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MODULATION OF THE INOSITOL 1,4,5-TRISPHOSPHATE RECEPTOR BY INOSITOL PHOSPHATES

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Abstract The mode of recognition at the binding site of the Ins(1,4,5)P₃ receptor was assessed by examining the structure-activity relationships of different Ca²⁺-mobilizing inositol phosphates.

Key Words: the Ins(1,4,5)P₃ receptor; inositol phosphates; calcium mobilization; rat brain microsomes; calcium signaling

INTRODUCTION

The pivotal role of D-*myo*-inositol 1,4,5-trisphosphate [Ins(1,4,5)P₃] in intracellular Ca²⁺ signaling is well recognized. In the cytosol, Ins(1,4,5)P₃ and its metabolites undergo extensive metabolism by the sequential actions of specific phosphatases and kinases, from which a plethora of inositol phosphates are produced (Figure 1).

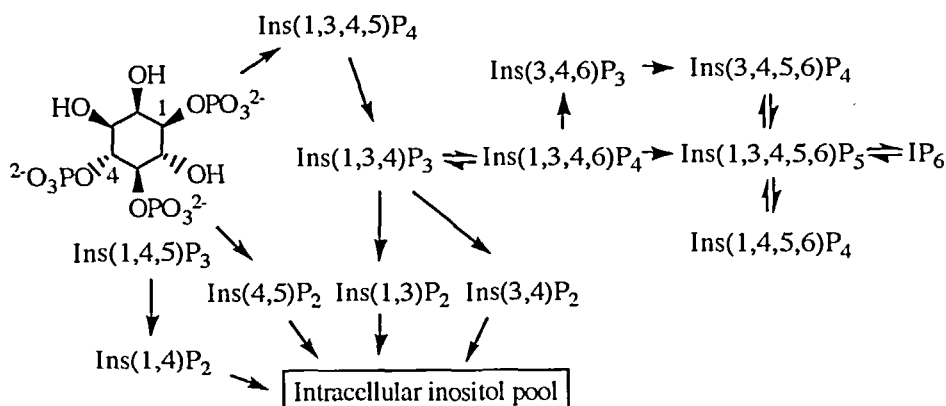


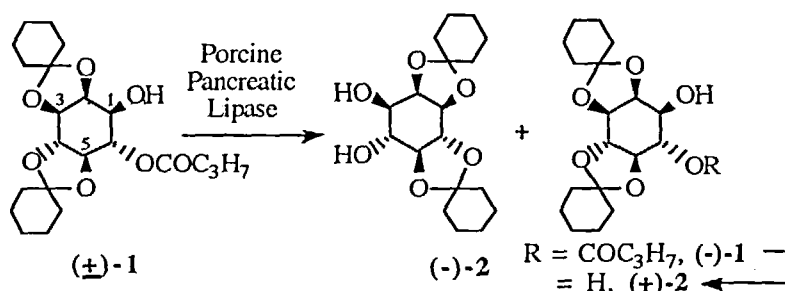
Figure 1 Metabolism of inositol phosphates

The rich diversity of phosphoinositols generated from such a complex metabolic network implies the physiological relevance of these molecules. Thus, examination of

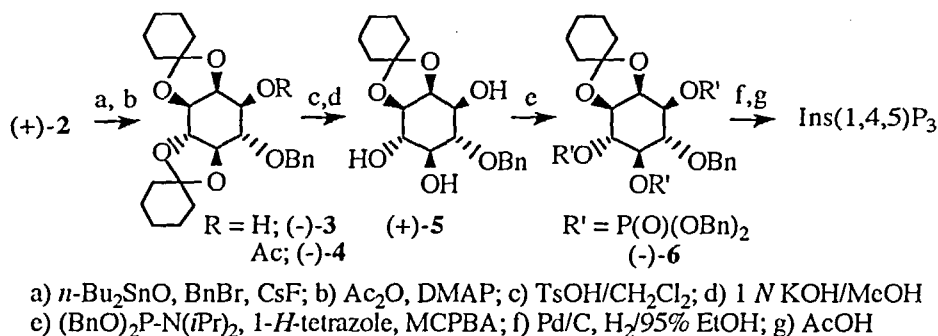
the second-messenger role of inositol phosphates in Ca^{2+} signaling constitutes one of our research foci. In this account, we report the systematic synthesis of phosphoinositol congeners and their interactions with the $\text{Ins}(1,4,5)\text{P}_3$ receptor.

CHEMOENZYMATIC SYNTHESIS OF INOSITOL PHOSPHATES

Our key strategy employed a pair of enantiomerically active 1,2:5,6-dicyclohexylidene-*myo*-inositols (**2**) as common precursors to the target molecules, which was prepared by a facile enzymatic method. The 4-butyryl monoester [(\pm)-**1**] was subjected to enantiospecific hydrolysis by porcine pancreatic lipase, which gave both product [($-$)-**2**] and substrate [($-$)-**1**] fractions with satisfactory optical purity (e.e. ≥ 0.98) after recrystallization.



The synthetic utility of **2** is illustrated by the example of $\text{Ins}(1,4,5)\text{P}_3$ synthesis.



Both enantiomerically active **2** allow the synthesis of eleven *D*-*myo*-inositol phosphates in fair yields, which included $\text{Ins}(1,4)\text{P}_2$, $\text{Ins}(4,5)\text{P}_2$, $\text{Ins}(1,3,4)\text{P}_3$, $\text{Ins}(1,4,5)\text{P}_3$, $\text{Ins}(1,5,6)\text{P}_3$, $\text{Ins}(1,2,5,6)\text{P}_4$, $\text{Ins}(1,3,4,5)\text{P}_4$, $\text{Ins}(1,3,4,6)\text{P}_4$, $\text{Ins}(1,4,5,6)\text{P}_4$, $\text{Ins}(3,4,5,6)\text{P}_4$, $\text{Ins}(1,3,4,5,6)\text{P}_5$.

INOSITOL PHOSPHATE-INDUCED CALCIUM RELEASE

Ca^{2+} -loaded rat brain microsomes were treated with individual inositol phosphates at 37 °C, and the released Ca^{2+} was monitored by bulk fluorimetry using Fura-2 as an indicator. Of the 12 phosphoinositols examined (the aforementioned 11 synthetic molecules and glycerophospho-D-*myo*-inositol 4,5-bisphosphate [GroPIns(4,5)P₂; purchased from Sigma], Ins(1,4,5)P₃, GroPIns(4,5)P₂, Ins(1,3,4,6)P₄, Ins(1,3,4,5)P₄, Ins(1,4,5,6)P₄, and Ins(4,5)P₂ exhibited Ca^{2+} -mobilizing activity in a dose-dependent manner (Figure 2), with apparent EC₅₀ values of 0.13, 1.3, 4.4, 8.2, 11.2, and 60 μM , respectively. Other inositol phosphates including Ins(1,4)P₂, Ins(1,5,6)P₃, Ins(1,3,4)P₃, Ins(3,4,5,6)P₄, Ins(1,2,5,6)P₄, and Ins(1,3,4,5,6)P₅ failed to exert appreciable Ca^{2+} release from the microsomal preparation, even at concentrations up to 100 μM .

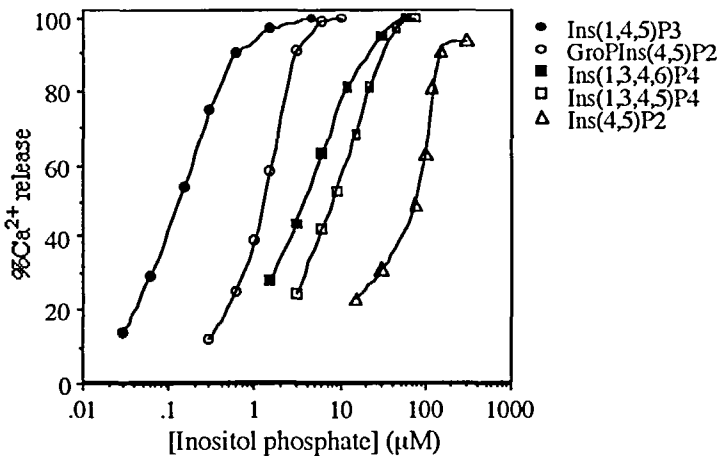


Figure 2 Inositol phosphate-induced Ca^{2+} release from rat brain microsomes.

BINDING AFFINITY OF INOSITOL PHOSPHATES WITH THE INS(1,4,5)P₃ RECEPTOR

To assess the binding of inositol phosphates to the Ins(1,4,5)P₃ receptor, displacement of specific [³H]Ins(1,4,5)P₃ binding was carried out using rat cerebellar membrane preparations. According to the displacement curves (not shown), the mean dissociation constants (K_d) for individual inositol phosphates were determined as follows ($n = 3$): Ins(1,4,5)P₃, 0.028 μM ; GroPIns(4,5)P₂, 0.92 μM ; Ins(1,3,4,5)P₄, 1.4 μM ; Ins(1,4,5,6)P₄, 2.1 μM ; Ins(1,3,4,6)P₄, 2.2 μM ; Ins(4,5)P₂, 24 μM ; Ins(1,3,4,5,6)P₅, 40 μM ; Ins(3,4,5,6)P₄, 56 μM ; Ins(1,2,5,6)P₄, 57 μM ; Ins(1,3,4)P₃, 146 μM ; Ins(1,4)P₂, 217 μM ; Ins(1,5,6)P₃, 454 μM . For the inositol phosphates capable of effecting Ca^{2+} mobilization, the relative potency of inhibiting [³H]Ins(1,4,5)P₃ binding to the receptor paralleled the order of the EC₅₀ values.

LIGAND RECOGNITION AT THE INS(1,4,5)P₃ RECEPTOR

Analysis of the structures of Ca²⁺-mobilizing inositol phosphates indicates that all these molecules assume conformations sharing or mimicking the structural features of the 4,5-bisphosphate 6-hydroxy and 1-phosphate motifs of Ins(1,4,5)P₃ (Figure 3)

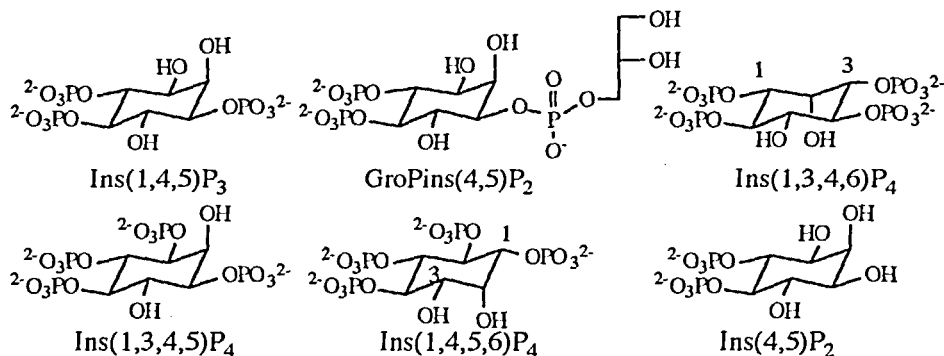


Figure 3 Structures of some inositol phosphates capable of eliciting Ca²⁺ release

On the basis of this finding, we propose a binding model to account for ligand recognition at the Ins(1,4,5)P₃ receptor (Figure 4). The binding site is presumably composed of two domains. The anchoring domain interacts with the 4,5-bisphosphate 6-hydroxy motif, attributing to the Ca²⁺-mobilizing activity. The auxiliary domain exerts long-range electrostatic interactions with the 1-phosphate group, which enhances the binding affinity. The stereochemical requirement for this phosphate recognition is, however, less stringent. The biochemical implication of the cross-reactivity of the Ins(1,4,5)P₃ receptor with a number of inositol phosphates besides Ins(1,4,5)P₃ remains unclear.

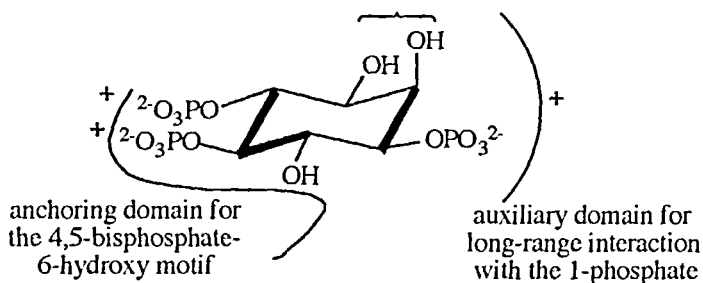


Figure 4. Ligand recognition at the Ins(1,4,5)P₃-binding site.

Reference

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