

PHOTOOXYGENATION OF 1,2-BIS(SILYLOXY)CYCLOALKENES

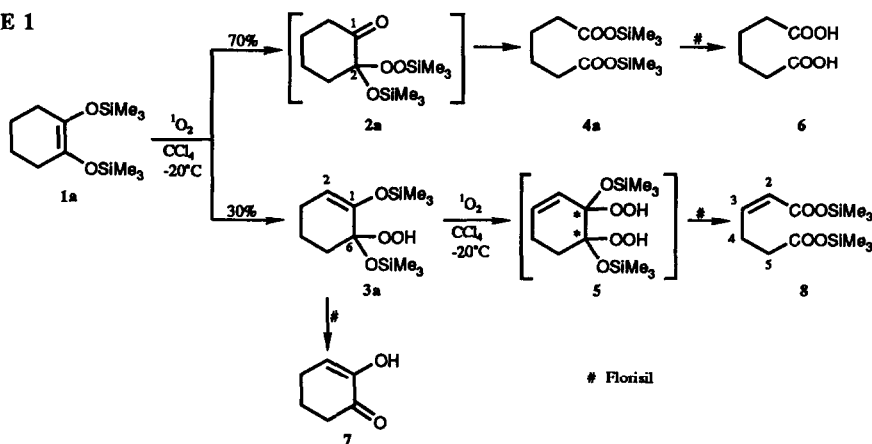
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Abstract: The reaction of singlet oxygen with O-silylated cyclic enediols **1a,b** afforded as ene products the hydroperoxy silyl enol ethers **3a,b** and as cleavage products the silyl esters **4a,b**; the latter presumably derived from rearrangement of the intermediary silylperoxy ketones **2a,b**.

Olefins containing allylic hydrogen react with singlet oxygen to produce ene products.¹ If the double bond is substituted with a silyloxy group, also silyl migration takes place, affording α -silylperoxy ketones.² In this paper we report on the photooxygenation of 1,2-bis(trimethylsilyloxy)cyclohexene (**1a**) and -cyclopentene (**1b**), showing that both the prototropic (hydrogen shift) and silatropic (silyl shift) ene reactions occur (Schemes 1, 2).

SCHEME 1

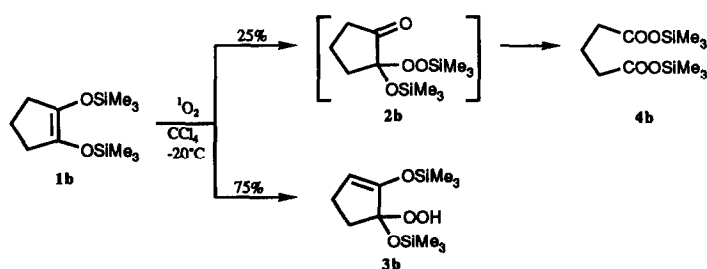


1,2-Bis(trimethylsilyloxy)cyclohexene **3** (**1a**) and 1O_2 in CCl_4 with TPP as photosensitizer at $-20^\circ C$ gave within 50 min the silatropic ene product **2a** ⁴ [characteristic ^{13}C signals: δ 205.0 (C-1), 104.1 (C-2)] and the prototropic ene product **3a** ⁵ [characteristic ^{13}C signals: δ 101.7 (C-6), 109.5 (C-2), 147.1 (C-1); characteristic 1H signals: δ 4.96 [s, 1H, C=CH], 8.6 (s, 1H, OOH)], the **2a** : **3a** ratio being 70 : 30. The α -silylperoxy cyclohexanone **2a** was quite unstable, but could be detected at subambient temperature ($-20^\circ C$) by 1H and ^{13}C NMR. When a sample of the product mixture was warmed up to $20^\circ C$ and further monitored by NMR, it was observed that the silatropic ene product **2a** rearranged gradually in a period of 1h completely into the cleavage product bis(trimethylsilyl) adipate (**4a**) ⁶, as evidenced in the ^{13}C NMR spectrum by the disappearance of the signals at δ 104.1 and 205.0 (specific for **2a**) and the appearance of the signal at δ 173.6 (specific for **4a**). During this period the signals of the prototropic ene product **3a** suffered no change.

On prolonged photooxygenation (ca. 3h, -20°C) NMR monitoring revealed new proton resonances at δ 5.5, 5.9, 9.0, 9.7, 10.1, and 10.2 (in the approximate relative ratios 2:2:1:1:1:1). These were assigned to the olefinic and hydroperoxy protons of the diastereomeric double ene products **5**. The solvent was removed by rotavaporation (0°C , 15 torr) and the residue chromatographed at -40°C on a Florisil column, eluting with petroleum ether / diethyl ether (9:1) to give as products the silyl adipate **4a**, adipic acid (**6**), the hydroxy-ene **7**, and the disilyl ester **8** in the relative proportion ca. 50 : 20 : 25 : 5, as well as hexamethylsiloxane. The physical constants and spectral data of these products, except those of the disilyl ester **8**, were identical with those of the authentic compounds. Although the ^1H NMR data of cleavage product **8** are not known, our spectral data, especially the chemical shifts δ 2.42 (t, $J_{5,4}=7.1\text{Hz}$, 2H, 5-H), 2.85 (tdd, $J_{4,3}=7.4\text{Hz}$, $J_{4,5}=7.1\text{Hz}$, $J_{4,2}=1.6\text{Hz}$, 2H, 4-H), 5.74 (dt, $J_{2,3}=11.4\text{Hz}$, $J_{2,4}=1.6\text{Hz}$, 1H, 2-H), and 6.19 (dt, $J_{3,2}=11.4\text{Hz}$, $J_{3,4}=7.4\text{Hz}$, 1H, 3-H), support the structure proposed for silyl ester **8**. On-column desilylation of disilyl adipate **4a** is a logical pathway to adipic acid (**6**), while desilylation of the hydroperoxy silyl enol ether **3a**, followed by loss of hydrogen peroxide, is a reasonable route to hydroxy-ene **7**. Disilyl ester **8** presumably arises via rearrangement of the novel double ene product **5** (Scheme 1). During NMR monitoring no dioxetane signals were observed.

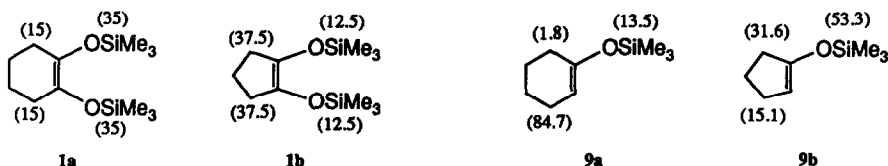
The photooxygenation of the disilyloxycyclopentene **1b**³ gave under the same conditions as employed for **1a** the bis(trimethylsilyl) glutarate (**4b**)⁷ and the hydroperoxy silyl enol ether **3b**⁸, as detected by ^1H and ^{13}C NMR at -20°C (Scheme 2). The structures of **3b** and **4b** were confirmed by comparison of the ^1H and ^{13}C NMR spectra with those of compounds **3a** and **4a**.

SCHEME 2



The differences in the behavior of the cyclopentene substrate **1b** versus the cyclohexene derivative **1a** towards singlet oxygen are: i) no silyl peroxy ketone **2b** was observed during NMR monitoring and ii) the proportion of **4b** to **3b** was ca. 25 : 75 compared to 70 : 30 for **4a** to **3a**, indicating that the prototropic ene reaction dominates for the cyclopentene system. Since five-membered ring-annulated 1,2-dioxetanes are more stable than six-membered analogues,⁹ it is unlikely that the disilyl diester **4a** arose from cleavage of the corresponding intermediary dioxetane because it should have survived subambient NMR monitoring. Moreover, since the disilyl diester **4a** resulted exclusively from rearrangement of the intermediary silylperoxy ketone **2a** (Scheme 1), it is more probable that also **4b** came from the corresponding silylperoxy ketone **2b**, except that the latter was too labile for NMR detection.

Accepting this hypothesis, it is of interest to compare the present prototropic (hydrogen migration) versus silylative (silyl migration) ene reactivities of the disilyloxycycloalkenes **1a,b** with those reported²ⁱ for the monosilyloxycycloalkenes **9a,b**. In view of the symmetric nature of the **1a,b** substrates, for better comparison



the observed proto- and silatropic ene reactivities have been divided equally between the two possible sites. Ignoring the regioselectivities exhibited for the ring protons of the unsymmetrical substrates **9a,b**, clearly a pronounced difference exists in the proto- versus silatropic ene reactivities between the disilyloxy and monosilyloxy sets **1a,b** and **9a,b**. For the latter, hydrogen migration dominates in the six-membered ring **9a** and silicon migration for the five-membered ring **9b**, while the reverse obtains for **1a,b**, i.e. silatropic ene reactivity is preferred for the six-membered ring **1a** and prototropic for the five-membered ring **1b**. This result is in keeping with the well established *cis*-effect,¹⁰ as exemplified by the pronounced exocyclic prototropic ene reaction for 1-methylcyclohexene but overwhelming endocyclic hydrogen abstraction for 1-methylcyclopentene.¹¹ Conformational control is responsible, in that for the cyclopentene the better perpendicular alignment of the allylic hydrogens relative to the molecular plane provides for a more favorable peroxide-like complexation, a requirement for the *cis*-effect.¹⁰

The rearrangement of the intermediary α -silylperoxy ketone **2a** into the cleavage product **4a** (and presumably also of **2b**, which was postulated in the scission of **1b** into **4b** in Scheme 2) is analogous to the Hock-Criegee rearrangement^{1, 12} of a variety of peroxides, except that here the silyl group rather than protonation promotes the 1,2-acyl shift. It is significant that such transpositions do not occur for the simpler α -silylperoxy ketones derived from the photooxygenation of the silyl enol ethers **9a,b**.²¹ Apparently the presence of the peroxy ketal moiety at C-2 in **2a** (Scheme 1) is a necessary condition for such rearrangements, as proposed in the enzymatic conversion of analogous catechol derivatives into the corresponding muconic acids by means of dioxygenases.^{13,14}

Although our initial goal of preparing the 1,2-dioxetanes of the 1,2-bis(trimethylsilyloxy)cycloalkenes **1a,b** via photooxygenation forfeited, and such dioxetanes are still challenging target molecules, novel rearrangements in the reaction of singlet oxygen with disilylated cyclic ene-1,2-diols were uncovered. The fact that silyloxy groups undergo ene reactions with ¹O₂ opens up interesting possibilities for synthetic applications and mechanistic studies.

Acknowledgements

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 1a: ^1H NMR (200 MHz, CDCl_3), δ 0.12(s, 18H), 1.56(m, 4H), 2.01(m, 4H);
 ^{13}C NMR (50 MHz, CDCl_3), δ 0.9(q), 23.2(t), 29.5(t), 132.0(s).
 1b: ^1H NMR (200 MHz, CDCl_3), δ 0.14(s, 18H), 1.73(m, 2H), 2.20(m, 4H);
 ^{13}C NMR (50 MHz, CDCl_3), δ 0.9(q), 16.6(t), 30.0(t), 130.2(s).
4. 2a: ^1H NMR (200 MHz, CDCl_3 , -20°C), δ 0.11(s, 9H), 0.17(s, 9H), 1.5 - 2.4(m, 6H), 2.74(m, 2H);
 ^{13}C NMR (50 MHz, CDCl_3 , -20°C), δ -1.0(q), 2.3(q), 22.5(t), 27.5(t), 39.3(t), 39.8(t), 104.1(s),
 205.0(s).
5. 3a: ^1H NMR (200 MHz, CDCl_3 , -20°C), δ 0.13(s, 9H), 0.19(s, 9H), 1.5 - 2.4(m, 6H),
 4.96(t, $J=4\text{Hz}$, 1H), 8.56(s, 1H);
 ^{13}C NMR (50 MHz, CDCl_3 , -20°), δ 0.4(q), 1.6(q), 19.7(t), 24.1(t), 33.6(t), 101.7(d), 109.5(s),
 147.1(s).
6. 4a: ^1H NMR (200 MHz, CDCl_3), δ 0.24(s, 18H), 1.59(m, 4H), 2.28(m, 4H);
 ^{13}C NMR (50 MHz, CDCl_3), δ -0.2(q), 24.4(t), 35.5(t), 173.6(s).
7. 4b: ^1H NMR (200 MHz, CDCl_3 , -20°C), δ 0.23(s, 18H), 2.20(m, 2H), 1.75(m, 4H);
 ^{13}C NMR (50 MHz, CDCl_3 , -20°C), δ -0.2(q), 19.6(t), 34.4(t), 173.6(s).
8. 3b: ^1H NMR (200 MHz, CDCl_3 , -20°C), δ 0.14(s, 9H), 0.19(s, 9H), 1.84(t, $J = 6\text{ Hz}$, 2H),
 2.36(m, 2H), 4.90(t, $J = 2\text{ Hz}$, 1H), 8.9(s, 1H);
 ^{13}C NMR(50 MHz, CDCl_3 , -20°C), δ 0.0(q), 1.6(q), 23.0(t), 33.7(t), 108.2(d), 111.3(s), 150.6(s).
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