.96, 131.25, 134.43, 136.07, 136.21, 138.73, 143.83 (one diastereomer). ¹³C NMR (MeOH- d_4): δ 17.38, 18.59, 31.51, 35.25, 35.87, 45.21, 60.52, 125.18, 125.31, 127.48, 129.66, 131.27, 134.49, 136.27, 136.30, 137.94, 143.58 (other diastereomer). Anal. RP-HPLC *Ib* (total run time 30 min): t_R = 12.89 min (purity 100%), *IIb*: t_R = 11.79, 12.79 min (purity 100%). HRMS (EI) *m/z* calcd for C₁₈H₂₁N: 251.1674; found: 251.1665.

6.1.13. 2-(2,4-Dimethoxybenzyl)-2,3-dihydro-1*H*-inden-1-amine hydrochloride (86)

Yield: 539 g (17% yield over five steps) of **86** as a 1:1 mixture of two diastereomers. ¹H NMR (MeOH- d_4): δ 2.58–3.14 (m, 5H), 3.78 and 3.79 (s, 3H), 3.81 and 3.82 (s, 3H), 4.43 and 4.69 (d, *J* = 4.1 and 5.8 Hz, 1H), 6.45–6.57 (m, 2H), 7.02–7.52 (m, 5H). ¹³C NMR (MeOH- d_4): δ 35.44, 35.79, 45.35, 54.82, 54.84, 54.91, 57.98, 98.55, 104.89, 119.62, 125.36, 125.41, 127.24, 129.95, 131.31, 138.83, 144.09, 158.89, 160.45 (one diastereomer). ¹³C NMR

(MeOH- d_4): δ 28.70, 32.75, 43.45, 54.82, 54.84, 54.91, 60.26, 98.50, 104.61, 119.77, 124.92, 125.59, 127.32, 129.80, 130.80, 138.12, 143.62, 158.54, 160.52 (other diastereomer). Anal. RP-HPLC *la* : $t_{\rm R}$ = 8.47 and 9.81 min (purity 100%), *IIIb*: $t_{\rm R}$ = 5.29 and 5.61 min (purity 100%). HRMS (EI) *m*/*z* calcd for C₁₈H₂₁NO₂: 283.1572; found: 283.1582.

6.1.14. 2-(2-Methoxybenzyl)-2,3-dihydro-1*H*-inden-1-amine hydrochloride (87)

Yield: 439 mg (17% yield over five steps) of one diastereomer of **87** as a white solid. ¹H NMR (MeOH-*d*₄): δ 2.52–2.80 (m, 2H), 2.85–3.15 (m, 3H), 3.85 (s, 3H), 4.71 (d, *J* = 5.9 Hz, 1H), 6.89–6.97 (m, 2H), 7.01–7.36 (m, 5H), 7.50 (d, *J* = 6.6 Hz, 1H). ¹³C NMR (MeOH-*d*₄): δ 30.37, 36.41, 44.21, 55.82, 59.00, 111.76, 121.82, 126.30, 126.43, 128.28, 128.54, 129.17, 131.01, 131.43, 139.80, 145.96, 158.71. RP-HPLC *Ic*: $t_{\rm R}$ = 4.19 min (purity 100%), *IIb*: $t_{\rm R}$ = 3.95 min (purity 100%). HRMS (EI) *m/z* calcd for C₁₇H₁₉NO: 253.1467; found: 253.1476.

6.1.15. 2-(4-Methoxybenzyl)-2,3-dihydro-1*H*-inden-1-amine hydrochloride (88)

Yield: 439 mg (19% yield over five steps) of **88** as a 1:1 mixture of two diastereomers. ¹H NMR (MeOH-*d*₄): δ 2.51–3.29 (m, 5H), 3.77 (s, 3H), 3.79 (s, 3H), 4.49 and 4.77 (d, *J* = 3.9 and 6.2 Hz, 1H), 6.80–6.94 (m, 2H), 7.08–7.52 (m, 6H). ¹³C NMR (MeOH-*d*₄): δ 35.89, 38.28, 46.58, 54.73, 60.39, 114.14, 125.03, 125.67, 127.44, 129.75, 130.06, 131.55, 137.93, 143.47, 158.92 (one diastereomer). ¹³C NMR (MeOH-*d*₄): δ 34.12, 35.33, 44.47, 54.73, 58.08, 114.11, 125.35, 125.46, 127.34, 129.92, 129.98, 131.49, 138.73, 143.82, 158.92 (other diastereomer). RP-HPLC *Ic*: $t_{\rm R}$ = 5.73 min (purity 100%), *IId*: $t_{\rm R}$ = 5.48 min (purity 96%). HRMS (EI) *m/z* calcd for C₁₇H₁₉NO: 253.1467; found: 253.1459.

6.1.16. 2-(4-Methoxy-3-methylbenzyl)-2,3-dihydro-1*H*-inden-1-amine hydrochloride (89)

Yield: 470 mg (20% yield over five steps) of one diastereomer of **89** as a white solid. ¹H NMR (MeOH- d_4): δ 2.42 (s, 3H), 2.42–3.09 (m, 5H), 3.80 (s, 3H), 4.77 (d, *J* = 6.0 Hz, 1H) 6.82–7.54 (m, 7H). ¹³C NMR (MeOH- d_4): δ 15.35, 34.14, 35.37, 44.48, 54.84, 58.09, 110.21, 125.30, 125.46, 126.72, 131.04, 127.05, 127.31, 129.96, 130.89, 138.78, 143.86, 156.94. Anal. RP-HPLC *Ib*: t_R = 4.56 min (purity 100%), *IIa*: t_R = 4.33 min (purity 99%). HRMS (EI) *m/z* calcd for C₁₈H₂₁NO: 267.1623; found: 267.1623.

6.1.17. *N*-(4-((1-Amino-2,3-dihydro-1*H*-inden-2-yl)methyl)phenyl)acetamide (90)

Yield: 581 mg (23% yield over five steps) of **90** as a 1:5 mixture of two diastereomers. (MeOH- d_4): δ 2.10 (s, 3H), 2.20–2.34 (m, 1H), 2.41–77 (m, 2H), 2.81–2.92 (m, 1H), 3.00–3.15 (m, 1H), 3.98 and 4.28 (d, *J* = 7.6 and 7.8 Hz, 1H), 6.98–7.51 (m, 8H). ¹³C NMR (MeOH- d_4): δ 22.73, 36.38, 38.92, 52.76, 61.76, 120.45, 123.50, 124.48, 126.56, 127.41, 129.28, 136.94, 137.22, 141.91, 146.11, 170.55 (major diastereomer). ¹³C NMR (MeOH- d_4): δ 22.73, 34.10, 35.19, 46.63, 58.46, 120.32, 124.23, 124.65, 124.77, 127.63, 129.17, 136.82, 137.34, 141.91, 146.11, 170.55 (minor diastereomer). RP-HPLC *le* (total run time 30 min): *t*_R = 10.42 min (minor diastereomer), 13.62 min (major diastereomer) (purity 100%), *Ilf*: *t*_R = 5.21 min (minor diastereomer), 6.22 min (major diastereomer) (purity 100%). HRMS (EI) *m/z* calcd for C₁₈H₂₀N₂O: 280.1576; found: 280.1571.

6.1.18. 2-Benzyl-2,3-dihydro-1*H*-inden-1-amine hydrochloride (91)

Yield: 350 mg (14% yield over five steps) of one diastereomer of **91** as a white solid. ¹H NMR (MeOH- d_4): δ 2.52–2.66 (m, 1H), 2.70–3.05 (m, 3H), 3.09–3.20 (m, 1H), 4.79 (d, *J* = 6.4 Hz, 1H), 7.25–7.55

(m, 9H). ¹³C NMR (MeOH- d_4): δ 35.03, 35.29, 44.29, 58.13, 125.29, 125.47, 125.61, 127.37, 128.73, 128.80, 130.02, 138.68, 139.59, 143.77. RP-HPLC *Ic*: t_R = 5.52 min (purity 100%), *IId*: t_R = 5.94 min (purity 97%), *IIIa*: t_R = 3.37 min (purity 99%). HRMS (EI) *m/z* calcd for C₁₆H₁₇N: 223.1361; found: 223.1363.

6.1.19. General method for the synthesis of target compounds 93–98

(i) 2,4-Dichlorobenzaldehyde (17.5 g, 100 mmol) and 1-indanone **81** (13.2 g, 100 mmol) were added to a saturated solution of anhydrous K_2CO_3 in EtOH (250 mL). The resulting reaction mixture was stirred vigorously for 18 h and 37% HCl (20 mL) was added. The precipitate was isolated by filtration, washed with water and dried under vacuum to give 25.8 g(89%) of 2-(2,4-dichlorobenzylid-ene)-2,3-dihydro-1*H*-inden-1-one as white crystals. ¹H NMR (CDCl₃): δ 3.92 (s, 2H), 7.12–7.66 (m, 5H), 7.80–7.99 (m, 2H).

(ii) A suspension of 2-(2,4-dichlorobenzylidene)-2,3-dihydro-1*H*-inden-1-one (11.6 g, 40.1 mmol) and 10% Pd/C (40 mg) in MeOH (100 mL) was hydrogenated with H₂ (atmospheric pressure) for 6 h at a temperature of 50 °C. The reaction mixture was filtrated over Hyflo and the filtrate was evaporated in vacuo. Recrystallization from EtOH afforded 4.6 g (39%) of 2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-one as white crystals. ¹H NMR (CDCl₃): δ 2.65–2.88 (m, 2H), 2.95–3.20 (m, 2H), 3.27–3.40 (m, 1H), 7.06–7.20 (m, 2H), 7.26–7.40 (m, 3H), 7.56 (t, *J* = 7.4 Hz, 1H), 7.75 (d, *J* = 7.6 Hz, 1H).

(iii) A solution of 2-(2,4-dichorolbenzyl)-2,3-dihydro-1*H*-inden-1-one (1.86 g, 6.40 mmol) and NaBH₄ (0.17 g, 4.59 mmol) in MeOH (40 mL) was stirred for 3 h at room temperature. The reaction mixture was evaporated in vacuo, water was added (20 mL) and the water layer was extracted with EtOAc (3×15 mL). The combined organic extracts were washed with water (3×25 mL) and brine (25 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give 1.84 (98%) of 2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-ol as a 1:1 mixture of two diastereomers. ¹H NMR (CDCl₃): δ 1.72 (br s, 1H), 2.41–3.02 (m, 5H), 4.92 (br s, 1H), 7.16–7.38 (m, 7H).

(iv) Thionyl chloride (0.75 mL, 10.3 mmol) was added to a solution of 2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-ol (1.84 g, 6.28 mmol) in toluene (30 mL) at 0 °C. The reaction mixture was stirred at room temperature for 30 min, heated to 55 °C and stirred for 1 h. The solution was cooled to room temperature, washed with ice-water (2 × 15 mL), dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to give 1.88 g (96%) of 1-chloro-2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-indene **92** as a 1:1 mixture of two diastereomers. ¹H NMR (CDCl₃): δ 2.55–3.29 (m, 5H), 5.10 and 5.27 (d, *J* = 5.3 and 4.8 Hz, 1H), 7.08–7.41 (m, 7H).

(v) A solution of piperidine (70 μ L, 0.71 mmol), NaI (75 mg, 0.50 mmol), and Na₂CO₃ (106 mg, 1.0 mmol) in CH₃CN (10 mL) was heated to reflux temperature and 1-chloro-2-(2,4-dichloroben-zyl)-2,3-dihydro-1*H*-indene **92** (0.16 g, 0.51 mmol) in CH₃CN (1 mL) was added. The reaction mixture was refluxed overnight, the solvent was removed in vacuo and the residue was diluted with water (15 mL) followed by an extraction with DCM (3 \times 10 mL). The combined organic layers were dried over anhydrous NaSO₄, filtered, evaporated in vacuo, and subsequently purified by flash chromatography (25% EtOAc in DCM) and recrystallized from EtOAc.

6.1.20. 1-(2-(2,4-Dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-yl)piperidine (94)

Yield: 90 mg (49%) of **94** as white crystals. ¹H NMR (CDCl₃): δ 1.29–1.51 (m, 6H), 2.29–2.70 (m, 6H), 2.76–2.99 (m, 3H), 3.95 (d, *J* = 3.4 Hz, 1H), 7.03–7.22 (m, 5H), 7.29–7.41 (m, 3H). ¹³C (CDCl₃): δ 24.61, 26.41, 37.00, 38.18, 38.80, 50.19, 76.04, 124.50, 125.88, 126.09, 126.65, 127.38, 129.12, 131.78, 132.15, 134.78, 137.23, 142.58. Anal. RP-HPLC *Ic* (total run time 30 min): $t_{\rm R}$ = 19.88 min (purity 100%), *Ilb* (total run time 30 min): $t_{\rm R}$ = 13.77 min (purity 100%). HRMS (EI) *m/z* calcd for C₂₁H₂₃Cl₂N: 359.1208; found: 359.1199.

6.1.21. 4-(4-Chlorophenyl)-1-(2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-yl)-1,2,3,6-tetrahydropyridine hydrochloride (97)

Yield: 50 mg (19%) of one diastereomer of **97** as a light brown solid. ¹H NMR (MeOH-*d*₄): δ 2.61–2.92 (m, 6H), 3.15–3.39 (m, 3H), 3.51–3.81 (m, 2H), 4.61–4.69 (m, 1H), 6.01–6.07 (m, 1H), 7.31–7.54 (m, 10H), 7.67 (d, *J* = 7.1 Hz, 1H). ¹³C (MeOH-*d*₄): δ 26.22, 37.74, 38.29, 40.69, 47.72, 47.83, 75.74, 118.08, 127.30, 127.70, 128.68, 128.96, 129.74, 130.48, 132.14, 134.00, 134.59, 135.07, 135.83, 136.27, 136.78, 138.51, 146.52. Anal. RP-HPLC *lb* (total run time 20 min): $t_{\rm R}$ = 13.04 min (purity 99%), *Ilb*: $t_{\rm R}$ = 6.78 min (purity 96%). HRMS (EI) *m*/*z* calcd for C₂₇H₂₄Cl₃N: 469.0945 (second isotope); found: 469.0931.

6.1.22. 4-(4-Chlorophenyl)-1-(2-(2,4-dichlorobenzyl)-2,3dihydro-1*H*-inden-1-yl)piperidin-4-ol hydrochloride (93)

Yield: 61 mg (23%) of one diastereomer of **93** as a white solid ¹H NMR (MeOH- d_4): δ 1.81–1.92 (m, 4H), 2.50 (br s, 1H), 2.70–3.12 (m, 5H), 3.15–3.36 (4H), 4.41–4.47 (m, 1H), 7.15–7.48 (m, 10H), 8.00 (s, 1H), 12.08 (br s, 1H). ¹³C (MeOH- d_4): δ 35.03, 35.26, 37.25, 37.94, 38.73, 44.31, 45.84, 46.37, 69.58, 75.31, 125.81, 126.00, 127.64, 127.97, 128.67, 128.76, 129.60, 131.01, 132.58, 132.80, 133.54, 133.63, 134.80, 134.83, 144.42, 144.51. Anal. RP-HPLC *Ib* : t_R = 6.19 min (purity 100%), *IIa* (total run time 30 min): t_R = 13.45 min (purity 100%). HRMS (EI) *m/z* calcd for C₂₇H₂₆Cl₃NO: 485.1080; found: 485.1073.

6.1.23. 1-(2-(2,4-Dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-yl)-4-phenylpiperazine dihydrochloride (96)

Yield: 48 mg (22%) of one diastereomer of **96** as a white solid. ¹H NMR (MeOH- d_4): δ 2.72–2.97 (m, 3H), 3.05–3.55 (m, 8H), 3.69–3.88 (m 2H), 4.64–4.72 (m, 1H), 6.88–7.08 (m, 2H), 7.19– 7.54 9 m, 9H), 7.74 (d, *J* = 7.2 Hz, 1H). ¹³C (MeOH- d_4): δ 37.56, 38.05, 40.90, 50.58, 76.79, 118.22, 122.92, 127.46, 128.73, 128.78, 129.44, 130.41, 130.51, 132.52, 133.79, 134.00, 134.66, 136.28, 136.55, 146.84, 150.63. Anal. RP-HPLC *Ib*: t_R = 7.07 min (purity 100%), *IIa*: t_R = 4.13 min (purity 98%), *IIb* (total run time 30 min): t_R = 19.11 min (purity 98%). HRMS (EI) *m*/*z* calcd for C₂₆H₂₆Cl₂N₂: 436.1473; found: 436.1471.

6.1.24. 1-(2-(2,4-Dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-yl)-4-methylpiperazine dihydrochloride (95)

Yield: 105 mg (46%) of one diastereomer of **95** as a light yellow solid. ¹H NMR (MeOH- d_4): δ 2.61–3.03 (m, 4H), 2.93 (s, 5H), 3.12–3.87 (m, 8H), 4.55–4.64 (m, 1H), 7.15–7.49 (m, 6H), 7.72 (d, *J* = 7.1 Hz, 1H). ¹³C (MeOH- d_4): δ 37.59, 38.05, 40.99, 43.36, 47.18, 51.93, 77.02, 127.37, 128.72, 129.35, 130.46, 132.33, 133.98, 134.55, 136.27, 136.76, 146.61. Anal. RP-HPLC *lb* (total run time 20 min): t_R = 12.99 min (purity 95%), *lld*: t_R = 12.70 min (purity 96%). HRMS (EI) *m/z* calcd for C₂₁H₂₄Cl₂N₂: 374.1317; found: 374.1311.

6.1.25. *N*-Benzyl-2-(2,4-dichlorobenzyl)-2,3-dihydro-1*H*-inden-1-amine fumarate (98)

Yield: 20 mg (8%) of one diastereomer of **98** as a white solid. ¹H NMR (MeOH- d_4): δ 2.69–2.79 (m, 2H), 2.89–2.95 (m, 1H), 3.02–3.08 (m, 1H), 3.17–3.25 (m, 1H), 4.14–4.21 (m, 2H), 4.62 (d,

J = 6.1 Hz, 1H), 6.68 (s, 2H), 7.27–7.53 (m, 12H). ¹³C NMR (MeOH*d*₄): δ 32.41, 36.00, 44.64, 51.56, 65.52, 126.53, 126.60, 127.91, 128.49, 129.71, 130.01, 130.30, 130.33, 130.51, 133.42, 134.08, 135.48, 135.89, 136.00, 137.77, 144.58, 170.70. Anal. RP-HPLC *lb*: *t*_R = 8.22 min (purity 100%), *lla*: *t*_R = 12.37 min (purity 97%). HRMS (ESI) *m/z* calcd for C₂₇H₂₅Cl₂NO₄: 381.1051; found: 381.1046.

6.2. Pharmacology

Pharmacological characterization of nonpeptidergic ligands was performed as previously described.¹¹

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