New anthracycline disaccharides. Synthesis of L-daunosaminyl- $\alpha(1 \rightarrow 4)$ -2-deoxy-L-rhamnosyl and of L-daunosaminyl- $\alpha(1 \rightarrow 4)$ -2-deoxy-L-fucosyl daunorubicin analogues



Fabio Animati, Federico Arcamone,* Marco Berettoni, Amalia Cipollone, Maurizio Franciotti and Paolo Lombardi

Menarini Ricerche S.p.A., Via Tito Speri 10, 00040, Pomezia, Rome, Italy

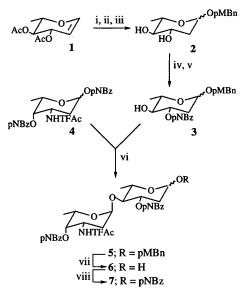
The synthesis of the new disaccharide anthracyclines 20, 21, 24 and 25, where the daunosamine moiety is separated from the aglycone by either a rhamnose or a fucose residue, performed following a convergent procedure, gives insight into the configurational requirement of the first sugar residue and opens the way to a new class of antitumour anthracyclines.

Anthracycline glycosides containing a disaccharide or trisaccharide chain reported in the literature ¹ bear the aminosugar directly attached to the anthracyclinone. According to X-ray analysis of drug–DNA complexes² and to theoretical calculations,³ the amino group in the first residue dictates the presence of an AT base pair in the position adjacent to the intercalation site, contributing to the DNA sequence specificity of the anthracyclines. In the course of our programme aimed at new antitumour anthracyclines, we became interested in the evaluation of the antitumour properties of anthracycline disaccharides *not* containing the amino group in the aglycone bound sugar residue.

We report here the synthesis of disaccharide daunorubicin analogues 20, 21, 24 and 25 having either a 2-deoxy-Lrhamnose or a 2-deoxy-L-fucose moiety directly linked to the aglycone, where the second sugar residue, daunosamine, is bound to the first one via an $\alpha(1\rightarrow 4)$ linkage. Anthracycline glycosides bearing such sugar sequences have not yet been synthesized, although recently the preparation of a (1-3)linked α -L-daunosaminyl 2-deoxy-L-rhamnosyl daunorubicin analogue has been disclosed.⁴

A convergent synthetic strategy, implying the separate construction of the two disaccharide moieties 7 (Scheme 1) and 15 (Scheme 2) and their successive coupling with both the aglycones 16 and 17, was adopted (Scheme 3) in order to avoid transglycosidation products 5 which may arise during the glycosylation of the anthracycline monosaccharide intermediate with the terminal sugar unit in the alternative stepwise strategy.

Starting materials for the synthesis of the p-nitrobenzoyl 3-Op-nitrobenzoyl-4-O-(3-N-trifluoroacetyl-4-O-p-nitrobenzoyl-a-L-daunosaminyl)-2-deoxy-L-rhamnoside 7 (Scheme 1) were 1,4-O-bis-p-nitrobenzoyl-3-N-trifluoroacetyldaunosamine 4. obtained by p-nitrobenzoylation of 3-N-trifluoroacetyl-daunosamine,⁶ and p-methoxybenzyl 3-O-p-nitrobenzoyl-2deoxy-L-rhamnoside 3. This was synthesized from 3,4-di-Oacetyl-L-rhamnal 1⁷ by reaction with *p*-methoxybenzyl alcohol (pMBnOH) and N-iodosuccinimide, followed by reduction with tributyltin hydride and de-O-acetylation to give 2. Selective protection in position C-3 was performed via formation of a stannylene cyclic acetal that exists in a dimeric form preferentially cleaved at the apical position,⁸ therefore affording 3 (72%) when treated with p-nitrobenzoyl chloride (pNBzCl) in pyridine. The reaction between 3 and 4, carried out in the presence of trimethylsilyltriflate (TMSOTf)⁶ gave the disaccharide 5 in high yield (79%) and with the desired α

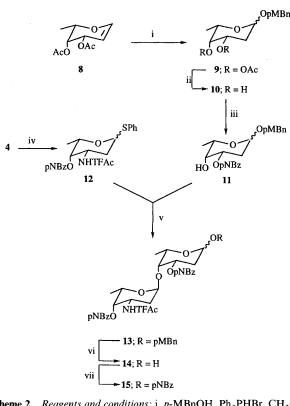


Scheme 1 Reagents and conditions: i, $(p-MeO)C_6H_4CH_2OH$ (p-MBnOH), N-iodosuccinimide (NIS), MeCN, -50 °C, 1 h; ii, Bu₃SnH, AIBN, reflux, 30 min; iii, MeONa, MeOH, room temp., 1 h; iv, Bu₂SnO, toluene, reflux, 6 h; v, $(p-NO_2)C_6H_4COCI$ (p-NBzCI), Et₃N, toluene, -40 °C, 2 h; vi, TMSOTf, CH_2Cl_2 , -15 °C, 40 min; vii, $Ce(NH_4)_2(NO_3)_6$, MeCN-H₂O, room temp., 1 h; viii, p-NBzCI, pyridine, CH_2Cl_2 , 0 °C, 2 h

configuration. The same disaccharide had been previously obtained as the methyl glycoside via Koenigs-Knorr glycosidation and subsequent alkoxyhalogenation and reduction of the disaccharide glycal intermediate.⁴ Our approach has the advantage that 4, more stable and easier to handle than the Ldaunosamine glycal derivative, allows a faster entry to the disaccharide 5. Quantitative removal of the anomeric pmethoxybenzyl group in 5 with ceric ammonium nitrate⁹ gave 6, that was activated (90% yield) at the anomeric position as the p-nitrobenzoyl ester 7. This was coupled with the aglycones 16¹⁰ and 17¹¹ according to a known procedure⁶ to give the corresponding α glycosides 18 and 19 in 60 and 57% yield, respectively (Scheme 3). Deprotection with 0.2 mol dm⁻³ Ba(OH)₂ afforded the final aminoglycosides 20 { $\delta_{\rm H}$ [(CD₃)₂SO] 5.2, d, 1 H, $J_{1',2'ax} = 4$ Hz} and 21 { $\delta_{\text{H}}[(\text{CD}_3)_2\text{SO}]$ 5.13, br s, 1 H, width $\frac{1}{2} = 9$ Hz}, \dagger isolated as hydrochlorides, in approximately 70% yield.

Starting materials for the synthesis of the activated protected disaccharide unit *p*-nitrobenzoyl 3-O-p-nitrobenzoyl-4-O-(3-N-trifluoroacetyl-<math>4-O-p-nitrobenzoyl- α -L-daunosaminyl)-2-

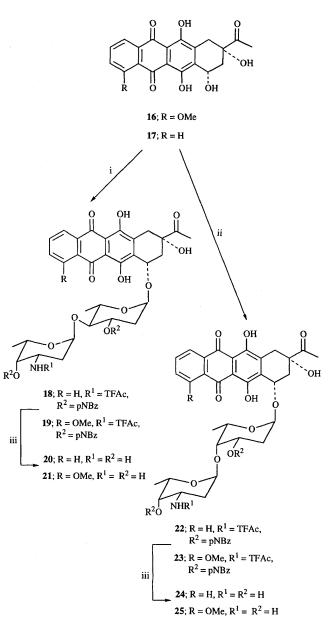
[†] NMR spectra were recorded on a Varian Gemini 300 MHz spectrometer. Supplementary data: see Instructions for Authors in the January issue.



Scheme 2 Reagents and conditions: i, p-MBnOH, Ph₃PHBr, CH₂Cl₂, room temp., 1 h; ii, MeONa 1 mol dm⁻³, MeOH, room temp.; iii, p-NBzCl, pyridine, CH₂Cl₂, -15 °C, 1 h; iv, PhSSiMe₃, CF₃SO₃SiMe₃, CH₂Cl₂, room temp., 12 h; v, Iodonium dicollidine perchlorate (IDCP), (CH₂Cl)₂-Et₂O, 4 Å molecular sieves, 5-10 °C, 2 h; vi, Ce(NH₄)₂(NO₃)₆, MeCN-H₂O, room temp., vii, p-NBzCl, pyridine, CH₂Cl₂, 0 °C, 2 h

deoxy-L-fucoside 15 (Scheme 2) were phenyl 4-O-pnitrobenzoyl-3-N-trifluoroacetyl-1-thiodaunosaminide 12, obtained from 4 upon treatment with (phenylsulfanyl)trimethylsilane and TMSOTf,¹² and p-methoxybenzyl 3-O-pnitrobenzoyl-2-deoxyfucoside 11. This derivative was prepared by the addition of p-methoxybenzyl alcohol to 3,4-di-O-acetyl-L-fucal 8^{13} in the presence of a Lewis acid catalyst to give 9, followed by de-O-acetylation and selective p-nitrobenzoylation of the resulting 10. Various glycosylation reactions using different methods 1d,14 for the activation of the glycosylating daunosamine derivative failed or gave poor yields of the desired disaccharide derivative 13. This known difficulty in the glycosylation of the C-4 axial hydroxy group¹⁵ of the sugar acceptor was overcome in the reaction of 11 with 12, carried out in the presence of iodonium dicollidine perchlorate¹⁶ to afford 13 in moderate yield. Conversion of 13 to the glycosylating disaccharide intermediate 15 was performed as above via oxidative cleavage of the p-methoxybenzyl group, followed by activation of 14 by p-nitrobenzoylation in practically quantitative overall yield. Finally, glycosylation of aglycones 16 and 17 to 22 and 23 followed by the deprotection step as above afforded the desired 4'-epimeric anthracycline disaccharides 24 and 25 in similar yield (40%) after purification as hydrochlorides by preparative reversed-phase HPLC (Scheme 3).

Biological evaluation of the four aminoglycosides showed that 20, 21 and 25 were less potent than clinically useful idarubicin and daunorubicin, whereas 24, possessing the 'natural' configuration at C-4' and lacking the 4-methoxy group at C-4, was endowed with a particularly high activity in inhibiting the growth of human tumour cells in culture.¹⁷ Therefore, our results open the way to an interesting new series of synthetic analogues of the antitumour anthracyclines that can be obtained in reasonable yields for further developments within this class of important pharmacological agents. We have



Scheme 3 Reagents and conditions: i, disaccharide 7, $CF_3SO_3SiMe_3$, CH_2Cl_2 -Et₂O, -20 °C, 1 h; ii, disaccharide 15, $CF_3SO_3SiMe_3$, CH_2Cl_2 -Et₂O, -20 °C, 1 h; iii, 0.2 mol dm⁻³ Ba(OH)₂, MeOH, 0 °C, 4 h

now found that the 14-hydroxy derivative of **24**, *i.e.* 3'-deamino-3'-hydroxy-4'-O- α -L-daunosaminyl-4-demethoxydoxorubicin, shows, according to our experience, unprecedented activity when compared with doxorubicin in different human cancers implanted in immunodepressed mice.¹⁸ The synthesis and the biological properties of the first doxorubicin disaccharide analogue will be reported in due course.

Acknowledgements

Financial support by A. Menarini (Industrie Farmaceutiche Riunite), Bristol-Myers Squibb (Italia) and Istituto Mobiliare Italiano (Grant No. 53658) is gratefully acknowledged.

References

 (a) F. Arcamone, Doxorubicin, Anticancer Antibiotics, Med. Chem. Series, ed. G. de Stevens, Academic Press, New York, 1981, vol. 17;
(b) S. Penco, G. Franchi and F. Arcamone, GP 2,751,395, May 18, 1978;
(c) H. Tanaka, T. Yoshioka, Y. Shimauchi, T. Oki, T. Ishikura, H. Umezawa and T. T. Takeichi, EP Appl., 63,776, Nov. 3, 1982;
(d) C. Kolar, G. Kneissl, H. Wolf and T. Kampchen, Carbohydr. Res., 1990, 208, 111;
(e) J. Boivin, C. Monneret and M. Païs, Tetrahedron, 1981, 37, 4219; (g) H. S. El Khadem and

- D. Matsuura, *Carbohydr. Res.*, 1982, **101**, C1. 2 A. H.-J. Wang, G. Ughetto, G. J. Quigley and A. Rich, *Biochemistry*, 1987, **26**, 1152.
- 3 N. Gresh, D. Pullman, F. Arcamone, M. Menozzi and R. Tonani, Mol. Pharmacol., 1989, 35, 251.
- 4 D. Horton, W. Priebe, M. L. Sznaidman and O. Varela, J. Antibiot., 1993, **46**, 1720.
- 5 H. S. El Khadem and A. Liav, Carbohydr. Res., 1979, 74, 199.
- 6 Y. Kimura, M. Suzuki, T. Matsumoto, R. Abe and S. Terashima, Bull. Chem. Soc. Jpn., 1986, 59, 423.
- 7 M. W. Bredenkamp, C. W. Holzapfel and F. Toerien, Synth. Commun., 1992, 22, 2459.
- 8 Y. Tsuda, M. E. Haque and K. Yoshimoto, Chem. Pharm. Bull. Jpn., 1983, 31, 1612.
- 9 B. Clason, P. J. Garegg and B. Samuelsson, Acta Chem. Scand., 1984, B38, 419.
- 10 F. Arcamone, G. Cassinelli, G. Franceschi, R. Mondelli, P. Orezzi and S. Penco, *Gazz. Chim. Ital.*, 1970, 100, 949.
- 11 W. Cabri, S. De Bernardinis, F. Francalanci and S. Penco, J. Chem. Soc., Perkin Trans. 1, 1990, 428.
- 12 S. Nambiar, J. F. Daeuble, R. J. Doyle and K. Grant Taylor, Tetrahedron Lett., 1989, 30, 2179.

- 13 B. Iselin and T. Reichstein, Helv. Chim. Acta, 1944, 27, 1200.
- 14 D. Kahne, S. Walker, Y. Chengand and D. van Engen, J. Am. Chem. Soc., 1989, 111, 6881.
- 15 J. Thiem and W. Klaffke, J. Org. Chem., 1989, 54, 2006.
- 16 P. Smid, G. A. de Ruiter, G. A. van der Marel, F. M. Rombouts and J. H. van Boom, J. Carbohydr. Chem., 1991, 10, 833. 17 F. Animati, F. Arcamone, M. Bigioni, M. Berettoni, A. Cipollone,
- M. Franciotti, P. Lombardi, G. Pratesi, R. Supino and F. Zunino, presented at the Ninth NCI-EORTC Symposium on New Drugs in Cancer Therapy, Amsterdam, 12-15 March, 1996, Comm. No. 382.
- 18 G. Pratesi, M. Bigioni, M. De Cesare, R. Supino, F. Zunino, F. Animati, F. Arcamone, M. Berettoni, A. Cipollone, M. Franciotti and P. Lombardi, presented at the Ninth NCI-OERTC Symposium on New Drugs in Cancer Therapy, Amsterdam, 12-15 March, 1996, Comm. No. 135.

Paper 6/01014H Received 12th February 1996 Accepted 25th April 1996