

A PREGNANE ESTER GLYCOSIDE FROM *PERIPLOCA CALOPHYLLA*

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Abstract—A new pregnane ester genin, plocigenin, and a new pregnane ester diglycoside, plocin, have been isolated from the dried twigs of *Periploca calophylla*. The chemical and spectroscopic properties are consistent with the structures 12,20-di-*O*-benzoyl drevogenin-D and 12,20-di-*O*-benzoyl drevogenin-D-3-*O*- β -D-oleandropyranosyl (1 \rightarrow 4)-*O*- β -D-oleandropyranoside, respectively.

INTRODUCTION

The members of the Asclepiadaceae are known to be rich in cardenolide and pregnane derivatives [1, 2]. The twigs of *Periploca calophylla* L. contain a 2-deoxyhexose, cymarose, a cardenolide, periplogenin [3], and a pregnane glycoside, calocin [4]. In a re-investigation of their chemical constituents, the shade-dried twigs of the plant were extracted using an earlier method [5, 6] to give a novel glycoside which we have named plocin (1). This paper describes the structure elucidation of this glycoside.

RESULTS AND DISCUSSION

Plocin (1), $C_{49}H_{66}O_{13}$, mp 148–150°, $[\alpha]_D + 40^\circ$. A positive Liebermann–Burchardt [7] colour reaction and characteristic colour tests for 2-deoxy sugars in the xanthidrol [8, 9] and Keller–Kiliani [10] reactions indicated it to be a steroidal 2-deoxyglycoside.

Mild acid hydrolysis of 1 with 0.025 M sulphuric acid afforded a new genin, plocigenin (2) and a chromatographically pure reducing sugar (5). Comparison of the optical rotation value and co-chromatography (PC) with an authentic sample identified the sugar as D-oleandrose [11, 12] (2,6-dideoxy-3-*O*-methyl-D-arabino-hexose). Bromine–water oxidation of 5 gave a lactone (6), which with phenyl hydrazine afforded the known D-oleandronic acid phenyl hydrazide [11, 12].

The difference, $C_{14}H_{24}O_6$, between the molecular formulae of plocin (1) ($C_{49}H_{66}O_{13}$) and plocigenin (2) ($C_{35}H_{42}O_7$) clearly indicated that plocin contained two oleandrose units and was in fact a di-oleandroside. For convenience, the two D-oleandrose units of 1 were designated as S_1 and S_2 .

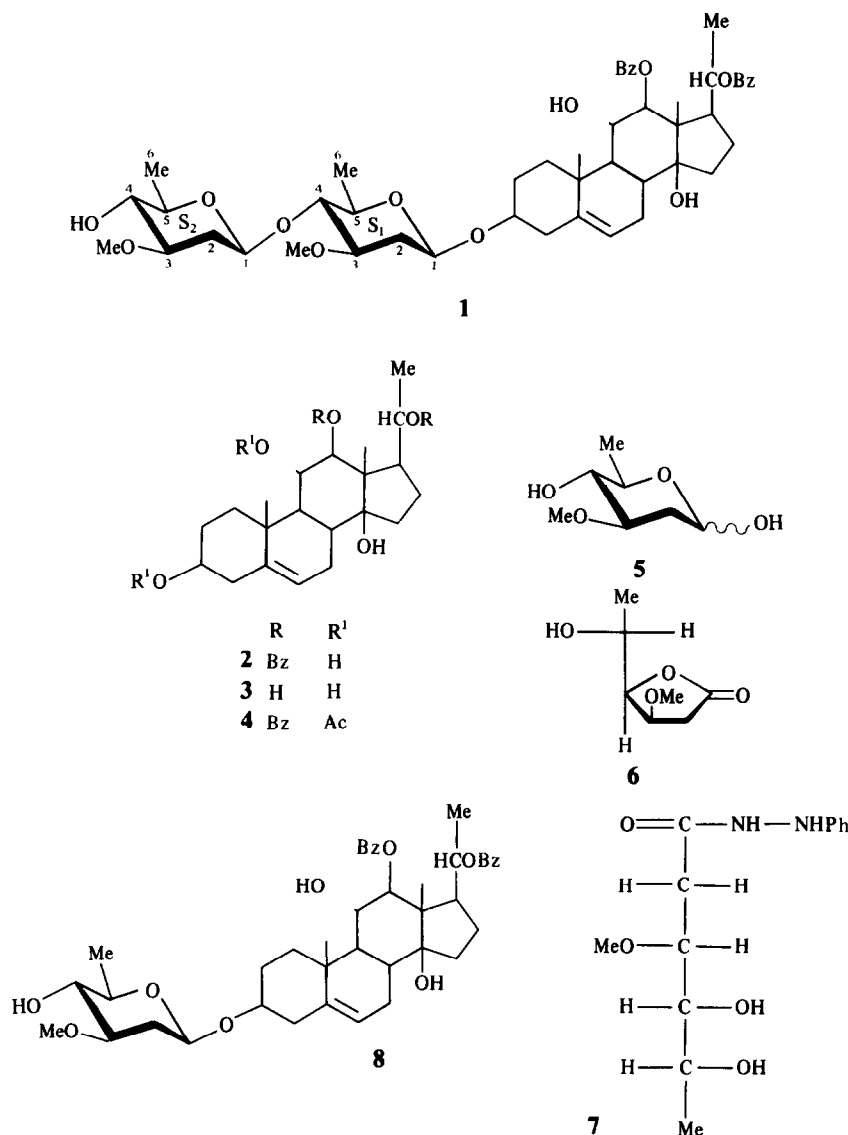
Although the genin 2 was inert to sodium periodate, its deacylated product (3), $C_{21}H_{34}O_5$, reacted with this reagent which suggested a vicinal diol arrangement in the molecule. This pregnane genin (C_{21}) was characterized as drevogenin-D [13] ($3\beta,11\alpha,12\beta,14\beta,20R$ -pentahydroxy- Δ^5 -pregnane) by co-chromatography (TLC) and mmp with an authentic sample of 2. The ability of plocin (1) and its aglycone (2) to undergo alkaline hydrolysis indicated the presence of ester functions. The difference ($C_{14}H_{24}O_6$) between the formulae of aglycone 2 and its deacylated product 3, supplemented by the UV absorption maximum

at 282 nm ($\log \epsilon$ 3.25), indicated the presence of two benzoyl groups, suggesting that the aglycone was a dibenzoyl drevogenin-D. As both glycoside 1 and its aglycone, di-*O*-benzoyl drevogenin-D (2), were inert to sodium periodate, it was inferred that one of the vicinal hydroxyl groups was benzoylated at C-11 or C-12 whereas the other benzoyl group could be at any of the remaining three secondary carbinol groups (C-3, C-12 or C-11, C-20) of drevogenin-D.

In the 1H NMR ($CDCl_3$) spectrum of 1, the chemical shift of a one-proton sharp doublet at δ 4.96 ($J = 8$ Hz) and another one-proton multiplet at δ 4.8–4.6 in the lower field indicated these methine protons to carry ester functions, which were assigned to C-12 and C-20 methine protons, respectively. Plocigenin was thus characterized as 12,20-di-*O*-benzoyl drevogenin-D and is reported for the first time from a natural source. Confirmation of this conclusion was forthcoming from the acetylation of 2 with acetic anhydride in pyridine which afforded an amorphous product, 4, $[\alpha]_D - 21^\circ$, which contained the expected two acetyl groups as three-proton singlets at δ 1.97 and 1.93 in its 80 MHz 1H NMR spectrum.

For further confirmation of 1 being a diglycoside, it was subjected to very mild acid hydrolysis (2.5 mM sulphuric acid). After 3 days, the partially hydrolysed reaction mixture showed two new spots besides some unreacted starting material (PC). The more polar spot had the same mobility as oleandrose (5) whereas the other more mobile vanillin–perchloric acid positive spot (R_{Ole} 2.3) was presumed to be the monoglycoside 8. The hydrolysis was complete in 85 hr and gave two new spots which were identical with the genin 2 and oleandrose (5) on TLC and PC, confirming that 1 was the dioleandroside of 12,20-di-*O*-benzoyl drevogenin-D.

The 250 MHz 1H NMR ($CDCl_3$) spectrum of 1 was in full agreement with the derived structure. It contained ten proton signals in the aromatic region in two sets of multiplets at δ 7.98–7.9 (4H) and δ 7.58–7.20 (6H) for the two phenyl groups of the benzoates. It also contained two methoxyl group singlets at δ 3.42 and 3.40, two secondary methyl doublets at δ 1.32 ($J = 6$ Hz) and 1.26 ($J = 6$ Hz), and four protons of 2-deoxymethylene of both the sugar units S_1 and S_2 as two sets of multiplets in the region δ 2.52–2.22 and 1.98–1.88 for equatorial and axial protons,



respectively. Two double-doublets ($J = 8$ and 3 Hz) of one proton each at δ 4.84 and 4.52 were attributed to the anomeric protons of two oleandrose units in the molecule. The large coupling constant (8 Hz) of both these anomeric protons, typical of their axial configuration in a 2-deoxyhexopyranose moiety in 4C_1 (D) conformation, suggested that both sugars were linked in a β -D-glycosidic linkage [14]. The low-field chemical shift of a one-proton doublet ($J = 8$ Hz) at δ 4.96 and a one-proton multiplet centred at δ 4.7 was evidently due to the methine protons of esterified carbon atoms respectively in the pregnane moiety. The C-11 methine proton carrying the hydroxyl group appeared at a high field as a triplet ($J = 8$ Hz) centred at δ 3.34. In addition, the spectrum also contained other appropriate proton signals of drevogenin-D and sugar residues.

In the light of all this evidence, the structure of plocin (**1**) is established as 12,20-di-*O*-benzoyl drevogenin-D-3-*O*- β -D-oleandropyranosyl (1 \rightarrow 4)-*O*- β -D-oleandropyranoside.

EXPERIMENTAL

Mps uncorr. PC: toluene-BuOH (4:1) saturated with H_2O . Sugars were visualized with 50% aq. H_2SO_4 (TLC) or vanillin- $HClO_4$ reagent (PC). 1H NMR: $CDCl_3$, TMS as internal standard.

Extraction. Shade-dried twigs (5 kg) of *P. calophylla* were extracted and fractionated with solvents of different polarities as reported earlier [4] to afford a petrol extract (2.5 g), an Et_2O extract (1 g), a $CHCl_3$ extract (20 g), a $CHCl_3$ - $EtOH$ (4:1) extract (7 g) and a $CHCl_3$ - $EtOH$ (3:2) extract (5.2 g), respectively. The $CHCl_3$ - $EtOH$ (3:2) extract was chromatographed over silica gel to give plocin (**1**) (64 mg).

Plocin (1). Mp 148–150° (MeOH- Et_2O), $[\alpha]_D^{25} + 40.2^\circ$ (MeOH, c 0.11) (Found: C, 71.38, H, 6.82. $C_{49}H_{66}O_{13}$ requires C, 71.45, H, 6.87). It gave a pink colour in the xanthidrol and a blue colour in Keller-Kiliani reactions. UV λ_{max}^{EtOH} nm (log ϵ): 282 (3.25). 1H NMR (250 MHz): δ 7.98–7.9 (m, 4H, aromatic),

7.58–7.20 (m, 6H, aromatic), 5.36 (m, 1H, H-6), 4.96 (d, 1H, $J = 8$ Hz, H-12), 4.84 (dd, 1H, $J = 8$ and 3 Hz, H-1 in S_1 or S_2), 4.80–4.64 (m, 1H, H-20), 4.52 (dd, 1H, $J = 8$ and 3 Hz, H-1 in S_1 or S_2), 3.84–3.74 (m, 2H, H-5 in S_1 and S_2), 3.66–3.50 (m, 2H, H-3 in S_1 and S_2), 3.42 (s, 3H, OMe), 3.40 (s, 3H, OMe), 3.34 (t, 1H, $J = 8$ Hz, H-11), 3.24–3.14 (m, 2H, H-4 in S_1 and S_2), 2.52–2.22 (m, 2H, H-2_{eq} in S_1 and S_2), 1.99–1.88 (m, 2H, H-2_{ax} in S_1 and S_2), 1.66–1.5 (methylenes of aglycone), 1.32 (d, 3H, $J = 6$ Hz, 6-Me in S_1 or S_2), 1.29 (d, 3H, $J = 6$ Hz, 21-Me), 1.26 (d, 3H, $J = 6$ Hz, 6-Me in S_1 or S_2), 1.24 (s, 3H, 18-Me), 1.12 (s, 3H, 19-Me).

Mild acid hydrolysis of 1 To a soln of crystalline **1** (18 mg) in 80% aq 1,4-dioxane (1.3 ml) was added 0.05 M H_2SO_4 (1.2 ml) and the solution warmed for 30 min at 50°. Dioxane was then removed under reduced pressure. The aq portion was repeatedly extracted with $CHCl_3$ (97.3) and the organic layer was washed successively with H_2O , Na_2CO_3 and H_2O , dried over Na_2SO_4 and evapd to afford genin **2** (8 mg), crystallized from MeOH–petrol, mp 187–189°, $[\alpha]_D^{25} -18.6^\circ$ (MeOH, c 0.18) ($C_{35}H_{42}O_7$). The aq hydrolysate was neutralized with freshly prepared $BaCO_3$, filtered and concd under reduced pressure to afford the syrupy sugar **5** (6 mg), $[\alpha]_D^{25} -14.2^\circ$ (H_2O , c 0.12), which gave a pink coloration in the xanthidrol and a blue coloration in the Keller–Kiliani reactions. Comparison of its optical rotation and co-chromatography (PC) identified **5** as D-oleandrose.

Oxidation of 5 with Br_2-H_2O A soln of **5** (4 mg) in H_2O (0.6 ml) was mixed with Br_2 (10 μ l) and shaken in a stoppered flask in the dark for 24 hr at room temp. Excess Br_2 was then removed under reduced pressure. The acidic mixture was made neutral with freshly precipitated Ag_2CO_3 and the suspension filtered, H_2S was passed through the filtrate to remove Ag^+ ions, and the suspension filtered. The filtrate was evapd to dryness under reduced pressure yielding syrupy lactone **6** (3.2 mg).

D-Oleandronic acid phenyl hydrazide (7) A soln of lactone **6** (3 mg) in EtOH (0.05 μ l) was mixed with freshly distilled phenyl hydrazine (0.05 ml) and the mixture heated for 30 min at 100°. The viscous mass was cooled and repeatedly triturated with Et₂O (to remove excess phenyl hydrazine) yielding D-oleandronic acid phenyl hydrazide (2.2 mg), which crystallized from MeOH–Et₂O as colourless needles (1.2 mg), mp 132–134°. It had superimposable IR and gave no depression in mp when mixed with authentic material (lit mp 135°).

Very mild acid hydrolysis of 1 To a soln of **1** (15 mg) in 80% aq 1,4-dioxane (2.5 ml) was added 5 mM H_2SO_4 (2.5 ml) and the soln was kept at room temp. After 3 days, the reaction mixture showed two new vanillin–perchloric acid positive spots on PC besides a spot of some unreacted starting material. The more polar spot (R_{Ole} 1.0) was identified as oleandrose (**5**) by co-chromatography with an authentic sample, while the less polar spot (R_{Ole} 2.3) was presumably the monoglycoside **8**. The hydrolysis was complete in 85 hr (TLC) and work-up of the

hydrolysate followed by CC over silica gel afforded a viscous syrup (4.5 mg), $[\alpha]_D^{25} -14.6^\circ$, and a crystalline compound (6.8 mg), mp 186–188°. The crystalline compound was identical with the known 12,20-di-O-benzoyl drevogenin-D (**2**) (mmp, $[\alpha]_D$) and the viscous syrup was identified as D-oleandrose by comparison with an authentic sample ($[\alpha]_D$ and PC).

Alkaline hydrolysis of 2 Compound **2** (4 mg) was dissolved in 5% methanolic KOH (1 ml) and refluxed for 2 hr. After addition of H_2O (0.5 ml), the MeOH was removed under reduced pressure. The aq concentrate was extracted with $CHCl_3$ –MeOH (9:1), dried over Na_2SO_4 , filtered and evapd to dryness yielding **3** (1.8 mg), which was crystallized from Me₂CO–Et₂O, mp 225–228°, $[\alpha]_D^{25} -10.2^\circ$ (MeOH, c 0.12) ($C_{21}H_{34}O_5$). No depression in mmp, comparison of TLC, and optical rotation with authentic sample confirmed **3** as drevogenin-D.

Acetylation of 2 Compound **2** (6 mg) on acetylation with Ac₂O (0.4 ml) in pyridine at 100° for 8 hr afforded **4** as an amorphous residue (6 mg), $[\alpha]_D^{25} -21.5^\circ$ (MeOH, c 0.11). It showed ¹H NMR (80 MHz) signals at δ 1.97 (s, 3H, OAc), 1.93 (s, 3H, OAc).

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