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Enantiopure (9-Anthryl)(2-piperidyl)- and (9-Anthryl)(2-pyridyl)methanols – Their Use as Chiral Modifiers for Heterogeneous Hydrogenation of Keto Esters over Pt/Al₂O₃

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A route toward the synthesis of the *erythro* isomer of (9-anthryl)(2-piperidyl)methanol is presented as well as resolution and assignment of the structure (through NMR). The use of both the *erythro* and *threo* enantiopure isomers of this new amino alcohol, and its precursor [(9-anthryl)(2-pyridyl)methanol], as chiral modifiers for the Pt/Al_2O_3 hydrogenation of ethyl lactate showed that the *erythro* isomer is not necessarily the most efficient chiral modifier. This is probably because of the 9-anthryl group. The enantioselectivities that this compound provides are not, as one would expect, higher than those observed with the naphthyl group.

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cinchonidine **1** (which is also an *erythro* isomer). Until 2002, it also seemed that the anthryl group had a tendency

to provide higher enantioselectivities than the naphthyl

group (when located on the carbon bearing the hydroxy

2 cinchonine

(8R,9S) erythro

5 : Ar = P

1 cinchonidine

(-)-(8S,9R) erythro

erythro and three

ervthro

Introduction

Asymmetric heterogeneous hydrogenation of a-keto esters over supported platinum catalysts with the use of cinchona alkaloids (Orito reaction^[1]) has been known for about thirty years and has been the subject of numerous reviews.^[2] When we started our work in this field in 1998.^[3] it was already known that cinchonidine 1 was the most efficient chiral modifier^[2] and apart from cinchonidine or cinchonine derivatives^[4] the only different modifiers tested were type 3 amino alcohols (1994)^[5] and type 4 amines (1995).^[6a,6b] Our target was then amino alcohol **5a**^[3] which had the same -CH(OH)-CH(N)- link as cinchona alkaloids and one chiral carbon less. Moreover, both enantiomers of both the erythro and threo diastereomers could be available which would provide straightforward conclusions. With the use of 5a, for the first time we clearly showed (2001) that the erythro isomer was a more efficient chiral modifier than the *threo* isomer^[3] and almost as efficient as

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a1 R=H, a, R=alkyl,...

b R=H

group). Therefore, we decided to synthesize and study *erythro* and *threo* amino alcohols **5b**. By the time this work was completed, other results appeared that dealt with the comparison of anthryl and naphthyl groups in the case of amines $4^{[6c-6e]}$ and amino alcohols $6^{[7]}$

We present here the synthesis and resolution of both the *erythro* and *threo* diastereomers of anthryl amino alcohol **5b**, a synthesis of the pure *erythro* isomer (lower overall yield but no difficulty with the separation of diastereomers), the use of **5b** as a chiral modifier for the heterogeneous hydrogenation of ethyl pyruvate over Pt/Al_2O_3 , as well as the use of precursor 7 to test the effect of the possible in situ hydrogenation of 7 into **5b** during hydrogenation of pyruvate.

Synthesis

Condensation of the 9-anthryl Grignard reagent with 2pyridinecarboxaldehyde proceeded smoothly in THF to provide anthrylpyridyl alcohol 7 in 80% yield (Scheme 1). Heterogeneous hydrogenation of 7 with the use of PtO₂ (from Aldrich) in EtOH/HCl (1 equiv.)^[8,9] then provided pure *erythro* **5b-II** in 40% yield (**5b-I/5b-II** = 0/100).



Scheme 1.

It is worth noting that *threo* isomer **5b-I** (necessary for structure determination) can be obtained (as the major isomer) from formamidine **8**, Scheme 2, by following Meyers' procedures,^[10,11] (overall yield 50%, after chromatographic purification).

Table 1. NMR spectroscopic data for *threo* and *erythro* configurations.

Amino alcohol	³ J ₁₋₂ CDCl ₃ [Hz]	$\Delta^3 J$ [Hz]	δ(1-H) [ppm]	Configuration
5-I (Ar = Ph)	7.5	2.5	4.36	threo ^[a]
5-II (Ar = Ph)	5.0		4.56	erythro ^[a]
(1 <i>S</i> ,2 <i>S</i>)-Pseudoephedrine	8.2	4.3		threo ^[a]
(1R,2S)-Ephedrine	3.9			erythro ^[a]
5a-I (67%)	6.0	2	5.24	threo ^[a]
5a-II (33%)	4.0		5.44	erythro ^[a]
5b-I (80%)	9.5	2	6.03	threo
5b-II (20%)	7.5		6.10	erythro

[a] Known compounds, cf. refs.^[12,13]

The ratios of **I/II** have been determined with the use of the 1-H proton doublets (between $\delta = 6.0$ ppm and 6.2 ppm, Table 1).

Determination of threolerythro Structure of 5b-I and 5b-II

threolerythro Structure from NMR

In amino alcohols having the CH(OH)–CH(NH) system and of known structure, Table 1, the *erythro* isomer has the smallest coupling constant. Therefore, the *erythro* structure has been assigned to isomer **5b-II**, which exhibits the smallest coupling constant: 7.5 Hz versus 9.5 Hz for isomer **5b-I**.

Moreover, the 1-H doublets in the *erythro* isomers are always the most deshielded, which is the case here too ($\delta = 6.10$ ppm compared to $\delta = 6.03$ ppm for the *threo* isomer in this case).

erythro Structure from Model

The *erythro* structure assigned by NMR to isomer II, the only isomer obtained from hydrogenation over PtO_2 , is consistent with the model shown in Scheme 3, as it is reasonable to postulate that the hydrogen (located on the catalyst surface) approaches from the less hindered side of the H-bonded C1 conformer of substrate 7 (1-H side).



Scheme 3.

Resolution of Piperidyl Alcohol 5b-I and 5b-II

Diastereomeric separation of the *threo* isomer from the *erythro* isomer was performed first by preparative supercritical fluid chromatography with a Chromegabond Amine column (25 cm \times 23 mm ID, 5 µm 60 Å, ES Industries) at a flow rate of 50 mL/min with a mobile phase of 25% methanol in carbon dioxide (100 bar outlet pressure) with UV detection at 257 nm.

Analytical separation of racemic pure *threo* isomer **5b-I** was further performed with a CHIRALCEL OD-RH column (150×4.6 mm, 5 µm) with acetonitrile/ethanolamine, 100:0.1, as the mobile phase (flow rate = 0.5 mL/min) and UV detection at 235 nm.



Scheme 2.

On a preparative scale, a CHIRALCEL OD column $(250 \times 50 \text{ mm}, 20 \text{ }\mu\text{m})$ and the same mobile phase (100 mL/min) were then used. After recrystallization from acetonitrile (to completely remove ethanolamine as seen from ¹H NMR) of the two fractions having the highest *ee*, the specific rotations of enantiomers *threo* **5b-I1** and *threo* **5b-I2** were determined: *threo* **5b-I1**: $[a]_{\rm D} = +11.0$ (c = 0.45, CHCl₃), ee = 99%; *threo* **5b-I2**: $[a]_{\rm D} = -10.8$ (c = 0.50, CHCl₃), ee = 99%.

The *erythro* isomer (**5b-II**) was obtained pure and the enantiomers were then preparatively obtained by supercritical fluid chromatography with the use of a CHIRALPAK AD column (250×20 mm, 10μ m) at a flow rate of 50 mL/ min with a mobile phase of 16% methanol (containing 25 mM of isobutylamine) in carbon dioxide (100 bar outlet pressure) with UV detection at 257 nm, with the isolation of two fractions: *erythro* **5b-II1** negative CD at 263 nm, *ee* = 99%; *erythro* **5b-II2** positive CD at 263 nm, *ee* = 99%. [*a*]_D = +17 (*c* = 0.24, CHCl₃).

Resolution of Pyridyl Alcohol 7

Analytical separation of *rac*-7 was performed with a CHIRALCEL OD column (250×4.6 mm, 10μ m) with hexane/EtOH/diethylamine, 8:2:0.5, as the mobile phase (flow rate = 1 mL/min) and UV detection at 254 nm. On a preparative scale, a CHIRALCEL OD column (0.46×25 cm, 10μ m) and the same mobile phase (120 mL/min) were used with the isolation of two fractions: (–)-7, chemical purity >95% (by 400 MHz ¹H NMR). [a]_D = -171 (c = 0.36, CHCl₃), ee = 100% (from analytical chromatography); (+)-7, chemical purity >95% (by 400 MHz ¹H NMR). [a]_D = +97 (c = 0.37, CHCl₃), ee = 57% (from specific rotation).



Asymmetric Pt/Al₂O₃ Hydrogenation of Ethyl Pyruvate

After hydrogenation and distillation, the enantiomeric excesses of the ethyl lactate samples were determined by gas chromatography with a Cyclodex-B capillary column (30 m) with the use of an HP 5890 GC-FID instrument and the *ee* values were reproducible within 1%.

The well-defined catalyst 5% Pt/Al₂O₃ 4759 from Engelhard was activated before use (3 h, 300 °C, under H₂ flow); AcOH was used as the solvent. The reactions were run at room temperature for 2 h with stirring at a rate of ca. 500 rpm, and the pyruvate, purchased from Aldrich, was distilled before used. The results are given in Table 2.

Table 2. Ethyl pyruvate hydrogenation over 5% Pt/Al_2O_3 at room temperature for 2 h.

Conversion [%]	(R)/(S)	ee [%]
13	_	_
100	68:32	36
100	32.5:67.5	35
100	27:73	46
100	73.5:26.5	47
100	37:63	26
	Conversion [%] 13 100 100 100 100 100 100	Conversion (R)/(S) [%] - 13 - 100 68:32 100 32.5:67.5 100 27:73 100 73.5:26.5 100 37:63

[a] Cf. ref.^[3]

It is worth noting that, contrary to the cinchona alkaloids generally used, **5b-I1** and **5b-I2** (as well as **5b-II1** and **5b-II2**) are true enantiomers; therefore, the samples of **5b-I1** and **5b-I2** (as well as **5b-II1** and **5b-II2**) that were used that have identical enantiomeric purities (cf. above) provide identical (R)/(S) ratios to ethyl lactate, within the reproducibility of the method used, for the determination of these ratios (compare 68:32 with 32.5:67.5 and 27:73 with 73.5:26.5). It appears that (S)-lactate is obtained from (–)-**5b-II** (*erythro*) and (–)-**5b-I** (*threo*) while (+)-**5b-II** and (+)-**5b-II** provide (R)-lactate. It is also worth noting that the *threo* isomers provide slightly higher enantioselectivities than the *erythro* isomers (46–47% versus 35–36% *ee*).

Precursor 7, which under these conditions undergoes slow hydrogenation^[14] into desired **5b-II**, is less efficient probably because hydrogenation of pyruvate is too rapid and that 7 is a less efficient chiral modifier.

Conclusions

These results show that the *erythro* isomer is not necessarily the most efficient chiral modifier as was suggested from previous results obtained by using compound $5a^{[3]}$ and cinchona alkaloids^[4,15] (Table 3).

Table 3. Hydrogenation of ethyl pyruvate with the use of various chiral modifiers.

Modifier	Conversion [%]	(R)/(S)	ее [%]
Cinchonidine (9 <i>R</i> ,8 <i>S</i>) erythro	100	93.5:6.5	87
Epiquinidine (<i>threo</i>) ^[a]	?	1:1	0
(-)- $(1R,2S)$ - 5a-II erythro	100	86.5:13.5	73
(+)-(1 <i>S</i> ,2 <i>R</i>)- 5a-II erythro ^[b]	100	14:86	72
(-)-5a-I threo	100	27:73	46
(+)- 5a-I <i>threo</i> ^[b]	67	67.5:32.5	35 ^[c]
(+)-6a- $(1S,2R)$ erythro ^[d]	_	6:94	88
(+)- 6b -(1 <i>S</i> ,2 <i>R</i>) erythro ^[d]	_	10.5:89.5	79

[a] Cf. ref.^[15] [b] Cf. ref.^[3], note that the absolute configuration of *erythro* **5a-II** was determined from the stereo-outcome of the hydrogenation of pyruvate and confirmed by VCD (ref.^[13]). [c] This sample of the *threo* isomer that was used contained 1.8% of one enantiomer of the *erythro* isomer (ref.^[13]). [d] Cf. ref.^[7]

The results also corroborate that the anthryl group is not necessarily a more efficient modifier than the naphthyl group, and it may provide lower enantioselectivities for both *erythro* and *threo* isomers as in the case of type **5** amino alcohols (compare Table 2 and Table 3) or compara-

ble enantioselectivities in the case of type 6 *erythro* amino alcohol (Table 3).

As in the case of (-)-**5a-II**, for which the (1R,2S) configuration was determined from the stereo-outcome of the reaction (pyruvate hydrogenation) and corroborated by VCD, one could postulate from the stereo-outcome of the pyruvate hydrogenation reaction that (+)-**5b-II**, which also provides (*R*)-lactate, has the (1*R*,2*S*) configuration.

Experimental Section

General Procedure for Anthryl Grignard Addition onto 2-Pyridincarboxaldehyde: It is worth noting that the 9-anthryl grignard formed from 9-bromoanthacene must be generated in THF heated at reflux. A solution of bromoanthracene (3.36 g, 13 mmol) in anhydrous THF (80 mL) was added dropwise whilst stirring to a suspension of Mg (0.31 g, 13 mmol) in anhydrous THF (30 mL) and 1,2dibromoethane (0.2 mL). The mixture was heated at reflux and stirring was maintained until all of the Mg disappeared. After cooling to room temperature, a solution of 2-pyridincarboxaldehyde (0.7 g, 6.5 mmol) in anhydrous THF (30 mL) was added dropwise whilst stirring overnight. Then, a saturated aqueous solution of NH₄Cl was added, THF was evaporated under vacuum, and the aqueous phase was extracted with CH_2Cl_2 (5 × 20 mL). The combined organic phases were dried with Na₂SO₄, the solvent removed under reduced pressure, and the amino alcohol isolated and purified by chromatography on silica gel (ether/hexane, 3:2). 7: ¹H NMR (300 MHz, CDCl₃): δ = 5.9 (br. s, 1 H), 6.72 (d, ³J = 7.5 Hz, 1 H), 7.2 (dd, ${}^{3}J$ = 7.5, 5 Hz, 1 H), 7.29 (s, 1 H), 7.44 (m, 5 H), 8.02 (m, 2 H), 8.31 (m, 2 H), 8.49 (s, 1 H), 8.71 (d, ${}^{3}J = 5$ Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 69.7, 120.8, 122.1, 124.8, 125.0, 127.2, 128.9, 129.3, 130.7, 131.8, 132.3, 137.0, 147.6, 161.8 ppm. C₂₀H₁₅NO (285.22): calcd. C 84.17, H 5.30; found C 84.00, H 5.42.

General Procedure for PtO₂ Hydrogenation: To a solution of 7 (0.2 g, 0.7 mmol) in EtOH (10 mL) in a glass tube that exactly fits inside of a stainless steel autoclave were added successively concentrated HCl (0.05 mL, 0.7 mmol) and PtO₂ (30 mg). After being purged twice (vacuum/H₂ admission), the mixture was stirred under 50 bar H₂ and at 0 °C for 6 h. The catalyst was filtered out, NaOH (1 equiv.) and H₂O (5 mL) were added, the mixture was extracted with CH_2Cl_2 (10×10 mL), the combined organic phases were dried with Na₂SO₄, the solvent was removed under vacuum, and the amino alcohol was then purified by chromatography on silica gel (ether/hexane, 9:1). threo 5b-I: ¹H NMR (400 MHz, CDCl₃): $\delta = 0.81$ (br. d, ${}^{3}J = 12.0$ Hz, 1 H, 2-H_e), 1.0 (m, 1 H, 3-H), 1.10 (br. q, 1 H, 2-H_a), 1.36 (m, 1 H, 4-H), 1.48 (m, 2 H, 3-H, 4-H), 2.69 (td, ${}^{2}J = 12$ Hz, ${}^{3}J = 12$ Hz, 2.5 Hz, 1 H, 5-H_a), 3.09 (br. d, J = 12 Hz, 1 H, 5-H_e), 3.52 (td, ${}^{3}J = 9.5$ Hz, 9.5 Hz and 4 Hz, 1 H, 1-H), 3.60 (br. s, 1 H), 6.03 (d, ${}^{3}J$ = 9.5 Hz, 1 H), 7.47 (m, 4 H), 8.00 (m, 2 H), 8.40 (s, 1 H), 8.72 (br., 2 H)^[16] ppm. ^{13}C NMR (100 MHz, CDCl₃): δ = 24.5, 26.0, 29.4, 46.7, 62.0, 73.5, 125.1, 125.2, 125.7, 128.4, 129.6, 130.7, 132.0, 133.8 ppm. C₂₀H₂₁NO (291.22): calcd. C 82.44, H 7.26; found C 81.79, H 7.34. erythro 5b-II: All signals overlapped with those of threo 5b-I, except: $\delta = 2.72$ (br. t, ${}^{2}J = {}^{3}J = 12$ Hz, 1 H, 5-H_a) and 6.10 (d, ${}^{3}J =$ 7.5 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 24.1, 25.2, 28.2, 46.6, 62.2, 73.7, 124.8, 125.7, 128.4, 129.1, 130.2, 131.6, 132.1 ppm.

General Procedure for Aldolization of Formamidine 8: To a solution of piperidine formamidine 8 (2.0 g, 11.88 mmol, 1.0 equiv.) in an ether/THF mixture (15:4 mL) was added (at -78 °C) *tert*-butyllith-

ium (1.7 M, 8.5 mL, 14.25 mmol, 1.2 equiv.). The solution was stirred at -20 °C for 1 h, which resulted in the formation of a white precipitate. The reaction mixture was cooled to -78 °C, the desired aldehyde (1.1 equiv.) added, and the mixture slowly warmed up to 0 °C over 5 h. The solution was poured into 10% HCl (30 mL) and extracted with diethyl ether (2 × 10 mL). The aqueous layer was then made basic (pH 12) with 20% NaOH and extracted several times with dichloromethane. The combined organic phases were then concentrated under vacuum. A solution of the resulting crude product in MeOH (25 mL) and water (4 mL) and KOH (4 g) was heated at 60 °C for 24 h. After cooling, the mixture was extracted with dichloromethane (5 × 15 mL). The combined dichloromethane phases were dried with Na₂SO₄, the solvents removed under reduced pressure, and the amino alcohol isolated and purified by chromatography on silica gel (CH₂Cl₂/MeOH, 4:1).

General Procedure for Pt/Al₂O₃ Hydrogenation: A mixture of ethyl pyruvate (5 mL, 3500 equiv.) and the desired chiral modifier (1 equiv.), stirred for 30 min before used, was poured into a glass tube that exactly fits inside of a stainless steel autoclave. AcOH (10 mL) and the activated catalyst (50 mg, 1 equiv. in Pt) were then added successively. After being closed tightly, the device was purged three times (through successive vacuum-H₂ admission), the H₂ pressure was fixed at 40 bar, and the mixture was stirred for 2 h. The conversion was determined by using ¹H NMR of the crude products, then ethyl lactate was separated from other components through distillation and the *ee* was determined by chromatography (cf. text above). 9: ¹H NMR (300 MHz, CDCl₃): $\delta = 1.23$ (t, ³*J* = 7 Hz, 3 H), 1.35 (d, ³*J* = 7 Hz, 3 H) 3.15 (br. s, 1 H, OH), 4.17 (2 H, CH₂ and 1 H, CH overlapped q).

Resolution of Piperidyl Alcohols 5b-I and 5b-II and Pyridyl Alcohol 7: Diastereomeric separation of the threo isomer from the erythro isomer was first necessary (cf. above). Analytical separation of racemic pure *threo* isomer **5b-I** was then performed further with a CHI-RALCEL OD-RH (150×4.6 mm, 5 µM) column with acetonitrile/ ethanolamine, 100:0.1 as the mobile phase (flow rate = 0.5 mL/min), the chromatographic parameters obtained are: R_{t_1} = 9.15 min, $R_{t_2} = 11.04$ min, $k'_1 = 1.54$, $k'_2 = 2.07$, alpha = 1.34. Then, on a preparative scale, 483 mg of rac-5b-I was resolved by using a larger CHIRALCEL OD column (250×50 mm, 20 µm) and the same mobile phase (100 mL/min), with isolation of three fractions whose enantiomeric purities were determined by using the analytical conditions (93% of the starting material was recovered): 250 mg (52%) three: enantiomer 1 (5b-I1), chemical purity = 95%, ee = 100%; 90 mg (19%) threo: enantiomer 2 (**5b-I2**), ee = 86.8%; 109 mg (22%) *threo*: enantiomer 2 (**5b-I2**), chemical purity = 94%, ee = 98.5%. The enantiomers of **5b-II** were preparatively obtained by supercritical fluid chromatography by using a CHIRALPAK® AD column (250×20 mm, 10μ m) from Chiral Technologies, Inc. and recovery of 81% of the starting material (the enantiomeric purities of the fractions have been determined by using the analytical conditions): erythro **5b-II1** (70 mg), first eluted: $R_t = 16.21$ min, negative CD at 263 nm, ee = 99%; erythro 5b-II2(108 mg), second eluted: $R_t = 17.31$ min, positive CD at 263 nm, ee = 99%. Analytical and preparative separation of rac-7 were performed with CHI-RALCEL OD columns with different sizes (either 250×4.6 mm, $10\,\mu\text{m}$ or $0.46\times$ 25 cm, $10\,\mu\text{m})$ and the same mobile phase. The analytic chromatographic parameters obtained are: R_{t_1} = 14.50 min, $R_{t_2} = 16.00$ min. On the preparative scale, 167 mg of rac-7 was resolved with isolation of two fractions and recovery of 74% of the starting material (the enantiomeric purities of the fractions have been determined by using the analytical conditions): 52 mg enantiomer (–)-7, chemical purity >95% (by 400 MHz ¹H

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NMR); 71 mg enantiomer (+)-7, chemical purity >95% (by 400 MHz ¹H NMR).

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