

Stereoselective Synthesis of α -C-Glycopyranosyl Isoprenoid Compounds

Anne Jégou, Carole Pacheco, Alain Veyrières*

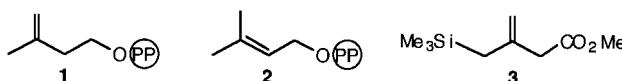
Laboratoire de Synthèses et Activations de Biomolécules, CNRS ESA 6052, Ecole Nationale Supérieure de Chimie de Rennes, Campus de Beaulieu, 35700 Rennes, France

Fax + 33(2)99871348

Received 20 October 1997

Abstract: Addition of the allylsilane $\text{Me}_3\text{SiCH}_2\text{C}(\text{CH}_2\text{CH}_2\text{CO}_2\text{Me})_2$ (**3**) to D-gluco and D-galactopyranosyl derivatives gives in good yields 3-(α -C-glycopyranosylmethyl)-but-3-enotes which can undergo oxidation to β -ketoesters, electrophile-promoted cyclizations or double bond migration.

Isopentenyl pyrophosphate (**1**) and its isomerization product, dimethylallyl pyrophosphate (**2**), are well-known¹ five-carbon building blocks involved in the terpenoid biosynthesis. These activated forms of isoprene can be chemically mimicked^{2,3} by the allylsilane **3** which is easily obtained^{2,4} by reaction of diketene with the Grignard reagent of chloromethyltrimethylsilane, followed by diazomethane esterification.



Addition of **3** to carbonyl compounds² or iminium ions⁵ occurs in high yields, the presence of a carboxylate function then allowing an easy double bond migration to yield a conjugated product.

Herein we report the stereoselective addition of **3** to oxocarbenium species⁶ generated from pyranose sugars (Table 1) and various transformations of the resulting C-glycosides.⁷

The stereoselective synthesis of α -C-allyl glycopyranosides from peracyl hexopyranoses has been described.⁸ However, β -D-glucopyranose pentaacetate (**4**) reacted sluggishly with **3** and $\text{BF}_3\cdot\text{OEt}_2$ in acetonitrile to give a 6:1 mixture of α - and β -C-glycosides **5** in only 24% yield. More reactive perbenzylated fluorides^{9,10} **6** and **9** gave C-glycosides **7** and **10**, respectively in much higher yields and stereoselectivities. ¹H NMR analysis of **7** and **10** in CDCl_3 at room temperature showed complex mixtures of conformers at the pyranose ring. The expected multiplicity of anomeric H-5 (ddd, $J_{4a,5}$ 11, $J_{4b,5}$ 3.3 and $J_{5,6}$ 5.6 Hz) was however observed in $\text{DMSO}-d_6$ at 80°C.¹¹ Pertrimethylsilylated sugars have been rarely used^{12,13} for C- and O-glycoside synthesis. Silylation of methyl α -D-glucopyranosides gave **12** and **14** which reacted with silane **3** and trimethylsilyl trifluoromethanesulfonate

Table 1. Addition of allylsilane **3** (3–5 eq) to D-gluco-(**4**, **6**, **12**) and D-galacto-(**9**, **14**)-pyranose derivatives

Starting material	Product	Method ^a	Yield ^b (%)	dr ^c $\alpha:\beta$
4	5	A	24%	6:1
6 ^d	7 X = CH ₂	B	86%	95:5
	8 X = O (64%)			
9 ^d	10 X = CH ₂	B	90%	1:0
	11 X = O (65%)			
12 ^f	13	C	74%	9:1
14 ^f	15	C	75%	1:0

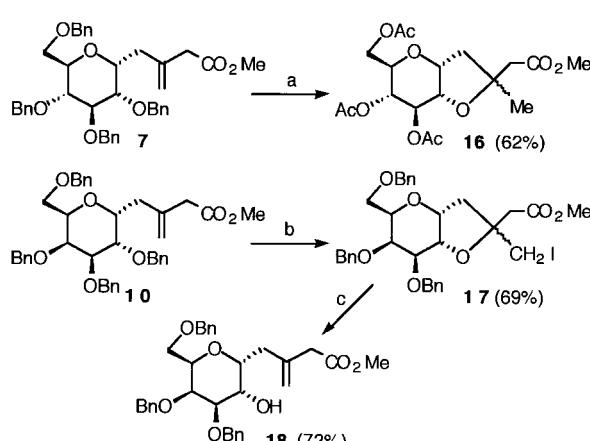
^aMethod A: $\text{BF}_3\cdot\text{OEt}_2$ (5 eq), CH_3CN , rt, 45h. Method B: $\text{BF}_3\cdot\text{OEt}_2$ (1.2 eq), CH_3CN , -30 → 0°C, 1h. Method C: i) TMSOTf (3 eq), CH_3CN , 0°C, 6h; ii) MeOH, 0°C, then Dowex 1 (OH^-), rt. ^bAfter column chromatography. ^cDiastereomeric ratios (dr) were determined by ¹H NMR analysis of chromatographed mixtures.

^dRef 9. ^ei) O_3 , CH_2Cl_2 , -78°C, 5 min; ii) PPh_3 (3 eq), -78 → 20°C, 4h. ^fMethyl α -D-glucopyranosides and D-galactopyranosides were silylated by Me_3SiCl (5.2 eq) and NEt_3 (5.2 eq) in DMF, 0 → 20°C, 4h, according to ref 13.

(TMSOTf) to give after desilylation unprotected C-glycosides **13** and **15**¹⁴ respectively in good yields and selectivities.

Brief treatment of methylene compounds **7** and **10** by ozone at -78°C, followed by reaction with triphenylphosphine gave β -ketoesters **8**¹⁵ and **11** respectively. A minor product resulting from regioselective oxidation of the 7-O-benzyl group into a benzoate was isolated in 6% yield after ozonolysis of the D-gluco compound **7**. γ -Furanosyl and pyranosyl- β -ketoesters have been previously prepared¹⁶ by a non stereoselective Wittig reaction of phosphorane $\text{Ph}_3\text{P}=\text{CHC(OCH}_2\text{CO}_2\text{R}$ with a sugar hemiacetal.

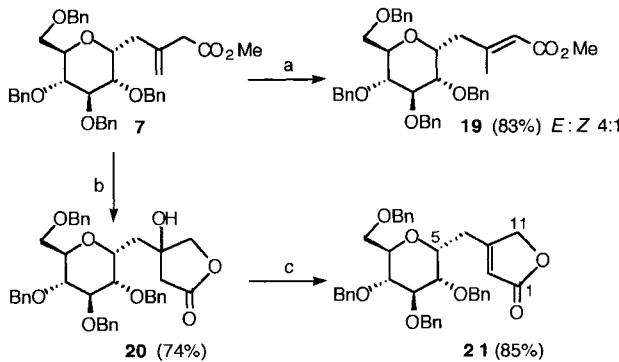
Attempted debenzylation of **7** by anhydrous ferric chloride in dichloromethane¹⁷, followed by acetylation, led to the bicyclic product **16** in 62% yield (Scheme 1). Activation of the double bond of **10** with iodine also led to the addition of O-6 with loss of the benzyl group. Treatment



Scheme 1. a) FeCl_3 (12 eq), CH_2Cl_2 , rt, 5h, then Ac_2O , pyridine, rt, 16h. b) I_2 (3 eq), CH_3CN , 0 → 20°C, 16h. c) Zn (10 eq), 10:10:1 MeOH -THF- AcOH , rt, 16h

of iodoether **17** (3:2 mixture of *C*-2 epimers) by zinc and acetic acid gave the *C*-glycoside **18** selectively debenzylated at *O*-6 (Scheme 1).¹⁸ A stereoselective access to α -C-glycoside of D-galactosamine from **18** should then be possible.¹⁹

Conjugation of the double bond in **7** was accomplished by treatment with piperidine in refluxing tetrahydrofuran²⁰ (Scheme 2). A 4:1 mixture of *E* and *Z* isomers **19** was obtained in 83% yield.



Scheme 2. a) piperidine (2 eq), THF, reflux, 120h. b) OsO_4 cat, NMO (1.1 eq), pyridine (1 eq), 2:1:3 THF- H_2O -tBuOH, reflux, 24h. c) MsCl (4 eq), NEt_3 (8 eq), CH_2Cl_2 , rt, 3h

Assignment of the stereochemistry²¹ was made on the basis of ^1H and ^{13}C chemical shifts of Me and CH_2 -4 groups.²² In the *E* isomer these signals occur respectively at δ_{H} 2.16, δ_{C} 18.56 and $\delta_{\text{H}-4a}$ 2.58, $\delta_{\text{H}-4b}$ 2.48, δ_{C} 36.18, whereas in the *Z* isomer the following values are found: δ_{H} 1.92, δ_{C} 25.55 and $\delta_{\text{H}-4a}$ 3.34, $\delta_{\text{H}-4b}$ 2.95, δ_{C} 28.69.

Finally, *C*-glycoside **7** was treated with a catalytic amount of osmium tetroxide and *N*-methylmorpholine oxide (NMO)²³ to give the lactonized diol **20** as a 7:3 mixture of epimers at *C*-3. Treatment of **20** with methanesulfonyl chloride and triethylamine gave the butenolide **21**²⁴ in a 63% overall yield from **7**.

References and Notes

- Hanson, J.R. In *Comprehensive Organic Synthesis*, Vol.5; Barton, D.; Ollis, W.D.; Haslam, E., Eds; Pergamon Press: Oxford, 1979; p 989.
- Itoh, K.; Fukui, M.; Kurachi, Y. *J. Chem. Soc., Chem. Commun.* **1977**, 500.
- Itoh, K.; Yogo, T.; Ishii, Y. *Chem. Lett.* **1977**, 103; Nishiyama, H.; Itagaki, K.; Takahashi, K.; Itoh, K. *Tetrahedron Lett.* **1981**, 22, 1691.
- Armstrong, R.J.; Weiler, L. *Can. J. Chem.* **1983**, 61, 2530.
- Aratani, M.; Sawada, K.; Hashimoto, M. *Tetrahedron Lett.* **1982**, 23, 3921; Oumoch, S.; Rousseau, G. *Bioorg. Med. Chem. Lett.* **1994**, 4, 2841.
- Ethoxycarbenium ion related to squaric acid has been allylated by **3** in 82% yield: Yamamoto, Y.; Ohno, M.; Egushi, S. *Chem. Lett.* **1995**, 525.
- Pseudomonoc acids, a group of compounds with antimicrobial activity produced by a strain of *Pseudomonas fluorescens*, bear a five-carbon unit, 3-methyl-but-2-enoate, linked at *C*-2 of a tetrahydropyran unit. Biosynthetic studies showed that a 3-methyl-but-3-enoyl CoA is attached to a polyketide chain which subsequently undergoes a cyclization to the tetrahydropyran ring: Feline, T.C.; Jones, R.B.; Mellows, G.; Phillips, L. *J. Chem. Soc., Perkin Trans. I* **1977**, 309.
- Giannis, A.; Sandhoff, K. *Tetrahedron Lett.* **1985**, 26, 1479; Horton, D.; Miyake, T. *Carbohydr. Res.* **1988**, 184, 221; Garcia Martin, M.de G.; Horton, D. *Carbohydr. Res.* **1989**, 191, 223.
- Glucopyranosyl (**6**, α : β 4:1) and galactopyranosyl (**9**, α : β 9:1) fluorides were prepared from the corresponding β -thiophenyl pyranosides: Nicolaou, K.C.; Caulfield, T.J.; Kataoka, H. *Carbohydr. Res.* **1990**, 202, 177.
- Nicolaou, K.C.; Dolle, R.E.; Chucholowski, A.; Randall, J.L. *J. Chem. Soc., Chem. Commun.* **1984**, 1153.
- 7a:** mp 42–45°C; $[\alpha]_D$ +20 (c 1, CHCl_3); ^1H NMR (400 MHz, $\text{DMSO}-d_6$, 80°C) δ 7.34–7.21 (m, 20H, 4 Ph), 5.03 and 4.94 (2 bd s, 2H, $\text{CH}_2=$), 4.83 and 4.73 (2 d, 2H, J 11.5 Hz, CH_2Ph), 4.73 and 4.55 (2 d, 2H, J 11.3 Hz, CH_2Ph), 4.63 (m, 2H, CH_2Ph), 4.51 and 4.46 (2 d, 2H, J 12.1 Hz, CH_2Ph), 4.23 (ddd, 1H, $J_{4a,5}$ 11, $J_{4b,5}$ 3.3, $J_{5,6}$ 5.6 Hz, H-5), 3.78 (dd, 1H, $J_{6,7}$ 8.8, $J_{7,8}$ 8.2 Hz, H-7), 3.70 (m, 1H, H-9), 3.65 (dd, 1H, H-6), 3.62–3.58 (m, 2H, H-10a,10b), 3.60 (s, 3H, CO_2Me), 3.46 (dd, 1H, $J_{8,9}$ 9.4 Hz, H-8), 3.14 (bd s, 2H, H-2a,2b), 2.60 (dd, 1H, $J_{4a,4b}$ 15.4 Hz, H-4a), 2.41 (dd, 1H, H-4b); ^{13}C NMR (100.6 MHz, CDCl_3) δ 171.90 (C-1), 139.37, 138.80, 138.24, 138.23 and 138.08 (C-3, 4 C quat. arom.), 128.51–127.69 (20 C arom.), 116.64 ($\text{CH}_2=$), 82.42, 79.96, 78.03, 72.68 and 71.26 (C-5,6,7,8,9), 75.60, 75.22, 73.54 and 73.05 (4 CH_2Ph), 68.85 (C-10), 51.92 (CO_2CH_3), 41.31 (C-2), 30.75 (C-4). Calcd for $\text{C}_{40}\text{H}_{44}\text{O}_7$: C, 75.45; H, 6.96. Found: C, 75.44; H, 7.13.
- Bennek, J.A.; Gray, G.R. *J. Org. Chem.* **1987**, 52, 892.
- Uchiyama, T.; Hindsgaul, O. *Synlett* **1996**, 499.
- 15a:** mp 103–5°C; $[\alpha]_D$ +73 (c 1, MeOH); ^1H NMR (400 MHz, CD_3OD) δ 5.05 and 4.96 (2 bd s, 2H, $\text{CH}_2=$), 4.14 (ddd, 1H, $J_{4a,5}$ 11.2, $J_{4b,5}$ 3.1, $J_{5,6}$ 5.6 Hz, H-5), 3.94 (m, 1H, H-8), 3.91 (dd, 1H, $J_{6,7}$ 9.2 Hz, H-6), 3.76–3.64 (m, 4H, H-7,9,10a,10b), 3.67 (s, 3H, CO_2Me), 3.18 (m, 2H, H-2a,2b), 2.55 (dd, 1H, $J_{4a,4b}$ 15.3 Hz, H-4a), 2.43 (dd, 1H, H-4b); ^{13}C NMR (100.6 MHz, CD_3OD) δ 174.00 (C-1), 141.60 (C-3), 116.75 ($\text{CH}_2=$), 74.79, 73.88, 71.82, 70.04 and 69.99 (C-5,6,7,8,9), 61.96 (C-10), 52.36 (CO_2CH_3), 42.08 (C-2), 32.36 (C-4).
- 8a:** $[\alpha]_D$ +33 (c 1, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.33–7.24 (m, 20H, 4 Ph), 4.89 and 4.79 (2 d, 2H, J 11.2 Hz, CH_2Ph), 4.79 and 4.56 (2 d, 2H, J 11.7 Hz, CH_2Ph), 4.69 (m, 1H, H-5), 4.61 and 4.47 (2 d, 2H, J 11.2 Hz, CH_2Ph), 4.59 and 4.46 (2 d, 2H, J 12.2 Hz, CH_2Ph), 3.78–3.55 (m, 6H, H-6,7,8,9,10a,10b), 3.68 (s, 3H, CO_2Me), 3.48 and 3.41 (2 d, 2H, $J_{2a,2b}$ 15.8 Hz, H-2a,2b), 3.01 (dd, 1H, $J_{4a,4b}$ 15.6, $J_{4a,5}$ 5.8 Hz, H-4a), 2.82 (dd, 1H, $J_{4b,5}$ 8.1 Hz, H-4b); ^{13}C NMR (100.6 MHz, CDCl_3) δ 200.15 (C-3), 167.42 (C-1), 138.47, 138.01, 137.87 and 137.67 (4 C quat. arom.), 128.52–127.68 (20 C arom.), 81.87, 79.06, 77.57, 72.55 and 70.83 (C-5,6,7,8,9), 75.38, 75.00, 73.51 and 73.38 (4 CH_2Ph), 68.73 (C-10), 52.30 (CO_2CH_3), 49.61 (C-2), 40.24 (C-4). Calcd for $\text{C}_{39}\text{H}_{42}\text{O}_8$: C, 73.33; H, 6.63. Found: C, 73.25; H, 6.57.
- Lopez Herrera, F.J.; Uraga Baelo, C. *Carbohydr. Res.* **1985**, 139, 95; Lopez Herrera, F.J.; Uraga Baelo, C. *Carbohydr. Res.* **1985**, 143, 161; Sun, K.M.; Dawe, R.D.; Fraser-Reid, B. *Carbohydr. Res.* **1987**, 171, 35.
- Rodebaugh, R.; Debenham, J.S.; Fraser-Reid, B. *Tetrahedron Lett.* **1996**, 37, 5477.
- 18:** ^1H NMR (400 MHz, CDCl_3) δ 7.37–7.22 (m, 15H, 3 Ph), 5.02 and 4.97 (2 bd s, 2H, $\text{CH}_2=$), 4.73, 4.72, 4.56 and 4.54 (4 d, 4H, J 11.7 Hz, 2 CH_2Ph), 4.52 and 4.48 (2 d, 2H, J 11.7 Hz, CH_2Ph), 4.23 (ddd, 1H, $J_{4a,5}$ 9.7, $J_{4b,5}$ ~5.6 ~4.6 Hz, H-5), 4.13–4.01 (m, 3H, H-8,10a,10b), 3.82 (dd, 1H, $J_{6,7}$ ~10 Hz, H-6), 3.71–3.60 (m, 2H, H-7,9), 3.64 (s, 3H, CO_2Me), 3.16 and 3.11 (2 d, 2H, J 15.8

- Hz, H-2a,2b), 2.47 (dd, 1H, $J_{4a,4b}$ 15.3 Hz, H-4a), 2.39 (dd, 1H, H-4b); ^1H NMR (400 MHz, $\text{CDCl}_3 + \text{CCl}_3\text{CONCO}$) δ 8.67 (bd s, 1H, NH), 7.39-7.26 (m, 15H, 3 Ph), 5.17 (m, 1H, H-6), 5.00 and 4.98 (2 bd s, 2H, $\text{CH}_2=$), 4.76 and 4.64 (2 d, 2H, J 12 Hz, CH_2Ph), 4.66 and 4.54 (2 d, 2H, J 11.7 Hz, CH_2Ph), 4.55 (s, 2H, CH_2Ph), 4.31 (m, 1H, H-5), 4.20 (m, 1H, H-9), 4.01 (dd, 1H, $J_{9,10a} \sim 8$, $J_{10a,10b}$ 11 Hz, H-10a), 3.94 (dd, 1H, $J_{7,8}$ 2.8, $J_{8,9}$ 4.7 Hz, H-8), 3.90 (dd, 1H, $J_{6,7}$ 5.7 Hz, H-7), 3.73 (dd, 1H, $J_{9,10b}$ 3.9 Hz, H-10b), 3.64 (s, 3H, CO_2Me), 3.14 and 3.08 (2 d, 2H, J 15.6 Hz, H-2a,2b), 2.46 (dd, 1H, $J_{4a,4b}$ 15.2, $J_{4a,5}$ 8.8 Hz, H-4a), 2.34 (dd, 1H, $J_{4b,5}$ 4.5 Hz, H-4b).
- (19) Cipolla, L.; Lay, L.; Nicotra, F. *Carbohydr. Lett.* **1996**, *2*, 131.
- (20) Miginiac, P.; Zamblouty, G. *J. Organomet. Chem.* **1975**, *96*, 163.
- (21) (*E*)-**19**: mp 76-8°C; $[\alpha]_D +49$ (*c* 0.98, CHCl_3); R_f 0.16 (19:1 toluene-EtOAc); ^1H NMR (400 MHz, CDCl_3) δ 7.34-7.11 (m, 20H, 4 Ph), 5.72 (bd s, 1H, H-2), 4.93, 4.81 and 4.46 (3d, 4H, J 11.2 Hz, 2 CH_2Ph), 4.73 and 4.60 (2 d, 2H, J 11.7 Hz, CH_2Ph), 4.62 and 4.43 (2 d, 2H, J 12.2 Hz, CH_2Ph), 4.24 (ddd, 1H, $J_{4a,5}$ 11.2, $J_{4b,5}$ 2.5, $J_{5,6}$ 5.6 Hz, H-5), 3.78-3.57 (m, 6H, H-6,7,8,9,10a,10b), 3.65 (s, 3H, CO_2Me), 2.58 (dd, 1H, $J_{4a,4b}$ 14.8 Hz, H-4a), 2.48 (dd, 1H, H-4b), 2.16 (d, 3H, $J_{2,\text{Me}}$ 1 Hz, Me); ^{13}C NMR (100.6 MHz, CDCl_3) δ 166.99 (C-1), 156.98 (C-3), 138.76, 138.23, 138.19 and 138.11 (4 C quat. arom.), 128.68-127.72 (20 C arom.), 117.60 (C-2), 82.46, 80.03, 78.04, 72.48 and 71.64 (C-5,6,7,8,9), 75.67, 75.26, 73.65 and 73.63 (4 CH_2Ph), 68.80 (C-10), 50.97 (CO_2CH_3), 36.18 (C-4), 18.56 (Me). (*Z*)-**19**: $[\alpha]_D +46$ (*c* 0.99, CHCl_3); R_f 0.25 (19:1 toluene-EtOAc); ^1H NMR (400 MHz, CDCl_3) δ 7.40-7.13 (m, 20H, 4 Ph), 5.76 (bd s, 1H, H-2), 4.95 and 4.47 (2 d, 2H, J 10.7 Hz, CH_2Ph), 4.81 (m, 2H, CH_2Ph), 4.79 and 4.69 (2 d, 2H, J 11.7 Hz, CH_2Ph), 4.57 and 4.43 (2 d, 2H, J 12.2 Hz, CH_2Ph), 4.40 (ddd, 1H, $J_{4a,5}$ 11.7, $J_{4b,5}$ 3.1, $J_{5,6}$ 5.6 Hz, H-5), 3.86-3.75 (m, 3H, H-6,7,9), 3.69-3.55 (m, 3H, H-8,10a,10b), 3.64 (s, 3H, CO_2Me), 3.34 (dd, 1H, $J_{4a,4b}$ 14.2 Hz, H-4a), 2.95 (dd, 1H, H-4b), 1.92 (s, 3H, Me); ^{13}C NMR (100.6 MHz, CDCl_3) δ 166.55 (C-1), 157.86 (C-3), 138.82, 138.42, 138.37 and 138.08 (4 C quat. arom.), 128.37-127.55 (20 C arom.), 117.59 (C-2), 82.28, 80.29, 78.07, 73.70 and 71.80 (C-5,6,7,8,9), 75.53, 74.92, 73.40 and 73.23 (4 CH_2Ph), 69.00 (C-10), 50.88 (CO_2CH_3), 28.69 (C-4), 25.55 (Me).
- (22) Crimmin, M.J.; O'Hanlon, P.J.; Rogers, N.H. *J. Chem. Soc., Perkin Trans. I* **1985**, 541.
- (23) Crich, D.; Lim, L.B.L. *J. Chem. Soc., Perkin Trans. I* **1991**, 2209.
- (24) **21**: mp 113-5°C; $[\alpha]_D +44$ (*c* 0.99, CHCl_3); ^1H NMR (400 MHz, C_6D_6) δ 7.10-6.75 (m, 20H, 4 Ph), 5.41 (bd s, 1H, H-2), 4.63 and 4.26 (2 d, 2H, J 11.2 Hz, CH_2Ph), 4.57 (m, 2H, CH_2Ph), 4.16 and 3.91 (2 d, 2H, J 11.6 Hz, CH_2Ph), 4.06 and 3.98 (2 d, 2H, CH_2Ph), 3.93 and 3.81 (2 dd, 2H, $J_{2,11a} \sim J_{2,11b} \sim 1$, $J_{11a,11b}$ 17.5 Hz, H-11a,11b), 3.48 (dt, $J_{4,5} \sim 7.2$, $J_{5,6} \sim 5.9$ Hz, H-5), 3.38-3.13 (m, 6H, H-6,7,8,9,10a,10b), 1.91 (d, 2H, H-4a,4b); ^{13}C NMR (100.6 MHz, C_6D_6) δ 171.81 (C-1), 164.88 (C-3), 137.93, 137.66, 137.30 and 137.15 (4 C quat. arom.), 126.96-126.44 (20 C arom.), 116.18 (C-2), 80.76, 78.70, 77.11, 71.26 and 70.99 (C-5,6,7,8,9), 73.93, 73.72, 72.32, 72.29 and 71.43 (4 CH_2Ph , C-11), 23.39 (C-4).