## Total Synthesis of Grahamimycin A<sub>1</sub>

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(3R)- and (3S)-3-Hydroxybutanoates were respectively converted into (2E,5R)- and (2E,5S)-5-t-butyldimethylsiloxy-2-hexenoic acids, corresponding to the O-1–C-6 fragment of grahamimycin  $A_1$  (1). The O-7–C-14 fragment [(4S,5S,7R)- or (4R,5R,7R)-7-hydroxy-4,5-isopropylidenedioxyoctanoate] was synthesized from 4,6-dideoxy- $\alpha$ -D-xylo-hexopyranoside. Condensation of the both fragments, followed by deprotection at the terminal hydroxyl- and carboxyl-functions afforded the seco acids of the precursors of 1. While the reaction of the seco acids having 13S-configuration with diethyl azodicarboxylate and triphenylphosphine afforded the corresponding lactones in low yields, macrolactonization of the seco acid having 13R-configuration by Yamaguchi procedure gave the desired lactone in 56% yield. Deprotection of vic-diol moiety of the lactone indicated the completion of relay synthesis of 1.

Macrocyclic dilactones can be divided into unsymmetrical macrodiolides such as grahamimycins  $A_1$  (1), 1) A (2), 2) B (3), 2) and colletodiol (4) 3) and  $C_2$ -symmetrical macrodiolides representatives pyrenophorin (5), 4) vermiculine (6), 5) conglobatin (7), 6) and elaiophylin (8), 7) (Scheme 1). The synthetic study of the latter class of compounds has been extensively carried out since 1970s and a variety of approaches have been reported. 4–7) On the other hand, unsymmetrical macrodiolides went largely unnoticed until 1980 when grahamimycin  $A_1$  (1) isolated from the fungus Cytospora was reported to exhibit significant antibacterial activity against a variety of pathogenic microorganisms. 1a)

The structural elucidation of the former class of compounds has been mainly established in past decade. Thus, the absolute configuration of colletodiol (4) originally proposed by MacMillan and Simpson,<sup>3d)</sup> has been revised by Seebach and co-workers based on X-ray crystallographic analysis in 1981.<sup>3c)</sup> The structure of grahamimycin A (2) and B (3) has been determined by Gurusiddaiah and Ronald in 1981.<sup>2)</sup> In 1982, Seebach established the absolute configuration of grahamimycin A<sub>1</sub> (1) through the total synthesis of (S, S)-(+)-grahamimycin A<sub>1</sub>, unnatural inactive enantiomer of 1.<sup>1b)</sup> (S,S)-(+)-Grahamimycin A<sub>1</sub> has also been prepared by Ghiringhelli in 1983.<sup>1f)</sup>

Confirmation of the absolute structures of colletodiol and grahamimycin  $A_1$  reveals that the two macrolides could be correlated by redox reactions. Thus, oxidation of the vic-diol system (C-11 and C-12) and reduction of C-9–C-10 double bond of colletodiol formally yields grahamimycin  $A_1$ .

The total synthesis of (R,R)-(-)-grahamimycin  $A_1$  (1) has been reported by Hillis and Ronald in  $1985^{1c}$  and Bestmann and Schobert in 1987, while colletodiol (4) was synthesized by Seebach et al. (1984), so Keck et al. (1989), and by us (1984). We wish to report herein relay synthesis of grahamimycin  $A_1$  (1).

One of the most crucial problems in the synthesis of

macrolide antibiotics and analogues is selection of enantiomerically pure starting materials having a suitable constitution and a functional group pattern which are readily incorporated into the chiral target structure.<sup>8)</sup> Seidel and Seebach have utilized 3-(S)-hydroxybutanal (9a) and (S)-methyloxirane (10a) as starting materials for the preparation of the O-1-C-6 fragment and the O-7-C-14 fragments of (S,S)-grahamimycin  $A_1$ . 1b) 3-(S)-Hydroxybutanal was used by Ghiringhelli in his synthesis of (S,S)-grahamimycin- $A_1$ . (R)-Methyloxirane (10b) has been correlated to the C-2 and C-8 asymmetric carbon atoms of (R,R)-grahamimycin  $A_1$  by Hillis and Ronald, 1c) while Bestmann and Schobert have selected 3-(R)-hydroxybutanal (9b) as common chiral source of both asymmetric centers of grahamimycin  $A_1^{1d}$  (Scheme 2). 3-(S)- and 3-(R)-Hydroxybutanal can be readily prepared by the reduction of the corresponding 3-hydroxybutanoates.9)

Grahamimycin A<sub>1</sub> has only two asymmetric carbon atoms, the structure being unique in containing vicdiketo group which could be formally derived from the corresponding vic-diol system. Although it was difficult to estimate in a priori fashion which macrocyclization sequence would be more successful for the construction of grahamimycin A<sub>1</sub> skeleton, we selected the preparation and union of two appropriately protected hydroxy acids.<sup>10)</sup> Scheme 3 represents the retrosynthetic analysis and strategic bond disconnections defining the key compounds 13 and 14 for an eventual total synthesis of 1. In view of successful synthesis of colletodiol, 3f) the bond forming of both fragments would be accomplished by esterification (O-1-C-2; grahamimycin A<sub>1</sub> numbering) and macrolactonization (O-7-C-8 or C-6-O-7; grahamimycin  $A_1$  numbering).

Apart from the strategies for constructing carbonchain framework with appropriate sense of chirality, the choice of protecting groups is also an important problem for the success of the synthesis. In order to avoid the difficulty of protecting group manipulation experienced in the synthesis of colletodiol,<sup>3f)</sup> t-butyldi-

Scheme 1.

OR 
$$H_3C$$
  $H_3C$   $H_3C$ 

methylsilyl (TBDMS) group and 2-(p-tolylsulfonyl)ethyl (PTSE) group were chosen as protecting groups for hydroxyl function of **13** and carboxyl function of **14** so as to remove simultaneously after connection of both segments.<sup>11)</sup>

(2E,5R)-5-Hydroxy-2-hexenoic acid (13a) and its enantiomer (13b) would be respectively prepared from readily available (R)- and (S)-3-hydroxybutanoic acids (15a and 15b) which have been utilized as chiral building blocks in synthesis of a variety of natural products.  $^{8a,9)}$ 

4,5,7-Trihydroxyoctanoic acids (14) would be generated from polyoxygenated substrates such as carbohydrates. The sense of chirality at the C-7 of 14 coincides with C-5 of D-aldohexose; hence hydroxyl groups at C-2 and C-3 of the hexose can be correlated with C-4 and C-5 of 14. Thus, deoxygenation at the positions 4 and 6 of the hexose and two carbon elongation at C-1 affords 14. In order to asses the any effect of the configuration at the C-2 and C-3 on the macrolactonization, 2-(p-tolylsulfonyl)ethyl (4S,5S,7R)- and (4R,5R,7R)-7-hydroxy-4, 5-isopropylidenedioxyoctanoates (14a, 14b) were prepared from xylo-hexopyranose and arabino-hexopyranose derivatives, respectively. The advantage of the use of D-hexose as starting material is taken of the fact that the structure of the D-hexose is independent to the target molecule because the hydroxyl groups at C-2 and C-3 are oxidized to vic-diketo functionality in the course of the synthesis. We postponed the diketone formation to the last step of the synthesis.

## Results and Discussion

Preparation of O-1-C-6 Fragment. The O-1-C-6 fragment, (2E,5R)- or (2E,5S)-5-t-butyldimethylsiloxy-2-hexenoic acid (17a, 17b), was synthesized from (3R)- or (3S)-3-hydroxybutanoic acid ester (18a, 18b). Thus, ethyl (R)-3-hydroxybutanoate (18a) prepared from poly(3-hydroxybutanoic acid) by known procedure<sup>9a,9b)</sup> reacted with t-butyldimethylsilyl (TBDMS) chloride to give the corresponding TB-DMS ether 19a in 89% yield. 12a) Reduction of 19a with diisobutylaluminium hydride (DIBAH)<sup>12b)</sup> and subsequent oxidation of the resulting alcohol 20a by pyridinium chlorochromate (PCC) in the presence of AcONa afforded the corresponding aldehyde 21a in 69% vield. 13,14a) Aldehyde 21a reacted with methoxycarbonylmethylenetriphenylphosphorane (22) giving methyl (2E,5R)-5-t-butyldimethylsiloxy-2-hexenoate (23a) in 74% yield along with the corresponding Zisomer (23a'). Hydrolysis of 23a with LiOH (0.1 M,  $1 \text{ M}=1 \text{ mol dm}^{-3}$ ) in THF-H<sub>2</sub>O (1:1) at room temperature proceeded smoothly to give (2E,5R)-5-t-buthyldimethylsiloxy-2-hexenoic acid (17a) in nearly quantitative yield (Scheme 4).

By a similar manner, t-butyl (S)-3-hydroxybutanoate (18b) gave TBDMS ether 19b which was converted into (3S)-3-(t-butyldimethylsiloxy)butanal (21b) in 67% yield via alcohol 20b. Alternatively, when 19b reacted with DIBAH in toluene at  $-70^{\circ}$ C for 3 h, aldehyde 21b was isolated in 76% yield. The reaction of 21b with 22 gave 23b and 23b' in 54 and 10% yields, respectively. Saponification of 23a under the conditions described above gave (2E,5S)-5-t-butyldimethylsiloxy-2-hexenoic acid (17b) in quantitative yield (Scheme 5).

Synthesis of O-7–C-14 Fragment. The O-7–C-14 fragments, 2-(p-tolylsulfonyl)ethyl esters of (4S, 5S, 7R)- and (4R, 5R, 7R)-7-hydroxy-4,5-isopropylidenedioxyoctanoates (24a, 24b) were synthesized from 4,6-dideoxy- $\alpha$ -D-xylo-hexopyranoside (25). Hydrolysis of 25 afforded the correspond-

rt, 18 h, quant. CH<sub>3</sub> 17a

0.1M LiOH in THF-H<sub>2</sub>O (1:1)

Scheme 4.

ing free sugar **26a** in 84% yield. The reaction of **26a** with 2-(p-tolylsulfonyl)ethoxycarbonylmethylenetriphenylphosphorane (**27**),<sup>16</sup> followed by acetonidation afforded 2-(p-tolylsulfonyl)ethyl (2E, 4S, 5S,7R)-7-hydroxy-4,5-isopropylidenedioxy-2-octenoate (**28a**) in 41% yield from **26a**. Hydrogenation of **28a** afforded the desired O-7-C-14 fragment **24a** in quantitative yield (Scheme 6).

In order to prepare appropriately protected (4R,5R,7R)-4,5,7-trihydroxyoctanoate **24b**, the configurations at the C-2 and C-3 of **25** were inverted via 2,3-anhydropyranoside. To sylation of **25** afforded 2-O-tosyl derivative **29** and 3-O-tosyl derivative **29**' in 68% yield in a ratio of 5.8:1. The mixture reacted with MeONa in methanol to yield methyl 2,3-anhydro- $\alpha$ -D-lyxo-hexopyranoside (**30**) in 50% isolated yield. The compound **30** was treated with 1 M KOH under reflux for 2 h, the de-

sired 4,6-dideoxy- $\alpha$ -D-arabino-hexopyranoside (31) being obtained in 91% yield without any detectable formation of 25. Following the procedure described in the preparation of 24a, 31 was converted into 2-(p-tolyl-sulfonyl)ethyl (2E,4R,5R,7R)-7-hydroxy-4,5-isopropylidenedioxy-2-octenoate (24b) in 12% overall yield from 31 (Scheme 6).

Synthesis of Seco Acids. In the course of our synthetic study of colletodiol and related compounds, intermolecular dehydration between  $\alpha,\beta$ -unsaturated carboxylic acid and long chain secondary alcohol with reactive esterification reagents was studied. Thus, when 2-butenoic acid or 2-pentenoic acid was allowed to react with 2-octanol and 1-ethyl-2-bromopyridinium tetrafluoroborate in the presence of tributylamine, the corresponding 1-methylheptyl 3-alkenoate 32 or 33 was obtained in 51 or 70% yield, respectively, rather than

expected  $\alpha$ ,  $\beta$ -unsaturated esters. The migration of the double bond could be explained by assuming the intermediacy of ketene **34** (Scheme 7).<sup>18,19)</sup> Although the reaction would be utilized for the preparation of  $\beta$ , $\gamma$ -unsaturated carboxylic ester from a thermodynamically more stable  $\alpha$ , $\beta$ -conjugated acid, the procedure could not be applicable to the present purpose.

When (2E,5S)-5-tetrahydropyranyloxyhexenoic acid (35) reacted with t-butyl (4R,5R,7R)-7-hydroxy-4, 5-isopropylidenedioxyoctanoate (36) under Stegrich conditions [dicyclohexylcarbodiimide (DCC; 1.5 molar amount) with catalytic amount of 4-dimethylaminopyridine (DMAP)],<sup>20)</sup> the expected ester 37 was obtained in 27% yield along with considerable amount of N-acylurea (38) (Scheme 7).<sup>18,21)</sup>

In the condensation of unreactive  $\alpha,\beta$ -unsaturated carboxylic acid with bulky secondary alcohol, therefore, the reaction system must be chosen such that the intermediary acylating reagent formed do not collapse to less reactive species under the reaction conditions used.

The reaction system of choice was Yamaguchi method,<sup>22)</sup> because this procedure was successfully utilized in a variety of macrolactonization under rather drastic conditions (toluene reflux for several hours). Thus, **17a** was treated with 2,4,6-trichlorobenzoyl chloride in the presence of triethylamine in THF at room temperature for 2 h, filtered, and the filtrate was evaporated. The resulting mixed anhydride was allowed to react with **24a** in the presence of DMAP in toluene at room temperature for 24 h to afford protected seco acid (**39a**) in 50% yield. Under the same conditions, fully protected seco acids **39b** and **39c** were prepared in 95 and 85% yields, respectively (Scheme 8).

Although the protecting groups of the carboxyl and hydroxyl functionalities of 39a, 39b, or 39c would

be removed by a single operation, the reactivities toward fluoride ion are different. Thus, the silyl ether of secondary alcohol is hardly cleaved under the conditions where PTSE group is completely removed [3 molar amount of tetrabutylammonium fluoride (TBAF) in THF, 0°C, 1 hl.23) Metcalf et al. have demonstrated that LiBF4 is effective for the cleavage of t-butyldimethylsilyl (TBDMS) secondary alkyl ether.<sup>24)</sup> Thus, fully protected seco acid 39c was treated with LiBF<sub>4</sub> (1.5 molar amount) in THF at room temperature for 25 h where the silyl group was completely removed but PTSE group still remained. Therefore the resulting mixture was treated with TBAF (3 molar amount) for 18 h at room temperature to give seco acid 40c and 2-(p-tolylsulfonyl)ethyl ester 41 in 50 and 50% yields, respectively (Table in Scheme 9; Entry 1). When **39c** was treated with TBAF (1.5 molar amount) for 1.5 h to remove PTSE group and any silyl group, followed by LiBF<sub>4</sub>(3 molar amount), it took 64 h to complete removal of the remaining silvl group (Table in Scheme 9; Entry 2). It was found that both PTSE group and TBDMS group simultaneously removed when **39c** was treated with TBAF (3 molar amount) in THF at room temperature for 64 h to afford the desired seco acid 40c in nearly quantitative yield (Table in Scheme 9; Entry 3). Seco acids **40a** and **40b** were also synthesized by the treatment of 39a and 39b with TBAF in THF at room temperature for 72 h (Table in Scheme 9; Entries 4 and 5). In these experiments, no ester bond cleavage could be detected. $^{25)}$ 

**Lactonization.** Since the configuration of the reaction site of seco acids  $\mathbf{40a}$  and  $\mathbf{40b}$  is S, the configuration of the chiral center must be inverted. This event would be carried out by the reaction of the seco acid with diethyl azodicarboxylate (DEAD) and triphenyl-

Scheme 6.

phosphine (TPP) where lactonization with inversion at the chiral center of the seco acid could be expected. Thus, the lactonization of the appropriately protected seco acid of colletodiol with 4R,5R,7R,13S configuration (42) by the use of DEAD and TPP afforded the fully protected colletodiol 43 in 45% yield after recrystallization (Scheme 10). 3f)

In view of the fact, **40a** was allowed to react with DEAD and TPP in toluene at  $-10^{\circ}$ C for 24 h and then at room temperature for 42 h in which lactone **44a** was obtained but the yield was unexpectedly low (12% isolated yield; Scheme 11). The difference between **40a** and **42** is the partial structure of C-2-C-5 segments. Thus, in the hope to obtain any information about the effect of configurations at C-4 and C-5 on lactonization, the reaction of **40b** with DEAD and TPP was attempted but the yield of the desired lactone **44b** was again low (4% isolated yield) (Scheme 11).

Although no side products could be isolated and identified in the lactonization of **40a** and **40b**,  $\beta$ -elimination would be the most likely side reaction.<sup>26)</sup> This assumption would be supported by the reaction of 2-

(p-tolylsulfonyl)ethyl (S)-5-hydroxyhexenoate (45) with (4S,5S,7R)-octanoic acid (46), DEAD, and TPP where 2-(p-tolylsulfonyl)ethyl 2,4-hexadienoic acid was obtained in 46% yield rather than expected ester 47 (Scheme 12). $^{27,28}$ )

The successful lactonization of **42** suggests that the hydroxyl group and carboxyl group of **42** are brought so close together as to facilitates intramolecular lactonization. In contrast, structural change at the C-2 and C-3 separates the reaction sites of seco acids **40a** and **40b** causing lactonization difficult at least at low temperature. At the present stage of investigation, however, the principal factor decisive in the ease of lactonization is not clear.<sup>29)</sup>

Based on the consideration described above, in order to cyclize a conformationally rigid substrate, the lactonization procedure must be selected such that the reaction could be carried out at elevated temperature. Thus, lactonization by Yamaguchi procedure was next examined. Yamaguchi esterification proceeds with retention of configuration at the hydroxyl functions, seco acid **40c** was treated with 2,4,6-trichlorobenzoyl chlo-

Scheme 7.

**39a**: 4S,5S,13S (50%) **39b**: 4R,5R,13S (95%) **39c**: 4S,5S,13R (86%)

Scheme 8.

Entry	substrate	conditions	product(s) / yield
1	39c	1) LiBF <sub>4</sub> (1.5 molar amount), THF, rt, 25 h 2) TBAF (3 molar amount), THF, rt, 18 h	<b>40c</b> /50% and <b>41</b> /50 %
2		1) TBAF (1.5 molar amount), THF, rt, 1.5 h 2) LiBF <sub>4</sub> (3 molar amount), THF, rt, 64 h	<b>40c</b> /quant.
3		TBAF (3 molar amount), THF, rt, 64 h	40c/quant.
4	39a	TBAF (3 molar amount), THF, rt, 72 h	<b>40a</b> /quant.
5	39b	TBAF (3 molar amount), THF, rt, 72 h	40b/quant.

Scheme 10.

Scheme 11.

ride and triethylamine in THF at room temperature for 2 h. Triethylamine hydrochloride was filtered off and the filtrate was diluted with toluene. The resulting solution was added dropwise to toluene containing DMAP (7 molar amount) over a period of 5.5 h at 95°C and the mixture was stirred for 30 min at this temperature to give lactone  $\bf 44a$  in 56% yield (Scheme 13). Methanolysis of the acetonide group of  $\bf 44a$  gave diol  $\bf 48$  in 78% yield. Since conversion of  $\bf 48$  into grahamimycin  $\bf A_1$  has already been reported by Bestmann and Schobert, 1d) the preparation of  $\bf 48$  indicate completion of relay synthesis of grahamimycin  $\bf A_1$  (Scheme 13).

## Experimental

General. Melting points were measured on a micro melting point apparatus (Yanagimoto Seisakusyo) and were uncorrected. Nuclear magnetic resonance (NMR) spectra were recorded on JEOL-GX270 spectrometer in CDCl<sub>3</sub>. Chemical shifts were reported in ppm relative to tetramethylsilane (0 ppm) as the internal standard. For the compounds having t-butyldimethylsilyl group, CHCl<sub>3</sub> (7.26 ppm) in CDCl<sub>3</sub> was used as the internal standard. Data were reported as follows: chemical shift (multiplicity, integrated intensity, coupling constants). Following abbreviation are used for spin multiplicity; s=singlet, d=doublet. t=triplet, q=quartet, qu=quintet, sx=sextet, br=broad, m=multiplet. Mass spectra were recorded on a JEOL-JMS-SX102 mass spectrometer. Optical rotations were measured on a JASCO-DIP-370 photoelectric polarimeter. Silica gel column chromatography were performed on Merck Kieselgel 60 (Art 7734) or Wakogel C-300. In general, organic solvents were purified and dried by the appropriate procedure. Air and moisture sensitive reactions were carried out under  $N_2$  or Ar atmosphere. t-Butyl 3-(S)-hydroxybutanoate was obtained from Chisso Co., Ltd. Poly-(R)-3-hydroxybutanoic acid was obtained commercially from Fluka.

Methoxycarbonylmethylenetriphenylphosphorane (22). This compound was prepared according to the previously outlined procedure  $^{30)}$  in 41% yield (11.2 g): Mp 170—173°C. Lit,  $^{30b)}$  mp 169—169.5°C.

2- (p- Tolylsulfonyl)ethoxycarbonylmethylenetriphenylphosphorane (27). This compound was prepared according to the reported procedures. 16,31)

Ethyl (*R*)-3- Hydroxybutanoate (18a). This compound was prepared by reported procedure. <sup>9a,9b)</sup> Bp 66° C/8 mmHg (1 mmHg =133.322 Pa),  $[\alpha]_D^{22}$  -44.1° (*c* 1.10, CHCl<sub>3</sub>); Lit, <sup>9a)</sup> bp 86—86.5° C/22 mmHg,  $[\alpha]_D^{21}$  -43.4° (*c* 1.36, CHCl<sub>3</sub>).

Ethyl (*R*)-3-*t*-Butyldimethylsiloxybutanoate (19a). A solution of 18a (3.81 g, 28.9 mmol), *t*-butyldimethylsilyl chloride (5.25 g, 34.7 mmol), and imidazole (4.97 g, 73.0 mmol) in dimethylformamide (70 ml) was stirred at room temperature for 72 h. To the reaction mixture was added water and ether and phases were separated. The ether layer was washed with saturated aqueous NaHCO<sub>3</sub> solution, saturated aqueous NaCl solution, dried with MgSO<sub>4</sub>, and concentrated. The residue was distilled under reduced pressure (75°C/3—4 mmHg) to afford 19a in 89% yield (6.32 g):  $[\alpha]_{\rm D}^{18} - 26.9^{\circ}$  (*c* 1.02, CHCl<sub>3</sub>); <sup>1</sup>HNMR  $\delta$ =4.27 (dqd, 1H, C3–H, J=7.4, 6.3, 5.6 Hz), 4.12 (qd, 2H, Et–CH<sub>2</sub>, J=7.1, 1.9 Hz), 2.47 (dd, 1H, C2–H, J=14.4 Hz), 2.35 (dd, 1H, C2–H'), 1.26 (t, 3H, Et–CH<sub>3</sub>), 1.19 (d, 3H, C4–H), 0.86 (s, 9H, *t*-Bu), 0.06 (s, 3H, Si–CH<sub>3</sub>), 0.04 (s, 3H, Si–CH<sub>3</sub>').

(R)-3-t-Butyldimethylsiloxy-1-butanol (20a). a stirred solution of 19a (4.68 g, 19 mmol) in toluene was added 1 M diisobutylaluminium hydride in dichloromethane (47.5 ml, 47.5 mmol) at  $-10^{\circ}$ C over 1.5 h and the mixture was stirred at  $-10^{\circ}$ C for 1.5 h. The reaction mixture was quenched with aqueous saturated sodium tartrate solution and filtered. The filtrate was extracted with dichloromethane and dichloromethane layer was dried with  ${\rm MgSO_4}$  and concentrated. The residue was distilled under reduced pressure  $(82^{\circ}C/1.2 \text{ mmHg})$  to afford **20a** in 76% yield (2.74 g):  $[\alpha]_{\rm D}^{18} - 30.1^{\circ}$  (c 1.09, CHCl<sub>3</sub>). <sup>1</sup>H NMR  $\delta = 4.06$  (qud, 1H, C3-H, J=4.4, 6.3 Hz), 3.78 (dddd, 1H, C1-H, J=10.8, 4.6, 6.6 Hz), 3.67 (dddd, 1H, C1-H', J=7.7, 4.7 Hz), 2.80 (t, 1H, C1–OH, J=5.0 Hz), 1.73 (ddt, 1H, C2–H, J=14.2, 4.4 Hz), 1.59 (dddd, 1H, C2-H', J=6.3 Hz), 1.16 (d, 3H, C4-H), 0.85 (s, 9H, t-Bu), 0.05 (s, 3H, Si-CH<sub>3</sub>), 0.04 (s, 3H, Si- $\mathrm{CH_{3}^{\prime}}$ ). Lit, <sup>14a)</sup> bp (racemic form) 39—42°C/0.02 mmHg.

(R)-3-t-Butyldimethylsiloxybutanal (21a). To a suspension of pyridinium chlorochromate (PCC: 6.44 g, 27.8

Scheme 12.

Scheme 13.

mmol) and sodium acetate (570 mg, 6.95 mmol) in dichloromethane (30 ml) was added **20a** (2.84 g, 13.9 mmol) in dichloromethane (30 ml) and the mixture was stirred at room temperature for 5 h. To the reaction mixture was added ether and filtered through a Florisil® and the filtrate was concentrated to afford **21a** as syrup in 91% yield (2.57 g). The aldehyde was used without further purification:  $[\alpha]_{\rm D}^{16}$   $-14.4^{\circ}$  (c 1.05, CHCl<sub>3</sub>);  $^{1}{\rm H}$  NMR  $\delta$ =9.78 (t, 1H, C1–H, J=2.4 Hz), 4.34 (dtd, 1H, C3–H, J=7.0, 6.3, 5.1 Hz), 2.54 (ddd, 1H, C2–H, J=15.7 Hz), 2.44 (ddd, 1H, C2–H'), 1.23 (d, 3H, C4–H), 0.86 (s, 9H, t-Bu), 0.06 (s, 3H, Si–CH<sub>3</sub>), 0.05 (s, 3H, Si–CH<sub>3</sub>). Lit,  $^{14a}$  bp (racemic form) 80°C/0.02 mmHg.

Methyl (2E,5R)-5-t-Butyldimethylsiloxy-2-hexenoate (23a). This compound was prepared by the modified procedure reported for the preparation of the corresponding racemic form. A solution of 21a (1.19 g, 5.9 mmol) and 22 (2.17 g, 6.5 mmol) in benzene (80 ml) was refluxed for 19 h. The reaction mixture was concentrated and chromatographed on silica gel with ether and petroleum ether (1:40, v/v) to afford 23a as syrup in 74% yield (1.13 g) and methyl (2Z, 5R)-5-t-butyldimethylsiloxy-2-hexenoate (23a')

in 7% yield (100 mg):  $^1\text{H}$  NMR E-isomer:  $\delta = 6.94$  (dt, 1H, C3–H, J = 15.7, 1.4 Hz), 5.82 (dt, 1H, C2–H, J = 1.4 Hz), 3.91 (sx, 1H, C5–H, J = 6.0 Hz), 3.71 (s, 3H, COOCH<sub>3</sub>), 2.30 (ddt, 2H, C4–H, J = 1.4 Hz), 1.14 (d, 3H, C6–H), 0.86 (s, 9H, t-Bu), 0.03 (s, 3H, Si–CH<sub>3</sub>), 0.02 (s, 3H, Si–CH'<sub>3</sub>). Lit,  $^{14a}$  bp (racemic form) 75°C/0.02 mmHg. Z-isomer:  $\delta = 6.36$  (dt, 1H, C3–H, J = 11.5, 7.3 Hz), 5.84 (dt, 1H, C2–H, J = 1.7 Hz), 3.95 (dqd, 1H, C5–H, J = 5.3, 6.2, 6.3 Hz), 3.69 (s, 3H, COOCH<sub>3</sub>), 2.82 (dddd, 1H, C4–H, J = 15.2 Hz), 2.75 (dddd, 1H, C4–H'), 1.15 (d, 3H, C6–H), 0.87 (s, 9H, t-Bu), 0.04 (s, 3H, Si–CH<sub>3</sub>), 0.03 (s, 3H, Si–CH'<sub>3</sub>).

(2E,5R)-5-t-Butyldimethylsiloxy-2-hexenoic Acid (17a). This compound was prepared by the modified procedure reported for the preparation of the corresponding racemic form. To a suspension of 23a (258 mg, 1 mmol) in tetrahydrofuran (THF) and water (1:1, 30 ml) was added lithium hydroxide monohydrate (126 mg, 3 mmol) and the mixture was stirred at room temperature for 18 h. The reaction mixture was washed with chloroform and the aqueous layer was acidified to pH 3.5 with Dowex-50W(H<sup>+</sup>). The solution was extracted with chloroform, dried with MgSO<sub>4</sub> and concentrated to afford 17a as syrup in quantitative

yield (277 mg). This was used without further purification:  $^1{\rm H\,NMR}$   $\delta{=}9{-}11$  (br, 1H, COOH), 7.08 (dt, 1H, C3–H,  $J{=}15.5,\,7.3$  Hz), 5.84 (dt, 1H, C2–H,  $J{=}1.3$  Hz), 3.95 (sx, 1H, C5–H,  $J{=}6.2,\,6.0$  Hz), 2.35 (ddt, 2H, C4–H,  $J{=}1.3$  Hz), 1.17 (d, 3H, C6–H), 0.88 (s, 9H, t-Bu), 0.05 (s, 3H, Si–CH<sub>3</sub>), 0.04 (s, 3H, Si–CH<sub>3</sub>). Lit,  $^{14\rm a)}$  bp (racemic form) 120°C/0.01 mmHg.

t- Butyl (S)- 3- t- Butyldimethylsiloxybutanoate A solution of 18b (8.01 g, 50 mmol), t-butyl-(19b). dimethylsilyl chloride (9.04 g, 60 mmol) and imidazole (8.5 g, 125 mmol) in dimethylformamide (80 ml) was stirred at room temperature for 72 h. To the reaction mixture was added water and ether and phases were separated. The ether layer was successively washed with saturated aqueous NaHCO<sub>3</sub> solution, saturated aqueous NaCl solution, dried with MgSO<sub>4</sub>, and concentrated. The residue was distilled under reduced pressure (79°C/1 mmHg) to afford 19b in 91% yield (12.5 g):  $[\alpha]_D^{20} + 21.6^{\circ}$  (c 1.03, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta = 4.25$  (dqd, 1H, C3-H, J = 6.8, 5.9, 6.3 Hz), 2.43 (dd, 1H, C2-H, J=14.8 Hz), 2.28 (dd, 1H, C2-H'), 1.46 (s, 9H, COOt-Bu), 1.17 (d, 3H, C4-H), 0.89 (s, 9H, Si-t-Bu), 0.08 (s, 3H,  $Si-CH_3$ ), 0.07 (s, 3H,  $Si-CH_3$ ).

(S)-3-t-Butyldimethylsiloxy-1-butanol (20b). a stirred solution of 19b (11.0 g, 40 mmol) in toluene was added 1 M diisobutylaluminium hydride in dichloromethane (100 ml, 100 mmol) at  $-10^{\circ}$ C over 1.5 h and the mixture was stirred at  $-10^{\circ}$ C for 1.5 h. The reaction mixture was quenched with aqueous saturated sodium tartrate and filtered. The filtrate was extracted with dichloromethane. Dichloromethane layer was dried with MgSO<sub>4</sub> and concentrated. The residue was distilled under reduced pressure  $(80^{\circ}\text{C}/2 \text{ mmHg})$  to afford **20b** in 81% yield (6.59 g):  $[\alpha]_D^{24} + 29.8^{\circ}$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta = 4.10$  (qud, 1H, C3-H, J=6.5, 4.2 Hz), 3.83 (ddd, 1H, C1-H, J=10.9, 7.9, 4.1 Hz), 3.71 (ddd, 1H, C1-H', J=6.2, 6.3 Hz), 2.38 (br, 1H, C1-OH), 1.78 (ddt, 1H, C2-H, J = 14.2, 4.2 Hz), 1.63 (dddd, 1H, C2-H', J=6.6 Hz), 1.19 (d, 3H, C4-H), 0.89 (s,9H, t-Bu), 0.08 (s, 3H, Si-CH<sub>3</sub>), 0.07 (s, 3H, Si-CH<sub>3</sub>').

(S)-3-t-Butyldimethylsiloxybutanal (21b). To a suspension of PCC (14.3 g, 62 mmol) and sodium acetate (1.27 g, 15.5 mmol) in dichloromethane (60 ml) was added 20b (6.33 g, 31.0 mmol) in dichloromethane (60 ml) and the mixture was stirred at room temperature for 6 h. To the reaction mixture was added ether and filtered through Florisil® and the filtrate was concentrated to afford 21b as syrup in 83% yield (5.23 g). The aldehyde was used without further purification:  $^{1}$ H NMR  $\delta$ =9.8 (t, 1H, C1-H, J=2.6 Hz), 4.36 (qud, 1H, C3-H, J=7.0, 5.9, 5.0 Hz), 2.56 (ddd, 1H, C2-H, J=15.7 Hz), 2.46 (ddd, 1H, C2-H'), 1.24 (d, 3H, C4-H), 0.87 (s, 9H, t-Bu), 0.08 (s, 3H, Si-CH<sub>3</sub>), 0.06 (s, 3H, Si-CH<sub>3</sub>).

Methyl (2*E*,5*S*)-5-*t*-Butyldimethylsiloxy-2-hexenoate (23b). A solution of 21b (5.23 g, 25.6 mmol) and 22 (10 g, 30 mmol) in benzene (200 ml) was refluxed for 18 h. The reaction mixture was concentrated and chromatographed on silica gel with ether and petroleum ether (1:20, v/v) to afford 23b as syrup in 54% yield (3.57 g) and methyl (2*Z*,5*S*)-5-*t*-butyldimethylsiloxy-2-hexenoate (23b'): For *E*-isomer, [α]<sub>D</sub><sup>18</sup> +8.94° (*c* 1.01, CHCl<sub>3</sub>); <sup>1</sup>H NMR *E*-isomer:  $\delta$ =6.95 (dt, 1H, C3–H, J=15.5, 7.5 Hz), 5.83 (dt, 1H, C2–H, J=1.5 Hz), 3.92 (sx, 1H, C5–H, J=6.0, 5.9 Hz), 3.72 (s, 3H, COOCH<sub>3</sub>), 2.31 (ddt, 2H, C4–H, J=1.5 Hz), 1.15 (d,

3H, C6–H), 0.87 (s, 9H, *t*-Bu), 0.04 (s, 3H, Si–CH<sub>3</sub>), 0.03 (s, 3H, Si–CH<sub>3</sub>'). For *Z*-isomer:  $\delta$ =6.36 (dt, 1H, C3–H, *J*=11.5, 7.3 Hz), 5.84 (dt, 1H, C2–H, *J*=1.7 Hz), 3.95 (dqd, 1H, C5–H, *J*=5.3, 6.2, 6.3 Hz), 3.69 (s, 3H, COOCH<sub>3</sub>), 2.82 (dddd, 1H, C4–H, *J*=15.2 Hz), 2.75 (dddd, 1H, C4–H'), 1.15 (d, 3H, C6–H), 0.87 (s, 9H, *t*-Bu), 0.04 (s, 3H, Si–CH<sub>3</sub>), 0.03 (s, 3H, Si–CH<sub>3</sub>').

(2E,5S)-5-t-Butyldimethylsiloxy-2-hexenoic Acid (17b). To a suspension of 23b (258 mg, 1 mmol) in THF and water (1:1, 30 ml) was added lithium hydroxide monohydrate (126 mg, 3 mmol) and the mixture was stirred at room temperature for 18 h. The reaction mixture was washed with chloroform and the aqueous layer was acidified to pH 3.5 with Dowex-50W(H<sup>+</sup>). The solution was extracted with chloroform, dried with MgSO<sub>4</sub>, and concentrated to afford 17b as syrup in quantitative yield (250 mg). This was used without further purification:  ${}^{1}\text{H NMR }\delta = 9-12$ (br, 1H, COOH), 7.08 (dt, 1H, C3-H, J=15.5, 7.7 Hz), 5.84 (dt, 1H, C2-H, J=1.2 Hz), 3.95 (sx, 1H, C5-H, J=6.0, 6.1 Hz), 2.35 (ddt, 2H, C4-H, J=1.1 Hz), 1.17 (d, 3H, C6-H), 0.88 (s, 9H, t-Bu), 0.05 (s, 3H, Si-CH<sub>3</sub>), 0.04 (s, 3H, Si- $CH_3'$ ).

Methyl 4,6-Dichloro-4,6-dideoxy-α-D-galactopyranoside. This compound was prepared by a literature procedure<sup>15a)</sup> in 36% yield (22.2 g): Mp 157—159°C; [α]<sub>D</sub><sup>17</sup>+179° (c 0.99, H<sub>2</sub>O); <sup>1</sup>H NMR δ=4.85 (d, 1H, C1–H, J=5.8 Hz), 4.53 (dd, 1H, C4–H, J=3.5, 0.8 Hz), 4.15 (td, 1H, C5–H, J=6.6 Hz), 4.00 (ddd, 1H, C3–H, J=3.6, 6.8 Hz), 3.85 (td,1H, C2–H, J=9.2 Hz), 3.68 (d, 2H, C6–H), 3.48 (s, 3H, C1–OCH<sub>3</sub>), 2.61 (d, 1H, C3–OH), 2.20 (d, 1H, C2–OH). Lit, <sup>15a)</sup> mp 158°C; [α]<sub>D</sub><sup>20</sup>+179° (c=2.1, H<sub>2</sub>O).

Methyl 4,6-Dideoxy-α-D-xylo-hexopyranoside (25). This compound was prepared according to reported procedure<sup>15b)</sup> in 79% yield (4.32 g): Mp 100—101°C;  $[\alpha]_D$  +171° (c 0.4, MeOH); <sup>1</sup>H NMR δ=4.74 (d, 1H, C1-H, J=3.8 Hz), 3.90 (ddd, 1H, C5-H, J=2.0, 11.9, 6.3 Hz), 3.83 (tdd, 1H, C3-H, J=9.2, 3.0, 4.9, 9.2 Hz), 3.40 (s, 3H, C1-OCH<sub>3</sub>), 3.39 (td, 1H, C2-H, J=9.4 Hz), 3.25 (d, 1H, C3-OH), 2.78 (d, 1H, C2-OH), 1.98 (ddd, 1H, C4-H', J=12.7 Hz), 1.36 (q, 1H, C4-H), 1.21 (d, 3H, C6-H). Lit, <sup>15b)</sup> mp 105—108°C:  $[\alpha]_D^{2D}$ +171° (c 1.0, MeOH).

Methyl 4,6-Dideoxy-2-O-tosyl- $\alpha$ -D-xylo-hexopyranoside (29). To a solution of 25 (4.87 g, 30 mmol) in pyridine (100 ml) was added tosyl chloride (6.86 g, 36 mmol) in pyridine (30 ml) at 0°C and the solution was stirred for 48 h at room temperature. To the solution was added water and extracted with chloroform. The organic layer was successively washed with 10% H<sub>2</sub>SO<sub>4</sub> and saturated aqueous NaHCO<sub>3</sub>. The organic layer was dried with MgSO<sub>4</sub> and concentrated. The residue was chromatographed on silica gel with benzene-ethyl acetate (7:1, v/v) to afford the mixture (6.41 g) of **29** and 3-O-tosyl derivative **29**' as syrup. The yields of 29 and 29' were estimated to be 58 and 10% by <sup>1</sup>H NMR spectroscopy. This was used without further purification. <sup>1</sup>H NMR for **29**  $\delta$ =7.84 (d, 2H, Aryl-H, J=8.1 Hz), 7.35 (d, 2H, Aryl-H'), 4.63 (d, 1H, C1-H, J=3.6 Hz), 4.23 (dd, 1H, C2-H, J=9.5 Hz), 4.09 (ddd, 1H, C3-H, J=11.3, 5.2 Hz), 3.90 (ddd, 1H, C5-H, J=11.5, 2.2, 6.2 Hz), 3.25 (s,3H, C1-OCH<sub>3</sub>), 2.44 (s, 3H, tolyl-CH<sub>3</sub>), 2.04 (ddd, 1H, C4-H', J=13.0 Hz), 1.86 (br, 1H, C3-OH), 1.38 (ddd, 1H, C4-H), 1.17 (d, 3H, C6-H).

Methyl 2,3-Anhydro-4,6-dideoxy-α-D-lyxo-hexopy-

ranoside (30). To a solution of **29** and **29'** (6.42 g, 20.3 m)mmol) in methanol (60 ml) was added 1 M sodium methoxide in methanol (24.3 ml) and the mixture was refluxed for 5 h. To the cooled solution was added water and the mixture was extracted with chloroform. The organic layer was dried with MgSO<sub>4</sub> and concentrated. The residue was chromatographed on silica gel with petroleum ether-ether (5:1, v/v) and distilled under reduced pressure (60°C/12 mmHg) to afford **30** in 50% yield (1.46 g):  $[\alpha]_D + 69.2^{\circ}(c)$ 0.6, MeOH); <sup>1</sup>H NMR  $\delta$ =4.88 (s, 1H, C1-H), 3.84 (dqd, 1H, C5-H, J=11.1, 4.4, 6.3 Hz), 3.44 (s, 3H, C1-OCH<sub>3</sub>), 3.33 (dd, 1H, C3-H, J=3.9, 15.3 Hz), 2.97 (d, 1H C2-H), 1.91 (ddd, 1H, C4-H'), 1.74 (dd, 1H, C4-H), 1.13 (d, 3H, C6-H). Lit. 32) bp 45—80°C/16 mmHg; mp 28—30°C;  $[\alpha]_D^{20} + 70^\circ$  (c 1.0, MeOH).

Methyl 4,6-Didexoy-α-D-arabino-hexopyranoside (31). A suspension of 30 (1.69 g, 11.7 mmol) in 1 M aqueous KOH (35 ml) was refluxed for 2 h. The cooled mixture was neutralized with glacial acetic acid and concentrated. To the residue was added chloroform and the mixture was filtered through Hyflo Super-Cel<sup>®</sup>. The filtrate was dried with MgSO<sub>4</sub>, concentrated, and distilled under reduced pressure (95°C/0.8 mmHg) to afford 31 in 91% yield (1.36 g):  $[\alpha]_D$ +101° (c 0.6, MeOH); <sup>1</sup>H NMR δ=4.68 (s, 1H, C1-H), 4.11 (dqd, 1H, C5-H, J=10.2, 3.3, 6.3 Hz), 3.87 (m, 1H, C3-H, J=3.5, 9.9, 3.3, 3.5 Hz), 3.61 (dd, 1H, C2-H, J=7.6 Hz), 3.49 (d, 1H, C3-OH), 3.44 (s, 3H, C1-OCH<sub>3</sub>), 2.42 (d, 1H, C2-OH), 1.82 (ddd, 1H, C4-H, J=14.2 Hz), 1.70 (dt, 1H, C4-H'), 1.24 (d, 3H, C6-H).

**4,6-Dideoxy-D-***xylo***-hexopyranose (26a).** A suspension of **25** (4 g, 24.7 mmol) and Dowex-50W(H<sup>+</sup>) (12 g) in water (64 ml) was refluxed for 8.5 h, cooled and concentrated. The residue was recrystallized from ethanol—petroleum ether to afford **31** in 84% yield (3.07 g): Mp 137—140°C;  $[\alpha]_D + 39.9^\circ$  (c 0.5, H<sub>2</sub>O). Lit, <sup>15b)</sup> mp 136—137°C;  $[\alpha]_D^{20} + 100^\circ \rightarrow +34^\circ (c$  1.0, H<sub>2</sub>O).

**4,6-Dideoxy-D-***arabino***-hexopyranose (26b).** A suspension of **31** (2.38 g, 14.7 mmol) and Dowex-50W(H<sup>+</sup>) (7 g) in water (32 ml) was refluxed for 8.5 h, cooled and concentrated to afford **26b** as syrup in quantitative yield (2.26 g). This was used without further purification.

2-(p-Tolylsulfonyl) ethyl (2E,4S,5S,7R)-7-Hydroxy-4,5-isopropylidenedioxy-2-octenoate (28a). tion of 26a (1.08 g, 7.2 mmol) and 27 (6.1 g, 10.8 mmol) in benzene (50 ml) was refluxed for 21.5 h. The reaction mixture was cooled and concentrated. The residue was chromatographed on silica gel with chloroform-methanol (20:1, v/v) to afford crude 2-(p-tolylsulfonyl)ethyl (2E,4S,5S,7R)-4,5,7-trihydroxy-2-octenoate. The crude product was dissolved in acetone (20 ml) and to the solution was added 2, 2-dimethoxypropane (3.58 g, 34.4 mmol) and p-toluenesulfonic acid (0.18 g, 1 mmol). The mixture was stirred for 77h at room temperature. The reaction mixture was cooled to  $0^{\circ}\mathrm{C}$  and to the mixture was added 0.3 M solution of sodium methoxide in methanol until a neutral solution was obtained. The mixture was filtered through Hyflo Super-Cel® and the filtrate was concentrated. The residue was chromatographed on silica gel with ether and recrystallized form hexane-ethyl acetate to afford 28a in 41% yield (1.22 g): Mp 78—79°C;  $[\alpha]_{\rm D} - 17.5^{\circ}$  (c 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta = 7.80$  (d, 2H, Ph-H, J=8.1 Hz), 7.37 (d, 2H, Ph-H'), 6.79 (dd, 1H, C3-H, J=15.7, 5.4 Hz), 5.92 (dd, 1H, C2-H, J=1.3 Hz), 4.47 (t,

2H, COOCH<sub>2</sub>, J=6.1 Hz), 4.18 (ddd, 1H, C4–H, J=8.5 Hz), 4.00—4.11 (m, 1H, C7–H, J=4.0, 6.3 Hz), 3.93 (ddd, 1H, C5–H), 3.47 (t, 2H, SO<sub>2</sub>–CH<sub>2</sub>), 2.46 (s, 3H, Ph–CH<sub>3</sub>), 2.14 (d, 1H, C7–OH), 1.69—1.74 (m, 2H, C6–H), 1.45 (s, 3H, isop-CH<sub>3</sub>), 1.41 (s, 3H, isop-CH'<sub>3</sub>), 1.24 (d, 3H, C8–H). Found: C, 58.31; H, 6.84%. Calcd for C<sub>20</sub>H<sub>28</sub>O<sub>7</sub>S: C, 58.30; H, 6.84%.

2- (p- Tolylsulfonyl)ethyl (2E, 4R, 5R, 7R)- 7- Hydroxy-4,5-isopropylidenedioxy-2-octenoate (28b). A solution of **26b** (0.17 g, 1.1 mmol) and **27** (0.61 g, 1.2 mmol) in benzene (25 ml) was refluxed for 18 h. The reaction mixture was cooled and concentrated. The residue was chromatographed on silica gel with chloroform-methanol (7:1, v/v) to afford crude 2-(p-tolylsulfonyl)ethyl (2E,4S,5S,7R)-4,5,7-trihydroxy-2-octenoate. The crude product was dissolved in acetone (3.7 ml) and to the solution was added 2, 2-dimethoxypropane (0.69 g, 6.7 mmol) and p-toluenesulfonic acid (0.023 g, 0.13 mmol). The mixture was stirred for 64 h at room temperature. The reaction mixture was cooled to 0°C and to the mixture was added 1 M solution of sodium methoxide in methanol until a neutral solution was obtained. The mixture was filtered through Hyflo Super-Cel® and the filtrate was concentrated. The residue was chromatographed on silica gel with ether and recrystallized from hexane-ethyl acetate to afford 28b in 12% yield (0.13 g): Mp 86°C;  $[\alpha]_D + 7.7^\circ$  (c 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta = 7.80$ (d, 2H, Ph-H, J=8.1 Hz), 7.37 (d, 2H, Ph-H'), 6.77 (dd, 1H, C3-H, J=15.7, 1.5 Hz), 5.93 (dd, 1H, C2-H, J=1.5Hz), 4.48 (t, 2H, COOCH<sub>2</sub>, J=6.1 Hz), 4.15 (ddd, 1H, C4-H, J=8.4 Hz), 3.93-4.07 (m, 1H, C7-H, J=5.9 Hz), 3.93(td, 1H, C5-H), 3.47 (t, 2H, SO<sub>2</sub>-CH<sub>2</sub>), 2.81 (br, 1H, C7-OH), 2.46 (s, 3H, Ph-CH<sub>3</sub>), 1.64—1.76 (m, 2H, C6-H), 1.45 (s, 3H, isop-CH<sub>3</sub>), 1.42 (s, 3H, isop-CH<sub>3</sub>), 1.22 (d, 3H, C8-H). Found: C, 58.18; H, 6.96%. Calcd for C<sub>20</sub>H<sub>28</sub>O<sub>7</sub>S: C, 58.30; H. 6.84%.

2-(p-Tolylsulfonyl) ethyl (4S,5S,7R)-7-Hydroxy-4, 5-isopropylidenedioxyoctanoate (24a). pound 28a (0.41 g, 1 mmol) in THF (50 ml) was hydrogenated in the presence of 10%-Pd/C (0.1 g) under 20 kg cm<sup>-2</sup> of H<sub>2</sub> at room temperature for 3 h in an autoclave. The reaction mixture was concentrated to afford 24a as syrup in quantitative yield (0.42 g). This was used without further purification:  ${}^{1}HNMR$   $\delta = 7.80$  (d, 2H, Ph-H, J=8.1 Hz), 7.38 (d, 2H, Ph-H'), 4.40 (t, 2H, COO-CH<sub>2</sub>, J=6.2 Hz), 4.05 (dqd, 1H, C7-H, J=4.2, 7.2, 6.3 Hz), 3.84 (td, 1H, C5-H, J=8.3, 7.2, 4.4 Hz), 3.63 (td, 1H, C4-H,J=3.2, 8.3 Hz), 3.44 (t, 2H, SO<sub>2</sub>-CH<sub>2</sub>), 2.46 (s, 3H, Ph-CH<sub>3</sub>), 2.2—2.4 (br. 1H, C7-OH), 2.34 (ddd, 1H, C2-H, J=16.9, 8.7, 8.8 Hz), 2.24 (ddd, 1H, C2-H', J=6.9, 8.4Hz), 1.85 (dddd, 1H, C3-H, J=13.9 Hz), 1.57—1.74 (m, 3H, C3-H' and C6-H), 1.37 (s, 3H, isop-CH<sub>3</sub>), 1.36 (s, 3H, isop-CH<sub>3</sub>), 1.23 (d, 3H, C8-H).

2-(p-Tolylsulfonyl)ethyl (4R,5R,7R)-7-Hydroxy-4, 5-isopropylidenedioxyoctanoate (24b). The compound 28b (0.38 g, 0.93 mmol) in THF (50 ml) was hydrogenated in the presence of 10%-Pd/C (0.2 g) under 20 kg cm<sup>-2</sup> of H<sub>2</sub> at room temperature for 3 h in an autoclave. The reaction mixture was concentrated to afford 24b as syrup in quantitative yield (0.42 g). This was used without further purification:  $^1$ H NMR  $\delta$ =7.79 (d, 2H, Ph-H, J=8.2 Hz), 7.37 (d, 2H, Ph-H'), 4.41 (td, 2H, COO-CH<sub>2</sub>, J=6.2, 1.2 Hz), 4.00 (dqd, 1H, C7-H, J=3.0, 9.2, 6.3 Hz),

3.71 (td, 1H, C5–H, J=9.2, 8.2, 3.4 Hz), 3.59 (td, 1H, C4–H, J=3.2, 8.2 Hz), 3.42 (t, 2H, SO<sub>2</sub>–CH<sub>2</sub>), 3.06 (br, 1H, C7–OH), 2.45 (s, 3H, Ph–CH<sub>3</sub>), 2.34 (ddd, 1H, C2–H, J=16.8, 8.8, 6.1 Hz), 2.25 (ddd, 1H, C2–H', J=6.9, 8.2 Hz), 1.84 (dddd, 1H, C3–H, J=14.0 Hz), 1.51—1.73 (m, 3H, C3–H' and C6–H), 1.37 (s, 3H, isop-CH<sub>3</sub>), 1.36 (s, 3H, isop-CH'<sub>3</sub>), 1.20 (d, 3H, C8–H).

2- (p- Tolylsulfonyl) ethyl (10E, 4S, 5S, 7R, 13R)-, (10E, 4S, 5S, 7R, 13S)-, and (10E, 4R, 5R, 7R, 13S)-13-t-Butyldimethylsiloxy-4,5-isopropylidenedioxy-7methyl-9-oxo-8-oxa-10-tetradecenoates (39c, 39a, and 39b). To a solution of 2,4,6-trichlorobenzovl chloride (0.366 g, 1.5 mmol) in THF (1.5 ml) was added 17a (0.250 g, 1 mmol) and triethylamine (0.250 g, 1.5 mmol) in THF (1.5 ml) over 30 min and the mixture was stirred for additional 2 h at room temperature. The mixture was filtered under Ar and the filtrate was evaporated. The resulting anhydride was dissolved in toluene (1 ml). To the mixture was added 24a (0.415 g, 1 mmol) and DMAP (0.135 g, 1.1 mmol) in toluene (5 ml) and stirred for 22 h at room temperature. The reaction mixture was filtered through Hyflo Super-Cel<sup>®</sup> and concentrated. The residue was chromatographed on silica gel with benzene-ethyl acetate (20:1, v/v) to afford 39c as syrup in 86% yield.

The compound **39a** and **39b** were synthesized as syrup in a similar manner as above (50 and 95%, respectively).

39a:  $^{1}$ H NMR  $\delta$ =7.79 (d, 2H, Ph–H, J=8.3 Hz), 7.36 (d, 2H, Ph–H'), 6.93 (dt, 1H, C11–H, J=7.7, 15.6 Hz), 5.81 (dt, 1H, C10–H, J=1.2 Hz), 5.09 (dqd, 1H, C7–H, J=4.4, 6.2, 8.5 Hz), 4.39 (t, 2H, COOCH<sub>2</sub>, J=6.0 Hz), 3.91 (sx, 1H, C13–H, J=6.0 Hz), 3.61 (td, 1H, C2–H, J=2.5, 8.4 Hz), 3.51 (td, 1H, C2–H', J=2.9, 8.2 Hz), 3.43 (t, 2H, SO<sub>2</sub>–CH<sub>2</sub>), 2.45 (s, 3H, Ph–CH<sub>3</sub>), 2.15—2.36 (m, 4H, C4–H, C5–H, C12–H), 1.82 (ddd, 1H, C6–H, J=2.5, 8.5, 14.2 Hz), 1.65 (ddd, 1H, C6–H', J=4.4, 8.9 Hz), 1.73—1.86 (m, 1H, C3–H), 1.55—1.70 (m, 1H, C3–H'), 1.33 (s, 3H, isop-CH<sub>3</sub>), 1.30 (s, 3H, isop-CH<sub>3</sub>'), 1.28 (d, 3H, C7–CH<sub>3</sub>), 1.15 (d, 3H, C13–CH<sub>3</sub>), 0.87 (s, 9H, t-Bu), 0.04 (s, 3H, Si–CH<sub>3</sub>), 0.03 (s, 3H, Si–CH<sub>3</sub>').

**39c:** <sup>1</sup>H NMR  $\delta$ =7.77 (d, 2H, Ph–H, J=8.2 Hz), 7.35 (d, 2H, Ph–H'), 6.90 (dt, 1H, C11–H, J=6.9, 15.5 Hz), 5.79 (dt, 1H, C10–H, J=1.4 Hz), 5.02 (dqd, 1H, C7–H, J=4.5, 6.3, 8.4 Hz), 4.37 (t, 2H, COOCH<sub>2</sub>, J=6.2 Hz), 3.91 (sx, 1H, C13–H, J=6.0 Hz), 3.59 (td, 1H, C2–H, J=2.5, 8.5 Hz), 3.50 (td, 1H, C2–H', J=3.0, 8.3 Hz), 3.41 (t, 2H, SO<sub>2</sub>–CH<sub>2</sub>), 2.43 (s, 3H, Ph–CH<sub>3</sub>), 2.13—2.34 (m, 2H, C4–H, C5–H), 2.29 (ddd, 2H, C12–H, J=1.4 Hz) 1.80 (ddd, 1H, C6–H, J=2.4, 8.4, 14.2 Hz), 1.63 (ddd, 1H, C6–H', J=4.5, 9.0 Hz), 1.73—1.83 (m, 1H, C3–H), 1.53—1.64 (m, 1H, C3–H'), 1.31 (s, 3H, isop-CH<sub>3</sub>), 1.28 (s, 3H, isop-CH<sub>3</sub>), 1.27 (d, 3H,

C7-CH<sub>3</sub>), 1.14 (d, 3H, C13-CH<sub>3</sub>), 0.85 (s, 9H, t-Bu), 0.02 (s, 3H, Si-CH<sub>3</sub>), 0.01 (s, 3H, Si-CH<sub>3</sub>).

(10E,4S,5S,7R,13S)-, (10E,4R,5R,7R,13S)-, and (10E,4S,5S,7R,13R)-13-Hydroxy-4,5-isopropylidene-dioxy-7-methyl-9-oxo-8-oxa-10-tetradecenoic acids (40a, 40b, and 40c). To a solution of 39c (1.4 g, 2.1 mmol) in THF (30 ml) was added 1 M TBAF in THF (6.3 ml, 6.3 mmol) and the mixture was stirred for 72 h at room temperature. The reaction mixture was extracted with 0.1 M aqueous NaHCO<sub>3</sub> and water layer was acidified to pH 4 with Dowex-50W(H<sup>+</sup>). The solution was extracted with chloroform and organic layer was dried with MgSO<sub>4</sub> and concentrated to afford 40c as syrup in quantitative yield (665 mg). This was used without further purification.

The compound **40a** and **40b** were prepared as syrup in a similar manner as above in quantitative yield.

Macrolactonization Using DEAD-TPP System: (3E,6R,11S,12S,14R)- or (3E,6R,11R,12R,14R)-11, 12-Isopropylidenedioxy-6,14-dimethyl-1,7-dioxa-3-cyclotetradecene-2,8-tetrone (44a, 44b). To a solution of 40a (44 mg, 0.13 mmol) in toluene (5 ml) was added triphenylphosphine (68 mg, 0.26 mmol) in toluene (5 ml) and diethyl azodicarboxylate (45 mg, 0.26 mmol) at -10°C. The mixture was stirred at -10°C for 24 h then 0°C for 42 h. The reaction mixture was evaporated and chromatographed on silica gel with ether-petroleum ether (1:1, v/v) to afford 44a (crystals, mp 57—59°C) in 12% yield (5 mg).

The isomer **44b** (syrup) was prepared in a similar manner as above in 4% yield.

44b: <sup>1</sup>H NMR δ=6.78 (ddd, 1H, C4–H, J=7.5, 8.4, 15.8 Hz), 5.80 (dt, 1H, C3–H, J=1.0 Hz), 5.06—5.18 (m, 2H, C6–H, C14–H, J=3.2, 4.3, 4.5, 11.2 Hz), 3.84 (td, 1H, C11–H, J=2.9, 7.1 Hz), 3.73 (ddd, 1H, C12–H, J=3.1, 7.2, 9.2 Hz), 2.55 (ddd, 1H, C9–H, J=5.4, 11.6, 14.0 Hz), 2.28 (dddd, 1H, C5–H, J=11.2 Hz), 2.09 (ddd, 1H, C5–H'), 2.04 (ddd, 1H, C9–H', J=3.1, 6.4 Hz), 1.82—1.94 (m, 2H, C13–H), 1.58—1.70 (m, 2H, C10–H), 1.38 (s, 3H, isop-CH<sub>3</sub>), 1.36 (s, 3H, isop-CH<sub>3</sub>), 1.33 (d, 3H, C6–CH<sub>3</sub>), 1.30 (d, 3H, C14–CH<sub>3</sub>).

Macrolactonization Using Yamaguchi Method: (3E, 6R, 11S, 12S, 14R)-11, 12- Isopropylidenedioxy-6, 14- dimethyl-1, 7- dioxa-3- cyclotetradecene-2, 8-tetrone (44a). To a solution of 40c (585 mg, 1.7 mmol) in THF (2 ml) was added triethylamine (344 mg, 3.4 mmol) in THF (4 ml) and 2,4,6-trichlorobenzoyl chloride (829 mg, 3.4 mmol) in THF (6 ml). The mixture was stirred for 2 h at room temperature and filtered under Ar. The filtrate was diluted with toluene (900 ml) and the mixture was added

dropwise to a solution of DMAP (1.454 g, 11.9 mmol) in toluene (200 ml) at  $90^{\circ}$ C over a period of 5 h. The mixture was stirred at  $90^{\circ}$ C for 1 h and cooled to room temperature. The mixture was diluted with ether (500 ml) and washed successively with saturated aqueous citric acid, saturated aqueous NaHCO<sub>3</sub> and water. The organic layer was dried with MgSO<sub>4</sub>, evaporated, and chromatographed on silica gel with ether–petroleum ether (1:5, v/v) to afford **44a** in 56% yield (306 mg).

(3E, 6R, 11S, 12S, 14R)- 11, 12- Dihydroxy- 6, 14- dimethyl-1, 7-dioxa-3-cyclotetradecene-2, 8-tetrone (48).To a solution of 44a (81 mg, 0.3 mmol) in methanol (5 ml) was added 42% aqueous HBF<sub>4</sub> (0.1 ml) and the mixture was stirred at 0°C for 6 h. The reaction mixture was added saturated aqueous NaHCO3 and extracted with dichloromethane. The organic layer was dried with MgSO<sub>4</sub>, evaporated, and chromatographed on silica gel with dichloromethane-methanol (20:1, v/v) to afford 48 as white needles (mp 90—92°C) in 78% yield (28 mg).  $[\alpha]_D - 48^\circ$  (c 0.3, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $\delta$ =6.78 (dt, 1H, C4–H, J=8.1, 15.8 Hz), 5.80 (dt, 1H, C3-H, J=1.2 Hz), 5.19 (dqd, 1H, C6-H, J=2.6, 6.3, 11.3 Hz), 5.06 (dqd, 1H, C14-H, J=3.2, 6.4, 10.7 Hz), 3.42 (ddd, 1H, C12-H, J=0.3, 5.0, 8.3 Hz), 3.22 (td, 1H, C11-H, J=7.2 Hz), 2.66-2.58 (br, 2H, C11-OH and C12-OH), 2.49 (ddd, 1H, C9-H, J=2.0, 11.3, 17.4 Hz). 2.41 (dddd, 1H, C5-H, J=13.5 Hz), 2.35 (dddd, 1H, C5-H'), 2.30 (ddd, 1H, C9-H', J=2.2, 7.0 Hz), 1.96 (ddd, 1H, C13-H, J=14.4 Hz), 1.92 (ddd, 1H, C10-H, J=14.5 Hz), 1.79 (dtd, 1H, C10-H'), 1.71 (ddd, 1H, C13-H'), 1.33 (d, 3H, C14-CH<sub>3</sub>), 1.29 (d, 3H, C6-CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ =21.0 (q, 2C), 27.7 (t), 30.2 (t), 39.2 (t), 40.5 (t), 68.3 (d), 69.7 (d), 70.3 (d), 73.1 (d), 124.2 (d), 144.6 (d), 166.1 (s), 174.2 (s); FAB MS (75 eV) m/z 287 (M+H)<sup>+</sup>; HRMS Calcd for  $(C_{14}H_{22}O_6+H)^+$ : M, 287.1495. Found: m/z 287.1533.

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