

PII: S0960-894X(97)10061-0

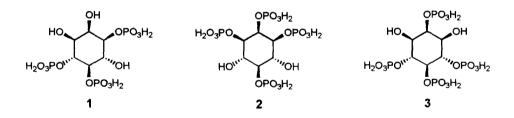
## SYNTHESES OF *MYO*-INOSITOL-1,2,3,5- AND -2,4,5,6-TETRAKISPHOSPHATES, UNUSUAL INHIBITORS OF *MYO*-INOSITOL-1,4,5-TRISPHOSPHATE 3-KINASE

Sung-Kee Chung\* and Young-Tae Chang

Department of Chemistry, Pohang University of Science & Technology, Pohang 790-784, Korea.

Abstract: D-myo-Inositol 1,4,5-trisphosphate [D-I(1,4,5)P<sub>3</sub>], a calcium mobilizing second messenger, is converted to D-I(1,3,4,5)P<sub>4</sub> by D-I(1,4,5)P<sub>3</sub>-3-kinase. Efficient syntheses of  $I(1,2,3,5)P_4$  (2) and  $I(2,4,5,6)P_4$  (3), novel 3-kinase inhibitors, are reported.© 1997 Elsevier Science Ltd.

Since the discovery that D-mvo-inositol 1.4.5-trisphosphate  $[I(1,4.5)P_3, 1]$  plays a pivotal role as a second messenger in the transmembrane signaling, thus mobilizing calcium ions from the intracellular storage, its interactions with the  $I(1.4,5)P_3$  receptor and metabolic enzymes have been widely studied.<sup>1</sup> One of the major metabolic pathways involves a specific phosphorylation of  $I(1,4,5)P_3$  to  $I(1,3,4,5)P_4$ , by  $I(1,4,5)P_3-3$ kinase  $[IP3K]^2$  It has been suggested that  $I(1,3,4,5)P_4$  also acts as a second messenger mediating the entry of extracellular  $Ca^{2+}$  through plasma membrane ion channel.<sup>3</sup> although the detailed mechanistic understanding has not yet been achieved. The other major metabolic pathway involves Ins(1,4,5)P<sub>3</sub> 5phosphatase to yield  $Ins(1,4)P_2$ .<sup>1</sup> Thus, IP3K not only occupies a central position in regulating the availability of the two Ca<sup>2+</sup> mobilizing second messangers but also provides an important branching point in the diverse pathways of the inositol polyphosphate metabolism. In the preceding paper,<sup>4</sup> we have reported the inhibitory activities of all possible regioisomers of IP<sub>n</sub> on IP3K and proposed an active site model for the enzyme on the basis of the binding affinity data. However, we noted that  $Ins(1,2,3,5)P_4$  (2) and  $Ins(2.4,5.6)P_4$  (3) did show substantial inhibitory effects, although their structures do not contain the essential 1,4,5-trisphosphate motif of D-Ins $(1,4,5)P_3$ . In order to help understand the structural characteristics of these substances in inhibiting IP3K, we sought efficient synthetic routes to these compounds. We report herein practical syntheses of compounds 2 and 3.

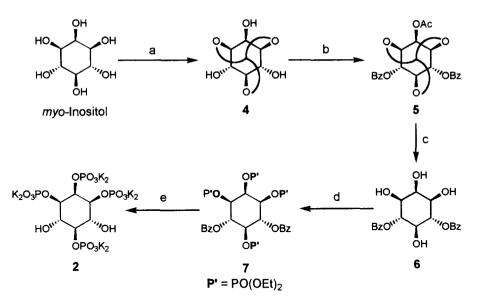


One of the key issues in the synthesis of inositol phosphates is to prepare suitable, selectively protected inositol intermediates. Inositol orthoformate, 4, which was proven to be a useful intermediate for the synthesis of various inositol phosphates by us and others.<sup>5</sup> was selected as the key intermediates for the synthesis of 2 and 3. The acetylation of compound 4 prepared from mvo-inositol<sup>6</sup> under the usual conditions employing AcCl in pyridine showed an initial acetylation of the 2-OH selectively. A preferential alkylation at 4- and 6-OH of 4 was previously reported under the conditions employing an alkyl halide and a metal hydride base.<sup>5c</sup> Although the enzyme assisted selective acetylation<sup>7</sup> and TBDMS silvlation<sup>5d</sup> of 4 at 2-OH are known, this is the first selective acetylation of 2-OH in 4 by a chemical method. Thus, successive treatments of 4 in pyridine with AcCl (1.6 eq., 1h) and then excess BzCl gave 5 as the major product together with a small amount of 2.4-diacetylated product (in ca. 3:1 ratio based on <sup>1</sup>H-NMR). A simple extractive work-up of the reation mixture was followed by an acid catalyzed hydrolysis to remove the acetyl and orthoester protecting groups. Pure 4,6-dibenzoated inositol, 6 was obtained by simple extraction in 56% yield over 3 steps from 4 and the by-product I(4)Bz was present exclusively in the water layer. Compound 6 was phosphorylated by successive treatments with diethyl chlorophosphite and diisopropylethylamine in DMF, and then 30%  $H_2O_2$  to afford compound 7<sup>8</sup> in 85% yield. In the final step. the protecting groups of 7 were removed by successive reactions with TMSBr and then LiOH. The target compound  $2^9$  was obtained after ion exchange chromatography on Dowex 50x8-100 (H<sup>+</sup> form), pH adjustment to 10 with KOH, and lyophilization (Scheme 1).

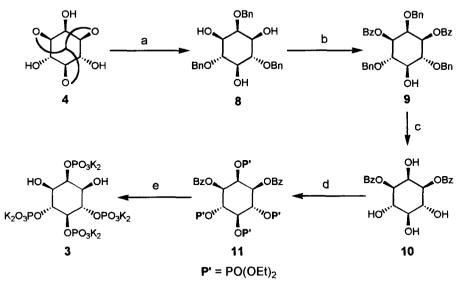
 $I(2,4,5,6)P_4$ , 3 was also synthesized conveniently from the intermediate 4 (Scheme 2). An exhaustive benzylation of 4 using excess amounts of BnBr and NaH was followed by an acid-catalyzed hydroylsis to obtain 8 in 94%. A selective benzoylation of 8 afforded the 1,3-dibenzoated product, 9 in 64% yield. The enhanced nucleophilic reactivity of 1- and 3-OHs toward BzCl might be related to the through-space  $\alpha$ effect caused by the *cis*-related 2-oxygen,<sup>10</sup> although the exact origin of this selectivity is not clear. Removal of the benzyl protecting groups in 9 by hydrogenolysis gave  $I(1,3)Bz_2$ , 10. Compound 10 was phosphorylated to give  $11^{11}$ , and the protecting groups of 11 were removed to give  $I(2,4,5,6)P_4$ ,  $3^{12}$  in good yield by the same procedures as described for  $I(1,2,3,5)P_4$ .

In conclusion, we successively prepared two novel IP3K inhibitors 2 and 3 in gram scales using inositol orthoformate as the key intermediate. These two routes represent the only synthetic pathways reported for  $I(1,2,3,5)P_4$  and  $I(2,4,5,6)P_4$  except the divergent total synthesis of all regioisomers of  $IP_4$ .<sup>13</sup>

Acknowledgement. This work was supported by the Korea Science & Engineering Foundation/Center for Biofunctional Molecules and the Korea Research Foundation.



Scheme 1. a. (EtO)<sub>3</sub>CH, pTSA, DMF, 89%. b.(i) AcCl (1.6 eq.), pyridine. (ii) BzCl (4 eq.). c. HCl-MeOH, 56% from 4. d.(i) (EtO)<sub>2</sub>P-Cl, iPr<sub>2</sub>NEt, DMF. (ii) H<sub>2</sub>O<sub>2</sub>, 85%. e.(i) TMSBr, CH<sub>2</sub>Cl<sub>2</sub>. (ii) LiOH, Δ.(iii) H<sup>+</sup> ion-exchange. (iv) KOH, pH 10, quant.



Scheme 2. a.(i) BnBr, NaH. (ii) HCl-MeOH, 94%. b. BzCl (2.5 eq), pyridine, 67%. c.  $H_2(1 \text{ atm})$ -Pd(OH)<sub>2</sub>, quant. d.(i) (EtO)<sub>2</sub>P-Cl, iPr<sub>2</sub>NEt, DMF. (ii) H<sub>2</sub>O<sub>2</sub>, 82%. e.(i) TMSBr, CH<sub>2</sub>Cl<sub>2</sub>. (ii) LiOH,  $\Delta$ .(iii) H<sup>+</sup> ion-exchange. (iv) KOH, pH 10, quant.

## **References and Notes**

- a) Majerus, P. W. Annu. Rev. Biochem. 1992, 61, 225-250. b) Berridge, M. J. Nature 1993, 361, 315-325.
- 2. Irvine, R. F.; Letcher, A. J.; Heslop, J. P.; Berridge, M. J. Nature 1986, 320, 631-634.
- 3. Irvine, R. F.; Cullen, P. J. Current Biology 1993, 3, 540-543.
- 4. Gildon Choi; Young-Tae Chang; Sung-Kee Chung; Kwan Yong Choi, preceding paper.
- a) Chung, S. K.; Chang, Y. T.; Sohn, K. H. J. Chem. Soc., Chem. Commun. 1996, 163-164. b) Ozaki,
  S.; Koga, Y.; Ling, L.; Watanabe, Y.; Kimura, Y.; Hirata, M. Bull. Chem. Soc. Jpn. 1994, 67, 1058-1063. c) Billington, D. C.; Baker, R.; Kulagowski, J. J.; Mawer, I.; Vacca, J. P.; deSolms, S. J.; Huff,
  J. R. J. Chem. Soc., Perkin Trans. 1 1989, 1423-1429. d) Baudin, G.; Glanzer, B. I.; Swaminathan, K.
  S.; Vasella, A. Helv, Chim. Acta 1988, 71, 1367-1378.
- 6. Chung, S. K.; Chang, Y. T.; Sohn, K. H. Kor. J. Med. Chem. 1994, 4, 57-65.
- 7. Andersch, P.; Schneider, M. P. Tetrahedron: Asymmetry 1993, 4, 2135-2138.
- 8. 7: mp 134-137 °C; <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$  0.74-1.43 (m, 24H, 8CH<sub>2</sub>CH<sub>3</sub>), 3.49-4.29 (m, 16H, 8CH<sub>2</sub>CH<sub>3</sub>), 4.72 (app. tt, J = 1.9, 10.1 Hz, 2H, H-1 & H-3), 4.91 (app. q, J = 9.4 Hz, 1H, H-5), 5.27 (dt, J = 2.3, 9.1 Hz, 1H, H-2), 5.92 (app. t, J = 10.0 Hz, 2H, H-4 & H-6), 7.42-8.18 (m, 10H, 2Ph); <sup>13</sup>C-NMR (CDCl<sub>3</sub>)  $\delta$  15.25-16.04 (8CH<sub>2</sub>CH<sub>3</sub>), 63.82-64.50 (8CH<sub>2</sub>CH<sub>3</sub>), [70.50(2C), 73.39(2C), 75.46, 76.57, inositol ring carbon], 128.36-133.29 (2Ph), 165.42 (2C, 2PhCO); <sup>31</sup>P-NMR (CDCl<sub>3</sub>)  $\delta$ -1.97, -0.78, -0.59 (2P).
- 9. **2**: <sup>1</sup>H-NMR (D<sub>2</sub>O, pH 10)  $\delta$  3.76 (q, J = 7.6 Hz, 1H, H-5), 3.81-3.90 (m, 4H, H-1, H-3, H-4, H-6), 4.52 (br d, J = 8.8 Hz, 1H, H-2); <sup>13</sup>C-NMR (D<sub>2</sub>O, pH 10)  $\delta$  74.71 (2C), 76.56 (2C), 78.32, 81.69; <sup>31</sup>P-NMR (D<sub>2</sub>O, pH 10)  $\delta$  3.41, 5.03, 5.75 (2P).
- Kim, K. S.; Cho, S. J.; Oh, K. S.; Son, J. S.; Kim, J.; Lee, J. Y.; Lee, S. J.; Lee, S.; Chang, Y. T.; Chung, S. K.; Ha, T. K.; Lee, B. S.; Lee, I. J. Phys. Chem. 1997, 101, 3776-3783.
- 11. 11: Oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 0.84-1.41 (m, 24H, 8CH<sub>2</sub>CH<sub>3</sub>), 3.64-4.31 (m, 16H, 8CH<sub>2</sub>CH<sub>3</sub>), 4.66 (app. q, J = 10.0 Hz, 1H, H-5), 5.09 (app. q, J = 9.3 Hz, 2H, H-4 & H-6), 5.24 (br d, J = 9.3 Hz, 1H, H-2), 5.34 (br d, J = 10.0 Hz, 2H, H-1 & H-3), 7.40-7.57 (m, 10H, 2Ph); <sup>13</sup>C-NMR (CDCl<sub>3</sub>) δ 16.06-16.79 (8CH<sub>2</sub>CH<sub>3</sub>), 64.54-65.19 (8CH<sub>2</sub>CH<sub>3</sub>), [70.84 (2C), 74.17, 74.24, 75.87 (2C), inositol ring carbon], 128.93-134.10 (2Ph), 166.15 (2C, 2PhCO); <sup>31</sup>P-NMR (CDCl<sub>3</sub>) δ -1.36 (2P), -1.18, -0.65.
- 12. **3**: <sup>1</sup>H-NMR (D<sub>2</sub>O, pH 10)  $\delta$  4.47 (m, 3H, H-4, H-5, H-6), 4.59 (br d, J = 10.6 Hz, H-2), 4.70 (br d, J = 11.9 Hz, 2H, H-1, H-3); <sup>13</sup>C-NMR (D<sub>2</sub>O, pH 10)  $\delta$  70.26, 75.01 (3C), 76.69 (2C); <sup>31</sup>P-NMR (D<sub>2</sub>O, pH 10)  $\delta$  3.82, 5.02 (3P).
- 13. Chung, S. K.; Chang, Y. T. J. Chem. Soc., Chem. Commun. 1995, 11-12.

(Received in Japan 20 August 1997; accepted 22 September 1997)