418 Communications Synthesis

have found by <sup>1</sup>H-NMR analysis that chlorotrimethylsilane in pyridine introduces only one silyl group on the hydroxylamine molecule.

In our one-pot procedure for the synthesis of N-substituted N-hydroxycarboxamides, N-substituted hydroxylamine 1 dissolved in dry pyridine is treated with a six-fold excess of chlorotrimethylsilane. N-Substituted O-trimethylsilylhydroxylamine 2 is directly acylated without isolation with mixed anhydride 3 (formed from an N-proteced amino acid or peptide and isobutylcarbonochloridate in the presence of N-methylmorpholine). Compounds 4, which are thus formed, are not isolated; the trimethylsilyl protecting group was immediately removed in the work-up procedure. The total yield of pure N-substituted N-hydroxycarboxamide 5, after all steps, was satisfactory to good (60-86%).

## A New Synthesis of N-Hydroxyamides Using Trimethylsilyl Protection

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The concept of transient hydroxylamine oxygen protection for the unambiguous synthesis of N-hydroxyamides has been applied in the amino acid field. First, hydroxylamines 1 were silylated in pyridine with chlorotrimethylsilane; then 2 was immediately N-acylated with mixed anhydride 3 of a protected amino acid or peptide; and finally, the O-trimethylsilyl protection was removed during the isolation procedure giving pure N-hydroxyamides 5.

Hydroxylamines are ambident reagents. In spite of this well known fact, simple condensing agents have been applied in the synthesis of N-hydroxycarboxamides (hydroxamic acids) and it is not surprising that the yields of hydroxamic acids were low. 1.2 Even with procedures with conditions specially chosen to favor the N-acylation process, as, for example, our method using N,N-dimethylchloromethaniminium chloride, 3 some undesirable O-acyl derivatives of hydroxylamine are formed in a few cases. We therefore concluded that a transient oxygen protection of the hydroxylamine function would be advantageous for an unambiguous, effective synthesis of N-hydroxycarboxamides. We chose the trimethylsilyl group since it is easily introduced and removed, and is stable under acylation conditions.

The idea of protection of the hydroxyl group by silylation is well documented; examples include that of serine for peptide synthesis, as well as sugar hydroxyl protection in nucleoside synthesis. Mono N-, O- and N,O-di trimethylsilyl derivatives of hydroxylamines have been described in the literature. 4.5.6 We

Yield, physical, and spectral data for the *N*-alkyl-*N*-hydroxy-carboxamides 5 obtained by this method, are reported in Tables 1 and 2.

Our experiments have shown, that the structure of the N-substituted-O-trimethylsilylhydroxylamine 2 does not have a significant effect on the yield of the synthesized N-hydroxy-carboxamides 5 (cf. 5a, b, c in Table 1). However, the synthesis of N-hydroxycarboxamides 5 with another tertiary amine (triethylamine) present during the generation of the mixed anhydride 3 results in smaller yields of products 5 (cf. 5a, b, c in Table 1 – yields in brackets), because N-hydroxyurea derivatives were formed, a result that has been confirmed by other research groups. 10.11

Silylated *N*-hydroxyamides have been obtained earlier by the reaction of *N*-hydroxyamides with silylating reagents<sup>12,13,14</sup> or in the reaction of acyl chlorides with *N*,*O*-bis(silyl-) or *N*,*N*,*O*-tris(silyl)hydroxylamine.<sup>12,15,16</sup> However, employment of silylated *N*-hydroxyamides for the synthesis of *N*-hydroxyamides, has until now been limited to the use of isolated *N*,*N*,*O*-tris(trimethylsilyl)hydroxylamine.<sup>15</sup>

We intend to extend our method to other acylations of N-substituted-O-trimethylsilylhydroxylamines 2. It will be applied to the synthesis of siderophores and their analogues, as well as to other ligands, forming complexes with other metals.<sup>17</sup>

All melting points are uncorrected. The IR and <sup>1</sup>H-NMR spectra were recorded on Jena-Zeiss UR-10 and Varian EM-360A instruments, respectively.

## N-Alkyl-N-hydroxycarboxamides (N-Alkyl-hydroxamic Acids) 5; General Procedure:

Chlorotrimethylsilane (3 ml, 0.024 mol) is added dropwise to a stirred solution of N-alkylhydroxylamine (1; 0.004 mol) in dry pyridine (12 ml)

Table 1. N-Alkyl-N-hydroxycarboxamides (Hydroxamic Acids) 5 Prepared

Product		Yield <sup>a</sup> (%)	m.p. (°C) (solvent)	Molecular Formula <sup>b</sup> or Lit. m.p. (°C)
5a	Z-L-Ala-N(OH)C <sub>2</sub> H <sub>5</sub>	85 (84) <sup>d</sup>	92 (CHCl <sub>3</sub> /hexane)	90-92 <sup>3</sup>
5b	Z-L-Ala-N(OH)CH(CH <sub>3</sub> )C <sub>6</sub> H <sub>5</sub>	85°(67) <sup>d</sup>	126–128 (ethyl acetate/hexane)	126-128 <sup>3</sup>
5c	Z-1-Ala-N(OH)C <sub>6</sub> H <sub>11</sub> -c	86°(62) <sup>d</sup>	104-105 (ethyl acetate/hexane)	$C_{17}H_{24}N_2O_4$ (320.3)
5d	BOC-Gly-N(OH)C <sub>6</sub> H <sub>11</sub> -c	86	85-86 (CHCl <sub>3</sub> /hexane)	$C_{13}H_{24}N_2O_4$ (272.3)
5e	Z-Gly-1-Ala-N(OH)C <sub>2</sub> H <sub>5</sub>	60	150-151 (ethyl acetate/hexane)	$C_{15}H_{21}N_3O_5$ (323.3)
5f	$\alpha$ -Z- $\epsilon$ -BOC-L-Lys-N(OH(C <sub>2</sub> H <sub>5</sub>	60	90-92 (ethyl acetate/hexane)	$C_{21}H_{33}N_3O_6$ (423.6)
5g	Z-DL-Phe-N(OH)C <sub>2</sub> H <sub>5</sub>	74	117-118 (ethyl acetate/hexane)	$C_{19}H_{22}N_2O_4$ (342.4)
5h	Z-L-Pro-N(OH)C <sub>2</sub> H <sub>5</sub>	66	87-89 (CHCl <sub>3</sub> /hexane)	$C_{15}H_{20}N_2O_4$ (292.3)
5i	OH OH OH C2H5	65	92-93 (ethyl acetate/hexane)	$C_{12}H_{24}N_2O_4$ (260.3)
5j	0 C <sub>6</sub> H <sub>5</sub>	80	7678 (CHCl <sub>3</sub> /hexane)	C <sub>23</sub> H <sub>23</sub> NO <sub>4</sub> (377.4)

<sup>&</sup>lt;sup>a</sup> Yield of crystallized product 5 based on 3.

<sup>b</sup> Satisfactory microanalyses obtained:  $C \pm 0.30$ ,  $H \pm 0.32$ ,  $N \pm 0.31$ .

Table 2. Spectral Data for N-Alkyl-N-hydroxycarboxamides 5

Table 2	Table 2. (Continue			
-				
Com-	IR	(KBr)		

	- IR (KBr) ad v (cm <sup>-1</sup> ) -CO-N(OH)-	<sup>1</sup> H-NMR (CDCl <sub>3</sub> /TMS) δ (ppm)
5a	1645	1.0-1.6 (m, 6H, 2CH <sub>3</sub> ); 3.7 (q, 2H, $J$ = 7 Hz, CH <sub>2</sub> N); 4.8 (m, 1H, CH); 5.0 (s, 2H, CH <sub>2</sub> C <sub>6</sub> H <sub>5</sub> ); 5.9 (m, 1H, NH); 7.3 (s, 5H,
5b	1610	$C_6H_5$ ); 8.9 (br s, 1H, OH) 1.35 (d, 3H, $J = 7$ Hz, $C_6H_5CHCH_3$ ); 1.65 (d, 3H, $J = 7$ Hz, $H_3CCHNH$ ); 5.0 (m, 1H, CH <sub>3</sub> CHNH); 5.25 (s, 2H, $L_2C_6H_5$ ); 5.8-
5e	1600	6.2 (m, 2H, $C_6H_5CHCH_3$ and NH); 7.5 (s, 10H, $2C_6H_5$ ); 9.0 (br s, 1H, OH) 1.3 (d, 3H, $J = 8$ Hz, $CH_3$ ); 1.6 (br s, 10H, $C_6H_{10+1}$ ); 4.2 (m, 1H, $C_6H_{10+1}$ ); 4.7 (m, 1H, $CHCH_3$ ); 5.0 (s, 2H, $CH_2C_6H_5$ ); 5.8 (m,
5d	1640	1H, NH); 7.2 (s, 5H, $C_6H_5$ ); 8.5 (s, 1H, OH) 1.37 [s, 9H, (CH <sub>3</sub> ) <sub>3</sub> ]; 1.6 (br s, 10H, $C_6H_{10+1}$ ); 3.9–4.33 (m, 3H, CH <sub>2</sub> and $C_6H_{10+1}$ ); 5.6 (m, 1H, NH); 8.77 (m, 1H,
5e	1620	OH)  1.2 (2t, 6H, $J = 6$ Hz, 2CH <sub>3</sub> ); 3.6 (q, 2H, $J = 7$ Hz, CH <sub>2</sub> N); 3.8 (d, 2H, $J = 6$ Hz, CH <sub>2</sub> CO); 4.8 (m, 1H, CHCH <sub>3</sub> ); 5.03 (s, 2H,
5f	1590	CH <sub>2</sub> C <sub>6</sub> H <sub>5</sub> ); 5.9 (br s, 1H, NH): 7.23 (s, 5H, C <sub>6</sub> H <sub>5</sub> ); 8.5 (m, 1H, OH) 1.1 (t, 3H, $J = 7$ Hz, CH <sub>3</sub> ); 1.37 [s, 15H, (CH <sub>3</sub> ) <sub>3</sub> and (CH <sub>2</sub> ) <sub>3</sub> ]; 3.0 (m, 2H, CH <sub>2</sub> NH); 3.6 [q, 2H, $J = 7$ Hz, CH <sub>2</sub> N(OH)]; 4.73 (m, 1H, CHNH); 4.95 (s, 2H, CH <sub>2</sub> C <sub>6</sub> H <sub>5</sub> ); 5.77
5 g	1620	(m, 2H, 2NH); 7.2 (s, 5H, $C_6H_5$ ); 8.93 (br s, 1H, OH) 1.0 (t, 3H, $J = 7$ Hz, CH <sub>3</sub> ); 3.0 (d, 2H, $J = 7$ Hz, CH <sub>2</sub> CH); 3.5 (q, 2H, $J = 7$ Hz, CH <sub>2</sub> N); 4.9 (s, 2H, CH <sub>2</sub> C <sub>6</sub> H <sub>5</sub> ); 5.1 (m, 1H, CHCO); 5.8 (m, 1H, NH); 7.1, 7.2 (2s, 10H, 2C <sub>6</sub> H <sub>5</sub> ); 8.4 (br s, 1H, OH)

	CO - N(OH) -	
5h	1635	1.1 (t, 3H, $J = 7$ Hz, CH <sub>3</sub> ); 2.0 [m, 4H, (CH <sub>2</sub> ) <sub>2</sub> ]; 3.53 (m, 4H, 2CH <sub>2</sub> N); 4.87 (m, 1H, CH); 5.07 (s, 2H, CH <sub>2</sub> O); 7.3 (s, 5H, C <sub>6</sub> H <sub>8</sub> ); 9.5 (m, 1H, OH)
5i i	1610, 1640	1.0–1.7 [m, 14H, 2CH <sub>3</sub> and (CH <sub>2</sub> ) <sub>4</sub> ]; 2.3 (m, 4H, 2CH <sub>2</sub> CO). 3.55 (q, 4H, $J = 7$ Hz, 2CH <sub>3</sub> N); 8.7 (m, 2H, 2OH)
<b>5</b> j 1	1600	1.1 (t, 3H, $J = 7$ Hz, CH <sub>3</sub> ); 3.5 (m, 2H, CH <sub>2</sub> N); 5.05, 5.15 (2s, 4H, 2CH <sub>2</sub> O); 7.1 (m, 3H, C <sub>6</sub> H <sub>3</sub> ); 7.3, 7.4 (2s, 10H, 2C <sub>6</sub> H <sub>5</sub> ); 8.5 (m, 1H, OH)

<sup>1</sup>H-NMR (CDCl<sub>3</sub>/TMS)

cooled to -10°C. After 15 minutes at room temperature the reaction mixture is again cooled to  $-10^{\circ}$ C. Then the mixed anhydride [3, 0.002 mol); also cooled to  $-10^{\circ}$ C; synthesized from N-protected amino acid or peptide (0.0021 mol) dissolved in dry dichloromethane (6 ml), N-methylmorpholine (0.22 ml, 0.002 mol) and isobutyl chloroformate (0.27 ml, 0.002 mol)] is added. The solution is stirred for several hours at room temperature. Then pyridine and dichloromethane are evaporated at reduced pressure. The residue is acidified with 2 normal hydrochloric acid (to pH 2) and extracted with ethyl acetate ( $2 \times 10$  ml). The ethyl acetate layer is washed with 0.5 normal hydrochloric acid (10 ml), water (20 ml), 3 % sodium hydrogen carbonate solution (10 ml) and dried with magnesium sulfate. After filtration and evaporation one obtains hydroxamic acids 5, which are purified by recrystallization (Tables 1 and 2). The purity of the prepared hydroxamic acids is checked by IR and <sup>1</sup>H-NMR spectroscopy and by TLC (silica gel, 2propanol/hexane, 1:9, and hexane/ethyl acetate/ethanol, 6:2:1).

<sup>&</sup>lt;sup>c</sup> After crystallization of compound 5, i-C<sub>4</sub>H<sub>9</sub>OCON(OH)CH(CH<sub>3</sub>)C<sub>6</sub>H<sub>5</sub> and i-C<sub>4</sub>H<sub>9</sub>OCON(OH)C<sub>6</sub>H<sub>11</sub>-c were found (TLC) in the mother liquor.

<sup>&</sup>lt;sup>d</sup> The yields of compound 5, obtained in the presence of triethylamine are given in parentheses.  $i-C_4H_9OCON(OH)C_2H_5$  (traces).  $i-C_4H_9OCON(OH)CH(CH_3)C_6H_5$  and  $i-C_4H_9OCON(OH)C_6H_{11}$ -c were found (TLC) in the mother liquor.

420 Communications Synthesis

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