Stereocontrolled Synthesis of the C(1)–C(17) Half of **Boromycin**

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The ionophore boromycin (1)1 consists of two stereochemically related halves linked head to tail to form a diolide, with a borate bridge spanning the macrocycle.² We recently described³ the

reconstitution of 1 from a degradation product 2,4 in which the macrodiolide nucleus was first converted to a borate and then selectively esterified with D-valine. In parallel with these efforts, we have pursued syntheses of the two halves of 25 via a route that permits convergence with lactones 3 and 4 derived by further degradation of 2.2.3 Hanessian et al.6 have independently reported chiral syntheses of these lactones, and recently, Corey et al.⁷ announced the total synthesis of aplasmomycin, a symmetrical macrocyclic borate structurally allied to 1.

A synthesis of the chiral C(11)-C(17) segment of 2 was achieved from 3-buten-2-ol via enantioselective epoxidation with tert-butyl hydroperoxide in the presence of titanium tetraisopropoxide and diisopropyl D-(-)-tartrate.9 The resulting epoxide 5, which possessed the desired (2R,3S) configuration, 10 was alkylated with the tetrahydropyranyl ether of propargyl alcohol (n-BuLi, THF, $-78 \rightarrow 25$ °C) to give 6 (91%). This acetylene was semihydrogenated (10% Pd/BaSO₄, quinoline, MeOH), affording cis olefin 7 (98%), and the latter was converted directly to 8 (91%) with 2,2-dimethoxypropane (p-TsOH, MeOH, C₆H₆). In preparation for coupling with the C(3)-C(10) segment, 8 was transformed to allylic chloride 9 (83%) with N-chlorosuccinimide and dimethyl sulfide.11

Synthesis of the C(3)-C(10) moiety began from 3,3-dimethoxy-2,2-dimethylpropanol (10),12 obtained via condensation of isobutyraldehyde with formaldehyde.¹³ Oxidation of 10 (PCC) provided the malondial dehyde derivative 11 (66%, bp 55 °C (17 mm)), which was alkylated with the dianion¹⁴ of tiglic acid (2.2 equiv of LDA, THF, $-78 \rightarrow 25$ °C, 24 h) to give 12 (95%). This hydroxy acid was hydrogenated (94%, 10% Pd/C, EtOAc) and lactonized (DCC, DMAP) to yield 13 (80%) as a 40:60 mixture of cis/trans isomers. 15 Without separation, this mixture was taken to aldehyde 14 (TiCl₄, AcCl, CH₂Cl₂, 0 °C, 0.5 h)¹⁶ and then to carboxylic acid 15 (RuCl₃, NaIO₄, CCl₄, CH₃CN, H₂O; 68% from 13), 17 which, upon esterification (CH_2N_2), furnished 16 (96%). Treatment of 16 with (2R,3R)-(-)-butanediol (camphorsulfonic acid, C_6H_6) afforded the ortho ester 17¹⁸ in 84% yield as a mixture of four diastereomers, in which the trans/cis ratio was 3.7:1.19 Condensation of this mixture with the dianion²⁰ of methyl phenyl sulfone (1.5 equiv, 3 equiv of n-BuLi, THF, 0 °C) gave a mixture of keto sulfones (92%), from which the desired diastereomer 18 (mp 96-98 °C) was obtained (35% from 16) by HPLC on μPorasil.

Alkylation of the enolate of 18 (1.06 equiv of n-BuLi, Me₂SO-THF) with 9 in the presence of KI yielded 19 (97%) as a pair of diastereomers. The sulfonyl group was removed (Al/Hg,

⁽¹⁾ Hütter, R.; Keller-Schierlein, W.; Knusel, F.; Prelog, V.; Rodgers, G., Jr.; Suter, P.; Vogel, G.; Voser, W.; Zahner, H. Helv. Chim. Acta 1967, 50, 1533

⁽²⁾ Dunitz, J. D.; Hawley, D. M.; Miklos, D.; White, D. N. J.; Berlin, Yu.; Marušić, R.; Prelog, V. Helv. Chim. Acta 1971, 54, 1709.

(3) Avery, M. A.; White, J. D.; Arison, B. H. Tetrahedron Lett. 1981, 22,

⁽⁴⁾ Marsh, W.; Dunitz, J. D.; White, D. N. J. Helv. Chim. Acta 1974, 57,

⁽⁵⁾ A portion of this work was communicated previously. See: White, J. D. "Abstracts of Papers", 179th National Meeting of the American Chemical Society, Houston, TX, March 24-28, 1980; American Chemical Society: Washington, DC, 1980; ORGN 48.

^{(6) (}a) Hanessian, S.; Tyler, P. C.; Demailly, G.; Chapleur, Y. J. Am. Chem. Soc. 1981, 103, 6243. (b) Hanessian, S.; Delorme, D.; Tyler, P. C.; Demailly, G.; Chapleur, Y. Can. J. Chem. 1983, 61, 634. (7) Corey, E. J.; Pan, B.-C.; Hua, D. H.; Deardorff, D. R. J. Am. Chem. Soc. 1982, 104, 6816. Corey, E. J.; Hua, D. H.; Pan, B.-C.; Seitz, S. P. Ibid.

^{1982, 104, 6818.}

⁽⁸⁾ Okazaki, T.; Kitahara, T.; Okami, Y. J. Antibiot. 1975, 28, 176. Sato, K.; Okazaki, T.; Maeda, K.; Okami, Y. Ibid. 1978, 31, 632.

⁽⁹⁾ Martin, V. S.; Woodard, S. S.; Katsuki, T.; Yamada, Y.; Ikeda, M.; Sharpless, K. B. J. Am. Chem. Soc. 1981, 103, 6237.

⁽¹⁰⁾ It was established by an independent synthesis of 5 that epoxidation had taken place in 91% enantiomeric excess. See: White, J. D.; Sheldon, B. G.; Kang, M.-c. Tetrahedron Lett., in press.
(11) Corey, E. J.; Kim, C. U.; Takeda, M. Tetrahedron Lett. 1972, 4339.

⁽¹²⁾ Spath, E.; Pallan-Raschik, L. Monatsh. Chem. 1948, 79, 447 (13) Hattori, S.; Tokizawa, M. Japan Pat. 21 687, 1970; Chem. Abstr. 1970, 73, 87460d.

⁽¹⁴⁾ Katzenellenbogen, J. A.; Crumrine, A. L. J. Am. Chem. Soc. 1976, 98, 4925

⁽¹⁵⁾ Treatment of this mixture with LDA-THF gave an equilibrium composition of 80:20 in favor of trans-13

⁽¹⁶⁾ Berkowitz, W. F.; Choudhry, S. C.; Hrabie, J. A. J. Org. Chem. 1982,

⁽¹⁷⁾ Carlsen, P. H. J.; Katsuki, T.; Martin, V. S.; Sharpless, K. B. J. Org. Chem. 1981, 46, 3936.

⁽¹⁸⁾ Saucy, G.; Borer, R.; Trullinger, D. P.; Jones, J. B.; Lok, K. P. J. Org. Chem. 1977, 42, 3206. (19) Configurational equilibrium at the center α to the lactone carbonyl

is probably attained in this reaction via a ketene acetal intermediate. (20) Kondo, K.; Tunemoto, D. Tetrahedron Lett. 1975, 1397.

THF-H₂O (10:1), 75 °C, 1 h) to furnish **20** as an oil ($[\alpha]^{23}$ _D +3.5°) in 75% yield from 18. Reduction of this ketone (NaBH₄, MeOH, 25 °C) gave a mixture of 21 and 22 (98%; 2:1 respectively), the acetates of which (Ac₂O, pyridine, DMAP) were easily separated. The 7S acetate 23 underwent hydrolysis (p-TsOH, THF-H₂O (4:1), 56 °C, 12 h) to 24, which was saponified (NaOH, THF-H₂O, 25 °C, 3 h) and acidified (5% aqueous HCl) to provide 4 (91% from 23), identical with the degradation product from boromycin.3 For a rigorous comparison with naturally derived material, synthetic 4 was converted to triacetate 25 (68%,

 $[\alpha]^{20}$ _D +14.2°; Ac₂O, pyridine, DMAP), which was spectroscopically identical with the substance $([\alpha]^{20}_D + 17.6^{\circ})^{6b}$ obtained from 1.

With the configuration at the five chiral centers in this segment authenticated, attention was turned to its homologation in order to complete the C(1)-C(17) perimeter of 2. After protection of 4 as its bis(tert-butyldimethylsilyl) ether 26 (excess TBDMSCl, imidazole, DMF, 48 h), the latter was treated with 2,2-dimethoxypropane (p-TsOH, C₆H₆-MeOH) to give a quantitative yield of 27. This ester was saponified (20% aqueous NaOH, MeOH, followed by 2% HCl, 0 °C), and the derived carboxylic acid 28 was converted to 29 (carbonyldiimidazole, THF). Acylation of the enolate of methyl methoxyisopropylglycolate21 (LDA, THF, -78 °C, 10 min) with 29 afforded 30 as a C(2) epimeric mixture in 35% overall yield from 27.²² Stereochemical inhomogeneity at this stage is probably of no consequence, since it has been demonstrated in the synthesis of aplasmomycin that borate formation from the macrocyclic tetraol is accompanied by epimer-

ization at C(2) to the natural R configuration.⁷

The synthesis of 30 permits access to a fully functionalized subunit of 2 with rigorously defined stereochemistry and also opens a prospective route to the second half of boromycin.

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New Mechanism-Based Serine Protease Inhibitors: Inhibition of Human Leukocyte Elastase, Porcine Pancreatic Elastase, Human Leukocyte Cathepsin G, and Chymotrypsin by 3-Chloroisocoumarin and 3,3-Dichlorophthalide

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Mechanism-based irreversible inhibitors, which have been reported for porcine pancreatic (PP) elastase and bovine pancreatic chymotrypsin A_a, include halo enol lactones and 6-chloropyrones. Human leukocyte (HL) elastase and cathepsin G are related serine proteases which are involved in the connective tissue destruction that occurs in emphysema and various inflammatory diseases. Both enzymes are inhibited reversibly by heterocyclic structures such as benzoxazinones² and benzisothiazolinones,³ and this

(2) Teshima, T.; Griffin, J. C.; Powers, J. C. J. Biol. Chem. 1982, 257,

5085-5091.

⁽²¹⁾ Prepared by exposing a mixture of methyl glycolate and 2-methoxy-

propene to the vapor of POCl₃ (Caution: exotherm).

(22) For a recent account of the elegant approaches by Hanessian to the two halves of boromycin, see: Hanessian, S.; Delorme, D.; Tyler, P. C.; Demailly, G.; Chapleur, Y. In "Current Trends in Organic Synthesis"; Nozaki, H., Ed.; Pergamon Press: Oxford, U.K., 1983; p 205.

⁽¹⁾ Westkaemper, R. B.; Abeles, R. H. Biochemistry 1983, 22, 3256-3264. Vilkas, M. In "Enzyme-Activated Irreversible Inhibitors"; Seiler, N., Jung, M. J.; Koch-Weser, J., Eds., Elsevier/North Holland Biochemical Press: New York, 1978; pp 323-335. White, E. H.; Jelinski, J. S.; Politzer, I. R.; Branchini, B. R.; Roswell, D. F. J. Am. Chem. Soc. 1981, 103, 4231-4239. Chakravarty, P. K.; Krafft, G. A.; Katzenellenbogen, J. A. J. Biol. Chem. 1982, 257, 610-612. Moorman, A. R.; Abeles, R. H. J. Am. Chem. Soc. 1982, 104, 6785-6786. Alazard, R.; Bechet, J.; Dupaix, A.; Yon, J. Biochim. Biophys. Acta 1973, 309, 379-396.