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Synthesis of two branched heptopyranoside- (L,D-Hepp)-containing trisaccharides of the inner-core region of *Citrobacter PCM* 1487

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Abstract. The diastereogenic addition of (phenyldimethylsilyl)magnesium chloride to methyl 3-O-allyl-2,4-di-O-benzyl- α -D-manno-hexodialdo-1,5-pyranoside gave a L-glycero-D-manno-heptopyranoside (L,D-Hepp) derivative having at C-7 the phenyldimethylsilyl function as a hydroxy-masking group. The L,D-Hepp derivative was converted into two acceptors which could be applied successfully for the preparation of the trisaccharides L- α -D-Hepp-(1 \rightarrow 3)-O-[L- α -D-Hepp-(1 \rightarrow 7)]-L- α -D-Hepp-O-Me 20 and α -D-Glcp-(1 \rightarrow 3)-O-[L- α -D-Hepp-(1 \rightarrow 7)]-L-D-Hepp-O-Me 26.

Introduction

Lipopolysaccharides (LPS) are constituents² of the outer membrane of Gram-negative bacteria. They are composed of a lipid part, the Lipid A moiety, and a polysaccharide, comprising an oligomeric repeating unit (the O-specific chain) and a core oligosaccharide. The core connects the O-specific chain to lipid A which tethers the LPS into the cell envelope. Core oligosaccharides have a rather conserved composition among different Gram-negative bacteria and are characterised by the presence of the uncommon seven- and eight-carbon sugars L-glycero-D-manno-heptopyranoside (L,D-Hepp see for derivatives e.g. 4, 9) and 3-deoxy-D-manno-2-octulosonic acid (KDO), respectively. The immunoreactivity of core oligosaccharides³ has received special attention in relation to endotoxin neutralization by antibodies directed against the inner-core LPS region.

As part of an ongoing program directed toward the synthesis of immunogenic inner-core fragments, we have developed convenient methods for the preparation of these rare sugars L,D-Hep $p^{3,4}$ and KDO⁵.

With the objective of widening the scope of our synthetic

All = allyl Bn = benzyl DMF = N,N-dimethylformamide Glcp = glucopyranoside KDO = 3-deoxy-D-manno-2-octulosonic acid 1.,D-Hepp = 1.-glycero-D-manno-heptopyranoside LPS = lipopolysaccharides PDMSi = phenyldimethylsilyl (IUPAC order: dimethylphenylsilyl) THF = tetrahydrofuran TLC = thin-layer chromatography triflate = trifluoromethanesulfonate route to L,D-Hepp building units, we now report in detail⁶ the assembly of the trisaccharides **20** and **26** which have been proposed⁷ (Figure 1) to be part of the inner-core region of *Citrobacter* PCM 1487.



Figure 1. Alternative locations of the branched terminal heptose are marked by dashed lines.

Results and discussion

The availability of suitably protected L,D-Hepp donors and acceptors is a key element in the assembly of trisaccharides 20 and 26. Recently, we showed⁵ that L,D-Hepp derivatives are easily accessible by a two-step stereoselective hydroxymethylation method. In this approach, anti-Cram addition⁸ of (phenyldimethylsilyl)methylmagnesium chloride (1) to the fully benzylated methyl x-D-manno-hexodialdo-1,5-pyranoside 2 constitutes the key carbon-bond-forming step: the resulting α -hydroxy silane adduct 3 can be converted to homologous L,D-Hepp 4 by oxidative generation⁹ of the hydroxy group at C-7 from the silvl moiety. On the other hand, unmasking of the phenyldimethylsilyl (PDMSi) moiety can also be executed at an advanced stage of the synthesis. The latter option opens the way to introduce glycosidic bonds at C-6 or C-7 of the L,D-Hepp acceptors. The usefulness of the above cited stereoselective L,D-Hepp synthesis can be extended further by using a D-manno-

synthesis can be extended further by using a D-manno--hexodialdo-1,5-pyranoside carrying different protecting groups in the carbon-bond-forming step: additional protective-group manipulations of the resulting Grignard addition

Abbreviations:

product may give access to L.D-Hep*p* acceptors which allow glycosylation of the ring and external hydroxy groups.

For example, the L,D-Hepp derivative 6, which can be obtained by condensing 1 with known¹⁰ methyl 3-O-allyl--2,4-di-O-benzyl-x-D-manno-hexodialdo-1,5-pyranoside (5), gives access to the acceptors 8 and 9 which, in turn, may serve as L,D-Hepp building units in the preparation of trisaccharides 20 and 26, respectively. In addition, the L,D-Hepp derivative 4 can be readily converted into L,D-Hepp derivative 17, which proved to be a suitable donor in the synthetic routes to the target trisaccharides. Thus, addition of 1 in 1,3-dioxolane to 5 at 0° occurred with high diastereofacial selectivity to give 6 (d.p. 95°_{o}) in 72°_{o} yield. Benzylation of 6 according to *Czernecki* et al.¹¹ gave 7. Deallylation¹² of 7 with palladium on carbon (10°_{0}) in the presence of p-toluenesulfonic acid in a mixture of MeOH + H₂O at 70° for 8 h gave the glycosyl acceptor 8 in 70°_{0} yield. In this respect, it is interesting to note that removal of the allyl ether was accompanied by the formation of a small amount (6°_{o}) of the *Peterson* elimination¹³ product 10. Moreover, the residual sulfur-containing impurities, resulting from the Swern oxidation of methyl 3-O--allyl-2,4-di-O-benzyl-x-D-mannopyranoside to give 5, impeded the deallylation of 7. Moreover, an increase in the formation of the unwanted product 10 was observed.



The second glycosyl acceptor **9** was isolated, following the procedure of *Flemming*⁹, in a good yield by generating the hydroxy group from the PDMSi moiety in **8** with peroxyacetic acid and potassium bromide in AcOH + NaOAc with the exclusion of light. On the other hand, glycosyl donor **17** was synthesised in an overall yield of 65°_{o} starting from **11**, which was obtained⁴ in three steps from **4**. Acetolysis (HOAc + H₂SO₄) of **11** gave **12**, which was quantitatively converted (HBr + AcOH) into the known⁴ α -glycosyl bromide **13**. Transformation of **13** into the α -glycosyl chloride **17** was executed via intermediates **14**, **15** and **16**, according to *Paulsen* et al.¹⁴.

Having the key L,D-Hepp donors and acceptors in hand, we now turned our attention to the assembly of the target trisaccharides **20** and **26**. In the first instance, the trisaccharide 20 was obtained by glycosylation of C-3,7 hydroxyls in the L,D-Hepp acceptor 9 with L,D-Hepp donor 17, followed by removal of all protective groups (Scheme 1). Thus, condensation of acceptor 9 with excess donor 17 under the conditions described by *Hanessian* and *Banoub*¹⁵ in the absence of 1,1,3,3-tetramethylurea gave a 71°_{0} yield of trisaccharide 18.



Scheme 1

Zemplén deacylation of **18** led to the isolation of **19** in 95° o yield. Hydrogenolysis of **19** in the presence of 10° o Pd-C in EtOH gave the target methyl *O*-L-glycero- α -D-manno-heptopyranosyl-(1 \rightarrow 3)-*O*-[L-glycero- α -D-manno-heptopyranoside-(1-7)-L-glycero-D-manno-heptopyranoside **20** in 85° o yield. The structural integrity of **20** was fully established by homoand heteronuclear-correlation NMR spectroscopy. In the next stage, the more complex trisaccharide **26** was assembled as outlined in Scheme 2. Thus, α -stereospecific coupling of glucosyl chloride **21**¹⁶ with L,D-Hepp acceptor **8** could be accomplished using the promoter silver triflate¹⁵ to give, after purification, anomerically pure disaccharide **22** in 90° o yield indicating that the PDMSi masking group in acceptor **8** was compatible with the glycosylation conditions. In order to introduce the required glycosidic bond at

C-7, the PDMSi function in 22 was unmasked by oxidation⁹



to give 23. Silver triflate promoted coupling of acceptor 23 with glycosyl chloride donor 17 gave the fully protected trisaccharide 24 in a yield of 89°_{0} . Deblockling of 24 to give the fully deprotected trisaccharide 26 was executed by Zemplén deacylation of 24 and hydrogenolysis of 25 to yield homogeneous 26, the ¹H- and ¹³C-NMR data of which were in complete accordance with the proposed structure.

In conclusion, the steroselective addition of (dimethylphenylsilyl)methylmagnesium chloride to properly protected α -D-manno-hexodialdo-1,5-pyranosides gives easy access to valuable L,D-Hepp units suitable for oligosaccharide synthesis.

Experimental

General methods and materials

Dichloromethane, 1,2-dichloroethane and toluene were distilled from P_2O_5 and stored over molecular sieves 4 Å. Diethyl ether and THF were distilled from CaH₂ and redistilled from LiAlH₄ before use. DMF was stirred with CaH₂ for 16 h, then distilled under reduced pressure and stored over molecular sieves 4 Å. Methanol was dried by refluxing with magnesium methoxide, distilled and stored over molecular sieves 3 Å. TiBr₄ was sublimed under reduced pressure and stored as a standard solution in dichloromethane. Silver triflate was purchased from Fluka and dried, with the exclusion of light, under reduced pressure for 18 h before use. Reactions were performed under strict anhydrous conditions unless noted otherwise. Evaporation was carried out below 40°C under reduced pressure.

Column chromatography was performed using silica gel (Merck, 70–230 mesh). Gel filtration was performed on Sephadex LH-20 (Pharmacia). TLC analyses were conducted on Schleicher and Schüll DC fertigfolien F 1500 LS 254. Compounds were visualized by UV light (254 nm) or by spraying with concd. H_2SO_4 /methanol (1/10, v/v) and charring at 140°C.

Optical rotation was determined with a Perkin–Elmer Model 241 polarimeter at 20°C. NMR spectra were recorded with a Yeol JNM-FX200 (¹³C and ¹H at 200 and 80.7 MHz, respectively), or a Bruker WM-300 spectrometer equipped with an Aspect-2000 computer (¹H, 300 MHz). Chemical shifts are given in ppm (δ) relative to TMS as internal standard.

Methyl 3-O-allyl-2,4-di-O-benzyl-7-(phenyldimethylsilyl)-7-deoxy-L--glycero-x-D-manno-heptopyranoside (6)

Aldehyde 5 (4.54 g, 11 mmol) was dissolved in THF (45 ml) and the mixture was added dropwise to a cooled (0°C) solution of Grignard reagent 1 (25 mmol) in THF (25 ml). After stirring the reaction mixture for 2 h at 0°C, TLC analysis (acetone/CH₂Cl₂, 1/19, v/v) indicated complete conversion of 5 into 6 (R_c 0.8). The mixture was slowly poured into aqueous ammonium chloride (50 ml, 20%) and extracted with CH₂Cl₂ (200 ml). The organic layer was washed with water (50 ml), dried over MgSO₄ and concentrated under reduced pressure. The residual oil was applied onto a column of silica gel (45 g) and eluted with CH₂Cl₂ followed by CH₂Cl₂/acetone (99/ $1 \rightarrow 49/\overline{1}$, v/v). Concentration of the appropriate fractions gave **6** as an oil. Yield 3.6 g (58%), R_f (light petro-leum/ether, 2/1, v/v), $[\alpha]_{15} + 25.0^\circ$ (c 0.8, CHCl₃). ¹H NMR (CDCl₃): δ 7.60–7.25 (m, 15H, H-arom), 5.95 (m, 1H, CH=, All), $5.30 (m, 2H, = CH_2, All), 4.95-4.60 (m, 4H, CH_2, Bn), 4.74 (s, 1H, CH_2, B$ H-1), 4.12 (2H, OCH₂, All), 4.08-3.29 (5H, H-2, H-3, H-4, H-5, H-6), 3.28 (s, 3H, OCH₃), 1.40 (dd, 1H, H-7a), 0.9 (dd, 1H, H-7b), 0.41 (2 × s, 6H, CH₃Si). ¹³C NMR (CDCl₃): δ 138.4, 138.3 (Cq, arom), 134.9 (CH=, All), 133.7-133.6 (CH, arom), 116.5 (=CH₂, All), 99.5 (C-1), 79.9, 75.3, 72.8, 71.0 (C-2, C-3, C-4, C-5), 67.2 (C-6), 69.7 (CH₂O, All), 54.6 (OCH₃), 21.8 (C-7), -2.0, -2.2 (SiCH₃). Anal. calcd. for C₃₃H₄₂O₆Si: C 70.43, H 7.52; found: C 70.38, H 7.50°

Methyl 3-O-allyl-2,4,6-tri-O-benzyl-7-(phenyldimethylsilyl)-7-deoxy--t-glycero- α -D-manno-heptopyranoside (7)

Tetra-n-butylammonium iodide (0.36 g, 0.97 mmol), sodium hydride (0.35 g, 14.5 mmol) and benzyl bromide (1.7 ml, 14.5 mmol) were added to a cooled $(0^{\circ}C)$ solution of 6 (5.45 g, 9.7 mmol) in MF (30 ml). After stirring the reaction mixture for 3 h at 20°C, TLC analysis (light-petroleum/ether, 2/1, v/v) indicated complete conversion of 6 into 7 (R_f 0.7). The reaction mixture was quenched by the addition of methanol (5 ml) and the solvents were evaporated. The residue was partitioned between CH₂Cl₂ (150 ml) and water (2 \times 50 ml). The organic layer was dried over MgSO₄ and concentrated to an oil, which was applied onto a column of silica gel (100 g). Elution of the column was effected with lightpetroleum/ether $(9/1 \rightarrow 4/1, v/v)$. The appropriate fractions were concentrated *in vacuo* to give 7 as an oil. Yield 6.12 g (97%); $R_{\rm f}$ 0.7 (light-petroleum/ether, 2/1, v/v), $[\alpha]_{\rm D}$ + 24.6° (c 1, CHCl₃). ¹H NMR (CDCl₃): δ 7.51–7.12 (m, 20H, H-arom), 5.90 (m, 1H, CH =, All), 5.16 (m, 2H, = CH_2 , All), 4.83-4.54 (m, 6H, CH_2 , Bn), 4.73 (s, 1H, H-1), 4.12 (2H, OCH2, All), 4.24-3.30 (5H, H-2, H-3, H-4, H-5, H-6), 3.26 (s, 3H, OCH₃), 1.42 (m, 2H, H-7a,b), 0.32 (s,

6H, CH₃Si). ¹³C NMR (CDCl₃): δ 138.9, 138.3 (Cq, arom), 134.9 (CH=, All), 129.3–127.6 (CH, arom), 116.4 (=CH₂, All), 99.1 (C-1), 80.1, 74.9, 74.5, 74.3, 72.9 (C-2, C-3, C-4, C-5, C-6), 74.3, 72.3, 70.7 (3 × CH₂, Bn), 69.8 (CH₂O, All), 54.9 (OCH₃), 17.1 (C-7), -2.0, -2.2 (SiCH₃). Anal. calcd. for C₄₀H₄₈O₆Si: C 73.59, H 7.41; found: C 73.48, H 7.52%.

Methyl 2,4,6-tri-O-benzyl-7-(phenyldimethylsilyl)-7-deoxy-t-glycero- $-\alpha$ -p-manno-heptopyranoside (8)

p-Toluenesulfonic acid (1.8 g) and palladium on active carbon (1.8 g, 10%) was added to 7 (6.12 g, 9.4 mmol) in a mixture of methanol/water (120 ml, 5/1, v/v). After heating the reaction for 8 h under reflux, TLC analysis (acetone/CH₂Cl₂, 1/99, v/v) showed the absence of 7. The reaction mixture was cooled to room temperature, neutralized with triethylamine, filtered over a pad of celite and the filtrate was concentrated under reduced pressure. The oily product was dissolved in CH₂Cl₂ (75 ml) and washed with water (25 ml). The organic phase was dried over MgSO4 and concentrated to an oil which was applied onto a column of silica gel (60 g). Elution of the column was effected with a mixture of lightpetroleum/ethyl acetate $(5/95, 15/85 \rightarrow v/v)$. The appropriate fractions were concentrated to give 8 as an oil. Yield 4.2 g (73°_{o}); $R_{\rm f}$ 0.6 (light-petroleum/ether, $2/\bar{1}$, v/v), $[\alpha]_{\rm D}$ + 21.6° (c 1, CHCl₃). ¹H NMR (CDCl₃): 87.52-7.20 (20H, H-arom), 4.90 (d, 1H, H-1. $J_{1,2}$ 1.8 Hz), 4.80-4.20 (m, 6H, 3 × CH₂, Bn), 4.02 (m, 1H, H-6. $J_{5,6}$ 1.3 Hz, $J_{6,7a}$ 1.7 Hz, $J_{6,7b}$ 4.0 Hz), 3.90 (bs, 1H, H-3), 3.77 (t, 1H, $J_{2,3}$, $J_{3,4}$ 9.2 Hz), 3.68 (dd, 1H, H-2, $J_{2,3}$ 3.7 Hz), 3.44 (dd, 1H, H-5), 3.29 (s, 3H, OCH₃), 2.25 (bs, 1H, OH), 1.43, 1.41 (2 × d, 2H. H-7a, H-7b). Anal. calcd. for C₃₇H₄₄O₆Si: C 72.52, H 7.24; found: C 72.61, H 7.48%.

Methyl 2,4,6-tri-O-benzyl-L-glycero- α -D-manno-heptopyranoside (9) Acetic acid (5 ml), sodium acetate (0.65 g) and potassium bromide (95 mg, 0.8 mmol) were added to 8 (0.39 g, 0.64 mmol) and the mixture was stirred until the salts were dissolved. The solution was cooled (10°C) and peracetic acid (3.3 ml, 30°_{o} in acetic acid) was added dropwise under exclusion of ight. During the addition, gas was liberated. After the mixture was stirred for 1 h at 20°C, TLC analysis (acetone/CH2Cl2, 3/97, v/v) indicated complete conversion of the starting material into a more hydrophilic product $(R_{\rm f} 0.2)$. The mixture was diluted with CH₂Cl₂ (25 ml) and poured in an aqueous solution of sodium thiosulfate (5 ml, 15°,). The organic layer was washed with aqueous sodium bicarbonate (10 ml, 10%) and water (10 ml), dried over MgSO₄ and concentrated under reduced pressure. The residue was dissolved in toluene $(2 \times 10 \text{ ml})$ and concentrated. The oily product was applied onto a column of silica gel (10 g) and elution was effected with CH₂Cl₂ followed by CH₂Cl₂/methanol (99/1 \rightarrow 49/1, v/v). The appropriate fractions were concentrated to give **9** as an oil. Yield 0.26 g (82%), $R_f 0.2$ (methanol/CH₂Cl₂, 3/97, v/v), $[\alpha]_D + 3.7^{\circ}$ (c 1, CHCl₃). ¹³C NMR (CDCl₃): δ 138.2–137.5 (Cq, arom), 128.4-127.3 (CH, arom), 97.7 (C-1), 80.0, 75.8, 71.7, 71.4 (C-2, C-3, C-4, C-5, C-6), 74.0, 72.4, 71.9 ($3 \times CH_2$, Bn), 61.9 (C-7), 54.6 (OCH₃). Anal. calcd. for C₂₉H₃₄O₇: C 70.43, H 6.93; found: C 70.56, H 6.93°

Methyl O-(2-O-acetyl-3,4,6,7-tetra-O-benzyl-L-glycero-D-manno-heptopyranosyl)-($1 \rightarrow 3$)-O-(2-O-acetyl-3,4,6,7-O-L-glycero-D-mannoheptopyranosyl($1 \rightarrow 7$)/-2,3,6-tri-O-benzyl-L-glycero- α -D-manno-heptopyranoside (18)

Activated powdered molecular sieves (0.5 g) was added to a solution of compound 9 (0.1 g, 0.17 mmol) in CH₂CCl₂ (3 ml) and the mixture was stirred for $2\frac{1}{2}h$ in an atmosphere of nitrogen. Silver triflate (0.15 g, 0.6 mmol) was added under the exclusion of light. The mixture was cooled to -30° C and chloride 17 (0.4 g, 0.64 mmol) in CH₂Cl₂ (2 ml) was added dropwise over a period of 30 min. The reaction mixture was stirred at -20° C for 2 h. The mixture was filtrated and the filtrate was diluted with CH₂Cl₂ (15 ml) and washed with aqueous sodium thiosulfate (10 ml), aqueous sodium bicarbonate (10 ml) and water (10 ml), dried over MgSO₄ and concentrated in vacuo. The residue was applied onto a column of silica gel (2 g) and eluted with light-petroleum/ethylacetate (85/15, v/v). Fractions containing the required dimer were concentrated under reduced pressure and the residual oil was applied on a LH-20 Sephadex column (eluent, CH_2Cl_2 /methanol, 1/1, v/v). Concentration of the appropriate fractions gave pure 18

in a yield of 0.205 g (71%). $R_{\rm f}$ 0.6 (acetone/CH₂Cl₂, 3/97, v/v), [α]_D + 31.6° (c 0.95, CHCl₃). ¹H NMR (CDCl₃): δ 7.35–7.05 (m, 55H, H-arom), 5.43 (t, 1H, H-2', $J_{2,3}$, $J_{1,2}$ 2.1 Hz), 5.26 (t, 1H, H-2", $J_{2,3}$, $J_{1,2}$ 2.5 Hz), 5.17 (d, 1H, H-1'), 4.82 (d, 1H, H-1"), 4.48 (d, H, H-1, $J_{1,2}$ 1.9), 3.91 (2× dd,, 2H, H-3', H-3"), 3.60 (dd, H-2, $J_{2,3}$ 3.2 Hz), 4.79–3.43 (m, 38H, 11× CH₂, Bn. H-3–H-7a,b, H-4'–H-7'a,b, H-4"–H-7"a,b), 3.09 (OCH₃), 2.10, 1.98 (2× s, 6H, CH₃, Ac). ¹³C NMR (CDCl₃): δ 170.1, 169.7 (C=O, Ac), 138.8–137.4 (Cq, arom), 128.3–137.4 (CH, arom), 100.1, 98.2 (C-1, C-1', C-1"), 78.4, 77.9, 76.9, 75.1, 74.8, 73.6, 71.8, 71.5, 86.3, (C-2 to C-6, C-2' to C-6', C-2" to C-6"), 74.5, 74.2, 73.2, 72.8, 72.6, 72.5, 72.2, 71.2, 70.3, 66.8 (CH₂, Bn, C-7, C-7", C-7"), 54.6 (OCH₃), 20.9, 20.8 (CH₃, Ac). Anal. calcd. for C₁₀₃H₁₁₀O₂₁: C 73.46, H 6.58; found: C 73.55, H 6.53%.

Methyl O-(3.4.6.7-tetra-O-benzyl-L-glycero- α -D-manno-heptopyranosyl)- $(1 \rightarrow 3)$ -O-[3.4.6.7-tetra-O-benzyl-L-glycero- α -D-manno-heptopyranosyl- $(1 \rightarrow 7)$]-2.4.6-tri-O-benzyl-L-glycero- α -D-manno-heptopyranoside (19)

Methanolic sodium methoxide (1M, 0.05 ml) was added to a solution of **18** (0.205 g, 0.12 mmol) in methanol (5 ml) and the reaction mixture was stirred for 2 h at 20 °C. The mixture was neutralized with Dowex 50 XW4 (H ⁺ form) resin (100–200 mesh), filtered and concentrated under reduced pressure. The oily residue was applied on to a column of silica gel (2 g) and eluted with CH₂Cl₂ followed by a mixture of acetone/CH₂Cl₂ (1/99 \rightarrow 3/97, v/v). Concentration of the appropriate fractions gave **19** as an oil. Yield 0.18 g (94%); *R*_f 0.5 (acetone/CH₂Cl₂, 3/97, v/v). ¹³C NMR (CDCl₃): δ 138.7–137.5 (Cq, arom), 128.3–127.0 (CH, arom), 99.8 (C-1, C-1', C-1'), 80.0, 75.1, 74.3, 73.4, 71.3, 71.0, 67.2, 67.8 (C-2–C-6, C-2' – C-6'), C-2" – C-6"), 74.4, 74.2, 73.2, 72.8, 72.4, 71.7, 71.4, 70.2, 66.88 (CH₂, Bn, C-7, C-7', C-7"), 54.6 (OCH₃). Anal. calcd. for C₉₉H₁₀₆O₁₉: C 74.32, H 6.68; found: C 74.35, H 6.63%.

Methyl (L-glycero- α -D-manno-heptopyranosyl)-($1 \rightarrow 3$)-O-[L-glycero- α -D-manno-heptopyranosyl-($1 \rightarrow 7$)]-L-glycero- α -D-manno-hepto-pyranosyl (20)

Compound **19** (0.18 g) was dissolved in methanol and after addition of 200 mg Pd/C (10%), the suspension was kept under an atmosphere of hydrogen during 42 h at 1 atmosphere. The catalyst was removed by filtration and washed with water. The combined filtrates were concentrated under reduced pressure to give **20** (50 mg) as an amorphous material. R_r 0.2 (methanol/ethyl-acetate/water, 3/5/2, v/v), $(\alpha]_{15} + 114.2^{\circ}$ (c 1, water). ¹H NMR (D₂O): δ 5.11 (d, 1H, H-1, $J_{1,2}$ 1.8 Hz), 4.87 (d, 1H, H-1", $J_{1,2}$ 1.7 Hz), 4.71 (d, 1H, H-1, $J_{1,2}$ 1.7), 4.17 (m, 1H, H-6'), 4.03 (dd, 1H, H-2", $J_{2,3}$ 3.0 Hz), 3.99 (dd, 1H, H-2', $J_{2,3}$ 3.0 Hz), 3.99 (dd, 1H, H-3'), 3.68 (t, 1H, H-4, $J_{4,5}$ 7.3), 3.54 (dd, 1H, H-5', $J_{5,6}$ 1.7 Hz), 3.36 (s, 3H, OCH₃). ¹³C NMR (CDCl₃): δ 103.4 (C-1), 101.9 (C-1'), 101.6 (C-1"), 72.4 (C-5'), 71.0 (C-2), 68.2 (C-2), 79.1, 72.7, 72.3, 71.7, 71.4, 70.0, 69.7, 67.0, 66.5, 63.9, 63.8 (C-3-C-5, C-7, C-2'-C-4', C-6', C-7', C-2'-C-7"), 55.7 (OCH₃). $J_{C-1,H-1}$ 172.9, $J_{C-1,H-1}$ 174.4 Hz, $J_{C-1",H-1"}$ 171.5 Hz.

Methyl 2.4,6-tri-O-benzyl-3-O-(2.3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)-7-deoxy-7-(phenyldimethylsilyl)-L-glycero-D-manno-heptopyranoside (**22**)

Activated powdered molecular sieves (0.5 g) was added to a solution of compound 8 (0.17 g, 0.28 mmol) in diethylether (2 ml) and the mixture was stirred for 2 h in an atmosphere of nitrogen. Silver triflate (0.13 g, 0.52 mmol) was added with the exclusion of light. The mixture was cooled to -30° C and the chloride 21 (0.44 mmol) in diethyl ether (2 ml) was added over a period of 30 min. The reaction mixture was stirred at -20° C for 16 h. The mixture was filtered and the filtrate was diluted with CH₂Cl₂ (25 ml) and washed with aqueous sodium thiosulfate (10 ml), aqueous sodium bicarbonate (10 ml), water (10 ml), dried over MgSO₄ and concentrated under reduced pressure. The residue was applied to a column of silica gel (3 g) and eluted with light-petroleum/ethyl-acetate $(95/5 \rightarrow 85/15, v/v)$. The dimer containing fractions were concentrated under reduced pressure and the residual oil was applied on a LH-20 Sephadex column (eluent, CH₂Cl₂/methanol, 1/1, v/v). Concentration of the appropriate fractions gave pure 22 in a yield of 0.29 g (90%). $R_{\rm f}$ 0.5 (acetone/CH₂Cl₂, 1/97, v/v): $[\alpha]_{\rm D}$ + 32.1° (c 0.95, CHCl₃). ¹H NMR (CDCl₃): δ 7.49–7.07 (40H, H arom), 5.12 (d, 1H, H-1', $J_{1,2}$ 3.4 Hz), 4.88 (d, 1H, H-1, $J_{1,2}$ 2.7), 4.15 (t, 1H, H-4, $J_{3,5}$, $J_{4,5}$ 11.8 Hz), 4.07 (dd, 1H, H-3), 4.01 (m, 1H, H-6), 3.89 (t, 1H, H-2, $J_{1,2}$ 2.3 Hz), 3.55 (dd, 1H, H-3), 3.49 (q, 1H, H-2'), 5.05–3.45 (19H, 8 × CH₂, Bn, H-3' to H-6'a,b), 3.27 (s, 3H, OCH₃), 1.42 (d, 2H, H-7a,b, $J_{6,7}$ 7.0 Hz), 0.33, 0.32 (2× s, 6H, CH₃ si). ¹³C NMR (CDCl₃): δ 139.1–137.8 (Cq, arom), 133.4–126.6 (CH, arom), 98.8, 97.0 (C-1, C-1'), 81.6, 79.6, 77.7, 77.4, 75.2, 74.6, 72.9, 70.8 (C-2–C-6, C-2'–C-5'), 75.2, 74.5, 73.7, 73.2, 72.6, 71.4, 69.6, 68.6 (CH₂, Bn, C-6'), 54.9 (OCH₃), 17.8 (C-7), – 2.3, – 2.4 (CH₃Si). Anal. calcd. for C₇₁H₇₈O₁₁: C 75.10, H 6.92; found: C 75.31, H 6.93%.

Methyl 2.4.6-tri-O-benzyl-3-O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)-L-glycero- α -D-manno-heptopyranoside (23)

Compound 22 (0.29 g, 0.25 mmol) was oxidized, as described earlier for $8 \rightarrow 9$, to give, after further work up and purification, homogeneous 23 in a yield of 140 mg (53%), $R_f 0.4$ (acetone/CH₂Cl₂, 1/97, v/v); $[\alpha]_D + 41.7^{\circ}$ (c 0.1, CHCl₃). ¹³C NMR (CDCl₃): δ 139.5–138.2 (Cq, arom), 129.8–126.9 (CH, arom), 98.8, 97.2 (C-1, C-1'), 81.6, 79.5, 77.7, 77.2, 75.7, 74.6, 73.9, 73.1, 70.9 (C-2–C-6, C-2'–C-5'), 75.3, 74.5, 73.3, 72.7, 71.7, 71.4 (CH₂, Bn), 68.6 (C-6'), 62.3 (C-7), 54.8 (OCH₃). Anal. calcd. for C₆₃H₆₈O₁₂: C 74.39, H 6.74; found: C 75.37, H 6.81%.

Methyl O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)-(1 \rightarrow 3)-O-[2--O-acetyl-3,4,6,7-tetra-O-benzyl- α -L-glycero-D-manno-heptopyranosyl-(1 \rightarrow 7)]-2,4,6-tri-O-benzyl-L-glycero- α -D-manno-heptopyranoside (24)

A mixture of compound 23 (0.140 mg, 0.09 mmol) and activated powdered molecular sieves (0.5 g) in CH₂Cl₂ (3 ml) was stirred for $2\frac{1}{2}$ h in an atmosphere of nitrogen. Under the exclusion of light, silver triflate (65 mg, 0.25 mmol) was added. The mixture was then cooled to -30° C and chloride 17 (0.135 g, 0.25 mmol) in CH₂Cl₂ (1 ml) was added over a period to 30 min. The reaction mixture was stirred at -20°C for 16 h. The mixture was filtered and the filtrate was diluted with CH₂Cl₂ (15 ml), washed with aqueous sodium thiosulfate (10 ml), sodium bicarbonate (10 ml) and water (10 ml). The organic phase dried over MgSO₄, was concentrated under reduced pressure and the residue was applied onto a column of silica gel (2 g). Elution of the column was effected with CH_2Cl_2 followed by a CH_2Cl_2 /acetone mixture (99/1 \rightarrow 97/2, v/v). Fractions containing the required dimer were concentrated under reduced pressure and the residual oil was applied on a LH-20 Sephadex column (eluent, CH₂Cl₂/methanol, 1/1, v/v). The appropriate fractions were concentrated to give pure 24 as an oil. Yield 0.20 g (89%); $R_{\rm f} 0.6$ (acetone/CH₂Cl₂, 3/97, v/v), $[\alpha]_{\rm D}$ + 34.7° (c 1.1, CHCl₃). ¹H NMR (CDCl₃): δ 7.40–7.10 (55H, H-arom), (c 1.1, CHC₁₃). ¹H MMR (CDC₁₃): 67.40-7.10 (33H, H-aroln), 5.35 (t, 1H, H-2", $J_{1,2}, J_{2,3}$ 2.3 Hz), 5.15 (d, 1H, H-1', $J_{1,2}$ 3.5 Hz), 4.90 (d, 1H, H-1', $J_{1,2}$ 1.9 Hz), 4.79 (s, 1H, H-1), 4.24 (t, 1H, H-4, $J_{3,4}, J_{4,5}$ 1.9 Hz), 4.23 (t, 1H, H-4, $J_{3,4}, J_{4,5}$ 9.4 Hz), 4.13 (dd, 1H, H-3, $J_{2,3}$ 2.9 Hz), 3.98 (dd, 1H, H3", $J_{3,4}$ 7.7 Hz), 3.90 (d, 1H, H-2, $J_{2,3}$ 2.3 Hz), 3.62 (d, 1H, H-5), 3.53 (dd, 1H, H-2'), $J_{2,3}$ 9.8 Hz), 4.91–3.49 (35H, 11× CH₂, Bn, H-6, H-7a,b, H-3'–H-6'a,b, H-4"-H-7"a,b), 3.21 (s, 3H, OCH₃), 2.15 (s, 3H, CH₃, Ac). ¹³C NMR (CDCl₃): δ 170.2 (C=O, Ac), 139.2–137.7 (Cq, arom), 128.5–127.1 (CH, arom), 99.0, 98.3 (C-1, C-1', C-1''), 81.9, 79.8, 78.3, 77.6, 75.0, 73.9, 71.5, 71.3, 68.5 (C-2–C-6, C-2'–C-5', C-2"–C-6"), 75.6, 74.8, 73.8, 73.5, 72.8, 72.6, 72.0, 71.7, 70.6, 70.2, 67.8 ($11 \times CH_2$, Bn, C-7, C-7", C-6'), 55.1 (OCH_3), 21.1 (CH_3 , Ac). Anal calcd. for C₁₀₀H₁₀₆O₁₉: C 74.51, H 6.63; found: C 74.39, H 6.61%.

Methyl O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O--[3,4,6,7-tetra-O-benzyl-L-glycero- α -D-manno-heptopyranosyl- $(1 \rightarrow 7)$]-L-glycero- α -D-manno-heptopyranoside (25)

Potassium *tert*-butoxide (5 mg) was added to a solution of compound **24** (0.150 g, 0.09 mmol) in methanol (3 ml). After stirred for 2 h, the reaction mixture was neutralized with Dowex 50 XW4 (H ' form) resin (100–200 mesh), filtered and concentrated under reduced pressure. The residue was purified by silica gel (3 g) column chromatography (eluent: CH_2Cl_2 followed by acetone/ CH_2Cl_2 , 1/99 \rightarrow 3/99, v/v). Concentration of the appropriate fractions gave **25** as an oil. Yield 0.115 g (81%); R_f 0.5

(acetone/CH₂Cl₂, 3/97, v/v); $[\alpha]_D$ +43.8° (c 1.05, CHCl₃). ¹³C NMR (CDCl₃): δ 139.0–137.6 (cq, arom), 128.4–126.9 (CH, arom), 99.7, 90.0, 97.8 (C-1, C-1', C-1''), 80.1, 79.5, 77.7, 77.3, 74.8, 73.8, 73.5, 71.2, 70.9, 67.9 (C-2–C-6, C-2'–C-5', C-2''–C-6''), 75.4, 74.5, 74.1, 73.4, 73.3, 72.6, 71.6, 71.4, 70.5, 68.5, 67.7 (CH₂, Bn. C-7, C-7'', C-6'), 54.9 (OCH₃). Anal. calcd. for C₉₈H₁₀₄O₁₈: C 74.98, H 6.68; found: C 74.87, H 6.74%.

Methyl O- $(\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O- $[\alpha$ -L-glycero-D-manno--heptopyranosyl- $(1 \rightarrow 7)$]-L-glycero- α -D-manno-heptopyranoside (26)

A mixture of **25** (0.115 g, 0.07 mmol) and 10% Pd-C (0.7 g) in methanol (3 ml) was stirred under an atmosphere of H₂ for 48 h at 20°C. The catalyst was removed by filtration and washed with water. The combined filtrates were concentrated under reduced pressure to give **26** (31 mg) as an amorphous material. $R_{\rm f}$ 0.3 (methanol/ethyl-acetate/water, 3/5/2, v/v/v), $[\alpha]_{\rm D}$ + 112.3° (*c* 1, water). ¹H NMR (D₂O): δ 5.19 (d, 1H, H-1', $J_{1,2}$ 3.9 Hz), 4.88 (d, 1H, H-1", $J_{1,2}$ 1.8 Hz), 4.71 (d, 1H, H-1, $J_{1,2}$ 1.9 Hz), 4.17 (ddd, 1H, H-6, $J_{5,6}$ 1.5 Hz, $J_{6,7a}$ 7.0 Hz, $J_{6,7b}$ 8.0 Hz), 4.06 (dd, 1H, H-2, $J_{2,3}$ 3.3 Hz), 4.02 (ddd, 1H, H-6", $J_{5,6}$ 5.0 Hz, $J_{6,7a}$ 1.5 Hz, $J_{6,7a}$ 7.0 Hz, $J_{6,7b}$ 8.0 Hz), 4.06 (dd, 1H, H-2, $J_{2,3}$ 3.3 Hz), 4.02 (ddd, 1H, H-6", $J_{5,6}$ 5.0 Hz, $J_{6,7a}$ 1.5 Hz, $J_{6,7b}$ 7.5 Hz), 4.01 (t, 1H, H-4, $J_{3,4}$, $J_{4,5}$ 9.8 Hz), 3.96 (dd, 1H, H-2", $J_{2,3}$ 3.0 Hz), 3.84 (dd, 1H, H-3), 3.82 (dd, 1H, H-3"), 3.78 (dd, 1H, H-5'', $J_{4,5}$ 1.1 3 Hz), 3.40 (t, 1H, H-4'), $J_{3,4}$, $J_{2,3}$ 9.7 Hz), 3.65 (dd, 1H, H-5", $J_{4,5}$ 1.1 3 Hz), 3.40 (t, 1H, H-4'), $J_{3,7}$ 3.59 (8H, H-7a,b, H-6'a,b, H-4" – H-7"), 3.36 (s, 3H, OCH₃). ¹³C NMR (D₂O): δ 101.6 (C-1), 101.4 (C-1"), 101.3 (C-1'), 79.7 (C-3), 72.5 (C-2"), 70.5 (C-2), 70.3 (C-4'), 68.2 (C-6), 73.6, 73.0, 72.2, 72.1, 69.5, 66.8, 66.0 (C-4, C-3', C-5', C-3" – C-6"), 69.8, 63.7, 61.2 (C-7, C-6', C-7"), 55.5 (OCH₃).

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References

¹ L. Kenne and B. Lindberg, in The Polysaccharides, G. O. Aspinall (Ed.), Vol. 2, 287-363, Academic Press, New York

(1983); H. Brade and E. Th. Rietschel, Eur. J. Biochem. 145, 231 (1984); H. Brade, U. Zahringer, E. Th. Rietschel, R. Christian, G. Schulz and F. Unger, Carbohydr. Res. 134, 157 (1984).

- ² H. Brade and C. Galanos, Infect. Immun. **42**, 250 (1983); O. Luderits, A. M. Staub and O. Westphal, Bacteriol. Rev. **30**, 192 (1966); A. D. Elibein and C. Heath, J. Biol. Chem. **240**, 1919 (1965).
- ³ G. J. P. H. Boons, P. A. M. van der Klein, G. A. van der Marel and J. H. van Boom, Recl. Trav. Chim. Pays-Bas 107, 507 (1988); G. J. P. H. Boons, G. A. van der Marel and J. H. van Boom, Recl. Trav. Chim. Pays-Bas 108, 339 (1989).
- ⁴ G. J. P. H. Boons, G. A. van der Marel and J. H. van Boom, Tetrahedron Lett. **30**, 229 (1989); G. J. P. H. Boons, M. Overhand, G. A. van der Marel and J. H. van Boom, Angew. Chem. Int. Ed. Engl. **28**, 1504 (1989); G. J. P. H. Boons, R. Steyger, M. Overhand, G. A. van der Marel and J. H. van Boom, J. Carbohydr. Chem., in the press.
- ⁵ G. J. P. H. Boons, P. A. M. van der Klein, G. A. van der Marel and J. H. van Boom, Recl. Trav. Chim. Pays-Bas 109, 255 (1990); P. A. M. van der Klein, G. J. P. H. Boons, G. H. Veeneman, G. A. van der Marel and J. H. van Boom, Tetrahedron Lett. 30, 5477 (1989).
- ⁶ G. J. P. H. Boons, M. Overhand, G. A. van der Marel and J. H. van Boom, Carbohydr. Res. **192**, c1-c4 (1989).
- ⁷ E. Romanowska, A. Gamian and J. Dabrowski, Eur. J. Biochem. 161, 557 (1986); E. Dabrowski and M. Hauck, Carbohydr. Res. 180, 163 (1988).
- ⁸ D. J. Cramm and D. R. Wilson, J. Am. Chem. Soc. 85, 1245 (1963).
- ⁹ *I. Feming, E. J. Philips* and *E. J. Sanderson*, Tetrahedron Lett. **28**, 4229 (1987).
- ¹⁰ K. Dziewiszek, A. Banaszek and A. Zamojski, Tetrahedron Lett. 28, 1569 (1987).
- ¹¹ S. Czernecki, G. Georgoulis and C. Provelenghion, Tetrahedron Lett. 17, 3535 (1976).
- ¹² *R. Boss* and *R. Scheffolo*, Angew. Chem. **15**, 578 (1976).
- ¹³ D. Peterson, J. Org. Chem. **33**, 780 (1968).
- ¹⁴ H. Paulsen and A. C. Heitman, Justus Liebigs Ann. Chem. 1061 (1988).
- ¹⁵ S. Hanessian and J. Banoub, Carbohydr. Res. **53**, c13-c16 (1977).
- ¹⁶ V. D. Grob, T. G. Squires and J. R. Vergellotti, Carbohydr. Res. **10**, 595 (1969).