

## 3'-Deoxy-3'-fluoro-xylo-Neplanocin A

Alan D Borthwick and Keith Biggadike

Department of Medicinal Chemistry,  
Glaxo Group Research, Greenford, Middlesex, UB6 0HE, UK.

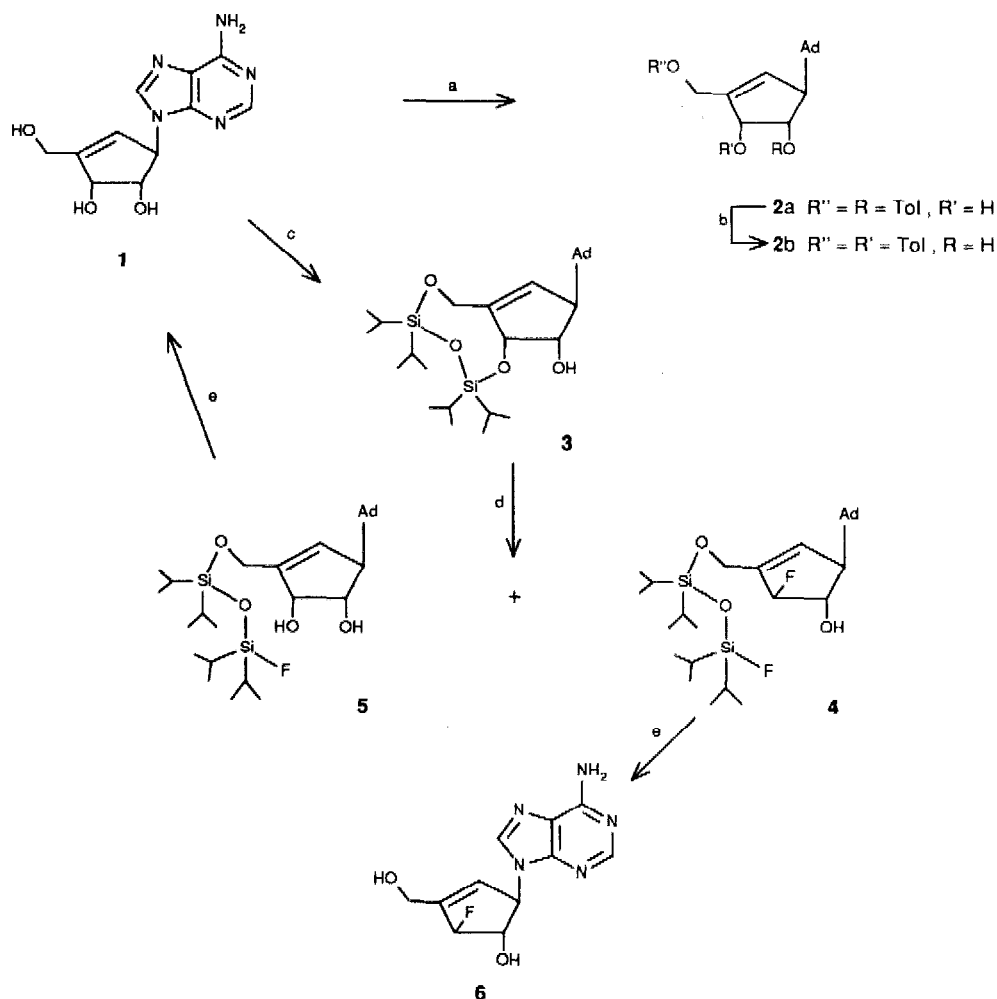
**Abstract:** Neplanocin A has been efficiently converted in three steps into its 3'-deoxy-3'-fluoro-xylo-analogue.

Fluorine substitution in nucleosides has a pronounced effect on their biological activity. The position and configuration of the fluorine substitution is crucial for the enhancement of their activity against Herpes and Retroviruses<sup>1</sup>. In the carbocyclic series<sup>2</sup>, we have shown recently that 2' $\beta$ <sup>3,4</sup>, 4' $\alpha$ <sup>5</sup> and 6' $\alpha$ <sup>3</sup> fluoro substitution of purine derivatives has a marked effect on increasing their antiviral activity. The unsaturated carbocyclic nucleoside neplanocin A **1** and its analogues<sup>6</sup> display antiviral and antitumour activity but, surprisingly, fluoro substitution in the cyclopentene ring has not been described. Having successfully<sup>4</sup> introduced a 2'-*ara*-fluoro substituent into aristeromycin we turned our attention to fluorination of neplanocin A.

Treatment of neplanocin A **1** with toluoyl chloride (2 equiv.) in pyridine gave a quantitative yield of the 2',5'-ditoluate **2a**. In the presence of SiO<sub>2</sub> this gave a mixture of the 2',5'-ditoluate **2a** and 3',5'-ditoluate **2b**, from which **2b** crystallised in 23% yield (m.p.118°C)(Scheme 1). However, treatment of the 3',5'-ditoluate **2b** with diethylaminosulphur trifluoride (DAST) (2 equiv., R.T., 1.5h) failed to give the required fluoro substitution and gave instead an equilibrium mixture of **2a** and **2b**. Because of the lability of the ester protecting groups in **2b**, we turned our attention to the TIPS protecting group which we had previously shown<sup>4,7</sup> was compatible with DAST.

Reaction of the 3',5'-*O*-TIPS derivative **3** of neplanocin A with DAST (2.4 equiv., 5°C, 1.5h) in methylene chloride gave a 1:1 mixture of the 3'-fluoro derivative **4**<sup>8</sup> and the diol **5**<sup>9</sup> in 38% yield. However, slow addition of **3** to a stirred solution of 3 equiv. of DAST/pyridine (1:1) in methylene chloride at 0°C during 45 mins followed by a further 15 mins at this temperature gave **4** in 65% yield together with a only a small quantity (5%) of the diol **5**. Deprotection of **4** with TBAF (R.T. 15min) gave in a quantitative yield 3'-deoxy-3'-fluoro-xylo-neplanocin A **6** [ $\alpha$ ]<sub>D</sub><sup>22</sup> -62° (c 0.5, MeOH), m.p. 225-227°C. The position and configuration of the fluorine atom in **6** was indicated by NMR<sup>10</sup> and was confirmed by X-Ray crystallography<sup>11</sup>. A similar deprotection of the diol **5** regenerated neplanocin A **1** in quantitative yield.

Scheme 1

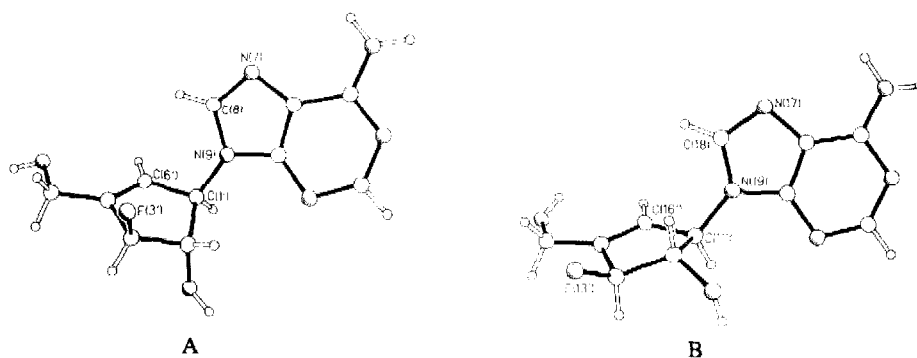


Reagents: (a) pMePhCOCl, py, RT, 1h 20min (b) SiO<sub>2</sub> (c) TIPS Cl, imidazole, DMF, RT, 1h (d) DAST/py, 0°C (e) TBAF

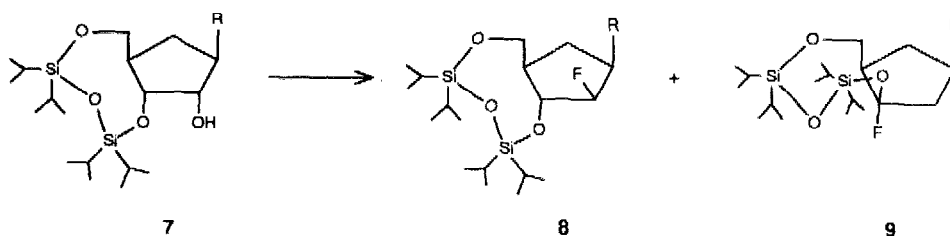
Interestingly, a single crystal X-Ray analysis of **6** reveals 2 crystallographically independent molecules A and B (Figure 1). Both have the adenine base in the usual anti-conformation. For molecule A the torsion angle about the N(9)-C(1') bond  $\phi[C(8)-N(9)-C(1')-C(6')]$  is +36.8° while the corresponding torsion angle for B is -12.5°. However, the pucker of the cyclopentene ring is quite different; while A has a C-2'exo,C-3'endo conformation as seen in neplanocin A **1**<sup>12</sup> and 2'-*ara*-mercapto-2'-deoxyneplanocin A<sup>13</sup>, the conformation of the cyclopentene ring in B is C-2'-endo,C-3'-exo.

We have previously shown<sup>4,7</sup> that the DAST reaction of a 2'-secondary hydroxyl adjacent to the TIPS

Figure 1



protecting group in a cyclopentane ring **7** gives a mixture of two products, the 2'-fluoride **8** with inversion and the 3'-fluoride **9** with retention via hydride transfer. The ratio of **8** to **9** depends on the substituent at the 1'-position.



However in the cyclopentene ring **3** (Scheme 1), the fluoride ion attacks the allylic 3'-position with inversion. A similar substitution in the DAST reaction on a 3'-*O*-*t*-butyldimethylsilyl derivative of adenosine has been reported by Herdewijn et al.<sup>14</sup>. Further studies are under way aimed at elucidating the mechanism of this reaction.

The recent interest in 3'-fluoro carbocyclic nucleosides as antiviral<sup>15</sup> and antitumour agents<sup>16</sup> together with the recent synthetic transformations<sup>2</sup> of carbocyclic adenine nucleosides into their imidazole, 8-*aza*-guanine, guanine and inosine analogues makes this efficient introduction of fluorine into the carbocyclic ring of neplanocin A a potentially useful transformation.

## References

1. Bergstrom, D. E.; Swartling, D. J. Fluorine Substituted Analogues of Nucleic Acid Components. In *Fluorine Containing Molecules*; Liebman, J. F.; Greenburg, A.; Dolbier, W. R. Eds.; VCH Publishers, New York, 1988, 259.
2. For a recent review see 'Synthesis of Chiral Carbocyclic Nucleosides' Borthwick, A. D.; Biggadike, K. *Tetrahedron* **1992**, *48*, 571.
3. Borthwick, A. D.; Kirk, B. E.; Biggadike, K.; Exall, A. M.; Butt, S.; Roberts, S. M.; Knight, D. J.; Coates, J. A. V.; Ryan, D. M. *J. Med. Chem.* **1991**, *34*, 907.

4. Biggadike, K.; Borthwick, A. D.; Exall, A. M.; Kirk, B. E.; Ward, R. A. *J. Chem. Soc., Chem. Commun.* **1988**, 899.
5. Biggadike, K.; Borthwick, A. D. *J. Chem. Soc., Chem. Comm.* **1990**, 1380.
6. (a) Marquez, V. E.; Lim, M.-I. *Med. Res. Rev.* **1986**, 6, 1. (b) De Clercq, E. *Biochem. Pharmacol.* **1987**, 36, 2567.
7. (a) Biggadike, K.; Borthwick, A. D.; Evans, D.; Exall, A. M.; Kirk, B. E.; Roberts, S. M.; Stephenson, L.; Youds, P. *J. Chem. Soc., Chem. Comm.* **1987**, 253. (b) Biggadike, K.; Borthwick, A. D.; Evans, D.; Exall, A. M.; Kirk, B. E.; Roberts, S. M.; Stephenson, L.; Youds, P. *J. Chem. Soc., Perkin Trans. 1* **1988**, 549.
8. 4: mp 104–105°C (MeOH);  $^1\text{H}$  NMR( $\text{CDCl}_3$ )  $\delta$  0.96–1.14 (28H, bs, 4  $\text{CHMe}_3$ ), 4.44–4.66 (3H, m, 2H-5', H-2'), 5.20 (1H, bs, H-1'), 5.51 (1H, dd,  $J_{\text{HF}}=56$  Hz, H-3'), 5.70 (2H, s,  $\text{NH}_2$ , exchange  $\text{D}_2\text{O}$ ), 5.82 (1H, s, OH, exchange  $\text{D}_2\text{O}$ ), 6.23 (1H, s, H-6'), 7.82 (1H, s, H-2), 8.34 (1H, s, H-8); MS(CI) (isobutane)  $m/z$ : 528 ( $\text{MH}^+$ ), 508 ( $\text{MH}^+ - \text{HF}$ ), 136 ( $\text{C}_5\text{H}_5\text{H}_4$ ); Anal. calcd. for  $\text{C}_{23}\text{H}_{39}\text{F}_2\text{N}_5\text{O}_3\text{Si}_2$ : C, 52.34; H, 7.45; N, 13.27; F, 7.20. Found: C, 52.25; H, 7.52; N, 13.02; F, 6.90.
9. 5: mp 179–181°C (MeOH);  $^1\text{H}$  NMR( $\text{Me}_2\text{SO}-d_6$ )  $\delta$  0.94–1.04 (28H, bs, 4  $\text{CHMe}_3$ ), 4.36 (1H, dd,  $J=5$ , 6.25 Hz, H-2'), 4.42–4.50 (3H, bs, H-3', 2H-5'), 5.02 (1H, d,  $J=6.25$  Hz, OH exchange  $\text{D}_2\text{O}$ ), 5.22 (1H, d,  $J=7.25$  Hz, OH exchange  $\text{D}_2\text{O}$ ), 5.37 (1H, bs, H-1'), 5.73 (1H, d,  $J=2.5$  Hz H-6'), 7.22 (2H, bs,  $\text{NH}_2$  exchange  $\text{D}_2\text{O}$ ), 8.04 (1H, s, H-2), 8.12 (1H, s, H-8); MS(CI) (isobutane)  $m/z$ : 526 ( $\text{MH}^+$ ), 506 ( $\text{MH}^+ - \text{HF}$ ), 136 ( $\text{C}_5\text{H}_5\text{H}_4$ ); HRMS(-FAB) calcd. for  $\text{C}_{23}\text{H}_{39}\text{N}_5\text{O}_4\text{Si}_2\text{F}$  [M-H] $^-$  524.2524, found 524.2506.
10. 6:  $^1\text{H}$  NMR ( $\text{Me}_2\text{SO}-d_6$ )  $\delta$  4.14 (2H, bs, 2H-5'), 4.69 (1H, dm,  $J_{\text{HF}}=24$  Hz, H-2'), 5.11 (1H, t,  $J=5.6$  Hz, 5'OH exchange  $\text{D}_2\text{O}$ ), 5.19 (1H, bs, H-1'), 5.38 (1H, dd,  $J=3.75$  Hz,  $J_{\text{HF}}=56$  Hz, H-3'), 5.85 (1H, bs, H-6'), 6.05 (1H, d,  $J=6.25$  Hz, 2'OH exchange  $\text{D}_2\text{O}$ ), 7.26 (2H, s  $\text{NH}_2$ ), 8.12, 8.14 (2H, 2s, H-8, H-2).  $^{13}\text{C}$  NMR ( $\text{Me}_2\text{SO}-d_6$ ) 157.4 (C-6), 153.8 (C-2), 150.9 (C-4), 146.4 ( $J_{\text{CF}}=20$  Hz, C-4'), 140.9 (C-8), 127.3 ( $J_{\text{CF}}=7$  Hz, C-6'), 120.5 (C-5), 100.9 ( $J_{\text{CF}}=172$  Hz, C-3'), 84.0 ( $J_{\text{CF}}=20$  Hz, C-2'), 63.8 ( $J_{\text{CF}}=6$  Hz, C-1'), 58.5 (C-5'); Anal. calcd. for  $\text{C}_{11}\text{H}_{12}\text{FN}_5\text{O}_2$ : C, 49.81; H, 4.56; N, 26.41; F, 7.12. Found: C, 49.66; H, 4.70; N, 26.54; F, 6.90.
11. *Crystal data*: **6**,  $\text{C}_{11}\text{H}_{12}\text{FN}_5\text{O}_2$ ,  $M = 265.25$ , monoclinic,  $a = 7.060(1)$ ,  $b = 12.955(3)$ ,  $c = 13.250(5)$  Å,  $\beta = 103.85(2)^\circ$ ,  $V = 1177(1)$  Å $^3$ ,  $\lambda = 1.54184$  Å, space-group  $P2_1$  (No. 4),  $Z = 4$ ,  $D_C = 1.50\text{ g cm}^{-3}$ ,  $F(000) = 552$ ,  $\mu(\text{Cu-K}\alpha) = 9.7\text{ cm}^{-1}$ ; crystal size:  $0.44 \times 0.32 \times 0.14$  mm. Nicolet R3m/V diffractometer, 1614 reflections measured ( $0 < 2\theta < 115^\circ$ ) of which, 1505 reflections had  $I > 3.0 \sigma(I)$ . Full matrix least-squares refinement with anisotropic thermal parameters was used for all non-hydrogen atoms. Hydrogen atoms were refined in riding mode. Individual weights were applied according to the scheme  $w = [\sigma^2(F_o) + 0.0038 |F_o|^2]^{-1}$  refinement converged at  $R$  0.039,  $R_w$  0.043, goodness-of-fit = 1.01. All computations were carried out using the SHELXTL PLUS ( $\mu$ -VAX II) system of programs (G.M. Sheldrick, SHELXTL release 3.4. Copyright 1988 Nicolet Instrument Corporation). Atomic parameters, bond lengths, and angles have been deposited at the Cambridge Data Centre.
12. Hayashi, M.; Yaginuma, S.; Yoshioka, H.; Nakatsu, K. *J. Antibiot.* **1981**, 34, 359.
13. Kinoshita, K.; Hayashi, M.; Hirano, T.; Nakatsu, K.; Fukukawa, K.; Ueda, T. *Nucleosides Nucleotides* **1983**, 2, 319.
14. Herdewijn, P.; Aerscoht, A. V.; Kerrimans, L. *Nucleosides Nucleotides* **1989**, 8, 65.
15. Koshida, R.; Cox, S.; Harmenberg, J.; Gilljam, G.; Wahren, B. *Antimicrob. Agents Chemother.* **1989**, 33, 2083.
16. Nakayama, T.; Morizawa, Y.; Matsumura, Y.; Yasuda, A.; Uchida, K. *Nucleic Acids Res. Symp. Ser.* **1989**, 21, 73.

(Received in UK 25 March 1992)