Studies on 2-Aziridinecarboxylic Acid. VII.¹⁾ Formation of Dehydroamino Acid Peptides *via* Isomerization of Peptides Containing 2-Aziridinecarboxylic Acid by Tertiary Amines

Kiichiro Nakajima,* Hitomi Oda, and Kenji Okawa Department of Chemistry, Faculty of Science, Kwansei Gakuin University, Nishinomiya 662 (Received March 19, 1982)

Peptides containing 1- $(\alpha$ -aminoacyl)-2-aziridinecarboxylic acid were isomerized into the corresponding dehydroamino acid peptides by treatment with the tertiary amines, triethylamine or 1,4-diazabicyclo[2.2.2]octane (Dabco). Dehydrohydantoin derivatives were also prepared by treatment of benzyloxycarbonyl-2-aziridinecarboxylic acid derivatives with tertiary amines.

Recently many biologically active peptides containing dehydroamino acid (DHA) have been isolated and much attention has been directed to their synthesis. Literature concerning the synthesis of DHA peptides has mainly dealt with β -elimination of the O-leaving group of β -hydroxy α -amino acid derivatives, ²⁾ and a more recent paper has reported Hofmann degradation reaction of 2,3-diaminopropionic acid derivatives. ³⁾

In the previous paper,⁴⁾ we reported the isomerization of peptides containing 2-aziridinecarboxylic acid (Azyline, Azy) and 3-methyl-2-aziridinecarboxylic acid (3-MeAzy) into the corresponding DHA peptides with NaI in an acetone solution. During our investigation on the reaction of Azy peptides with several amines, we

found that they react with primary and secondary amines to form the corresponding amine adducts, α,β -diaminopropionic acid derivatives,⁵⁾ and the *trans*-acylation reaction occurs due to peptide bond cleavage.⁵⁾ Also, 1-acyl- and 1-(α -aminoacyl)-2-aziridinecarboxylic acid peptides are isomerized into the corresponding DHA peptides upon treatment with tertiary amines as in the reaction with NaI.

In the present study the reactions of Azy peptides with the tertiary amines, triethylamine and Dabco were observed in detail. The Azy and 3-MeAzy peptides used as the starting materials were synthesized by the mixed anhydride method from N-protected amino acid and Azy derivatives according to the procedure described

Table 1. Yields and properties of Azy-peptides (1, 2, 5)

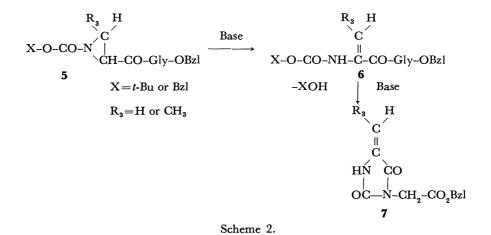
Agu pontido ⁸)	Viold/0/	M 0 /9C	F 793/0	Found (Calcd) (%)			
$egin{aligned} \mathbf{Azy} \ \mathbf{peptide^{a}} \ \downarrow \end{aligned}$	Yield/%	${ m Mp}\; heta_{ m m}/^{\circ}{ m C}$	$[\alpha]_D^{23}/^{\circ}$	$\overline{\mathbf{c}}$	H	N	
Z-Gly-Azy-NHCH ₃ (1a)	70	193 (decomp)	-38.0 (c 0.8, MeOH)	57 84 (57.72	5 81 5.88	14.38 14.43)	
Z–Gly–Azy–NHBzl (1b)	72	122.5—123.5	$5 - 90.0 (c 1.1, CHCl_3)$	66.42	5.53 5.58	11.11 11.08)	
Z–Phe–Azy–NHCH $_3$ (1c)	56	170—171	-66.9 (c 1.1, CHCl ₃)	66.08 (66.13	6.12 6.08	11.13 11.02)	
Z-Ala-Azy-OBzl (1d)	83	58—61	-50.9 (c 1.0, DMSO)	65.91 (65.96	5.73 5.80	7.29 7.33)	
Z–Phe–Azy–OBzl (1e)	72	68—69	-62.7 (c 1.0, DMSO)	70.69 (70.73	5.68 5.72	6.16 6.11)	
Z–Gly–Azy–Gly–OBzl (1f)	92	118—119	-52.5 (c 1.0, MeOH)	62.23 (62.16	5.40 5.45	9.92 9.88)	
Z-Phe-3-MeAzy-NHCH ₃ (2a)	77	129—130	-25.3 (c 1.1,CHCl ₃)	66.81 (66.82	$\begin{array}{c} 6.27 \\ 6.37 \end{array}$	10.69 10.63)	
Z-Phe-3-MeAzy-NHBzl (2b)	62	157—157.5	-35.1 (c 0.23, CHCl ₃)	71.28 (71.32	6.11 6.20	8.83 8.91)	
Z-Phe-3-MeAzy-OBzl (2c)	80	95—97	-61.1 (c 1.0, MeOH)	71.26 (71.17	6.02 5.97	5.89 5.93)	
Z–Gly–3-MeAzy–Gly–OBzl (2d)	90	123.5—124.5	5 –58.2 (c 1.0, AcOEt)	62.88 (62.86	5.70 5.73	9.68 9.56)	
Z-Ala-3-MeAzy-Gly-OBzl (2e)	93	110111	-59.7 (c 1.0, CHCl ₃)	63.48 (63.50	5.98 6.00	9.30 9.27)	
Z-Azy-Gly-OBzl (5a)	60	109—110	-50.6 (c 1.0, MeOH)	65.17 (65.21	5.50 5.47	7.71 7.60)	
Boc–Azy–Gly–OBzl (5b)	61	71—71.5	-80.4 (c 1.0, MeOH)	61.22 (61.07	$\begin{array}{c} 6.85 \\ 6.63 \end{array}$	8.35 8.34)	
Z-3-MeAzy-Gly-OBzl (5c)	73	62—64	-63.2 (c 1.0, MeOH)	65.95 (65.96	5.83 5.80	7.42 7.33)	
Boc-3-MeAzy-Gly-OBzl (5d)	79	89.5—90	-70.0 (c 1.0, MeOH)	66.21 (66.06	7.24 7.34	8.01 ['] 8.04)	

a) All Azy peptides were synthesized by the mixed anhydride method except **5a—d**. Arrow indicates final coupling position. Azy: (2S)-form, 3-MeAzy: (2S, 3S)-form.

Table 2. Yields and properties of DHA-peptides (3, 4)

Product (3 , 4)	Yield/% $^{ m Mp}_{ heta_{ m m}}$ °C	Base ^{a)}	Found	Found (Calcd) (%)		NMR data of DHA δ , DMSO- d_{6}				
			Ċ	H	N	NH		CH ₂	$^{\circ}$ CH $_{3}$	
Z-Gly-△Ala-NHCH ₃ (3a)	89	175 (decomp)	Et ₃ N(3)	57.84 57.72	5.81 5.88	14.38 14.43	9.01	5.58	6.20	
Z-Gly-⊿Ala-NHBzl (3b)	84	117.5—118.5	$\text{Et}_3N(3)$	66.42 66.48	5.53 5.58	11.11 11.08	9.07	5.60	6.24	
Z-Phe-⊿Ala-NHCH ₃ (3c)	100	syrup	Et ₃ N(6)	66.08 66.13	6.12 6.08	11.13 11.02	8.58	6.30	6.60	<u></u> b)
Z-Ala-∆Ala-OBzl (3d)	100	syrup	$\text{Et}_3N(3)$	65.91 65.96	5.73 5.80	$7.29 \\ 7.33$	8.34	5.92	6.56	b)
Z-Phe-⊿Ala-OBzl (3e)	100	syrup	$\mathrm{Et_3N}(3)$	70.69 70.73	5.68 5.72	6.16 6.11	8.12	5.92	6.60	b)
Z–Gly–∆Ala–Gly–OBzl (3f)	100	133—136	$\text{Et}_3N(6)$	62.23 62.16	5.40 5.45	9.92 9.88	8.99	5.57	6.31	-
Z-Phe-⊿Aba-NHCH ₃ (4a)	77	165—167	Dabco(24)	66.81 66.82	6.27 6.37	10.69 10.63	9.05		6.36q	1.40d
Z-Phe-⊿Aba-NHBzl (4b)	88	162.5—163.5	Dabco(24)	71.28 71.32	6.11 6.20	8.83 8.91	9.15	_	6.44q	1.40d
Z–Phe–⊿Aba–OBzl (4c)	90	151—152	Dabco(6)	71.26 71.17	6.02 5.97	$\frac{5.89}{5.93}$	9.20		6.67q	1.68d
Z-Gly-∆Aba-Gly-OBzl (4d)	87	101—102	Dabco(24)	62.88 62.86	5.70 5.73	$9.68 \\ 9.56$	9.01		6.55q	1.66d
Z–Ala–⊿Aba–Gly–OBzl (4e)	87	139—140.5	Dabco(24)	$63.48 \\ 63.50$	5.98 6.00	$9.30 \\ 9.27$	9.02	-	6.52q	1.65d

a) The reaction was carried out in a CHCl₃ solution at 35 °C and the base used was 2 equimolar against Azy peptide. The values in parenthesis are the reaction time/h. b) NMR spectrum in CDCl₃.



in our previous papers.⁵⁻⁷⁾ Their properties and yields are summarized in Table 1 and the reaction procedure of the Azy peptides with amines is shown in Scheme 1.

Azy and 3-MeAzy peptides (1a—f and 2a—e) were treated with 2 equimolar amount of tertiary amine (Et₃N or Dabco) in CHCl₃ solution at 35 °C for 3—24 h, and then the corresponding dehydroamino acid (DHA, Δ AA) derivatives were isolated by silica-gel column

chromatography. The DHA produced was identified by NMR.

In the NMR data, characteristic signals of the DHA structure appeared at δ 5.5—5.9 for the vinyl proton of dehydroalanine (Δ Ala), and at δ 1.4—1.7 as a doublet for the methyl proton and at δ 6.3—6.8 as a quartet for the β -proton of dehydro- α -aminobutyric acid (Δ Aba). Δ Aba was of the (Z)-configuration. The product (3, 4)

Table 3. Boc- and Z-△AA-GLY-OBZL (6) and dehydrohydantoin (7)

A	Base	React	Product yields/%		
Azy peptide (5)		time/h	DHA (6)	Hydantoin (7)	
Z-Azy-Gly-OBzl (5a)	Et ₃ N Dabco	12 6	_	75 80	
Boc-Azy-Gly-OBzl (5 b)	Et ₃ N Dabco	24 6	95 100		
Z-3-MeAzy-Gly-OBzl~(5c)	Et₃N Dabco	7 ^{a)} 3 ^{a)}		 28	
Boc-3-MeAzy-Gly-OBzl (5d)	Et ₃ N Dabco	7 ^{a)} 4 ^{a)}	 85		

a) Reaction time/d.

of the isomerization of the Azy peptides are summarized in Table 2.

The reaction results show that 1-acyl-2-aziridine-carboxylic acid peptides $(1\mathbf{a}-\mathbf{f})$ were easily isomerized into the corresponding Δ Ala peptides $(3\mathbf{a}-\mathbf{f})$ by the treatment of triethylamine, but contrary to our expectation, 1-acyl-3-methyl-2-aziridinecarboxylic acid peptides $(2\mathbf{a}-\mathbf{e})$ were not isomerized into the Δ Aba peptides $(4\mathbf{a}-\mathbf{e})$, even after 7 d of treatment. Use of the more basic Dabco as an isomerizing reagent was necessary to obtaine Δ Aba peptides $(4\mathbf{a}-\mathbf{e})$.

This isomerization reaction seems to be induced by a direct withdrawing of the α -proton of Azy or 3-MeAzy with base. The α -proton of unsubstituted Azy ($1\mathbf{a}$ — \mathbf{f}) could be easily withdrawn by Et₃N treatment, but not that of the substituted 3-MeAzy ($2\mathbf{a}$ — \mathbf{e}) because of the methyl group on the aziridine ring. Thus, the more basic Dabco was needed to withdraw the α -proton of 3-MeAzy for its isomerization.

We also examined the reaction of 1-benzyloxycarbonyl- and 1-t-butoxycarbonyl-2-aziridinecarboxylic acid peptides (5a—d) with amines. As shown in Scheme 2 and Table 3, the reaction of the Azy peptides blocked by urethane-type groups with tertiary amines afforeded not only the corresponding DHA peptides, but also dehydrohydantoin derivatives (7a, 7b). Again, the 3-MeAzy peptides (5c, 5d) were not isomerized with Et₃N and the benzyloxycarbonyl derivatives were more easily isomerized into the corresponding DHA peptides than the t-butoxycarbonyl peptides. The results of the dehydrohydantoin (7) formation reaction, which occurred via reaction of the DHA peptides produced by the reaction with tertiary amines, agreed very well with the results of the β -elimination reaction of O-activated Z-Ser- or Z-Thr- derivatives found by Campbell and Behr,8) us,9) and, Shin et al.10) Thus the benzyloxycarbonyl group of the aziridine is more labile than the t-butoxycarbonyl group in the basic conditions, that is, Z-Azy-Gly-OBzl (5a) and Z-3-MeAzy-Gly-OBzl (5c) isomerized more easily than Boc-Azy-Gly-OBzl (5b) and Boc-3-MeAzy-Gly-OBzl (5d) as shown in Table 3. The benzyloxycarbonyl group attached to the unsubstituted aziridinecarboxylic acid (5a) seemed to be very labile and Z-Ala-Gly-OBzl could not be isolated after the reaction of 5a with tertiary amine, even in $Et_3N.$

From the isomerization reaction of Azy and 3-MeAzy described above, it is concluded that Azy and 3-MeAzy

has a distinctly different reactivity against tertiary amines and the isomerization by the tertiary amines, which can be used to the selective synthesis of DHA peptides, differs from that by NaI or the synthesis of DHA peptides via β -elimination of β -hydroxy α -amino acid

Experimental

Uncorrected melting points are reported. The homogeneity of the products was checked by thin-layer chromatography on silica-gel plates. The optical rotations were determined at the D line on a Perkin-Elmer 141 polarimeter. The NMR spectra were obtained with a Hitachi R20B high-resolution NMR spectrometer, the chemical shifts being obtained using TMS as the internal reference. All the N-aminoacyl Azy and 3-MeAzy peptides used in this study were synthesized by the method described in literature, 6,7) and their properties and yield are summarized in Table 1.

(2S)-Z-Azy-Gly-OBzl (5a). To a solution of (2S)-H-Azy-Gly-OBzl¹¹⁾ (1.63 g, 6.96 mmol) and Et₃N (0.97 ml, 6.96 mmol) in CHCl₃ (15 ml), was added Z-ON¹⁾ (1.95 g, 6.96 mmol) with stirring at 0 °C. After being stirred overnight at 25 °C, the solvent was removed in vacuo and the residue was dissolved in ethyl acetate. The ethyl acetate solution was washed with 10% citric acid, 1 M NaHCO₃ and water, and dried over Na₂SO₄, concentrated in vacuo. The residue was crystallized from ethyl acetate-hexane, 1.53 g (60%) of 5a was obtained, mp 109—110 °C, [a]_D²³ -50.6° (c 1.0, MeOH). NMR (CDCl₃) δ: 2.38 (1H q, $J_{\rm gem}$ =0.8 Hz, $J_{\rm trans}$ =3.5 Hz, Azy β-proton), 2.53 (1H q, $J_{\rm gem}$ =0.8 Hz, $J_{\rm cis}$ =6.3 Hz, Azy β-proton), 3.07 (1H q, $J_{\rm trans}$ =3.5 Hz, $J_{\rm cis}$ =6.3 Hz, Azy α-proton). Z-ON can be replaced with Z-Cl.¹⁾

Found: C, 65.17; H, 5.50; N, 7.71%. Calcd for $C_{20}H_{20}-O_5N_2$: C, 65.21; H, 5.47; N, 7.60%.

(2S)-Boc-Azy-Gly-OBzl (5b). To a solution of (2S)-H-Azy-Gly-OBzl¹¹ (4.0 g, 17.1 mmol) and Et₃N (2.38 ml, 17.1 mmol) in CHCl₃ (50 ml), was added Boc-ON¹ (4.63 g, 18.8 mmol) with stirring at 0 °C. After the reaction mixture had been worked up as described above, the product was crystallized from ethyl acetate-hexane, 3.51 g (61.3%) of **5b** was obtained. mp 71.0—72.5 °C, [α]₂²³ -80.4° (c 1.0, MeOH). NMR (CDCl₃) δ: 2.30 (1H q, J_{gem} =0.8 Hz, J_{trans} =3.8 Hz, Azy β-proton), 2.49 (1H q, J_{gem} =0.8 Hz, J_{cis} =6.6 Hz, Azy g-proton), 3.03 (1H q, g_{trans}=3.8 Hz, g_{cis}=6.6 Hz, Azy g-proton).

Found: C, 61.22; H, 6.85; N, 8.35%. Calcd for $C_{17}H_{22}-O_5N_2$: C, 61.07; H, 6.63; N, 8.34%.

(2S, 3S)-Z-3-MeAzy-Gly-OBzl (5c). To a solution of (2S, 3S)-H-3-MeAzy-Gly-OBzl⁷⁾ (1.87 g, 7.54 mmol) and Et₃N (1.05 ml, 7.54 mmol) in CHCl₃ (15 ml), was added

Z–ON¹¹ (1.95 g, 7.54 mmol) with stirring at 0 °C. The reaction mixture was stirred overnight and worked up as described above. The product was crystallized from ethyl acetate–hexane, 2.10 g (72.8%) of **5c** was obtained, mp 62—64 °C, $[a]_{2}^{23}$ –63.2° (c 1.0, MeOH). NMR (CDCl₃) δ : 1.28 (3H d, J=5.5 Hz, CH₃), 2.79 (1H m, 3-MeAzy, β -proton), 3.14 (1H d, J=7.0 Hz, 3-MeAzy, α -proton). Z–ON can be replaced with Z–Cl.

Found: C, 65.96; H, 5.83; N, 7.42%. Calcd for $C_{21}H_{22}$ - O_5N_2 : C, 65.96; H, 5.80; N, 7.33%.

(2S, 3S)-Boc-3-MeAzy-Gly-OBzl (5d). To a solution of (2S, 3S)-H-3-MeAzy-Gly-OBzl⁷⁾ (1.25 g, 5.06 mmol) and Et₃N (0.70 ml, 5.06 mmol) in CHCl₃ (15 ml), was added Boc-ON¹⁾ (1.40 g, 6.07 mmol) with stirring at 0 °C. The reaction mixture was stirred overnight and worked up as described above. The product was crystallized from ethyl acetate-hexane, 1.38 g (78.5%) of 5d was obtained, mp 89.5—90.0 °C, [α]₂³³ -70.0° (c 1.0, MeOH). NMR (CDCl₃) δ: 1.27 (3H d, J=5.5 Hz, CH₃), 2.74 (1H m, 3-MeAzy β-proton), 3.09 (1H d, J=6.4 Hz, 3-MeAzy α-proton).

Found: C, 62.21; H, 6.88; N, 8.01%. Calcd for $C_{18}H_{24}-O_5N_2$: C, 62.06; H, 6.94; N, 8.04%.

Z–Gly– ΔA la–NHBzl (3b). General Isomerization Procedure of Azy with Et₃N: A solution of **1b** (100 mg, 0.27 mmol) and Et₃N (0.08 ml, 0.54 mmol) in CHCl₃ (5 ml) was stirred at 35 °C for 3 h. After the solvent had been removed in vacuo, the residue was dissolved in ethyl acetate. The ethyl acetate solution was washed with 10% citric acid and water, and then dried over Na₂SO₄, concentrated in vacuo. After the obtained residue was purified by the silica-gel column chromatography, the product was crystallized from CHCl₃–ether–hexane, 84.3 mg (84.3%) of **3b** was obtained, mp 117.5—118.5 °C.

Z-Gly-\(\Delta Aba-Gly-OBzl\) (4d). General Isomerization Procedure of 3-MeAzy with Dabco: A solution of 2d (200 mg, 0.45 mmol) and Dabco (102 mg, 0.91 mmol) in CHCl₃ (5 ml) was stirred at 35 °C for 24 h. After the reaction mixture had been worked up as described above, the residue was crystallized from ethyl acetate-hexane, 174 mg (87%) of 4d was obtained, mp 101—102 °C [lit, mp 102—104 °C⁴].

Benzyl 5- Methylenehydantoin - 3-acetate (7a). Using Et_3N : A solution of $\mathbf{5a}$ (200 mg, 0.54 mmol) snd Et_3N (0.15 ml, 1.09 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 12 h. After the reaction mixture had been worked up as described above, the residue was purified with silica-gel column chromatography and was crystallized from ethyl acetate—ether—hexane, 105 mg (75%) of $\mathbf{7a}$ was obtained, mp 171—172 °C. NMR (CDCl₃) δ : 4.32 (2H s), 4.92, (1H d, J=1.8 Hz), 5.15, 5.17 (4H 2s), 5.42 (1H d, J=1.8 Hz), 7.32 (5H s), 8.35 (1H bs).

Found: C, 60.05; H, 4.72; N, 10.72%. Calcd for $C_{13}H_{12}-O_4N_2$: C, 59.99; H, 4.65; N, 10.77%.

 $Z-\Delta Ala-Gly-OBzl$ (6a) could not be prepared by this procedure.

Using Dabco: A solution of **5a** (200 mg, 0.54 mmol) and Dabco (121 mg, 1.08 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 6 h. After the reaction mixture had been worked up as described above, 112 mg (80%) of **7a** was obtained. **6a** could not be prepared.

Boc-ΔAla-Gly-OBzl (6b). Using Et₃N: A solution of 5b (100 mg, 0.29 mmol) and Et₃N (0.08 ml, 0.58 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 24 h. After the reaction mixture had been worked up as described above, 95 mg (95%) of 6b was obtained as an oily material. NMR (CDCl₃) δ: 1.48 (9H s), 4.12 (2H d, J=5.0 Hz) 5.15 (1H m) 5.18 (2H s), 6.00 (1H d, J=1.8 Hz), 6.78 (1H bt), 7.25 (1H bs), 7.32 (5H s). Found: C, 61.15; H, 6.72; N, 8.28%. Calcd for C₁₇H₂₂-O₅N₂: C, 61.07; H, 6.63; N, 8.34%. 7b could not be prepared. Using Dabco: A solution of 5b (300 mg, 0.86 mmol) and

Dabco (194 mg, 1.73 mmol) in CHCl₃ (5 ml) was stirred at 35 °C for 6 h. After the reaction mixture had been worked up as described above, 300 mg (100%) of **6b** was obtained as an oily material. **7b** could not be prepared.

Z-ΔAba-Gly-OBzl (6c) and Benzyl 5-Ethylidenehydantoin-3-acetate (7c). Using Dabco: A solution of 5c (200 mg, 0.5 mmol) and Dabco (112 mg, 1 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 3 d. After the reaction mixture had been worked up as described above, the crude product of two components was subjected to silica-gel column chromatography developed by CHCl₃, yielding 136 mg (68%) of 6c, mp 99—100 °C, and 38 mg (28%) of 7c, mp 186—186.5 °C. 6c: NMR (CDCl₃) δ: 1.70 (3H d, J=7.0 Hz), 4.16 (2H d, J=5.5 Hz), 5.12 (2H s), 5.20 (2H s), 6.52 (1H q, J=7.0 Hz), 6.76 (1H bs), 7.30 (10H s), 7.00 (1H bs), 7.15 (1H bs). 7c: NMR (CDCl₃) δ: 1.83 (3H d, J=7.5 Hz), 4.35 (2H s), 5.19 (2H s), 5.98 (1H q, J=7.5 Hz), 7.32 (5H s), 9.05 (1H bs).

6c; Found: C, 66.02; H, 5.86; N, 7.23%. Calcd for $C_{21}H_{22}$ - O_5N_2 : C, 65.96; H, 5.80; N, 7.33%.

7c; Found: C, 61.28; H, 5.20; N, 10.32%. Calcd for $C_{14}H_{14}O_4N_2$: C, 61.31; H, 5.15; N, 10.21%.

Using Et₃N: A solution of **5c** (200 mg, 0.5 mmol) and Et₃N (0.14 ml, 1 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 7 d. No reaction occurred and only **5c** was recovered.

Boc-ΔAba-Gly-OBzl (6d). Using Dabco: A solution of 5d (200 mg, 0.57 mmol) and Dabco (128 mg, 1.14 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 4 d. After the reaction mixture had been worked up as described above, 170 mg (85%) of 6d was obtained, mp 119.5—120.5 °C. NMR (CDCl₃) δ: 1.45 (9H s), 1.75 (3H d, J=7.0 Hz), 4.14 (2H d, J=5.5 Hz), 5.20 (2H s), 6.49 (1H q, J=7.0 Hz), 6.69 (1H bs), 7.34 (5H s), 7.28 (1H bs).

Found: C, 62.18; H, 6.81; N, 8.12%. Calcd for $C_{18}H_{24}-O_{5}N_{2}$: C, 62.06; H, 6.94; N, 8.04%.

7d could not be prepared.

 U_{sing} Et_3N : A solution of **5d** (100 mg, 0.29 mmol) and Et_3N (0.08 ml, 0.58 mmol) in CHCl₃ (3 ml) was stirred at 35 °C for 7 d. No reaction occurred and only **5d** was recovered.

References

- 1) Part VI: K. Nakajima, M. Neya, S. Yamada, and K. Okawa, Bull. Chem. Soc. Jpn., 55, 3049 (1982). The abbreviations of the IUPAC-IUB Commission (J. Biol. Chem., 247, 977 (1972)) are used. Z: benzyloxycarbonyl, OBzl: benzyl ester, Z-Cl: benzyloxycarbonyl chloride, Z-ON: 2-benzyloxycarbonyloxyimino-2-phenylacetonitrile. Boc-ON: 2-t-butoxycarbonyloxyimino-2-phenylacetonitrile. "Azyline" is used as the name of 2-aziridinecarboxylic acid, "Azy" being its abbreviation. 3-MeAzy: (2S, 3S)-3-methyl-2-aziridinecarboxylic acid. △Ala: Dehydroalanine, △Aba: dehydro-2-aminobutylic acid.
- 2) A. Srinivasan, R. W. Stephanson, and K. Olsen, J. Org. Chem., 42, 2253 (1977); D. H. Rich and J. P. Tam, Tetrahedron Lett., 1975, 211 (1975); C. Shin, Y. Yonezawa, M. Takahashi, and J. Yoshimura, Bull. Chem. Soc. Jpn., 54, 1132 (1981); D. H. Rich and J. P. Tam, J. Org. Chem., 42, 3815 (1977); S. Konno and C. H. Stammer, Synthesis, 1978, 598 (1978).
- 3) S. Nomoto, A. Sano, and T. Shiba, Tetrahedron Lett., 1979, 521 (1979).
- 4) K. Okawa, K. Nakajima, T. Tanaka, and M. Neya, Bull. Chem. Soc. Jpn., 55, 174 (1982).
- 5) K. Nakajima, M. Iwai, and K. Okawa, "Peptide Chemistry 1980," ed by K. Okawa, Protein Research

Foundation, Osaka (1981), p. 5; K. Nakajima, T. Tanaka, K.

- Morita, and K. Okawa, *Bull. Chem. Soc. Jpn.*, **53**, 283 (1980).

 6) K. Nakajima, F. Takai, T. Tanaka, and K. Okawa, *Bull. Chem. Soc. Jpn.*, **51**, 1577 (1978).
- 7) K. Okawa, K. Nakajima, T. Tanaka, and Y. Kawana, Chem. Lett., 1975, 591.
- 8) R. D. Campbell and F. E. Behr, J. Org. Chem., 38, 1183 (1973).
- K. Nakajima, H. Kawai, M. Takai, and K. Okaw, Bull. Chem. Soc. Jpn., 50, 917 (1977).
- 10) C. Shin, Y. Yonezawa, T. Yamada, H. Hirano, and J. Yoshimura, 43rd National Meeting of the Chemical Society of Japan, Tokyo, April 1981, Abstr. No. 1G16.
- 11) K. Nakajima, K. Kanda, and Okawa, "Peptide Chemistry 1977," ed by T. Nakajima, Protein Research Foundation, Osaka (1977), p. 17.