

Carbohydrate Research 278 (1995) 351-362

CARBOHYDRATE RESEARCH

Note

Synthesis of 2-acetamido-2-deoxy- β -Dglucopyranosyl- $(1 \rightarrow 2) - \alpha$ -D-mannopyranosyl- $(1 \rightarrow 6) - \beta$ -D-mannopyranosyl- $(1 \rightarrow 4) - 2$ acetamido-2-deoxy-D-glucopyranose. Acceptor-substrate recognition by *N*-acetylglucosaminyltransferase-V(GnT-V) *

Shaheer H. Khan *,¹, Khushi L. Matta *

Department of Gynecologic Oncology, Roswell Park Cancer Institute, Elm and Carlton Streets, Buffalo, NY 14263, USA

Received 4 May 1995; accepted 23 June 1995

Keywords: N-Acetylglucosaminyltransferase-V; Glycosyltransferase; Glycosyltransferase assay; Acceptor analog; Oligosaccharide synthesis

The enzyme UDP-Glc pNAc: α -D-mannopyranosyl- $(1 \rightarrow 6)$ -N-acetyl- β -D-glucosaminyltransferase (Glc pNAc-transferase V or GnT-V, EC 2.4.1.155) catalyzes the transfer of a β -D-Glc pNAc residue to O-6 of the $(1 \rightarrow 6)$ -linked α -D-mannosyl residue that forms the part of the trimannosyl core of oligosaccharides having structure 1 (Fig. 1) [2,3]. Recent years have seen a great surge of interest in GnT-V because of its increased activity in cells transformed by viruses [4,5] or by oncogenes [6] and its involvement in cancer metastasis [7-9]. More recently interest became enhanced when this enzyme was shown to be substantially elevated in malignant human breast cancer biopsies [10]. This enzyme has therefore become an attractive target for the synthesis of glycosyltransferase inhibitors which might have the potential to prevent metastasis [11-18].

^{*} Synthetic studies in Carbohydrates, part 98. For part 97, see ref. [1].

^{*} Corresponding authors.

¹ Present address: Perkin-Elmer, Applied Biosystems Division, 850 Lincoln Centre Drive, Foster City, CA 94404, USA



Fig. 1. Natural heptasaccharide acceptor 1 for GnT-V compared with the potential synthetic acceptor 2.

Pioneering work by Tahir and Hindsgaul [19] has established that oligosaccharides much smaller than 1 can be effectively used to assay the activity of this enzyme [20,21]. Since then, this enzyme has been the subject of extensive investigations [22–24], and several laboratories including our own have reported the synthesis of various acceptor analogs in order to probe the enzyme's acceptor binding site [25–33]. Thus, in a continuing effort to shed more light on the substrate specificity of GnT-V, we describe herein the synthesis of title tetrasaccharide 2, which represents a part of the natural heptasaccharide acceptor (see structure 1, Fig. 1). It should be pointed out that compound 2 also proved useful [34] in specificity studies of β -(1 \rightarrow 4)-Nacetylglucosaminyltransferase (GnT-VI') acting on the α -3 and α -6 arms of N-linked oligosaccharides.

A retrosynthetic analysis of the target molecule 2 suggested, as the key intermediates, the disaccharide donor 14 and disaccharide acceptor 13. (See Schemes 1-3.) The synthesis of donor 14 has already been reported [29,31]. Therefore, a synthetic route



towards 13 was sought. The synthesis of 13 entailed two key problems; (i) the glycosylation of 6 at the C-4 hydroxyl which is known [35,36] to be a very unreactive position, and (ii) the formation of a β -D-mannopyranosyl linkage. The problem of synthesizing a β -(1 \rightarrow 4)-linkage was circumvented by the use of methods developed by Fraser-Reid et al. [37] and van Boom et al. [38] which give respectable yields of the (1 \rightarrow 4)-trans disaccharide. For the synthesis of a β -D-Man residue we employed a route that involves an oxidation-reduction sequence at O-2 of the corresponding β -D-Glc p derivative resulting in epimerization at C-2 [39]. Thus, the preparation of 12 was achieved as follows. The condensation of an α , β mixture of thioglycoside 4/5 with alcohol 6 [40] in the presence of iodonium ion (NIS-TfOH) gave the fully protected



disaccharide 7 (53%), and subsequent O-deacetylation of 7 gave 8 (93%). Thioglycoside 4/5 was readily prepared in 76% yield by treating its precursor 1,2,4,6-tetra-O-acetyl-3-O-benzyl- β -D-glucopyranose (3) [41] with phenyl thiotrimethylsilane in the presence of trimethylsilyl trifluoromethanesulfonate according to a literature procedure [42]. Benzylidenation of 8 with α , α -dimethoxytoluene in N,N-dimethylformamide and in the presence of 4-toluenesulfonic acid afforded the 4,6-O-benzylidene derivative 9 (84%)



which was oxidized by the procedure of Albright and Goldman [43] to give benzyl 3-O-benzyl-4,6-O-benzylidene- β -D-arabino-hexopyranosyl-2-ulose- $(1 \rightarrow 4)$ -2-acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranoside (10, 97%). Stereoselective reduction of 10 by the action of sodium borohydride gave 11 (66%), together with a small proportion of the D-glucose derivative 9 (11%), which was removed by column chromatography and recycled. Although various excellent methods for the synthesis of β -D-mannopyranosides have been reported [44-54] over the years, the method we applied here still remains one of the few practical methods of choice [39,55-61].

Acetylation of 11 (2:1 Py-Ac₂O) into the corresponding O-acetate 12 (98%), followed by cleavage of the benzylidine acetal group with hot, 60% aq acetic acid gave benzyl 2-O-acetyl-3-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-3,6-di-O-benzyl-2deoxy- α -D-glucopyranoside (13) (82%). Regioselective glycosylation of 13 with 2acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-acetyl- α -D-mannopyranosyl bromide (14) [26,28] under Helferich-type conditions afforded the partially protected tetrasaccharide 15 (56%). Zemplén transesterification of 15 gave 16 (63%) which was subjected to hydrogenolysis in glacial acetic acid and in the presence of 10% Pd-C to afford the title tetrasaccharide 2 (70%) as a white amorphous powder.

Preliminary evaluation [62] of tetrasaccharide 2 as acceptor for GlcNAcT-V shows that it is very poorly recognized by the enzyme.

1. Experimental

General methods.—Optical rotations were measured at $22 \pm 2^{\circ}$ with a Perkin-Elmer 241 polarimeter. TLC was conducted on aluminum sheets, precoated with 0.2-mm layers of Silica Gel 60F-254 (E. Merck); the compounds were located by UV light and/or by charring with 5% sulfuric acid. Column chromatography was performed on silica gel (Baker Analyzed, 60-200 mesh). The following solvent systems (v/v) were used for chromatography: A, 2:1 hexane-ethyl acetate; B, 1:1 hexane-ethyl acetate; C, 24:1 chloroform-methanol; D, 2:1 toluene-ethyl acetate; E, 97:3 dichloromethane-methanol; F, 19:1 dichloromethane-methanol; G, 9:1 chloroform-methanol; H, 26:12:1 chloroform-methanol-water; I, 13:6:1 chloroform-methanol-water; J, 10:9:1 chloroformmethanol-water; K, 5:4:1 chloroform-methanol-water. IR spectra were recorded with a Nicolet 20 SX FTIR spectrometer using thin film on NaCl plates. ¹H NMR spectra were recorded at 300 MHz (Bruker AM 300) for solutions in CDCl₂ or CD₂OD (internal Me₄Si, δ 0) or D₂O (internal acetone, δ 2.225). ¹³C NMR spectra were recorded at 75.5 MHz (Bruker AM 300) for solutions in CDCl₃ or CD₃OD (internal Me₄Si, δ 0) or D_2O (external 1% 1,4-dioxane in D_2O , δ 67.4). Only partial NMR data are reported, and the assignments of ¹³C chemical shifts are tentative. Fast atom bombardment mass spectra (FABMS) were obtained using an AEI MS-9 instrument with xenon as the bombarding gas and 5:1 1,4-dithiothreitol-1,4-dithioerythritol as matrix. Unless otherwise indicated, all reactions were carried out at ambient temperatures. Solutions were dried with Na₂SO₄ and concentrated at 40–50°C/2 kPa. Elemental analyses were performed by Robertson Laboratory, 29 Samson Ave., Madison, NJ 08940.

Phenyl 2,4,6-tri-O-acetyl-3-O-benzyl-1-thio- α - (4) and - β -D-glucopyranoside (5).— To a cold (0°C bath), stirred solution of 3 (5 g, 11.4 mmol) in dry dichloroethane (25 mL) were added trimethylsilyl trifluoromethanesulfonate (5.29 mL, 27.35 mmol) and phenyl thiomethylsilane (6.48 mL, 34.2 mmol). After stirring at 0°C for 4 h, the mixture was allowed to warm to room temperature, and stirring was continued overnight. The mixture was then diluted with dichloromethane (100 mL), successively washed with water, satd NaHCO₃, and water, dried and concentrated to a syrup that was chromatographed (5 \rightarrow 20% ethyl acetate in hexane). Evaporations of earlier fractions gave a solid residue that was dissolved in ethyl acetate. Addition of ether-petroleum ether caused the precipitation of 4 (0.56 g, 10%) as an amorphous solid: $[\alpha]_D + 173.5^\circ$ (c 1.4, chloroform); TLC (solvent A): R_f 0.44; ¹H NMR (CDCl₃): δ 7.48–7.24 (m, 10 H, Ar), 5.92 (d, 1 H, $J_{1,2}$ 5.8 Hz, H-1), 5.14–5.05 (m, 2 H, H-2, H-4), 4.77, 4.65 (2d, 1 H each, J_{gem} 11.5 Hz, PhC H_2), 4.5–4.41 (m, 1 H, H-5), 4.22 (dd, 1 H, $J_{5,6a}$ 5.5, $J_{6a,6b}$ 12 Hz, H-6a), 4.02 (dd, 1 H, $J_{5,6a}$ 2.25, $J_{6a,6b}$ 12 Hz, H-6b), 3.92 (t, 1 H, $J_{2,3} = J_{4,5} = 9.5$ Hz, H-3), 2.1, 2.02, and 1.96 (s, 3 H each, 3 OAc); ¹³C NMR (CDCl₃): δ 170.64, 169.77, 169.47 (COCH₃), 85.24 (C-1), 77.83 (C-3), 75.06 (PhCH₂O), 73.26, 69.73, and 69.69 (C-2,4,5), 62.23 (C-6), 20.89, 20.74, and 20.70 (COCH₃). Anal. Calcd for C₂₅H₂₈O₈S (488.56): C, 61.46; H, 5.78. Found: C, 61.55; H, 5.86.

Continued elution of the column and evaporation of solvent gave mixed fractions (4 and 5, 1.43 g, 25.7%), followed by a pure solid residue that was dissolved in ethyl acetate. Addition of ether-petroleum ether caused the precipitation of 5 (2.23 g, 40%) as an amorphous solid: $[\alpha]_D - 18.2^{\circ}$ (c 1.3, chloroform); TLC (solvent A): R_f 0.33; ¹H NMR (CDCl₃): δ 7.54-7.17 (m, 10 H, Ar), 5.12-5.01 (m, 2 H, H-2, H-4), 4.64 (d, 1 H, $J_{1,2}$ 10 Hz, H-1), 4.6, 4.57 (2d, 1 H each, J_{gem} 11 Hz, PhC H_2), 4.16 (m, 2 H, H-6a, H-6b), 3.72 (t, 1 H, $J_{2,3} = J_{4,5} = 9.5$ Hz, H-3), 3.68-3.59 (m, 1 H, H-5), 2.08, 2.03, and 1.96 (s, 3 H each, 3 OAc); ¹³C NMR (CDCl₃): δ 170.67, 169.33, 169.22 (COCH₃), 86.26 (C-1), 81.56 (C-3), 74.24 (PhCH₂O), 76.14, 71.39, and 69.67 (C-2,4,5), 62.59 (C-6), 20.97, and 20.77 (2C) (COCH₃). Anal. Calcd for C₂₅H₂₈O₈S (488.56): C, 61.46; H, 5.78. Found: C, 61.66; H, 5.84.

Benzyl 2,4,6-tri-O-acetyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-acetamido-3,6di-O-benzyl-2-deoxy- α -D-glucopyranoside (7).—A mixture of thioglycosides 4 and 5 (2.98 g, 6.1 mmol) and alcohol 6 (2 g, 4.1 mmol) was dissolved in dry dichloromethane (21 mL). Pulverized activated molecular sieves (4 Å, 1.5 g) and N-iodosuccinimide (2.27 g, 10.1 mmol) were added, and the mixture was stirred for 30 min in the dark in an atmosphere of argon. The mixture was then cooled (0°C bath) and a solution of trifluoromethanesulfonic acid (42 μ L) in dichloromethane (42 mL) was added dropwise, and the stirring was continued for 1.5 h. It was then diluted with dichloromethane (200 mL), and the solids were filtered off (Celite bed) and washed with dichloromethane. The filtrate and washings were combined, successively washed with water, aq NaHCO₃, aq Na₂S₂O₃, and water, dried and concentrated to dryness. Evaporation of the solvent and purification of the residue by chromatography $(15 \rightarrow 50\%)$ ethyl acetate in hexane) gave first unchanged 6 (0.52 g). Continued elution of the column and concentration of the fractions corresponding to the product gave a solid that was dissolved in a little methanol. Addition of ether-hexane caused the precipitation of 7 (1.85 g, 52.3%) as an amorphous solid: $[\alpha]_D + 64^\circ$ (c 1.7, chloroform); TLC (solvent B): $R_f 0.1$; ¹H NMR (CDCl₃): δ 7.43–7.19 (m, 20 H, Ar), 4.92 (d, 1 H, $J_{1,2}$ 4 Hz, H-1), 1.97, 1.94, 1.92 (s, 3 H each, 3 OAc), and 1.73 (s, 3 H, NAc); ¹³C NMR (CDCl₃): δ 170.74, 169.73, 169.29, 168.97 (COCH₃), 100.45. (C-1'), 97.08 (C-1), 80.43 (C-4), 74.06, 73.86, 73.66, 69.93 (PhCH₂O), 67.61 (C-6), 61.98 (C-6'), 52.26 (C-2), and 23.25, 20.91, 20.77, 20.67 (COCH₃). Anal. Calcd for C₄₈H₅₅NO₁₄ (869.97): C, 66.27; H, 6.37; N, 1.61. Found: C, 66.55; H, 6.76; N, 1.91.

Benzyl 3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-acetamido-3,6-di-O-benzyl-2deoxy- α -D-glucopyranoside (8).—Compound 7 (1.51 g, 1.74 mmol) was taken up in 0.1 M methanolic sodium methoxide (33 mL) and stirred overnight at room temperature. The base was neutralized by Amberlite IR-120 (H⁺) cation-exchange resin. The resin was filtered off (Celite bed), thoroughly washed with methanol, and the filtrate and washings were combined and concentrated to give **8** (1.2 g, 93%) as an amorphous solid: $[\alpha]_D$ + 80° (c 0.7, chloroform); TLC (solvent C): R_f 0.56; ¹H NMR (CDCl₃): δ 7.40–7.20 (m, 20 H, Ar), 5.32 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 4.90 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 4.28 (dt, $J_{2,3}$ 10.0 Hz, H-2), and 1.80 (s, 3 H, NAc); ¹³C NMR (CDCl₃): δ 169.80 (COCH₃), 102.64 (C-1'), 97.20 (C-1), 74.72, 74.23, 73.79, 69.92 (PhCH₂O), 68.43 (C-6), 62.41 (C-6'), 52.38 (C-2), and 23.36 (COCH₃). Anal. Calcd for C₄₂H₄₉NO₁₁ (743.86): C, 67.82; H, 6.64; N, 1.88. Found: C, 67.85; H, 6.76; N, 1.91.

Benzyl 3-O-benzyl-4,6-O-benzylidene-β-D-glucopyranosyl-(1 → 4)-2-acetamido-3,6di-O-benzyl-2-deoxy-α-D-glucopyranoside (9).—To a stirred solution of 8 (1.5 g, 2.02 mmol) in N,N-dimethylformamide (35 mL) were added 4-toluenesulfonic acid (50 mg, 0.26 mmol) and α,α-dimethoxytoluene (1.52 mL, 10.12 mmol), and the stirring was continued overnight at room temperature. The acid was then neutralized with a little triethylamine, and the solution was concentrated to a syrup that was dissolved in ethyl acetate. Addition of ether-hexane caused the precipitation of 9 (1.4 g, 83.5%): $[\alpha]_D$ +74° (c 1, chloroform); TLC (solvent D): R_f 0.2; IR (neat): ν_{max} 3465 (OH), 3320 (NH), 1648, 1550 (NCO), 733, and 696 cm⁻¹ (Ph); ¹H NMR (CDCl₃): δ 7.50–7.3 (m, 25 H, Ar), 5.46 (s, 1 H, PhCH), 5.30 (d, 1 H, J_{NH,2} 9.0 Hz, NH), 4.91 (d, 1 H, J_{1,2} 3.5 Hz, H-1), 4.27 (dt, J_{2,3} 10.0 Hz, H-2), and 1.79 (s, 3 H, NAc); ¹³C NMR: (CDCl₃): δ 169.72 (COCH₃), 103.42 (PhCH₂O₂), 101.25 (C-1'), 97.25 (C-1), 74.59 (C-2'), 74.21, 73.64, 70.89, 69.89 (C₆H₅CH₂O), 68.70, 68.22 (C-6,6'), 52.40 (C-2), and 23.38 (COCH₃). Anal. Calcd for C₄₉H₅₃NO₁₁ (831.97): C, 70.74; H, 6.42; N, 1.68. Found: C, 70.89; H, 6.55; N, 1.71.

Benzyl 3-O-benzyl-4,6-O-benzylidene- β -D-mannopyranosyl- $(1 \rightarrow 4)$ -2-acetamido-3,6di-O-benzyl-2-deoxy- β -D-glucopyranoside (11).—A solution of 9 (1.35 g, 1.62 mmol) in 1:2 acetic anhydride-dimethyl sulfoxide (45 mL) was stirred overnight at room temperature. The solvents were evaporated and the residue was chromatographed (solvent E) to give 10 as a white solid (1.3 g, 96.5%): $[\alpha]_D + 69^\circ$ (c 1.8, chloroform); TLC (solvent E): R_f 0.33; IR (neat): ν_{max} 3308 (NH), 1745 (C=O), 1648, 1545 (NCO), 735, and 696 cm⁻¹ (Ph); ¹H NMR (CDCl₃): δ 7.52–7.15 (m, 25 H, Ar), 5.45 (s, 1 H, PhCH), 5.31 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 4.91 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 4.27 (dt, $J_{2,3}$ 10.0 Hz, H-2), and 1.79 (s, 3 H, NAc); ¹³C NMR (CDCl₃): δ 197.22 (C-2' as carbonyl), 169.71 (COCH₃), 101.95 (PhCH₂O₂), 101.06 (C-1'), 97.26 (C-1), 74.73, 73.47, 73.31, 69.97 (PhCH₂O), 68.46, 68.11 (C-6,6'), 52.46 (C-2'), 23.36 (COCH₃).

A solution of 10 (1.2 g, 1.45 mmol) in 1:1 dichloromethane-methanol (94 mL) was treated with sodium borohydride (0.62 g, 16.4 mmol) for 4 h at room temperature. The mixture was diluted with chloroform (150 mL), and successively washed with water, 5% citric acid solution, aq KHCO₃, and water. Evaporation of the solvent and purification of the residue by chromatography (solvent B) first gave 9 (0.13 g, 10.8%), followed by 11 as a white solid (0.79 g, 65.7%): $[\alpha]_D + 62^\circ$ (c 1, chloroform); TLC (solvent D): R_c 0.13; IR (neat): ν_{max} 3468 (OH), 3298 (NH), 1651, 1553 (NCO), 731, and 695 cm⁻¹ (Ph); ¹H NMR (CDCl₃): δ 7.52-7.18 (m, 25 H, Ar), 5.51 (s, 1 H, PhCH), 5.35 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 4.95 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 4.59 (br s, 1 H, H-1'), 4.29 (dt, 1 H, $J_{2,3}$ 10.0 Hz, H-2), 3.96 (br t, 1 H, $J_{2',3'}$ 2.8 Hz, H-2'), 3.42 (dd, $J_{3',4'}$ 9.5 Hz, H-3'), and

1.84 (s, 3 H, NAc); ¹³C NMR (CDCl₃): δ 169.74 (COCH₃), 101.53 (C₆H₅CH₂O₂), 100.77 (C-1'), 97.31 (C-1), 69.54 (C-2'), 74.39, 73.65, 72.46, 69.95 (C₆H₅CH₂O), 68.55, 68.40 (C-6,6'), 52.38 (C-2), and 23.38 (COCH₃). Anal. Calcd for C₄₉H₅₃NO₁₁ (831.97): C, 70.74; H, 6.42; N, 1.68. Found: C, 70.80; H, 6.39; N, 1.70.

Benzyl 2-O-acetyl-3-O-benzyl-4,6-O-benzylidene- β -D-mannopyranosyl- $(1 \rightarrow 4)$ -2acetamido-3,6-di-O-benzyl-2-deoxy- β -D-glucopyranoside (12).—Compound 11 (0.6 g, 0.72 mmol) was dissolved in a mixture of acetic anhydride (5 mL) and pyridine (10 mL) and stirred overnight at room temperature. Acetic anhydride and pyridine were evaporated under diminished pressure, and the residue was chromatographed (solvent B) to afford 11 (0.62 g, 98%) as an amorphous solid: $[\alpha]_D + 42^\circ$ (c 1, chloroform); TLC (solvent D): R_f 0.32; ¹H NMR (CDCl₃): δ 7.54–7.14 (m, 25 H, Ar), 5.52 (s, 1 H, PhCH), 5.42 (dd, 1 H, $J_{1',2'}$ 1, $J_{2',3'}$ 3.5 Hz, H-2'), 5.28 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 4.94 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 4.57 (br s, 1 H, H-1'), 3.43 (dd, 1 H, $J_{3',4'}$ 9.8 Hz, H-3'), 2.18 (s, 3 H, OAc) and 1.82 (s, 3 H, NAc); ¹³C NMR: (CDCl₃): δ 170.31, 169.63 (COCH₃), 101.48 (PhCH₂O₂), 99.42 (C-1'), 97.16 (C-1), 74.25, 73.52, 71.64, 69.92 (PhCH₂O), 68.44, 68.28 (C-6,6'), 52.32 (C-2), and 23.32, 21.11 (COCH₃). Anal. Calcd for C₅₁H₅₅NO₁₂ (874.01): C, 70.09; H, 6.34; N, 1.60. Found: C, 70.00; H, 6.30; N, 1.56.

Benzyl 2-O-acetyl-3-O-benzyl-β-D-mannopyranosyl-(1 → 4)-2-acetamido-3,6-di-Obenzyl-2-deoxy-α-D-glucopyranoside (13).—Compound 12 (0.65 g, 0.74 mmol) was taken up in 60% aq acetic acid (40 mL) and heated for 2 h at 70°C. The acetic acid was evaporated under reduced pressure to leave a residue that was chromatographed (solvent E) to give 13 (0.48 g, 82%): $[\alpha]_D + 34^\circ$ (c 1.6, chloroform); TLC (solvent F): R_f 0.26; ¹H NMR (CDCl₃): δ 7.44–7.20 (m, 20 H, Ar), 5.36 (br d, 1 H, $J_{2',3'}$ 3.5 Hz, H-2'), 5.27 (d, 1 H, $J_{NH,2}$ 9.0 Hz, NH), 4.92 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 4.56 (s, 1 H, H-1'), 2.12 (s, 3 H, OAc) and 1.80 (s, 3 H, NAc); ¹³C NMR (CDCl₃): δ 170.32, 169.72 (COCH₃), 98.52 (C-1'), 97.13 (C-1), 74.40, 73.58, 71.35, 69.94 (PhCH₂O), 68.36 (C-6), 62.34 (C-6'), 52.31 (C-2), and 23.33, 21.07 (COCH₃). Anal. Calcd for C₄₄H₅₁NO₁₂ (785.90): C, 67.25; H, 6.54; N, 1.78. Found: C, 67.14; H, 6.34; N, 1.72. Benzyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy-β-D-glucopyranosyl-(1 → 2)-3,4,6-tri-

O-acetyl- α -D-mannopyranosyl- $(1 \rightarrow 6)$ -2-O-acetyl-3-O-benzyl- β -D-mannopyranosyl-

 $(1 \rightarrow 4)$ -2-acetamido-3,6-di-O-benzyl-2-deoxy- α -D-glucopyranoside (15).—A stirred mixture of diol acceptor 13 (0.5 g, 0.64 mmol) powdered Hg(CN)₂ (0.17 g, 0.67 mmol) and 4 Å molecular sieves (0.5 g) in 1:1 benzene-nitromethane (80 mL) was boiled until 20 mL of the solvent had distilled off. After cooling to room temperature bromide 14 (0.57 g, 0.82 mmol) in 1:1 benzene-nitromethane (9 mL) was added and stirring was continued overnight at 40-45°C. The mixture was filtered (Celite), the solids were thoroughly washed with benzene, and the filtrate and washings were combined and diluted with benzene to a total volume of 200 mL. This solution was successively washed with water, M KI, aq NaHCO₃, and water, dried, and concentrated to give a solid residue. TLC (solvent G) of the crude mixture showed the presence of a major product, slightly faster migrating than 13. Small proportions of some faster and some slower migrating contaminants, as well as of 13, were also revealed in TLC. The crude product was chromatographed ($0 \rightarrow 2\%$ methanol in chloroform), and concentration of the fractions corresponding to the product gave a solid that was dissolved in dichloromethane and precipitated by the addition of ether-hexane to furnish 15 (0.5 g,

56%) as an amorphous solid: $[\alpha]_D + 1.8^{\circ}$ (*c* 1, chloroform); TLC (solvent G): R_f 0.71; ¹H NMR (CDCl₃): δ 7.43–7.20 (m, 20 H, Ar), 5.75 (d, 1 H, $J_{NH,2}$ 8.5 Hz, NH), 2.15 (s, 3 H, OAc), 2.04, 2.03, (s, 6 H each, 4 OAc), 2.0, 1.96 (s, 3 H each, 2 OAc), 1.89, and 1.80 (s, 3 H each, 2 NAc); ¹³C NMR (CDCl₃): δ 170.82, 170.65, 170.58, 170.05, 169.82, 169.62, 169.44 (COCH₃), 99.50 (C-1'''), 98.96 (C-1'), 97.85 (C-1''), 97.17 (C-1), 80.03 (C-4), 78.91 (C-2''), 73.79, 73.23, 71.33, 69.66 (PhCH₂), 68.45 (C-6), 66.41 (C-6'), 62.59 (C-6''), 61.88 (C-6'''), 55.28 (C-2'''), 52.34 (C-2), 23.19, 23.12, 20.94, 20.69, 20.67, 20.63, and 20.55 (COCH₃). Anal. Calcd for C₇₀H₈₆N₂O₂₈ (1403.46): C, 59.91; H, 6.18; N, 2.00. Found: C, 59.64; H, 6.10; N, 2.11.

Benzyl 2-acetamido-2-deoxy-β-D-glucopyranosyl-(1 → 2)-α-D-mannopyranosyl-(1 → 6)-3-O-benzyl-β-D-mannopyranosyl-(1 → 4)-2-acetamido-3,6-di-O-benzyl-2-deoxy-α-D-glucopyranoside (16).—Compound 15 (0.38 g, 0.27 mmol) was O-deacetylated in 20 mM methanolic sodium methoxide (50 mL) exactly as described for the preparation of 8 to give after chromatography (10 → 20% methanol in chloroform) a solid residue that was dissolved in a small amount of methanol. Addition of ether caused the precipitation of 16 (0.19 g, 63%) as an amorphous solid: [α]_D + 2.4° (c 8.7, H₂O); TLC (solvent H): R_f 0.54; ¹H NMR (CD₃OD): δ 7.46-7.20 (m, 20 H, Ar), 4.78 (d, 1 H, $J_{1",2"}$ 1.8 Hz, H-1"), 4.55 (d, 1 H, $J_{1",2"}$ 8.0 Hz, H-1"), 4.42.(s, 1 H, H-1'), 1.90 (s, 6 H, 2 NAc); ¹³C NMR (CD₃OD): δ 174.38, 173.32 (COCH₃), 102.13, 101.95 (C-1', C-1"), 98.59 (C-1"), 97.94 (C-1'), 70.66 (C-6), 69.84 (C-6'), 63.15 (C-6"), 62.43 (C-6"), 57.22 (C-2"), 54.21 (C-2), 23.91, and 22.65 (COCH₃). Anal. Calcd for C₅₆H₇₂N₂O₂₁ (1109.20): C, 60.64; H, 6.54; N, 2.53. Found: C, 60.80; H, 6.56; N, 2.50.

2-Acetamido-2-deoxy- β -D-glucopyranosyl- $(1 \rightarrow 2)$ - α -D-mannopyranosyl- $(1 \rightarrow 6)$ - β -D-mannopyranosyl- $(1 \rightarrow 4)$ -2-acetamido-2-deoxy-D-glucoyranose (2).—A mixture of 16 (0.1 g, 90 μ mol) and 10% Pd–C (0.12 g) in glacial acetic acid (10 mL) was shaken under H₂ at 345 kPa for 2 days at room temperature. The suspension was filtered (Celite), the solid was thoroughly washed with glacial acetic acid and methanol (Caution! Fire hazard.), and the filtrate and washings were combined and concentrated. The residue was chromatographed (solvent I \rightarrow J \rightarrow K) to give a solid that was dissolved in a small volume of water and lyophilized to give 2 (47 mg, 69.6%): $[\alpha]_D + 5.4^{\circ}$ (initial) $\rightarrow +6.4^{\circ}$ (12 h, c 0.3, water); TLC (solvent K): R_f 0.1. FABMS: 749 (M + 1)⁺ and 771 (M + Na)⁺. ¹H NMR (D₂O): δ 5.19 (d, $J_{1,2}$ 2.5 Hz, H-1 α), 4.90 (d, $J_{1',2''}$ 1 Hz, H-1''), 4.71 (s, H-1'), 4.54 (d, $J_{1'',2'''}$ 8.5 Hz, H-1'''), 4.50 (d, $J_{1,2}$ 8.5 Hz, H-1 β), 4.02 (br t, $J_{2'',3''}$ 3.30 Hz, H-2''), 2.05 (NAc, C-2 α), 2.04 (NAc, C-2'') and 2.03 (NAc, C-2 β); ¹³C NMR (D₂O): δ 175.63 (COCH₃), 101.39 (C-1'''), 100.46 (C-1'), 99.96 (C-1''), 91.35 (C-1 α), 67.00 (C-6'), 62.46 (C-6''), 61.47 (C-6'''), 60.97 (C-6), 56.19 (C-2''''), 54.44 (C-2), 23.18 and 22.67 (COCH₃). Anal. Calcd for C₂₈H₄₈N₂O₂₁ · 2H₂O (784.73): C, 42.86; H, 6.68; N, 3.57. Found: C, 42.61; H, 6.78; N, 3.48.

Acknowledgements

This investigation was supported by Grant No. AI 29326 awarded by the NIAID, National Institutes of Health, and in part by Grant No CH 419 awarded by the American Cancer Society. The authors are thankful to Mr Conrad Piskorz and Mrs Humaira Bano for their valuable assistance in preparing the manuscript.

References

- [1] R.K. Jain, C.F. Piskorz, and K.L. Matta, Carbohydr. Res., 275 (1995) 231-243.
- [2] R.D. Cummings, I.S. Trowbridge, and S. Kornfeld, J. Biol. Chem., 257 (1982) 13421-13427.
- [3] H. Schachter, in M. Fukuda and O. Hindsgaul (Eds.), Molecular Glycobiology: Frontiers in Molecular Biology, Oxford University Press, Oxford, UK, 1994, pp 88-149.
- [4] K. Yamshita, Y. Tachibana, T. Ohkura, and A. Kobata, J. Biol. Chem., 260 (1985) 3963-3969.
- [5] J. Arango and M. Pierce, J. Cell. Biochem., 37 (1988) 225-231.
- [6] J.W. Dennis, K. Kosh, D.-M. Bryce, and M.L. Breitman, Oncogene, 4 (1987) 853-860.
- [7] J.W. Dennis, S. Laferté, C. Waghorne, M.L. Breitman, and R.S. Kerbel, Science, 236 (1987) 582-585.
- [8] J.W. Dennis, Cancer Surveys, 7 (1988) 573-594.
- [9] J.W. Dennis, in M. Fukuda (Ed.), Cell Surface Carbohydrates and Cell Development, CRC Press, Boca Raton, FL, 1991, pp 161-194.
- [10] J.W. Dennis and S. Laferté, Cancer Res., 49 (1989) 945-950.
- [11] M.M. Palcic, J. Ripka, K.J. Kaur, M. Shoreibah, O. Hindagaul, and M. Pierce, J. Biol. Chem., 265 (1990) 6759-6769.
- [12] O. Hindsgaul, K.J. Kaur, G. Srivastava, M. Blaszczyk-Thurin, S.C. Crawley, L.D. Heerze, and M.M. Palcic, J. Biol. Chem., 266 (1991) 17858-17862.
- [13] S.H. Khan and K.L. Matta, in H.J. Allen and E.C. Kisailus (Eds.), Glycoconjugates: Composition, structure, and function, Marcel Dekker, New York, 1992, pp 361–378.
- [14] S.H. Khan, O. Kanie, S.C. Crawley, and O. Hindagaul, J. Biol. Chem., 268 (1993) 2468-2473.
- [15] S.H. Khan and K.L. Matta, Carbohydr. Res., 243 (1993) 29-42.
- [16] S.H. Khan and O. Hindsgaul, in M. Fukuda and O. Hindsgaul (Eds.), Molecular Glycobiology: Frontiers in Molecular Biology, Oxford University Press, Oxford, UK, 1994, pp 206-229.
- [17] S.H. Khan, J.Ø. Duus, S.C. Crawley, M.M. Palcic, and O. Hindagaul. Tetrahedron Asymm., 5 (1994) 2415-2435.
- [18] I. Brockhausen, F. Reck, W. Kuhns, S.H. Khan, K.L. Matta, E. Meinjohanns, H. Paulsen, R.N. Shah, M.A. Baker, and H. Schachter, *Glycoconjugate J.*, 12 (1995) 371–379.
- [19] S.H. Tahir and O. Hindsgaul, Can. J. Chem., 64 (1986) 1771-1780.
- [20] M. Pierce, J. Arango, S.H. Tahir, and O. Hindsgaul, Biochem. Biophys. Res. Commun., 146 (1987) 679-684.
- [21] O. Hindsgaul, S.H. Tahir, O.P. Srivastava, and M. Pierce, Carbohydr. Res., 173 (1988) 263-273.
- [22] S.C. Crawley, O. Hindsgaul, G. Alton, M. Pierce, and M.M. Palcic, Anal. Biochem. 185 (1990) 112-117.
- [23] M. Shoreibah, O. Hindagaul, and M. Pierce, J. Biol. Chem., 267 (1992) 2920-2927.
- [24] M. Shoreibah, G-S. Perng, B. Adler, J. Weinstein, R. Basu, R. Cupples, D. Wen, J.K. Browne, P. Buckhaults, N. Fregien, and M. Pierce, J. Biol. Chem., 268 (1993) 15381-15385.
- [25] O.P. Srivastava, O. Hindsgaul, M. Shoreibah, and M. Pierce, Carbohydr. Res., 179 (1988) 137-161.
- [26] I. Lindh and O. Hindsgaul, J. Am. Chem. Soc., 113 (1991) 216-223.
- [27] O. Kanie, S.C. Crawley, M.M. Palcic, and O. Hindsgaul, Carbohydr. Res., 243 (1989) 139; Bioorg. Med. Chem., 2 (1994) 1231-1241.
- [28] T. Linker, S.C. Crawley, and O. Hindsgaul, Carbohydr. Res., 245 (1993) 323-331.
- [29] S.H. Khan, S.A. Abbas, and K.L. Matta, Carbohydr. Res., 193 (1989) 125-139.
- [30] S.H. Khan, S.A. Abbas, and K.L. Matta, Carbohydr. Res., 205 (1990) 385-397.
- [31] S.H. Khan and K.L. Matta, J. Carbohydr. Chem., 12 (1993) 335-348.
- [32] H. Paulsen and E. Meinjohanns, Tetrahedron Lett., 33 (1992) 7327-7330.
- [33] H. Paulsen and E. Meinjohanns, F. Reck, and I. Brockhausen, Liebigs Ann. Chem., (1993) 737-750.
- [34] I. Brockhausen, G. Möller, J-M. Yang, S.H. Khan, K.L. Matta, H. Paulsen, A.A. Grey, R.N. Shah, and H. Schachter, Carbohydr. Res., 236 (1992) 281-299.
- [35] F. Schmitt and P. Sinaÿ, Carbohydr. Res., 29 (1973) 99.
- [36] S. Sadeh, C.D. Warren, and R.W. Jeanloz, Carbohydr. Res., 123 (1983) 73-81.
- [37] P. Konradsson, D.R. Mootoo, R.E. McDevitt, and B. Fraser-Reid, J. Chem. Soc., Chem. Commun., (1990) 270-272.
- [38] V.H. Veeneman, S.H. van Leeuwen, and J.H. van Boom, Tetrahedron Lett., 31 (1990) 1331-1334.

- [39] C.D. Warren, C. Auge, M.L. Liver, S. Suzuki, D. Power, and R.W. Jeanloz, Carbohydr. Res., 82 (1980) 71-83.
- [40] P.J. Garegg and H. Hultburg, Carbohydr. Res., 93 (1983) C10-C11.
- [41] B.H. Koppen, Carbohydr. Res., 24 (1972) 154-158.
- [42] S. Nambir, J.F. Daeuble, R.J. Doyle, and K.G. Taylor, Tetrahedron Lett., 30 (1989) 2179-2182.
- [43] J.L. Albright and L. Goldman, J. Org. Chem., 30 (1965) 1107-1110; J. Am. Chem. Soc., 87 (1965) 4214-4216.
- [44] P.J. Garegg and T. Iversen, Carbohydr. Res., 70 (1979) C13-C14.
- [45] H. Paulsen, R. Lebuhn, and O. Lockhoff, Carbohydr. Res., 103 (1982) C7-C11.
- [46] P.J. Garegg and P. Ossowski, Acta. Chem. Scand., Ser. B, 37 (1983) 249-251.
- [47] D. Kahne, D.Y. Yang, J. Lim, R. Miller, and E. Paguaga, J. Am. Chem. Soc., 110 (1988) 8716-8717.
- [48] S. David, A. Malleron, and C. Dini, Carbohydr. Res., 188 (1989) 193-200.
- [49] H. Kunz and W. Gunther, Angew. Chem., Int. Ed. Engl., 27 (1988) 1086-1087.
- [50] F. Barresi and O. Hindsgaul, J. Am. Chem. Soc., 113 (1991) 9376-9377; Synlett, (1992) 759-761; Can. J. Chem., 72 (1994) 1447-1465.
- [51] G. Stork and G. Kim, J. Am. Chem. Soc., 114 (1992) 1765-1767.
- [52] Y. Ito and T. Ogawa, Angew. Chem., Int. Ed. Engl., 33 (1984) 1086-1087.
- [53] J. Brunckova, D. Crich, and Q. Yao, Tetrahedron Lett., 35, (1994) 6619-6622.
- [54] N. Yamazaki, E. Eichenberger, and D.P. Curran, Tetrahedron Lett., 35, (1994) 6623-6626.
- [55] G. Ekborg, B. Lindberg, and J. Lönngren, Acta. Chem. Scand., 26 (1972) 2639-2644.
- [56] G. Ekborg and C.P.J. Glaudemans, Carbohydr. Res., 129 (1984) 287-292.
- [57] H.H. Lee, L_N. Congson, and D.M. Whitfield, L.R. Radics, and J.J. Krepinsky, Can. J. Chem., 70 (1992) 2607-2617.
- [58] P.J. Garegg, Acc. Chem. Res., 25 (1992) 575-580.
- [59] J. Kerékgyártó, J.G.M. van der Ven, J.P. Kamerling, A. Lipták, and J.F.G. Vliegenthart, Carbohydr. Res., 238 (1993) 135-145.
- [60] E. Kaji and F.W. Lichtenthaler, Trends. Glycosci. Glycotechnol., 5 (1993) 121-142.
- [61] K.K.-C. Liu and S.J. Danishefsky, J. Org. Chem., 59 (1994) 1892-1894.
- [62] I. Brockhausen, S.H. Khan, and K.L. Matta, unpublished results.