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Stereocontrolled Aldol Additions to α -Methylene- β -Alkoxy Aldehydes: Application to the Synthesis of a $C_{13}-C_{25}$ Segment of Bafilomycin A₁.

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Abstract: A boron-mediated, syn-aldol coupling between ethyl ketone 8 and aldehyde 9, followed by directed hydrogenation at C_{16} and acetonide hydrolysis, gives the C_{13} - C_{25} segment 6 of bafilomycin A_1 .

Bafilomycin A₁ (1 in Scheme 1) belongs to a family of structurally related polyketide macrolide antibiotics, which include other bafilomycins,¹ the concanamycins² and the hygrolidins.³ First isolated in 1983 by Werner *et al.*,^{1a,b} bafilomycin A₁ is a potent and specific ATPase inhibitor⁴ which shows broad spectrum antibiotic activity. The stereochemistry, originally proposed by Corey⁵ from NMR analysis in combination with molecular modelling, was determined to be as shown in 1 by X-ray crystallography.^{1c} The first total synthesis of bafilomycin A₁ was recently completed by Evans and Calter,⁶ whilst a synthesis of a C₁₃–C₂₅ segment has been reported by the Roush group.⁷



As part of our work on polypropionate synthesis,⁸ we recently found that α -methylene- β -alkoxy aldehydes such as 2 produce useful levels of 1,3-asymmetric induction in aldol reactions with simple ketones. For example, Ti(IV), B and Sn(II) enolates 3 gave the 2,3-syn-3,5-anti adduct 4 preferentially, which can be stereoselectively hydrogenated, as in $4 \rightarrow 5$.^{8a} The macrolide bafilomycin A₁ appeared to be an ideal target to test this methodology, due to the occurrence of the same syn-anti-syn stereotetrad spanning C₁₅-C₁₈ (cf. 5). We now report a novel aldol construction of the C₁₃-C₂₅ bafilomycin A₁ segment 6 based on this approach.

Our synthetic strategy (Scheme 1) relied on: (i) introduction of the C_{16} stereocentre in 6 by hydroxyldirected hydrogenation of alkene 7; (ii) C_{17} - C_{18} bond formation by aldol coupling between ketone 8 and aldehyde 9. Achieving a useful level of remote stereocontrol (by 1,3-induction from 9 and/or 1,4-induction from 8) was an essential requirement for the aldol step. The synthesis of the coupling partners chosen for the critical C₁₇–C₁₈ aldol connection is summarised in Scheme 2. Ketone 8,⁹ with acetonide protection across the C₂₁ and C₂₃ hydroxyls, was efficiently prepared using an *anti* aldol reaction between the (*S*)-lactate-derived ketone 10 and isobutyraldehyde. Using our standard conditions,^{10a} the *E*-enol dicyclohexylborinate derived from 10 gave the *anti-anti* adduct 11 in 97% yield with 97% ds. After conversion^{10b} into aldehyde 12 (81%), a Felkin-Anh controlled addition of the allylsilane 13,¹¹ promoted by TiCl₄, gave an 84% yield of 14 with 97% ds. Alkene 14 then led to ketone 8, $[\alpha]_D^{20} = -9.1^{\circ}$ (*c* 3.0, CHCl₃), *via* a 3-step sequence (81% overall) of silyl ether deprotection, acetonide formation, and oxidative double bond cleavage. The synthesis of the aldehyde 9 relied on the Evans alkylation of the chiral glycolate 15.¹² Alkylation of the Ti(IV) enolate of 15 with BnOCH₂Cl gave 16 (85%) with high selectivity (97% ds). Transamination to the Weinreb amide,¹³ followed by addition of the organolithium reagent 17,¹⁴ then gave the enone 18 (75%). Luche reduction (NaBH₄, CeCl₃)¹⁵ of 18 proceeded with 98% ds in favour of the *anti* glycol derivative 19 (97%). Finally, silyl protection and acetal hydrolysis gave a 74% yield of the enal 9, $[\alpha]_D^{20} = +17.6^{\circ}$ (*c* 1.6, CHCl₃).



Scheme 2 (a) ^cHex₂BCl, Me₂NEt, Et₂O, 0 ^oC, 3 h; ⁱPrCHO, $-78 \rightarrow -20$ ^oC, 15 h; aq. MeOH, H₂O₂, 0 ^oC, 1 h. (b) TBSOTf, 2,6-lutidine, CH₂Cl₂ -78 ^oC, 2 h. (c) NaBH₄, MeOH, 0 ^oC, 30 min; K₂CO₃, MeOH, 20 ^oC, 3 h. (d) NalO₄, aq. MeOH, 20 ^oC, 30 min. (e) 13, TiCl₄, CH₂Cl₂, -94 ^oC, 10 min. (f) TBAF, THF, 20 ^oC, 30 min. (g) (MeO)₂CMe₂, PPTS, CH₂Cl₂, 20 ^oC, 4 h. (h) OsO₄, NMO, ^fBuOH/THF/H₂O, 20 ^oC, 4 h; NalO₄, pH 7 buffer, 10 min. (i) TiCl₄, ⁱPr₂NEt, CH₂Cl₂, 0 ^oC, 1 h; BnOCH₂Cl, 0 ^oC, 16 h. (j) MeONHMe.HCl, Me₃Al, CH₂Cl₂, -15 \rightarrow 20 ^oC, 20 h. (k) 17, THF, -78^oC, 2 h. (l), CeCl₃.7H₂O, NaBH₄, EtOH, -78 ^oC, 1.5 h. (m) TBSOTf, 2,6-lutidine, THF, 0 ^oC, 20 min; EtOH. (n) (CO₂H)₂, aq. THF, 20 ^oC, 20 h.

The π -facial selectivities of the ketone 8 and aldehyde 9 in aldol reactions were investigated separately (Scheme 3). As in our previous study,^{8a} the 1,3-asymmetric induction arising from the chiral aldehyde 9 was examined in reactions with the Sn(II), B and Ti(IV) enolates of diethylketone. In all cases, the major syn adduct 20, corresponding to si-face attack on the aldehyde was obtained,¹⁶ with good selectivity for both the Sn(II) and B enolates (20: 21 = ca. 4: 1). Minor amounts of anti adducts were also obtained. The sense of induction agreed with that from our earlier work (cf. 2 + 3 \rightarrow 4).^{8a} Contrary to our previous results, the TiCl₄-mediated reaction provided low facial selectivity here (20: 21 = 1.2: 1). The 1,4-induction in the syn aldol reaction of chiral ketone 8 with methacrolein was examined next. The Sn(II) enolate derived from 8 showed little selectivity for 22 vs 23. However, the corresponding Z-enol di-n-butylborinate gave improved results. With CH₂Cl₂ or Et₂O as solvent, the syn adduct 22, again corresponding to si-face attack on the aldehyde,¹⁶ predominated (22: 23 = 5: 1 in Et₂O). Cyclic protection of the 1,3-diol in 8 proved essential for good aldol stereocontrol.¹⁷

These studies suggested that matched double diastereodifferentiation should result from a boronmediated syn aldol coupling at C₁₇-C₁₈ of the two components. The desired (17*S*)-adduct 24 was accordingly obtained, with 82% ds, from addition of the Z-enol di-*n*-butylborinate of ketone 8 to aldehyde 9. Hydroxyldirected hydrogenation of alkene 24 using (Ph₃P)₃RhCl^{8a,18} introduced the C₁₆ stereocentre with excellent selectivity (>99 : 1 ds), giving 25 in 96% yield. This has the required syn-anti-syn C₁₅-C₁₈ stereotetrad of bafilomycin A₁. Under acidic conditions in methanol, the acetonide and silyl ether in 25 were removed and cyclisation occurred to give a 2 : 1 mixture of the spiroacetal 26⁹ and the hemiacetal 27 (a C₁₃-C₂₅ segment of L681,110 B₁)¹⁹ in 60% yield. Note that both these compounds had incorporated methanol at C₂₁ (presumably by dehydration to the enone and conjugate addition of MeOH).²⁰ NOE studies performed on 26 served to confirm the stereochemistry at C₁₆, C₁₇ and C₁₈. Careful acetonide hydrolysis and cyclisation of 25 under aqueous acidic conditions, however, provided an 83% yield of 6⁹, [α]²⁰_D = +8.0° (c 0.25, CHCl₃), corresponding to the required C₁₃-C₂₅ segment of bafilomycin A₁.



Scheme 3 (a) Sn(OTf)₂, Et₃N, CH₂Cl₂, -78 °C, 2 h; RCHO, -78 \rightarrow -25 °C, 3 h. (b) ^{*n*}Bu₂BOTf, ^{*i*}Pr₂NEt, Et₂O, -78 °C, 2 h; RCHO, -78 \rightarrow -25 °C, 16 h. (c) TiCl₄, CH₂Cl₂, -78 °C, 30 min; ^{*i*}Pr₂NEt, 1 h; RCHO, 1 h. (d) as for b, except reaction in CH₂Cl₂. (e) H₂ (15 bar), PhH, (Ph₃P)₃RhCl, 16 h. (f) conc. HCl, MeOH (1:10), -7 °C, 1.5 h. (g) 40% aq. HF, 4:1 MeCN:H₂O, 20 °C, 1 h.

In conclusion, the hemiacetal 6, which contains the C_{13} - C_{25} subunit of the macrolide bafilomycin A_1 , has been synthesised in 11 steps and 22% overall yield starting from ketone (S)-10. Largely due to the efficiency of the key aldol coupling/hydrogenation sequence, $8 + 9 \rightarrow 24 \rightarrow 25$, a high level of stereocontrol is realised (73% overall ds for the introduction of 9 stereocentres).

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- 9. All new compounds gave spectroscopic data in agreement with the assigned structures. Compound 6 had ¹H NMR &(400 MHz, CDC1₃) 7.27 7.35 (5H, m, Ph), 5.00 (1H, d, J = 1.9 Hz, CHO<u>H</u>), 4.57 (1H, d, J = 12.0 Hz, C<u>H</u>_AH_BPh), 4.52 (1H, m, H₁₇), 4.49 (1H, d, J = 12.0 Hz, CH_AH_BPh), 4.04 (1H, br d, CHO<u>H</u>), 3.96 (1H, dd, J = 6.1, 2.6 Hz, H₁₅), 3.75 (1H, d, J = 8.2 Hz, H_{13A}), 3.68 (1H, ddd, J = 10.8, 9.9, 4.7 Hz, H₂₁), 3.43 3.56 (3H, m, H_{13B}, H₁₄, H₂₃), 3.42 (3H, s, OMe), 2.25 (1H, dd, J = 11.9, 4.7 Hz, H_{20A}), 1.93 (1H, m, H₁₆), 1.91 (1H, m, H₂₄), 1.61 (1H, m, H₁₈), 1.32 (1H, m, H₂₂), 1.16 (1H, dd [partially obscured], H_{20B}), 1.02 (3H, d, J = 6.8 Hz, C₂₄-Me_A), 0.94 (3H, d, J = 6.5 Hz, C₂₂-Me), 0.93 (3H, d, J = 7.1 Hz, C₁₈-Me), 0.86 (9H, s, Me₃CSi), 0.84 (3H, d, J = 7.0 Hz, C₁₆-Me), 0.82 (3H, d, J = 6.8 Hz, C₂₄-Me_B), 0.11 (3H, s, Me_ASi), 0.05 (3H, s, Me₃Si); HRMS (CI, NH₃) calc. for C₃₁H₅₇O₇Si (M + H)⁺ 569.38736 found 569.3870. Compound 26 had ¹H NMR &(500 MHz, CDCl₃) 7.27 7.36 (5H, m, Ph), 4.60 (1H, d, J = 12.2 Hz, CH_AH_BPh), 4.48 (1H, d, J = 12.2 Hz, CH_AH_BPh), 3.98 (1H, dd, J = 9.5, 2.3 Hz, H₁₅), 3.87 (1H, dd, J = 10.5, 2.2 Hz, H_{13A}), 3.79 (1H, m, H₁₇), 3.53 (1H, dd, J = 10.5, 6.0 Hz, H_{13B}), 3.45 (3H, s, OMe), 3.40 (1H, obscured, H₁₄), 3.28 (3H, s, OMe), 3.07 (1H, m, H₂₁), 3.03 (1H, dd, J = 11.0, 2.2 Hz, H₂₃), 2.17 (1H, m, H₁₆), 2.05 (1H, dd, J = 15.1, 7.4 Hz, H_{20A}), 1.86 (1H, m, H₂₂), 1.77 (1H, dd, J = 15.1, 1.1 Hz, H_{20B}), 1.75 (1H, m, H₂₄), 1.54 (1H, m, H₁₈), 0.98 (3H, d, J = 6.8 Hz, C₂₄-Me_A), 0.93 (3H, d, J = 6.5 Hz, C₂₄-Me_B); HRMS (CI, NH₃) calc. for C₂₆H₄₆O₆N (M + NH₄)⁴ 468.33251 found 468.3325.
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