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Synthetic Studies on Vancomycin: Synthesis of Seco-Aglucovancomycins

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Abstract: Preparation of the vancomycin analogs (SAV-1 and SAV-2) is described. The key step is construction of a bicyclic diaryl ether residue by means of thallium (III) oxidation of the corresponding halogenated phenols.

The glycopeptide antibiotic vancomycin, isolated from *Streptomyces orientalis*, is one of the most challenging molecules for chemists, from the the viewpoint of its characteristic heptapeptide core involving diaryl ethers.¹ The potent bactericidal activities of this antibiotic, particularly against methicillin resistant *Staphylococcus aureus* (MRSA), are considered to be derived from its high affinity to the terminal D-Ala-D-Ala residue of peptidoglycan precursors in bacterial cell wall biosynthesis. However, difficulties of synthesis and derivation of this antibiotic prevent a drug-based analysis of a detailed mechanism of action.²



Since our own development of the phenolic oxidation methodology employing thallium (III) salts,³ natural products possessing a diaryl ether moiety have been extensively synthesized.^{3,4} A part of our continuous investigation has been focused on the 12-membered ring of 1, which consists of the amino acid residue 5-7, bearing one of five points of the plausible hydrogen bonds contributing to complexation with the D-Ala-D-Ala residue.⁵ To investigate the role of the ring in the interaction, we have synthesized two seco-aglucovancomycin derivatives [SAV-1 (3) and SAV-2 (4)]. These compounds are only examples of vancomycin analogs carrying a full peptide chain and a characteristic diaryl ether moiety.⁶ The structural similarity to the mother antibiotic will impart a comparable binding property to the bacterial cell wall or its model. Particularly, lack of a rigid biphenyl moiety in the molecule may also promise diverse possibilities of mode of action. We describe herein the synthesis of SAV-1 (3) and SAV-2 (4).

The synthesis was initiated with Strecker reaction applied to 3,5-dimethoxybenzaldehyde, followed by N-acylation, to give the racemic acetyl amino acid (5) in 70% yield (3 steps). Compound 5 was subjected to enzymatic hydrolysis,⁷ and protection to afford the optically active phenylglycine derivative (6). The

enantiomeric excess (>95% ee) was spectroscopically determined by employing the (+)-MTPA amide of 6. For the following synthetic process, 6 could be used for the common starting material towards 3 and 4.

Synthesis of SAV-1 (3) was continued by coupling of amine 6 with N-Boc-3,5-dibromotyrosine (7a) to furnish the corresponding dipeptide (8a). Further peptide chain elongation of 8a was effected by successive couplings with N-Boc-4-benzyloxyphenylglycine (9), and then with N-Boc-3,5-diiodo-4-hydroxyphenylglycine (11) to afford the desired tetrapeptide (12a) which was a substrate of the following phenolic oxidation.



Scheme 1 Reagents : a. 1) KCN - Na2S2O5 / NH4OH; 2) 6M - HCl; 3) Ac2O-K2CO3aq. (70% in 3 steps). b. 1) amino acylase ("Amano"); 2) Boc2O-K2CO3aq. ; 3) CH2N2; 4) TFA, then HCl-MeOH (42% from racemate 5). c. 1) 7a or 7b, EDC [1-(3-dimethylaminopropyl)-3-ethylcarbodiimide] - HOBt (1-hydroxybenztriazole), iPr_2EtN ; 2) TFA, then NaHCO3 (8a: 100%, 8b: 100% in 2 steps, respectively). d. 1) 9, EDC - HOBt; 2) TFA, then NaHCO3 (10a: 73%, 10b: 82% in 2 steps, respectively). e. 11, EDC - HOBt (12a: 91%, 12b: 78% in 2 steps, respectively). f. 1) TTN (2 eq) in THF-MeOH-CH(OMe)3 (8:1:1) (13a: 42%, 13b: 40%); 2) TFA, then NaHCO3 (14a: 81%, 14b: 61%). g. H2-Pd/C AcONa in MeOH (15a: 62%, 15b: ca. 50%).

Exposure of 12a to thallium (III) nitrate (TTN) underwent the desired cyclization to yield 13a in 42% yield. The reaction mechanism is assumed to be through a synchronized derivation of a thallium phenoxide or through a phenoxy radical intermediate.³ On addition of the oxidant, the reaction mixture immediately turned to a deep green solution. This observation suggested an electron transfer process, implying the presence of the latter intermediate. The structure of 13a was confirmed by characterization of 15a obtained by the usual deprotection and dehalogenation process.⁸

Compound 13a was deprotected with TFA (81% yield), then subjected to coupling with tripeptide $16a^9$ in the usual manner to yield the corresponding heptapeptide (17a) in 59% yield. Final ring closure was undertaken by the same oxidation of 17a as described in the case of 12a to provide the expected product (18a)

in 35% yield, which was successively submitted to deprotection and dehalogenation to give the target molecule SAV-1 (3),⁸ as can be seen in Scheme 2.



Scheme 2 Reagents: a. EDC-HOBt / DMF (17a: 59%, 17b: 53%). b. 1) TTN (4 eq) in THF-McOH-CH(OMe)3 (8:1:1); 2) Zn - AcOH (18a: 35%, 18b: 40% in 2 steps, respectively). c. H2-Pd/C in MeOH (19a: ca. 40%, 19b: ca. 40%). d. TFA (3: 61%, 4: 76%).

The synthesis of the tetrachloro derivative SAV-2 (4) was accomplished via a similar protocol starting from 6 (Scheme 1). The first peptide bond formation was performed by coupling with N-Boc-3,5-dichlorotyrosine (7b). The peptide elongation afforded the corresponding tetrapeptide (12b) in 64% yield (5 steps). The cyclized product (13b) was obtained by TTN oxidation in 40% yield. The structural confirmation of 13b was achieved by characterization of 15b⁸ obtained by the same procedure as that of 15a.

Compound 13b underwent a similar sequence to the case of 13a involving deprotection (61%) and coupling (53%) to provide the heptapeptide (17b). Final ring closure of 17b via the phenolic oxidation procedure afforded the desired product (18b), which was further converted to the target molecule SAV-2 (4)⁸ (Scheme 2).

In conclusion, we successfully synthesized seco-aglucovancomycins SAV-1 (3) and SAV-2 (4) bearing characteristic bicyclic diaryl ethers and a full peptide chain by means of thallium (III) oxidation methodology as a key step. Conformational analysis of the synthesized compounds and evaluation of their binding manner with the cell wall model will be discussed in the accompanying papers.¹¹

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- ¹H NMR data of 15a, 3 and 4 are cited in the accompanying paper (reference 11). 15b: ¹H NMR (CD₃OD) δ: 2.84 (1H, dd, J= 9.5, 13.6 Hz), ~3.3 (1H, complex, overlapped with solvent signal), 3.71 (3H, s), 3.78 (6H, s), 4.61 (1H, dd, J= 5.1, 9.5 Hz) 5.24 (1H, s), 5.34 (1H, s) 6.48 (1H, t, J= 2.2 Hz), 6.51 (2H, d, J=2.2 Hz), 6.55 (1H, d, J= 1.5 Hz), 6.72 (2H, d, J= 8.4 Hz), 6.93 (1H, d, J= 8.0 Hz), 7.02 (1H, dd, J= 8.0, 1.5 Hz), 7.05 (2H, d, J=8.4 Hz), 7.23 (1H, d, J= 1.8 Hz), and 7.51 (1H, d, J= 1.5 Hz).
- 9. Compounds 16a and 16b were synthesized by the following procedure.



Reagents: a. 1) EDC-HOB1; 2) NaOHaq-dioxane (91% yield for Br derivative, 82% yield for Cl derivative in 2 steps, respectively). b. 1) Asn-allyl ester / EDC-HOB1; 2) 0.5 mol% Pd(PPh3)4 - morpholine (16a: 65%, 16b: 63% in 2 steps, respectively).

- 10. Although N-methylation could be achieved via the corresponding methylene imine (1. CH₂=O; 2. NaBH₃CN *ca.* 70% in 2 steps), the NH₂ derivatives were used for conformation analysis. See the accompanying papers (reference 11).
- 11. Nakamura, K.; Nishiyama, S.; Yamamura, S. Tetrahedron Lett. the accompanying papers in this issue.

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