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Stereochemical assignment of topsentolide C₂ by stereodivergent synthesis of its four diastereomers



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tion of topsentolide C₂ as 8R, 11S, and 12S.

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ABSTRACT

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Topsentolide C₂ is a cytotoxic oxylipin isolated, together with six other congeners (topsentolides A₁, A₂, B₁, B₂, B₃, and C₁), by Jung and co-workers from the marine sponge Topsentia sp.^{1,2} The structure of topsentolide C_2 was proposed as A (Fig. 1) on the basis of extensive spectroscopic analyses including the modified Mosher method to determine its 12S stereochemistry, coupled with the assumption that it would share the same S configuration at the C8 stereocenter as structurally related natural fatty acid lactones (halicholactone and neohalicholactone),³ although the stereochemistry at the C11 position was left unassigned because of the paucity of the isolated material. The naturally rare nine-membered lactone unit embedded in common in the topsentolides and their significant cytotoxicity against five human solid tumor cell lines $(ED50 = 2.0-17.5 \,\mu g/mL)$ have attracted considerable attention from organic chemists, and five synthetic studies on topsentolides have been reported so far.^{4–7} Among them, the one disclosed by Watanabe and co-workers led to the determination of the stereochemistry of topsentolide A1 to be 8R, 11R, and 12S (structure B in Fig. 1) through comparison of the ¹H NMR spectra and specific rotations of two stereoisomers of B with those of natural topsentolide A1.⁴ This Letter made us suspect that the genuine stereochemistry at the C8 position of topsentolide C₂ might also be R, instead of S as originally proposed by Jung and co-workers on the basis of analogy with halicholactone and neohalicholactone.^{1,3}

Since the absolute configuration at the C12 position of topsentolide C_2 was unambiguously determined to be *S* by the modified

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Four diastereomers of topsentolide C_2 , a cytotoxic nine-membered lactone isolated from the marine

sponge Topsentia sp., were synthesized stereodivergently from a common chiral seco acid by the com-

bined use of the Yamaguchi and Mitsunobu lactonizations. Comparison of the NMR spectra of the four

diastereomers with those of an authentic sample of topsentolide C_2 led to the stereochemical determina-

Figure 1. Structure of topsentolide C_2 (A) proposed by Jung et al. and the absolute stereochemistry of topsentolide A₁; (B) determined by Watanabe et al.

Mosher method, its stereochemistry should be represented by one of the four structures, **1**, 8-*epi*-**1**, 11-*epi*-**1**, and 8,11-bis-*epi*-**1** (Fig. 2). The synthesis of the four diastereomers and comparison of their NMR data (or other physicochemical properties) with those of topsentolide C_2 would, therefore, give a decisive answer concerning the stereochemistry of topsentolide C_2 . From a viewpoint of accessibility, however, we chose to synthesize 8,12-bis-*epi*-**1** and 12-*epi*-**1** together with **1** and 8-*epi*-**1**, since 8,12-bis-*epi*-**1** and 12-*epi*-**1** are enantiomeric to 11-*epi*-**1** and 8,11-bis-*epi*-**1**, respectively, and therefore should provide the same NMR information as the latter two isomers. As part of our ongoing efforts toward the total synthesis of oxylipins⁸ as well as to unequivocally elucidate the stereochemistry of topsentolide C_2 , we conducted a stereodivergent synthesis of **1**, 8-*epi*-**1**, 8,12-bis-*epi*-**1**, and 12-*epi*-**1**.

Scheme 1 outlines our synthetic plan for the targeted four diastereomers. We envisaged that 1 and 8-*epi*-1 would be obtainable





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Scheme 1. Retrosynthetic analysis of 1, 8-epi-1, 12-epi-1, and 8,12-bis-epi-1.

by the Yamaguchi and Mitsunobu lactonization, respectively, from common seco acid **2**. Subjection of **1** and 8-*epi*-**1** to the Mitsunobu inversion reaction would then provide 12-*epi*-**1** and 8,12-bis-*epi*-**1**, respectively. The common hydroxy acid intermediate **2** should readily be prepared from aldehyde **3** and phosphonate **4** via the Horner–Wadsworth–Emmons olefination.

The preparation of the aldehyde intermediate **3** began with the conversion of known protected alcohol **5a**⁹ into the corresponding phosphonium ion **5c** via iodide **5b** in 82% yield for the two steps (Scheme 2). The Wittig reaction of **5c** with methyl 5-oxopentanoate afforded *Z*-olefin **6**; the corresponding *E* isomer was not detected by NMR analysis. Deprotection of the acetal group of **6** followed by bis-TBS protection of the resulting diol **7a** gave **7b**, the treatment of which with HF-Py in THF-Py effected selective unmasking of the primary hydroxy group,¹⁰ furnishing **7c**. Finally, exposure of **7c** to the Swern oxidation conditions afforded **3**.



Scheme 2. Preparation of 3. Reagents and conditions: (a) I_2 , Ph_3P , Imid, CH_2CI_2 , 0 °C, 8 h; (b) Ph_3P , MeCN, rt, 14 h, 82% from 5a; (c) NaHMDS, methyl 5-oxopentanoate, THF, -78 °C to rt, 12 h, 70%; (d) TsOH·H₂O, MeOH, reflux, 14 h, 96%; (e) TBSCI, Imid, DMF, 0 °C, 2 h, 98%; (f) HF·Py, THF/Py, 0 °C to rt, 7 h, 75%; (g) (COCI)₂, DMSO, Et₃N, CH₂CI₂, -78 to -30 °C, 3 h, 94%.

The other coupling partner **4** in the Horner–Wadsworth– Emmons reaction was obtained from known iodide $9^{8b,11}$ by a four-step sequence shown in Scheme 3. The Evans asymmetric alkylation of $8^{8b,12}$ with **9** gave **10** as an 15:1 mixture of diastereomers, from which pure **10** was isolated in 62% yield by SiO₂ column chromatography. Hydrolytic removal of the chiral auxiliary in **10** and subsequent Weinreb amide formation from the resulting carboxylic acid **11a** afforded **11b**, which was then treated with dimethyl lithiomethylphosphonate to furnish **4**.

With the two building blocks, 3 and 4, in hand, we proceeded to the final stage of our stereodivergent synthesis of the four diastereomers of topsentolide C₂ (Scheme 4). The E-selective Horner-Wadsworth-Emmons reaction between 3 and 4 under the Roush-Masamune conditions gave **12** in 86% yield;¹³ none of the corresponding Z isomer could be detected by NMR analysis. Reduction of 12 under Luche's conditions proceeded highly diastereoselectively,^{8b} furnishing Felkin-Ahn product **13a** in 85% isolated vield. O-Methylation of the alcohol 13a followed by removal of the TBS group of the resulting methyl ether **13b** gave ester **13c**, which was then saponified with aqueous LiOH to afford seco acid 2. The Yamaguchi lactonization of 2 followed by deprotection of the PMB group took place uneventfully, giving nine-membered lactone 1 in 90% yield for the two steps. Subjection of the alcohol 1 to the Mitsunobu inversion conditions using *p*-nitrobenzoic acid as a nucleophile gave a p-nitrobenzoate intermediate, albeit in a modest yield of 34% probably due to the elimination of H₂O to generate undesired tetraene lactones, as judged by ¹H NMR analysis. Methanolysis of the benzoate intermediate furnished 12-epi-1 in 63% yield along with the starting benzoate recovered in 34% yield. The Mitsunobu lactonization of 2, on the other hand, afforded 8-epi-1 in 56% yield, after removal of the PMB group. Finally,



Scheme 3. Preparation of **4.** Reagents and conditions: (a) NaHMDS, THF, $-78 \degree C$, 19 h, 62%; (b) LiOOH, THF/H₂O, 0 $\degree C$ to rt, 3 h; (c) NHMe(OMe)·HCl, DCC, DMAP, CH₂Cl₂, 0 $\degree C$ to rt, 4 h, 80% from **10**; (d) MePO(OMe)₂, *n*-BuLi, THF, $-78 \degree C$, 6 h, 87%.



Scheme 4. Synthesis of four diastereomers of topsentolide C₂: Reagents and conditions: (a) Et₃N, LiBr, THF, rt, 23 h, 86%; (b) NaBH₄, CeCl₃·7H₂O, MeOH, -78 °C, 1 h, 85%; (c) NaHMDS, MeI, THF, -78 °C to rt, 24 h, 83%; (d) TBAF, THF, 0 °C, 6 h, 98%; (e) LiOH·H₂O, THF/H₂O, rt to 40 °C, 12 h, quant; (f) Cl₃C₆H₂COCl, i-Pr₂NEt, THF, 0 °C to rt, then DMAP, toluene, 90 °C, 15 h, 90%; (g) DDQ, THF/H₂O, 0 °C to rt, 2 h, quant; (h) DEAD, Ph₃P, p-(NO₂)C₆H₄CO₂H, toluene, 0 °C to rt, 4 h, 34%; (i) K₂CO₃, MeOH, 0 °C, 7 h, 63% (95% brsm); (j) DEAD, Ph₃P, p-(NO₂)C₆H₄CO₂H, toluene, 0 °C to rt, 24 h, 56%; (k) DDQ, THF/H₂O, 0 °C to rt, 2 h, quant; (l) DEAD, Ph₃P, p-(NO₂)C₆H₄CO₂H, toluene, 0 °C to rt, 2 h, 56%; (k) DDQ, THF/H₂O, 0 °C to rt, 2 h, quant; (l) DEAD, Ph₃P, p-(NO₂)C₆H₄CO₂H, toluene, 0 °C to rt, 2 h, 56%; (m) K₂CO₃, MeOH, 0 °C, 8 h, 61% (96% brsm).

application of the two-step protocol, employed for the conversion of **1** into 12-*epi*-**1**, to 8-*epi*-**1** provided 8,12-bis-*epi*-**1**.¹⁴

With the four diastereomers in hand, we compared their spectral data with those of an authentic sample of topsentolide C_2 to elucidate its stereochemistry. Although the ¹³C NMR spectral data of the four diastereomers were very similar to one another, slight differences were observed, especially in the chemical shift difference between the C-8 and C-12 carbons: $\delta_{C-12}-\delta_{C-8}=0.8$ and 1.0 ppm for 1 and 8-epi-1 (11,12-syn isomers), respectively, 0.5 ppm for both 12-epi-1 and 8,12-bis-epi-1 (11,12-anti isomers), and 0.9 ppm for topsentolide C_2 (see Supplementary data). Moreover, the ¹H NMR spectra of 12-epi-**1** and 8,12-bis-epi-**1** (11,12-anti isomers) were clearly different from that of topsentolide C_2 ; the signals for the 11-H and 12-H of the 11,12-anti isomers were observed as two separate sets of peaks [δ 3.54 (1H, dd, J = 7.6, 4.3 Hz) and 3.63 (1H, dt, J = 7.6, 4.8 Hz) for 12-epi-1, and δ 3.54 (1H, dd, J = 7.6, 4.5 Hz) and 3.62 (1H, dt, J = 7.6, 4.8 Hz) for 8, 12-bis-epi-1, while those of the 11,12-syn isomers as well as topsentolide C₂ appeared as overlapping peaks at δ 3.47–3.55 (2H, m). From these results, coupled with the 12S configuration assigned by the modified Mosher method, we could conclusively determine the stereochemistry at the side chain moiety of topsentolide C₂ as 11S and 12S. The difference between the two 11,12-syn isomers in ¹H NMR was, on the other hand, quite subtle, but close inspection of the spectra of 1, 8-epi-1, and authentic topsentolide C₂ indicated some noticeable differences in the shape of peaks in the region of δ 2.3–2.5 ppm, and the peak appearance in that region of topsentolide C₂ was more similar to that of 8-*epi*-**1** rather than **1** (see Supplementary data). As a whole, we led to the conclusion that the absolute stereochemistry of topsentolide C₂ should be represented by structure 8-*epi*-**1**.^{15,16}

In conclusion, four diastereomers of topsentolide C_2 (1, 8-*epi*-1, 12-*epi*-1, and 8,12-bis-*epi*-1) were synthesized stereodivergently from the common chiral seco acid **2** by the combined use of the Yamaguchi and Mitsunobu lactonizations. Comparison of the NMR spectra of the four diastereomers with those of topsentolide C_2 indicated that the absolute stereochemistry of topsentolide C_2 should be represented by structure 8-*epi*-1 [(8*R*,11*S*,12*S*)-isomer].

Acknowledgements

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.tetlet.2013. 10.032. These data include MOL files and InChiKeys of the most important compounds described in this article.

References and notes

- Luo, X.; Li, F.; Hong, J.; Lee, C.-O.; Sim, C. J.; Im, K. S.; Jung, J. H. J. Nat. Prod. 2006, 69, 567–571.
- For recent reviews on oxylipins, see: (a) Andreou, A.; Brodhun, F.; Feussner, I. Prog. Lipid Res. 2009, 48, 148–170; (b) Böttcher, C.; Pollmann, S. FEBS J. 2009, 276, 4693–4704; (c) Brodhun, F.; Feussner, I. FEBS J. 2011, 278, 1047–1063.
- (a) Niwa, H.; Wakamatsu, K.; Yamada, K. Tetrahedron Lett. **1989**, 30, 4543–4546;
 (b) Kigoshi, H.; Niwa, H.; Yamada, K.; Stout, T. J.; Clardy, J. Tetrahedron Lett. **1991**, 32, 2427–2428;
 (c) Critcher, D. J.; Connolly, S.; Wills, M. J. Org. Chem. **1997**, 62, 6638–6657.
- Kobayashi, M.; Ishigami, K.; Watanabe, H. Tetrahedron Lett. 2010, 51, 2762– 2764.
- Sreedhar, E.; Venkanna, A.; Chandramouli, N.; Babu, K. S.; Rao, J. M. Eur. J. Org. Chem. 2011, 1078–1083.
- (a) Fernandes, R. A.; Kattanguru, P. Tetrahedron Lett. 2011, 52, 1788–1790; (b) Fernandes, R. A.; Kattanguru, P. Tetrahedron: Asymmetry 2011, 22, 1930–1935.
- Wetzel, I.; Krauss, J.; Bracher, F. Lett. Org. Chem. 2012, 9, 169–174.
 (a) Miura, A.; Kuwahara, S. Tetrahedron 2009, 65, 3364–3368; (b) Kurashina, Y.;
- (a) Milita, A., Ruwahara, S. Fertunearon 2009, 05, 5504–5508, (b) Rufashina, F., Miura, A.; Enomoto, M.; Kuwahara, S. *Tetrahedron* 2011, 67, 1649–1653; (c) Kurashina, Y.; Kuwahara, S. *Biosci. Biotechnol. Biochem.* 2012, 76, 605–607.
- (a) Honjo, E.; Kutsumura, N.; Ishikawa, Y.; Nishiyama, S. *Tetrahedron* 2008, 64, 9495–9506; (b) Jackson, S. K.; Karadeolian, A.; Driega, A. B.; Kerr, M. A. J. Am. Chem. Soc. 2008, 130, 4196–4201.
- 10. Ruiz, P.; Murga, J.; Carda, M.; Marco, J. A. J. Org. Chem. 2005, 70, 713–716.
- Singh, J.; Kaur, J.; Nayyar, S.; Bhandari, M.; Kad, G. L. J. Indian Chem., Sect B: Org. Chem. Incl. Med. Chem. 2001, 40B, 386–390.
- 12. Askin, D.; Reamer, R. A.; Joe, D.; Volante, R. P.; Shinkai, I. *Tetrahedron Lett.* 1989, 30, 6121–6124.
- 14. *Physical and spectral data* 1: $[\varkappa]_D^{28} 95.8$ (c 0.570, MeOH); ¹H NMR (400 MHz, CD₃OD) δ 0.90 (3H, t, *J* = 6.9 Hz), 1.25–1.40 (6H, m), 1.70–1.82 (1H, m), 2.00–2.24 (7H, m), 2.30–2.58 (4H, m), 3.29 (3H, s), 3.47–3.55 (2H, m), 5.25–5.31 (1H, m), 5.42–5.53 (4H, m), 5.69 (1H, ddd, *J* = 15.6, 7.7, 1.2 Hz), 5.87 (1H, dd, *J* = 15.6, 5.7 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 14.4, 23.6, 26.3, 27.6, 28.4, 30.4, 31.8, 32.7, 34.4, 35.6, 57.0, 74.0, 74.8, 85.7, 125.6, 126.6, 130.3, 132.7, 134.0, 136.2, 175.6; HRMS (FAB) *m/z* calcd for C₂₁H₃₄O₄Na ([M+Na]⁺) 373.2355, found 373.2351. 12–*epi*-11: $[\varkappa]_0^{28} 55 (c 0.47, MeOH); ¹H NMR (400 MHz, CD₃OD) <math>\delta$ 0.90 (3H, t, *J* = 6.9 Hz), 1.25–1.40 (6H, m), 1.70–1.82 (1H, m), 2.01–2.24 (7H, m), 2.25–2.41 (2H, m), 2.42–2.58 (2H, m), 3.28 (3H, s), 3.54 (1H, dd, *J* = 7.6, 4.3 Hz), 3.63 (1H, dt, *J* = 7.6, 4.8 Hz), 5.26–5.32 (1H, m), 5.40–5.53 (4H, m), 5.74 (1H, dd, *J* = 15.7, 7.6 Hz), 5.85 (1H, dd, *J* = 15.7, 5.4 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 14.4, 23.6, 26.3, 27.6, 28.5, 30.5, 31.8, 32.7, 34.4, 35.6, 56.9, 74.0, 74.5, 85.8, 125.6, 126.6, 129.7, 132.8, 134.4, 136.2, 175.7; HRMS (FAB) *m/z* calcd for C₂₁H₃₄O₄Na ([M+Na]⁺) 373.2355, found 373.2355, m/z, 11, 25–1.40 (6H, m), 1.70–1.82 (1H, m), 2.01–2.24 (7H, m), 2.01–2.24 (7H, m), 2.25–2.41 (2H, m), 2.42–2.58 (2H, m), 3.28 (3H, s), 3.54 (1H, dd, *J* = 7.6, 4.3 Hz), 5.26 (5.126.6, 129.7, 132.8, 134.4, 136.2, 175.7; HRMS (FAB) *m/z* calcd for C₂₁H₃₄O₄Na ([M+Na]⁺) 373.2355, found 373.2359. *B*-*epi*-1: $[\alpha]_1^{28}$ +86.8 (*c* 1.33, MeOH); ¹H NMR (400 MHz, CD₃OD) δ 0.90 (3H, *t, J* = 6.9 Hz), 1.25–1.40 (6H, m), 1.70–1.82 (1H, m), 2.00–2.24 (7H, m), 2.30–2.58 (4H, m), 3.29 (3H, s), 3.47

3.55 (2H, m), 5.24–5.30 (1H, m), 5.42–5.53 (4H, m), 5.70 (1H, ddd, *J* = 15.8, 7.4, 1.0 Hz), 5.87 (1H, dd, *J* = 15.8, 5.5 Hz); ¹³C NMR (100 MHz, CD₃OD) δ 14.4, 23.6, 26.3, 27.6, 28.4, 30.4, 31.8, 32.7, 34.4, 35.5, 57.0, 73.9, 74.9, 85.7, 125.6, 126.6, 130.1, 132.7, 134.0, 136.2, 175.7; HRMS (FAB) *m/z* calcd for C₂₁H₃₄QANa ([M+Na]⁺) 373.2355, found 373.2354. 8,12-bis-epi-1: $[\alpha]_{2}^{28}$ +66.2 (*c* 0.520, MeOH); ¹H NMR (400 MHz, CD₃OD) δ 0.90 (3H, t, *J* = 6.8 Hz), 1.25–1.40 (6H, m), 1.70–1.82 (1H, m), 2.01–2.24 (7H, m), 2.25–2.41 (2H, m), 2.42–2.58 (2H, m), 3.29 (3H, s), 3.54 (1H, dd, *J* = 7.6, 4.5 Hz), 362 (1H, dt, *J* = 7.6, 4.8 Hz), 5.25–5.31 (1H, m), 5.40–5.53 (4H, m), 5.73 (1H, dd, *J* = 15.8, 7.7 Hz), 5.85 (1H, dd, *J* = 15.8, 7.3 4.4, 35.6, 56.9, 74.0, 74.5, 85.9, 125.6, 126.6, 129.7, 132.8, 134.4, 136.2, 175.7; HRMS (FAB) *m/z* calcd for C₂₁H₃₄O₄Na ([M+Na]⁺) 373.2355, found 373.2353.

- 15. Topsentolide C_2 was suspected by Jung et al. to be an artifact derived from topsentolide A_2 (17,18-dhihydro-**B**, see Fig. 1) during its extraction with MeOH (see Ref. 1), and the absolute stereochemistry of topsentolide A_1 (**B**) was determined as dipicted in Figure 1 through synthetic studies by Watanabe et al. (see Ref. 4). Therefore, it would be natural to consider that topsentolide C_2 was formed by epoxide ring opening of topsentolide A_2 by MeOH at the allylic C11 position with inversion of configuration.
- 16. Unfortunately, the specific rotation values of 8-epi-1 $[\alpha]_D^{29}$ +86.8 (*c* 1.33, MeOH) and 1 $\{[\alpha]_D^{29} 95.8$ (*c* 0.570, MeOH) were both far different from that reported for topsentolide C₂ $\{[\alpha]_D^{23} + 14.5 (c 0.27, MeOH)\}$. Remeasurement of the specific rotation of authentic topsentolide C₂ is needed to solve the discrepancy.