Contents lists available at ScienceDirect

Tetrahedron: Asymmetry

journal homepage: www.elsevier.com/locate/tetasy

Enantioselective synthesis of beta-amino acids using hexahydrobenzoxazolidinones as chiral auxiliaries

Gloria Reyes-Rangel, Erika Jiménez-González, José Luis Olivares-Romero, Eusebio Juaristi *

Departamento de Química, Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, Apartado Postal 14-740, 07000 México, D. F., Mexico

ARTICLE INFO

Article history: Received 14 October 2008 Accepted 10 December 2008 Available online 24 January 2009

ABSTRACT

A practical synthetic route for the asymmetric synthesis of β^2 -amino acids is described. In the first step, the procedure involves the N-acylation of readily available, enantiopure hexahydrobenzoxazolidinone (4R,5R)-1 with 3-methylbutanoyl chloride 2, 4-methylpentanoic acid 3, and 3-(1-tert-butoxycarbonyl)-1H-indol-3-yl)propanoic acid 4 to afford derivatives 5a, 5b, and 5c, respectively, which were alkylated with high diastereoselectivity by means of reaction between their sodium enolates and benzyl bromoacetate. Removal of the chiral auxiliary from the alkylated products followed by hydrogenation and hydrolysis gave β^2 -amino acids (S)-10a, (S)-10b, and (S)-10c, which were N-protected with Fmoc. Enantiomeric (R)-10a-c were similarly prepared from the isomeric hexahydrobenzoxazolidinone (45.55)-1; thus, the route presented here provides access to both enantiomers of valuable highly enantioenriched β^2 -amino acids.

© 2009 Elsevier Ltd. All rights reserved.

1. Introduction

The development of synthetic procedures to obtain β-amino acids is a relevant endeavor since these homologues of α -amino acids play a significant role in medicinal chemistry.¹ Furthermore, β-amino acids have long been employed as precursors for β-lactams, and more recently have gained attention as building blocks for oligomers with well-defined folding behavior, 'foldamers'.² Moreover, pioneering works by Seebach et al.³ and Gellman et al.⁴ have demonstrated the folding propensity of oligomers of these compounds, leading to secondary and tertiary structures very similar to those of α -peptides, hence they are considered as excellent peptidomimetics.⁵ One can distinguish between the β^2 and β^3 -amino acids depending on the position of the side chain in the β -amino acid skeleton.⁶

Proteinogenic α -amino acids are constituents of all enzymes which control the function and metabolism in living organisms, whereas β-amino acids occur mainly as constituents of distinct natural products, such as peptides. The incorporation of β-amino acids instead of their α -analogues increases the stability of peptides against degradation by mammalian peptidases;³ therefore, β-amino acids are an important tool in the development of drugs capable of withstanding hydrolytic degradation for prolonged periods of time.7

For these reasons, numerous methodologies for the synthesis of racemic and enantiomerically pure β-amino acids have emerged.¹ Noticeably, the Arndt–Eistert homologation of α -amino acids allows the preparation of β^3 -amino acids bearing functional groups on their side chains in a few steps.⁸

The preparation of β^2 -amino acids is much more challenging, and several useful strategies are reported for the synthesis of β^2 amino acids bearing trivial⁹ or functionalized¹⁰ side chains.

The use of chiral auxiliaries is another methodology employed for the synthesis of enantiopure α - and β -amino acids. In this context, Juaristi et al.¹¹ reported a convenient procedure for the preparation of both pairs of enantiomeric hexahydrobenzoxazolidin-2-ones (R,R)-1 and (S,S)-**1** from inexpensive cyclohexene oxide and (S)- α -phenylethvlamine (Fig. 1). These authors have also described the use of (R,R)-1 and (S,S)-1 as effective chiral auxiliaries for the stereoselective alkylation, acylation, and aldol condensation of propionic and hydrocinnamic acids.¹² The application of hexahydrobenzoxazolidinones (R,R)-1 and (S,S)-1 as chiral sulfinyl transfer reagents has recently been reported.¹³ In addition, Juaristi et al.¹⁴ described the application of trans-hexahydrobenzoxazolidinones (R,R)-1 and (S,S)-1 in the asymmetric synthesis of aminophosphonic acids AP-3 and AP-4.

Herein we report the use of trans-hexahydrobenzoxazolidinones (R,R)-1 and (S,S)-1 in the enantioselective synthesis of both enantiomers of β^2 -homovaline, β^2 -homoleucine, and β^2 -homotryptophan.



Figure 1. Hexahydrobenzoxazolidinones (R,R)-1 and (S,S)-1.





^{*} Corresponding author. Tel.: +52 55 5747 3722; fax: +52 55 5747 3389. E-mail address: juaristi@relaq.mx (E. Juaristi).



Scheme 1. Synthesis of N-acylated hexahydrobenzoxazolidin-2-ones (R,R)-5a-c. A similar scheme can be applied to reactions carried out with (S,S)-1 as the starting material.

2. Results and discussion

2.1. N-Acylation of enantiopure hexahydrobenzoxazolidinone (R,R)-5

trans-Hexahydrobenzoxazolidinone (R,R)-**1** was N-acylated following the established protocol,¹⁵ by treatment with *n*-butyllithium at -78 °C, followed by the addition of 3-methylbutanoyl chloride **2** to generate derivative (R,R)-**5a**. By the same token, derivatives (R,R)-**5b** and (R,R)-**5c** were prepared by treatment of (R,R)-**1** with acid chlorides derived from carboxylic acids **3** and **4**, respectively, in the presence of pivaloyl chloride and triethyl amine at -30 °C (Scheme 1).

2.2. Diastereoselective alkylation of N-acylated hexahydrobenzoxazolidin-2-ones 5a-c

The sodium enolates derived from (*R*,*R*)-**5a**–**c** and (*S*,*S*)-**5a**–**c**, generated with sodium hexamethyldisilazide (NaHMDS), were treated with 2.0 equiv of benzyl bromoacetate at -78 °C for 5 h.¹⁶ Most relevant, ¹H NMR analysis of crude alkylated products

6a–c showed a single set of signals, indicating a diastereomeric purity greater than 98%.

The absolute configuration of the newly created stereogenic center at $C(\alpha)$ in products **6b** and **6c** was established as (*S*), whereas in **6a**, the configuration was assigned as (*R*) owing to a change in priority of the substituents at $C(\alpha)$. This result is consistent with the intermediacy of a (*Z*)-configured enolate,¹⁷ where the sodium cation is chelated by both carbonyl oxygens in the enolate, and the electrophile is incorporated from the less sterically hindered face (Scheme 2).

2.3. Hydrogenolisis of α -alkylated hexahydrobenzoxazolidin-2ones 6a–c and Curtius rearrangement of the derived carboxylic acids 7a–c

Compounds **6a**, **6b**, and **6c** were hydrogenated with Pd/C 10% at atmospheric pressure to afford carboxylic acid derivatives **7a–c**, which were subjected to Curtius rearrangement by treatment with diphenyl phosphoryl azide, in the presence of Et_3N and benzyl alcohol to give carbamates **8a–c** (Scheme 3).



Scheme 2. Diastereoselective alkylation of N-acylated hexahydrobenzoxazolidin-2-ones (R,R)-5a-c. A similar scheme can be applied to reactions carried out with (S,S)-5a-c.



Scheme 3. Hydrogenolysis and Curtius rearrangement of α -alkylated hexahydrobenzoxazolidin-2-ones (*R*,*R*)-**6a**-**c** to give carbamates **8a**-**c**. A similar scheme can be applied to reactions carried out with (*S*,*S*)-**6a**-**c**.

Crystallization of carbamate (R,R,S)-**8a** from ethyl acetate–hexane (1:9) afforded suitable crystals for X-ray diffraction structural analysis (Fig. 2).¹⁸



Figure 2. X-ray crystallographic structure and solid state conformation of (*R*,*R*,*S*)-8a.¹⁸

Examination of Figure 2 confirms that the alkylation reaction takes place on the less sterically hindered face of the enolates derived from N-acylated substrates **5a–c**, as discussed in Section 2.2.

2.4. Removal of the chiral auxiliary, hydrogenation, and protection with Fmoc

The removal of the oxazolidinone chiral auxiliary from compounds (*S*)-**8a** and (*S*)-**8b** was achieved with lithium hydroperoxide, as suggested by Evans et al.^{16,17} These reactions proceed in 53% and 70% yield, respectively, and the resulting products (*S*)-**9a** and (*S*)-**9b** were reduced by catalytic hydrogenation to afford β -amino acids (*S*)-**10a** and (*S*)-**10b** in 89% and 81% yield, respectively. Removal of the chiral auxiliary in compound (*S*)-**9c** was achieved by treatment with *n*-BuLi and BnOH to obtain the benzylated derivative in 47% yield, and the resulting adduct was hydrogenated to give the β -amino acid (*S*)-**10c** in 65% yield. Finally, the protection of β -amino acids (*S*)-**10a**-**c** was achieved by treatment with Fmoc-Osu, in acetone and sodium carbonate 0.15 M (1:1) to afford the *N*-Fmoc-protected β -amino acids (*S*)-**11a**-**c** in 45-65% yield (Scheme 4).

2.5. HPLC analysis

HPLC analysis confirmed the efficiency of the present approach for the preparation of enantiomerically pure β^2 -homovaline, β^2 homoleucine, and β^2 -homotryptophan, which were obtained in 99.4%, 96.0%, and 90% enantiomeric excess, respectively (Fig. 3). Since the alkylation reaction of N-acylated hexahydrobenzoxazolidin-2-ones **5a–c** proceeds with greater than 98% diastereoselectivity, we ascribed the lower enantiomeric excess in two cases to the presence of minor amounts of enantiomeric *trans*-hexahydrobenzoxazolidinone (*S*,*S*)-**1** in the starting substrate (*R*,*R*)-**1**, which at the end afforded a small amount of the undesired enantiomeric amino acid.

3. Conclusion

An efficient synthetic route that provides both enantiomers of β^2 -amino acids from hexahydrobenzoxazolidinones (*R*,*R*)-**1** and (*S*,*S*)-**1** was developed. In particular, N-acylated derivatives (*S*,*S*)-**6** and (*R*,*R*)-**6**, which were prepared by N-acylation of *trans*-hexahydrobenzoxazolidinones (*S*,*S*)-**1** and (*R*,*R*)-**1**, were alkylated with very high diastereoselectivity to afford products which were deprotected, subjected to Curtius rearrangement, and finally converted into the desired enantioenriched β^2 -homovaline, β^2 -homoleucine, and β^2 -homotryptophan.

4. Experimental

4.1. General

Flasks, stirring bars, and hypodermic needles used for the generation and reactions of organometallic compounds were dried for ca. 12 h at 120 °C and allowed to cool in a desiccator over anhydrous CaSO₄. Anhydrous solvents were obtained by distillation from benzophenone/ketyl radical. *n*-Butyllithium was titrated according to the method of Juaristi et al.¹⁹ TLC: Merck DC-F254 plates, detection by UV light, iodine vapor, or ninhydrin spray. Flash chromatography: Merck silica gel (0.040–0.063 mm). Melting points: Melt Temp apparatus, not corrected. ¹H NMR spectra: Jeol Eclipse-400 (400 MHz), Bruker Ultra Shield (300 MHz), and Jeol GSX-270 (270 MHz) spectrometers; ¹³C NMR spectra: Jeol Eclipse-400 (100 MHz) and Bruker Ultra Shield (75 MHz); ³¹P NMR spectra: Jeol Eclipse-400 (162 MHz) and Bruker Ultra Shield



Scheme 4. Reagents and conditions: (a) Method A: LiOH·H₂O, THF-H₂O (1:1), H₂O₂. Method B: (a) BnOH, *n*-BuLi, THF; (b) Pd/C, H₂, EtOH; (c) acetone-Na₂CO₃ 0.15 M (1:1), Fmoc-Osu. A similar scheme can be applied to reactions carried out with (*S*,*S*,*R*)- 8a-c).



Figure 3. HPLC (*Chiralcel OD*, λ = 210 nm, elution system: *i*-PrOH-hexane–TFA; 10:90:0.1, for (S)-**11a** and (S)-**11b**. *Chiralcel OD-H*, λ = 210 nm, elution system: *i*-PrOH-hexane–TFA; 20:80:0.1, for (S)-**11c**).

(121 MHz) spectrometers. Chemical shifts δ are given in parts per million relative to Me₄Si as an internal reference; coupling constants are given in *J* (Hertz). Mass spectra were obtained in a Hewlett–Packard HP-5986 instrument. High-resolution mass spectra (HRMS) were obtained on an HPLC 1100 coupled to MSDTOF Agilent Technologies mod. 1969A.

4.2. N-Acylation procedure for the preparation of 5a

n-BuLi (2.35 M, 28.3 mmol, 1 equiv) was added to a cold (-20 °C) solution of hexahydrobenzoxazolidin-2-one (*R*,*R*)- or (*S*,*S*)-1 (28.3 mmol, 1 equiv) in THF (100 mL) and the resulting mixture was stirred for 30 min at -20 °C. The corresponding acid chloride (28.3 mmol, 1 equiv) was added for 15 min and the reaction mixture was stirred for 12 h at room temperature, before the addition of saturated aqueous NH₄Cl (100 mL), H₂O (80 mL), and EtOAc (150 mL). The organic phase was separated and washed with brine (100 mL), whereas each aqueous fraction was re-extracted with EtOAc (2×80 mL). The combined organic extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo.

4.2.1. (4*R*,5*R*)-3-(3-Methylbutanoyl)hexahydrobenzoxazolidin-2-one, (*R*,*R*)-5a

The general procedure described above was followed and the crude product was purified by flash chromatography using ethyl acetate–hexane (80:20) as eluent to give 98% yield of the desired product, mp 89–90 °C. $[\alpha]_D^{25} = -72$ (*c* 1.02, CHCl₃). IR (KBr, cm⁻¹): v_{max} 2994, 2954, 1798, 1690, 1368, 1332, 1214, 1150. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.94 (d, 3H, *J* = 3.2), 0.96 (d, 3H, *J* = 3.2), 1.42 (m, 3H), 1.66 (m, 1H), 1.85 (m, 2H), 2.18 (m, 2H), 2.78 (m, 3H), 3.54 (ddd, 1H, $J_1 \approx J_2 = 10.6$, $J_3 = 3.0$), 3.86 (ddd, 1H, $J_1 \approx J_2 = 11.5$, $J_3 = 3.6$). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 22.8(2xC), 23.9, 24.1, 25.6, 28.8, 29.2, 45.4, 63.4, 81.6, 153.1, 175.3; MS (20 eV): *m*/*z* 225 (M⁺), 210, 183, 166, 142, 123, 110, 98, 81, 69, 57, 41. Elemental Anal. Calcd for C₁₂H₁₉O₃N: C, 63.97; H, 8.50; N, 6.22. Found: C, 64.24; H, 8.39; N, 6.07.

4.2.2. (4\$,5\$)-3-(3-Methylbutanoyl)hexahydrobenzoxazolidin-2-one, (\$,\$)-5a

The general procedure described above was followed, and the crude product was purified by flash chromatography using ethyl acetate–hexane (80:20) as eluent to give 94% yield of the desired product, mp 90–91 °C. $[\alpha]_D^{25} = +72$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 2954, 2868, 1798, 1690, 1470, 1370, 1332, 1312, 1254, 1214. The ¹H and ¹³C NMR data were identical to those recorded for enantiomeric (*R*,*R*)-**5a**. MS (20 eV): *m*/*z* 225 (M⁺), 210, 183, 166, 142, 123, 110, 98, 81, 69, 57, 41. Elemental Anal. Calcd for C₁₂H₁₉O₃N: C, 63.97; H, 8.50; N, 6.22. Found: C, 63.62; H, 8.40; N, 6.12.

4.3. N-Acylation procedure for the preparation of 5b and 5c

To a solution of carboxylic acid **3** or **4** (10.4 mmol, 1.05 equiv) in THF (70 mL), were added Et₃N (27 mmol, 2.6 equiv) and pivaloyl chloride (10.8 mmol, 1.05 equiv) at -30 °C. The resulting white suspension was stirred at -30 °C for 90 min, before the addition of LiCl (11.9 mmol, 1.15 equiv) and hexahydrobenzoxazolidin-2-one **1** (10.4 mmol, 1 equiv) in THF (30 mL). The resulting mixture was allowed to warm to room temperature and stirred for 12 h. The mixture was treated with saturated aqueous NH₄Cl (100 mL), H₂O (50 mL), and EtOAc (150 mL). The organic phase was separated, washed with brine (100 mL), and each aqueous fraction was re-extracted with EtOAc (80 mL). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated under vacuum.

4.3.1. (4*R*,5*R*)-3-(4-Methylpentanoyl)hexahydrobenzoxazolidin-2-one, (*R*,*R*)-5b

The crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) to give 92% yield of the desired product, mp 68–69 °C. [α]_D²⁵ = -74 (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): ν_{max} 2952, 2868, 2372, 1796, 1692, 1472, 1378, 1338, 1310, 1268, 1214, 1148. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 3.89–3.80 (dt, 1H, J_1 = 3.6 Hz J_2 = 11.4 Hz), 3.56–3.47 (dt, 1H, J_1 = 3.3 Hz J_2 = 10.9 Hz), 2.97–2.74 (m, 3H), 2.23–2.17 (m, 1H), 1.93–1.82 (m, 2H), 1.68– 1.30 (m, 7H), 0.89 (d, 6H, J = 6.3 Hz). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 176.2, 155.1, 81.6, 63.5, 29.24, 28.81, 28.1, 24.1, 23.9, 22.7 (2 × C). MS (20 eV): m/z 240 (M + H)⁺, 196, 183, 152, 142, 110, 98, 81, 71, 55, 43. HR-ESI-TOF Calcd for C₁₃H₂₁O₃N [M+H]⁺: 240.15942; found: 240.15961.

4.3.2. (4*S*,5*S*)-3-(4-Methylpentanoyl)hexahydrobenzoxazolidin-2-one, (*S*,*S*)-5b

The crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give 98% yield of the desired product, mp 68–69 °C. $[\alpha]_D^{25} = +77$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): ν_{max} 2952, 2868, 2362, 1796, 1692, 1472, 1378, 1310, 1268, 1214. The ¹H and ¹³C NMR data were identical to those recorded for enantiomeric (*R*,*R*)-**5b**. MS (20 eV): *m*/*z* 240 (M+H)⁺, 196, 183, 152, 142, 110, 98, 81, 71, 55, 43. Elemental Anal. Calcd for C₁₃H₂₁O₃N: C, 65.25; H, 8.84; N, 5.85; found: C, 65.48; H, 9.21; N, 5.65.

4.3.3. (4*R*,5*R*)-3-{1-[(*tert*-Butoxy)carbonyl]-1*H*-indol-3-yl}-1-oxopropyl-hexahydrobenzoxazolidin-2-one, (*R*,*R*)-5c

Following the procedure described above, the crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give 76% yield of the desired product, mp 127–128 °C. [α]_D²⁵ = -28 (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): ν_{max} 2942, 2366, 1788, 1724, 1696, 1454, 1390, 1364, 1312, 1156. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 8.14 (d, 1H, *J* = 6.8 Hz), 7.6 (d, 1H, *J* = 7.7 Hz), 7.43 (s, 1H), 7.35–7.23 (m, 2H), 3.95–3.81 (dt, 1H, *J*₁ = 3.3 Hz, *J*₂ = 11.4 Hz), 3.61–3.54 (dt, 1H, *J*₁ = 3.2 Hz *J*₂ = 10.8 Hz), 3.45–3.34 (m, 1H), 3.27–3.13 (m, 1H), 3.10–2.99 (m, 2H), 2.85–2.82 (m, 1H), 2.25–2.18 (m, 1H), 1.95–1.75 (m, 2H), 1.68 (s, 10H), 1.50–1.25 (m, 3H). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 174.7, 154.8, 149.9, 135.6, 130.5, 124.5, 122.9, 122.6, 119.6, 119.1, 115.4, 83.5, 81.6, 63.3, 36.3, 28.9, 28.6, 28.4, 23.8, 23.7, 19.9; MS (20 eV): *m*/*z* 412 (M⁺) 412, 356, 312, 171, 130, 57. HR-ESI-TOF Calcd for C₂₃H₂₈O₅N₂ [M⁺+Na]: 435.188904; found: 435.18897.

4.3.4. (4*S*,5*S*)-3-{1-[(*tert*-Butoxy)carbonyl]-1*H*-indol-3-yl}-1-oxopropyl-hexahydrobenzoxazolidin-2-one, (*S*,*S*)-5c

Following the procedure described above, the crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give 73% yield of the desired product, mp 123–124 °C. $[\alpha]_D^{25} = +24$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 2936, 1796, 1724, 1694, 1458, 1384, 1352, 1308, 1260, 1210. The ¹H and ¹³C NMR data were identical to those recorded for enantiomeric (*R*,*R*)-**5c**. MS (20 eV): *m*/*z* 412 (M⁺), 412, 356, 312, 171, 130, 57. Elemental Anal. Calcd for C₂₃H₂₈O₅N₂: C, 66.97; H, 6.84; N, 6.79. Found: C, 67.05; H, 6.85; N, 6.70.

4.4. General procedure for the alkylation of 5a-c

At first, NaHMDS (1 M solution, 8.98 mmol, 1.1equiv) was added over 15 min to a cold (-78 °C) solution of compound **5** (8.16 mmol, 1 equiv) in THF (80 mL), and the resulting mixture was stirred for an additional 60 min at -78 °C before the slow addition of a solution of benzyl bromoacetate (16.3 mmol, 2 equiv) in THF (20 mL). The reaction mixture was stirred for 5 h at -78 °C, before the addition of saturated aqueous NH₄Cl (100 mL), and the

resulting mixture was warmed to room temperature, diluted with H_2O (50 mL) and EtOAc (2 \times 80 mL). The combined organic phases were dried over anhydrous Na_2SO_4 and concentrated in vacuo.

4.4.1. Benzyl-3(*R*)-(isopropyl)-4-[(4*R*,5*R*)-2-oxo-hexahydrobenz-oxazolidin-3-yl]-4-oxobuta-noate, (*R*,*R*,*P*)-6a

Following the general procedure, the crude product was purified by flash chromatography using hexane–ethyl acetate (90:10) as eluent to give 71% yield of the alkylated product, mp 61–63 °C. $[\alpha]_D^{25} = -5.6 \ (c \ 1.0, \text{CHCl}_3)$. IR (KBr, cm⁻¹): v_{max} 2954, 2874, 2358, 1784, 1740, 1688, 1386, 1334, 1186, 1038. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.92 (d, 3H, *J* = 6.9), 1.01 (d, 3H, *J* = 6.8), 1.34 (m, 3H), 1.62 (m, 1H), 1.83 (m, 2H), 2.10 (m, 2H), 2.83 (m, 3H), 3.59 (m, 2H), 3.83 (m, 1H), 5.06 (s, 2H), 7.36 (m, 5H). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 18.3, 21.3, 23.9, 24.1, 28.7, 29.0, 29.6, 33.4, 45.7, 63.7, 66.9, 81.7, 128.8, 129.0, 136.2, 154.8, 172.7, 177.2. MS (20 eV): *m/z* 373 (M⁺), 331, 319, 272, 266, 232, 204, 196, 161, 142, 107, 91, 70, 43. Elemental Anal. Calcd for C₂₁H₂₇O₅N: C, 67.54; H, 7.29; N, 3.75. Found: C, 67.86; H, 7.00; N, 3.72.

4.4.2. Benzyl-3(*S*)-(isopropyl)-4-[(4*S*,5*S*)-2-oxo-hexahydrobenz-oxazolidin-3-yl]-4-oxobutanoate, (*S*,*S*,*S*)-6a

Following the general procedure, the crude product was purified by flash chromatography using hexane–ethyl acetate (90:10) as eluent to give 64% yield of the alkylated product, mp 63–64 °C. $[\alpha]_D^{D} = +6.3$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 2956, 2876, 1784, 1738, 1688, 1464, 1386, 1334, 1186. The ¹H and ¹³C NMR data were identical to those recorded for enantiomeric (*R*,*R*,*P*)-**6a**. MS (20 eV): *m*/*z* 373 (M⁺), 266, 232, 204, 196, 142, 107, 91, 70. Elemental Anal. Calcd for C₂₁H₂₇O₅N: C, 67.54; H, 7.29; N, 3.75. Found: C, 67.67; H, 7.90; N, 3.53.

4.4.3. Benzyl-3(*S*)-(isobutyl)-4-[(4*R*,5*R*)-2-oxo-hexahydrobenz-oxazolidin-3-yl]-4-oxobutanoate, (*R*,*R*,*S*)-6b

Following the general procedure, the crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give 82% yield of the desired alkylated product, mp 60–62 °C. $[\alpha]_D^{25} = +11$ (*c* 1.0, MeOH). IR (KBr, cm⁻¹): ν_{max} 2958, 2362, 1790, 1724, 1688, 1458, 1382, 1312, 1268, 1210. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 7.39–7.27 (m, 5H), 5.08 (s, 2H), 4.02–3.93 (m, 1H), 3.68–3.60 (dt, 1H, J_1 = 3.5 Hz, J_2 = 11.4 Hz), 3.53–3.45 (dt, 1H, J = 3.1 Hz, J_2 = 11.0 Hz), 2.89–2.59 (m, 3H), 2.19–2.15 (m, 1H), 1.94–1.76 (m, 2H), 1.70–1.53 (m, 3H), 1.42–1.13 (m, 4H), 0.94–0.91 (dd, 6H, J = 1.8 Hz). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 178.0, 172.2, 154.7, 128.8, 128.7, 128.6, 81.7, 66.9, 63.7, 40.4, 39.1, 36.9, 28.7, 25.9, 24.1, 23.8, 21.8. MS (20 eV): *m/z* 387 (M⁺), 280, 246, 218, 156, 142, 107, 91. HR-ESI-TOF Calcd for C₂₂H₂₉O₅N [M+Na]⁺: 410.19379; found: 410.19372.

4.4.4. Benzyl-3(*R*)-(Isobutyl)-4-[(4*S*,5*S*)-2-oxo-hexahydrobenz-oxazolidin-3-yl]-4-oxobutanoate, (*S*,*S*,*R*)-6b

Following the general procedure, the crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give 69% yield of the desired product, mp 63–64 °C. $[\alpha]_D^{25} = -14$ (*c* 1.0, MeOH). IR (KBr, cm⁻¹): v_{max} 2960, 2370, 1792, 1724, 1688, 1460, 1382, 1312, 1268, 1210. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*S*)-**6b**. MS (20 eV): *m*/*z* 387 (M⁺), 280, 246, 218, 156,142, 107, 91. HR-ESI-TOF Calcd for C₂₂H₂₉O₅N [M+Na]⁺: 410.19379; found: 410.19392.

4.4.5. Benzyl-3(*S*)-({1-[(*tert*-butoxy)carbonyl]-1*H*-indol-3-yl}methyl)-4-[(4*R*,5*R*)-2-oxohexa-hydrobenzoxazolidin-3-yl]-4oxobutanoate, (*R*,*R*,*S*)-6c

Following the general procedure, the crude product was purified by flash chromatography using hexane–ethyl acetate (90:10) as eluent to give 83% yield of the desired alkylated product, mp 81–82 °C. [α]_D²⁵ = -31.7 (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): ν_{max} 2948, 1730, 1692, 1458, 1368, 1258, 1198. ¹H NMR (CDCl₃, 300 MHz) *δ* (ppm): 8.14 (d, 1H, *J* = 7.0 Hz), 7.89 (d, 1H, *J* = 7.15 Hz), 7.48 (s, 1H), 7.36–7.26 (m, 7H), 5.02 (s, 2H), 4.28–4.21 (m, 1H), 3.78–3.67 (dt, 1H, *J*₁ = 3.6 Hz, *J*₂ = 11.3 Hz), 3.61–3.54 (dt, 1H *J*₁ = 3.0 Hz, *J*₂ = 11.3 Hz), 3.40–3.34 (dd, 1H, *J* = 3.7 Hz), 2.99–2.89 (dd, 1H, *J* = 10.9 Hz), 2.77–2.48 (m, 4H), 2.22 (d, 1H, *J* = 10.6), 1.94–1.24 (m, 14H). ¹³C NMR (CDCl₃, 75.5 MHz) *δ* (ppm): 177.3, 172.1, 154.9, 150.0, 136.0, 130.6, 129.0, 128.8, 128.7, 124.9, 124.8, 123.1, 120.2, 117.3, 115.5, 83.9, 83.4, 66.7, 63.8, 41.0, 36.5, 28.8, 28.6, 27.1, 24.0, 23.9. MS (20 eV): *m*/*z* 560 (M⁺), 460, 369, 311, 200, 130, 91, 41. HR-ESI-TOF Calcd for C₃₂H₃₆O₇N₂ [M+Na]⁺: 583.24147; found: 583.24189.

4.4.6. Benzyl-3(*R*)-({1-[(*tert*-butoxy)carbonyl]-1*H*-indol-3-yl}methyl)-4-[(4*S*,5*S*)-2-oxohexa-hydrobenzoxazolidin-3-yl]-4oxobutanoate, (*S*,*S*,*R*)-6c

Following the general procedure, the crude product was purified by flash chromatography using hexane–ethyl acetate (90:10) as eluent to give 63% yield of the desired alkylated product, mp 83–84 °C. [α]₀²⁵ = +34 (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): ν_{max} 2948, 1798, 1730, 1692, 1458, 1368, 1258, 1198. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*S*)-**6***c*. MS (20 eV): *m*/*z* 560 (M⁺), 460, 369, 311, 228, 200, 130, 91, 41. Elemental Anal. Calcd for C₃₂H₃₆O₇N₂: C, 68.55; H, 6.47; N, 5.00. Found: C, 68.65; H, 6.75; N, 4.98.

4.5. General procedures for the hydrogenolysis of alkylated products 6

4.5.1. Method A

A solution of compound **6** (12 mmol) in EtOAc (100 mL) was stirred under H_2 (1 atm) in the presence of 10% of Pd/C for 12 h. The catalyst Pd/C was filtered off, washed with EtOAc, and the filtrate was concentrated in vacuo.

4.5.1.1. 3(R)-(Isopropyl)-4-[(4R,5R)-2-oxo-hexahydrobenzoxaz-

olidin-3-yl]-4-oxobutanoic acid, (*R*,*R*,*R*)-7**a**. The crude product was purified by flash chromatography using ethyl acetate–hexane (1:1) as eluent to give the desired product as a yellow oil in 98% yield. $[\alpha]_D^{25} = -25.4$ (*c* 1.0, CHCl₃); IR (KBr, cm⁻¹): v_{max} 2958, 1788, 1696, 1466, 1370, 1308, 1252, 1204, 1144, 1116. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 0.87 (d, 3H, *J* = 8.8), 0.99 (d, 3H, *J* = 6.8), 1.41 (m, 3H), 1.63 (m, 1H), 1.85 (m, 2H), 2.09 (m, 1H), 2.22 (m, 1H), 2.54 (dd, 1H, *J* = 17.4), 2.84 (dd, 2H, *J* = 17.4), 3.56 (ddd, 1H, *J*₁ \approx *J*₂ = 11.0, *J*₃ = 3.4), 3.74 (m, 1H), 3.89 (ddd, 1H, *J*₁ \approx *J*₂ = 11.7, *J*₃ = 3.6). ¹³C NMR (CDCl₃, 100.5 MHz) δ (ppm): 17.9, 20.9, 23.6, 23.9, 28.5, 28.8, 29.1, 32.7, 45.2, 63.6, 81.8, 154.6, 176.8, 178.8. MS (20 eV): *m*/*z* 283 (M⁺), 241, 224, 206, 179, 142, 124, 112, 98, 70, 55, 43. HR-ESI-TOF Calcd for C₁₄H₂₁O₅N [M+Na]⁺: 306.1311, found: 306.1303.

4.5.1.2. 3(S)-(Isopropyl)-4-[(4S,5S)-2-oxo-hexahydrobenzoxaz-

olidin-3-yl]-4-oxobutanoic acid, (*S*,*S*)-7a. The crude product was purified by flash chromatography using ethyl acetate–hexane (1:1) as eluent to give the desired product as a yellow oil in 93% yield. $[\alpha]_D^{25} = +25.5$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 2956, 1790, 1704, 1468, 1360, 1306, 1252, 1204, 1144, 1116. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*R*)-7a. MS (20 eV): *m*/*z* 283 (M⁺), 241, 224, 206, 179, 142, 124, 112, 98, 70, 55, 43. HR-ESI-TOF calcd for C₁₄H₂₁O₅N [M+Na]⁺: 306.1311; found: 306.1303.

4.5.1.3. 3(S)-(Isobutyl)-4-[(4R,5R)-2-oxo-hexahydrobenzoxaz-

olidin-3-yl]-4-oxobutanoic acid, (*R*,*R*,*S*)-7b. The crude product was purified by flash chromatography using hexane–ethyl acetate

(1:1) as eluent to give 95% yield of the desired hydrogenolyzed product, mp 98–100 °C. $[\alpha]_{D}^{25} = +6 (c \ 1, MeOH)$. IR (KBr, cm⁻¹): v_{max} 3308, 2954, 2876, 2362, 1790, 1738, 1676, 1400, 1350. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 11.3 (br s, 1H), 3.92–3.84 (m, 2H), 3.58–3.51 (dt, 1H, J_1 = 2.9 Hz, J_2 = 10.4), 2.81–2.55 (m, 3H), 2.23–2.16 (m, 1H), 1.93–1.82 (m, 2H), 1.69–1.62 (m, 3H), 1.45–1.20 (m, 4H), 0.91 (d, 6H, J = 5.8 Hz). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 178.5, 177.7, 154.5, 81.8, 63.5, 39.9, 38.6, 36.2, 28.6, 28.5, 25.7, 23.8, 23.6, 21.4. MS (20 eV): m/z 297 (M⁺), 254, 241, 192, 142, 98, 69, 56, 43. HR-ESI-TOF Calcd for C₁₅H₂₃O₅N [M+Na]⁺: 298.16490, found: 298.16461.

4.5.1.4. 3(R)-(Isobutyl)-4-[(45,55)-2-oxo-hexahydrobenzoxaz-

olidin-3-yl]-4-oxobutanoic acid, **(***S***,***S***,***R***)-7b**. The crude product was purified by flash chromatography using hexane–ethyl acetate (1:1) as eluent to give 93% yield of the desired hydrogenolyzed product, mp 98–99 °C. $[\alpha]_D^{25} = -8$ (*c* 1, MeOH). IR (KBr, cm⁻¹): v_{max} 3306, 2954, 2876, 2362, 1790, 1738, 1676, 1400, 1312. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*S*)-**7b**. MS (20 eV): *m*/*z* 297 (M⁺+1), 254, 241, 192, 142, 98, 69, 56, 43. Elemental Anal. Calcd for C₁₅H₂₃O₅N: C, 60.59; H, 7.80; N, 4.71. Found: C, 60.28; H, 7.99; N, 4.65.

4.5.2. Method B

A solution of compound **6** (4.20 mmol) in THF (50 mL) was stirred under H_2 (1 atm) in the presence of 10% Pd/C for 2 h. The catalyst Pd/C was filtered off, washed with THF, and the filtrate was concentrated in vacuo.

4.5.2.1. 3(*S*)-({1-[(*tert*-Butoxy)carbonyl]-1*H*-indol-3-yl}methyl)-4-[(4*R*,5*R*)-2-oxo-hexahydrobenzoxazolidin-3-yl]-4-oxobuta-

noic acid, (R,R,S)-7c. Following the described procedure, the residue was purified by crystallization from dichloromethane-hexane (1:20) to give the desired hydrogenolyzed product in 95% yield, mp 80–82 °C. $[\alpha]_{D}^{25} = -49$ (*c* 1, CHCl₃). IR (KBr, cm⁻¹): v_{max} 3450, 2942, 1786, 1730, 1456, 1256, 1156. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 8.12 (d, 1H, J = 7.3 Hz), 7.86 (d, 1H, J = 7.4 Hz), 7.47 (s, 1H), 7.34-7.24 (m, 2H), 4.19–4.07 (m, 1H), 4.00–3.87 (dt, 1H, J₁ = 32 Hz, $I_2 = 11.4 \text{ Hz}$), 3.65–3.61 (m, 1H), 3.40–3.34 (dd, 1H, $I_1 = 3.8 \text{ Hz}$, J_2 = 13.9 Hz), 2.94–2.77 (m, 2H), 2.67–2.59 (dd, 1H, J_1 = 10.9 Hz, $J_2 = 13.8 \text{ Hz}$), 2.51–2.44 (dd, 1H, $J_1 = 3.5 \text{ Hz}$, $J_2 = 17.5 \text{ Hz}$), 2.29– 2.18 (m, 1H), 1.19-1.84 (m, 2H), 1.68 (s, 10H), 1.44-1.35 (m, 3H). ^{13}C NMR (CDCl₃, 75.5 MHz) δ (ppm): 177.9, 176.9, 154.7, 149.8, 135.7, 130.4, 124.7, 124.6, 122.9, 119.9, 116.9, 115.3, 83.8, 82.1, 63.6, 40.7, 35.9, 28.5 (2 × C), 28.4 (2 × C), 26.7, 23.8, 23.7. MS (20 eV): *m*/*z* 470 (M⁺), 370, 273, 229, 130, 57, 41. HR-ESI-TOF Calcd for C₂₅H₃₀O₇N₂ [M+Na]⁺: 493.19452, found: 493.19495.

4.5.2.2. 3(*R*)-({1-[(*tert*-Butoxy)carbonyl]-1*H*-indol-3-yl}methyl)-

4-[(45,55)-2-oxohexahydrobenzoxazolidin-3-yl]-4-oxobutanoic acid, (*S,S,R*)-**7c**. The residue was purified by crystallization using dichloromethane–hexane (1:20) as eluent to give 95% yield of the desired product, mp 82–83 °C. $[\alpha]_{2}^{D5} = +33$ (*c* 1, CHCl₃). IR (KBr, cm⁻¹): v_{max} 3450, 2948, 1786, 1730, 1454, 1256, 1158. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R,R,S*)-**7c**. MS (20 eV): *m/z* 470 (M⁺), 370, 273, 229, 130, 57, 41. HR-ESI-TOF Calcd for C₂₅H₃₀O₇N₂ [M+Na]⁺: 493.19452, found: 493.19394.

4.6. General procedure for the preparation of compounds 8a-c via Curtius rearrangement of 7a-c

To a stirred solution of compound **7** (7.1 mmol, 1 equiv) in toluene (50 mL) and Et₃N (14.2 mmol, 2 equiv), were added diphenyl-phosphoryl azide [(PhO)₂P(O)N₃, 8.53 mmol, 1.2 equiv)] and BnOH (14.2 mmol, 2 equiv). The resulting mixture was stirred for 1 h at

room temperature and then heated at reflux for an additional 2 h. The toluene solvent was evaporated in vacuo and the residue was dissolved in EtOAc (150 mL) and 2 M HCl (70 mL). The organic phase was separated, washed with saturated solution NaHCO₃, dried over anhydrous Na₂SO₄, and concentrated in vacuo.

4.6.1. (4*R*,5*R*)-3-[2(*S*)-({[(Benzyloxy)carbonyl]amino}methyl)-3-methyl-butanoyl] hexahydrobenzoxazolidin-2-one, (*R*,*R*,*S*)-8a

The crude product was purified by flash chromatography using dichloromethane–ethyl acetate (90:10) as solvent to give the desired product in 70% yield, mp 72–73 °C. $[\alpha]_D^{25} = -8$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 3356, 2962, 2870, 1764, 1700, 1528, 1260. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 0.97 (d, 3H, *J* = 3.1), 0.98 (d, 3H, *J* = 3.3), 1.35 (m, 3H), 1.59 (m, 1H), 1.78 (m, 1H), 1.88 (m, 1H), 2.07 (m, 1H), 2.17 (m, 1H), 2.75 (d, 1H, *J* = 10.8), 3.48 (m, 3H), 3.72 (m, 2H), 4.96 (s, 1H), 5.08 (dd, 2H, *J*₁ = 19.4 Hz, *J*₂ = 12.2 Hz), 7.34 (m, 5H). ¹³C NMR (CDCl₃, 100.5 MHz) δ (ppm): 19.1, 21.2, 23.5, 23.8, 28.4, 28.8, 41.0, 50.2, 63.3, 66.7, 81.2, 128.3, 128.6, 163.7, 154.5, 156.3, 177.2. MS (20 eV): *m/z* 388 (M), 297, 281, 247, 210, 166, 142, 112, 91, 70; Elemental Anal. Calcd for C₂₁H₂₇O₅N: C, 64.93; H, 7.27; N, 7.21; found: C, 65.25; H, 7.27; N, 7.00.

4.6.2. (45,55)-3-[2(R)-({[(Benzyloxy)carbonyl]amino}methyl)-3methylbutanoyl] hexahydrobenzoxazolidin-2-one, (*S*,*S*,*R*)-8a

The crude product was purified by flash chromatography using dichloromethane–ethyl acetate (90:10) as eluent to give the desired product in 61% yield, mp 72–73 °C. $[\alpha]_D^{25} = +9$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 3356, 2962, 2870, 1764, 1708, 1528, 1260. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*S*)-**8a**. MS (20 eV): *m*/*z* 388 (M⁺), 297, 281, 247, 210, 166, 142, 112, 91, 70. Elemental Anal. Calcd for C₂₁H₂₇O₅N: C, 64.93; H, 7.27; N, 7.21. Found: C, 65.23; H, 7.57; N, 7.16.

4.6.3. (4*R*,5*R*)-3-[2(*S*)-({[(Benzyloxy)carbonyl]amino}methyl)-4methyl-pentanoyl] hexahydrobenzoxazolidin-2-one, (*R*,*R*,*S*)-8b

The crude product was purified by flash chromatography using dichloromethane–ethyl acetate (90:10) as eluent to give the desired product in 64% yield, mp 72–73 °C. [α]_D²⁵ = +24 (*c* 1.0, MeOH). IR (KBr, cm⁻¹): ν_{max} 3338, 2938, 2872, 2374, 1796, 1720, 1686, 1530, 1232, 1144, 1036. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 7.34–7.27 (m, 5H), 5.08 (s, 2H), 4.03–3.95 (m, 1H), 3.76–3.67 (dt, 1H, J_1 = 3.4 Hz, J_2 = 11.4 Hz), 3.56–3.31 (m, 3H), 2.70–2.65 (m, 1H), 2.19–2.18 (m, 1H), 1.89–1.53 (m, 5H), 1.38–1.19 (m, 4H), 0.91 (t, 6H, J = 5.7 Hz). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 177.1, 156.3, 154.6, 136.7, 128.6, 128.2, 81.3, 66.7, 63.4, 43.4, 42.6, 37.6, 28.5, 25.8, 23.8, 23.5, 22.9, 22.5. MS (20 eV): *m/z* 402 (M⁺), 346, 295, 261, 208, 196, 126, 107, 91. HR-ESI-TOF Calcd for C₂₂H₃₀O₅N₂ [M+Na]⁺: 425.20469; found: 425.20488.

4.6.4. (4*S*,5*S*)-3-[2(*R*)-({[(Benzyloxy)carbonyl]amino}methyl)-4-methyl-pentanoyl] hexahydrobenzoxazolidin-2-one, (*S*,*S*,*R*)-8b

The crude product was purified by flash chromatography using dichloromethane–ethyl acetate (90:10) as eluent to give the desired product in 82% yield, mp 74–75 °C. [α]_D²⁵ = -28 (*c* 1.0, MeOH). IR (KBr, cm⁻¹): ν_{max} 3338, 2936, 2872, 2364, 1796, 1720, 1684, 1530, 1232, 1144, 1036. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*S*)-**8b**. MS (20 eV): *m*/*z* 402 (M⁺), 346, 295, 261, 208, 196, 126, 107, 91. HR-ESI-TOF calcd for C₂₂H₃₀O₅N₂ [M+Na]⁺: 425.20469; found: 425.20518.

4.6.5. (4*R*,5*R*)-3-[2(*S*)-({[(Benzyloxy)carbonyl]amino}methyl)-3-({1-[(*tert*-butoxy)carbonyl]-1*H*-indol-3-yl}-1-oxo-propyl)hexahydrobenzoxazolidin-2-one, (*R*,*R*,*S*)-8c

The crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give the desired product in 88% yield, mp 99-100 °C. $[\alpha]_{D}^{25} = +7.4$ (*c* 1.01, Acetone). IR (KBr,

cm⁻¹): ν_{max} 3410, 3346, 2944, 2366, 1790, 1732, 1456, 1368. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 8.12 (d, 1H, *J* = 7.3 Hz), 7.65 (d, 1H, *J* = 7.4 Hz), 7.35–7.19 (m, 7H), 5.07 (s, 2H), 4.38–4.29 (m, 1H), 3.80–3.71 (dt, 1H, *J*₁ = 11.4, *J*₂ = 3.3), 3.58–3.41 (m, 3H), 3.22 (dd, 1H, *J*₁ = 14.7, *J*₂ = 5.9 Hz), 2.86–2.71 (m, 2H), 2.18 (m, 2H), 1.91–1.50 (m, 12H), 1.43–1.14 (m, 3H). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 175.9, 156.4, 154.7, 149.8, 136.7, 135.6, 128.7, 128.2, 124.6, 124.1, 122.8, 119.3, 117.2, 116.9, 115.3, 83.7, 81.6, 66.8, 63.5, 44.3, 42.6, 28.6, 28.5 (2 × C), 28.3, 24.3, 23.7. MS (20 eV): *m*/*z* 575 (M⁺) 475, 367, 324, 170, 130, 41. HR-ESI-TOF Calcd for C₃₂H₃₇O₇N₃ [M+Na]⁺: 598.25237; found: 598.25346.

4.6.6. (4*S*,5*S*)-3-[2(*R*)-({[(Benzyloxy)carbonyl]amino}methyl)-3-({1-[(*tert*-Butoxy)carbonyl]-1*H*-indol-3-yl}-10xo-propyl)hexahydrobenzoxazolidin-2-one, (*S*,*S*,*R*)-8c

The crude product was purified by flash chromatography using hexane–ethyl acetate (80:20) as eluent to give the desired product in 60% yield, mp 100–101 °C. [α]_D²⁵ = -5.5 (*c* 1.0, acetone); IR (KBr, cm⁻¹): ν _{max} 3410, 3346, 2944, 2362, 1792, 1730, 1522, 1458, 1372. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*R*,*R*,*S*)–**8c**. MS (20 eV): *m*/*z* 575 (M⁺), 475, 367, 324, 170, 130, 41. Elemental Anal. Calcd for C₃₂H₃₇O₇N₃: C, 66.77; H, 6.48; N, 7.30. Found: C, 67.10; H, 6.31; N, 7.27.

4.7. General procedures for the preparation of derivatives 9

4.7.1. Method A

The reaction was carried out in two steps:

Flask A: To a stirred solution of LiOH·H₂O (1.5 mmol, 2 equiv) in 8 mL of a mixture of THF–H₂O (1:1) was added H₂O₂ (30% aq solution; 2.98 mmol, 4 equiv) at 0 °C. The resulting mixture was stirred at 0 °C for 3 min.

Flask B: A stirred solution of compound **8** (0.75 mmol, 1 equiv) in THF (8 mL) was cooled to 0 °C before the solution previously prepared in flask A was added via cannula. The reaction mixture was stirred at 0 °C for 30 min before the addition of NaSO₃ (2.68 mmol, 4 equiv) in H₂O (4 mL). The mixture was stirred a 0 °C for 30 additional minutes, treated with H₂O (10 mL), and extracted with EtOAc (2 × 20 mL). The aqueous phase was acidified to pH 2 with 1 M HCl at 0 °C. A white solid precipitated, which was extracted with EtOAc (3 × 25 mL), the organic phase was separated, washed with a saturated solution of sodium and potassium tartrate, and dried over anhydrous Na₂SO₄ and concentrated in vacuo.

4.7.1.1. 2(S)-([{(Benzyloxy)carbonyl}amino]methyl)-3-methyl-

butanoic acid, (*S*)-9a. The general procedure (Method A) was followed and the crude product was purified by flash chromatography using dichloromethane–methanol (90:10) as eluent to give the desired product as a colorless oil in 53% yield. $[\alpha]_D^{25} = +18$ (c 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 3332, 3034, 2964, 1728, 1724, 1720, 1586, 1266. ¹H NMR (DMSO-*d*₆, 270 MHz, 120 °C) δ (ppm): 0.93 (d, 3H, *J* = 6.9), 0.96 (d, 3H, *J* = 6.9), 1.89 (ddd, 1H, *J*₁ \approx *J*₂ = 13.3, *J*₃ = 6.7), 2.35 (m, 1H), 3.24 (m, 2H), 5.04 (s, 2H), 6.62 (s, 1H), 7.33 (m, 5H). ¹³C NMR (DMSO-*d*₆, 67.9 MHz, 120 °C) δ (ppm): 179.8, 158.1, 136.3, 128.5, 128.1, 127.8, 67.4, 52.4, 40.5, 28.7, 20.2 (2 × C). MS (20 eV): *m*/*z* 265 (M⁺), 250, 247, 173, 141, 108, 91, 70. HR-ESI-TOF: Calcd for C₁₄H₁₉O₄N [M+Na]⁺: 288.12063; found: 288.12027.

4.7.1.2. 2(*R*)-([{(Benzyloxy)carbonyl}amino]methyl)-3-methylbutanoic acid, (*R*)-9a. The general procedure (Method A) was followed and the crude product was purified by flash chromatography using dichloromethane–methanol (90:10) as eluent to give 36% yield of the desired product as a colorless oil. $[\alpha]_D^{25} = -18.8$ (*c* 1.0, CHCl₃). IR (KBr, cm⁻¹): v_{max} 3332, 3034, 2964, 1724, 1720,

1526, 1266. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**9a**. MS (20 eV): m/z 265 (M⁺), 250, 247, 173, 141, 108, 91, 70. HR-ESI-TOF: Calcd for C₁₄H₁₉O₄N [M + Na]⁺: 288.12063; found: 288.12089.

4.7.1.3. 2(S)-([{(Benzyloxy)carbonyl}amino]methyl)-4-methyl-

pentanoic acid, (*S***)-9b.** The general procedure (Method A) was followed and the crude product was purified by flash chromatography using dichloromethane–methanol (90:10) as eluent to give 70% yield of the desired product, mp 74–76 °C. $[\alpha]_D^{25} = +12.5$ (*c* 1.0, MeOH). IR (KBr, cm⁻¹): v_{max} 3348, 3186, 2956, 2360, 1718, 1536, 1462. ¹H NMR (DMSO-*d*₆, 300 MHz, 100 °C) δ (ppm): 7.61–7.28 (m, 5H), 6.8 (br s, 1H), 5.04 (s, 2H), 3.29–3.10 (m, 2H), 2.60–2.49 (m, 1H), 1.66–1.23 (m, 3H), 0.87 (d, 6H, *J* = 6.5 Hz). ¹³C NMR (DMSO-*d*₆, 75.5 MHz, 100 °C) δ (ppm): 176.3, 156.9, 138.2, 129.1, 128.4, 128.3, 66.2, 44.6, 39.5, 26.5, 23.5, 22.8. MS (20 eV): *m/z* 279 (M⁺), 261, 223, 115, 108, 91, 79, 57, 43. HR-ESI-TOF: Calcd for C₁₅H₂₁O₄N [M+H]⁺: 280.15433; found: 280.15414.

4.7.1.4. 2(R)-([{(Benzyloxy)carbonyl}amino]methyl)-4-methyl-

pentanoic acid, (*R***)-9b.** The general procedure (Method A) was followed and the crude product was purified by flash chromatography using dichloromethane–methanol (90:10) as eluent to give the desired product in 63% yield, mp 88–89 °C. $[\alpha]_D^{25} = -11$ (*c* 1.0, MeOH). IR (KBr, cm⁻¹): v_{max} 3310, 3178, 2956, 2366, 1720, 1550, 1462. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**9b.** MS (20 eV): *m*/*z* 279 (M⁺), 261, 223, 115, 108, 91, 79, 57, 43. HR-ESI-TOF: Calcd for C₁₅H₂₁O₄N [M+H]⁺: 280.15433; found: 280.15473.

4.7.2. Method B

The reaction was carried out in two steps:

Flask A: To a stirred solution of benzyl alcohol (4.1 mmol, 2.36 equiv) in THF (7 mL) at -78 °C was added *n*-BuLi (2.21 M in hexane, 2.71 mmol, 1.56 equiv) and the resulting mixture was stirred for 5 min.

Flask B: The solution mixture prepared in flask A was added at -78 °C to a stirred solution of compound **8** (1.74 mmol, 1 equiv) in THF (20 mL). The reaction mixture was stirred for 15 min at -78 °C and then the temperature was allowed to rise to 0 °C. Stirring was continued for 2 h before saturated aqueous NH₄Cl (20 mL) was added, and the mixture was diluted with H₂O (5 mL) and EtOAc (60 mL). The organic phase was separated and washed with brine solution, which was re-extracted with EtOAc (2 × 30 mL). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated in vacuo.

4.7.2.1. Benzyl-2(S)-({[(Benzyloxy)carbonyl]amino}methyl)-3-{1-[(tert-butoxy)carbonyl]-1H-indol-3yl}-propanoate, (S)-9c. The general procedure (Method B) was followed and the crude product was purified by flash chromatography using hexane-ethyl acetate (60:40) as eluent to give the desired product as an oil in 47% yield. $[\alpha]_{D}^{25} = +6.5$ (*c* 1.85, MeOH). IR (KBr, cm⁻¹): v_{max} 3362, 2930, 2360, 1732, 1518, 1454, 1370, 1160. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 8.15 (bs, 1H), 7.51-7.18 (m, 14H), 5.09 (d, 2H, J = 2.2 Hz), 5.04 (s, 2H), 3.60-3.41 (m, 2H), 3.18-3.06 (m, 2H), 2.98-2.90 (m, 1H), 1.66 (s, 9H). $^{13}\mathrm{C}$ NMR (CDCl_3, 100.5 MHz) δ (ppm): 174.18, 156.5, 149.7, 136.5, 135.6, 130.3, 128.6, 128.4, 128.3, 128.2, 128.1, 124.6, 123.8, 122.6, 118.9, 116.9, 115.4, 83.6, 66.9, 66.8, 45.8, 42.3, 28.3, 25.3. MS (20 eV): m/z 542 (M⁺), 442, 378, 334, 290, 243, 200, 130, 91, 57. HR-ESI-TOF: Calcd for C₃₂H₃₄O₆N₂ [M+H]⁺: 543.24896; found: 543.24867.

4.7.2.2. Benzyl-2(R)-({[(benzyloxy)carbonyl]amino}methyl)-3-{1-[(*tert***-butoxy)carbonyl]-1***H***-indol-3yl}-propanoate, (***R***)-9c. The general procedure (Method B) was followed and the crude product** was purified by flash chromatography using hexane–ethyl acetate (60:40) as eluent to give the desired product as an oil in 31% yield. $[\alpha]_D^{25} = -8.65$ (*c* 1.85, MeOH). IR (KBr, cm⁻¹): v_{max} 3368, 3032, 2976, 2936, 1728, 1520, 1452, 1372, 1254. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**9c**. EM (20 eV): m/z 542 (M⁺), 442, 378, 334, 290, 243, 200, 130, 91, 57. HRMS (FAB): Calcd for C₃₂H₃₄O₆N₂ (M⁺): 542.2410; found: 542.2417.

4.8. General procedure for the preparation of $\beta\mbox{-amino}$ acids 10a–c

The corresponding substrate **9** (4.26 mmol,1 equiv) was dissolved in EtOH (75 mL) and stirred with 10% Pd/C under an H₂ atmosphere (1 atm) at room temperature for 4 h. The catalyst Pd/ C was filtered off, washed with EtOH, and the filtrate was concentrated in vacuo.

4.8.1. 2(S)-([Amino]methyl)-3-methylbutanoic acid, (S)-10a

The general procedure was followed and the crude product was purified by flash chromatography using isopropyl alcohol–methanol–ammonium hydroxide (5:2:1) as eluent to give the desired product in 89% yield, mp 228–230 °C. $[\alpha]_D^{25} = +13 \ (c \ 1.0, \ H_2O)$ for 98% ee, Chirobiotic T, MeOH–H₂O (80:20), 1 mL/min, 205 nm, t = 8.18 min; [Lit.²⁰ mp 228–230 °C. $[\alpha]_D^{25} = -11.4 \ (c \ 1.0, \ H_2O)$, (*R*)]. IR (KBr, cm⁻¹): v_{max} 2968, 2664, 2204, 1550, 1408. NMR ¹H (D₂O, 270 MHz) δ (ppm): 0.89 (d, 3H, J = 6.7), 0.93 (d, 3H, J = 6.9), 1.94 (ddd, 1H, $J_1 \approx J_2 = 13.5$, $J_3 = 6.9$), 2.3 (m, 1H), 3.09 (m, 2H); NMR ¹³C (D₂O, 67.9 MHz) δ (ppm): 18.7, 20.0, 28.6, 39.0, 52.0, 180.5. EM (20 eV): m/z 131 (M⁺), 89, 83, 70, 55, 44; HR-ESI-TOF: Calcd for C₆H₁₃O₂N [M+H]⁺: 132.10191, found: 132.10228.

4.8.2. 2(R)-([Amino]methyl)-3-methylbutanoic acid, (R)-10a

The general procedure was followed and the crude product was purified by flash chromatography using isopropyl alcohol–methanol–ammonium hydroxide (5:2:1) as eluent to give the desired product in 70% yield, mp 238–240 °C. $[\alpha]_D^{25} = -11.4$ (c 1.0, H₂O) for 100% ee, Chirobiotic T, MeOH–H₂O (80:20), 1 mL/min., 205 nm, t = 7.27 min, [Lit.²⁰ mp 228–230 °C. $[\alpha]_D^{25} = -11.4$ (c 1.0, H₂O)]. IR (KBr, cm⁻¹): v_{max} 2968, 2664, 2204, 1550, 1408. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**10a**. MS (20 eV): m/z 131 (M⁺), 89, 87, 70, 55, 41. HR-ESI-TOF: Calcd for C₆H₁₃O₂N [M+H]⁺: 132.10191; found: 132.10211.

4.8.3. 2(S)-([Amino]methyl)-4-methylpentanoic acid, (S)-10b

The general procedure was followed and the crude product was purified by flash chromatography using isopropyl alcohol–methanol–ammonium hydroxide (5:2:1) as eluent to give the desired product in 81% yield, mp 226–228 °C. $[\alpha]_D^{25} = -4$ (*c* 1.0, H₂O) for 94% ee, Chirobiotic T, MeOH–H₂O (80:20), 1 mL/min, 205 nm, *t* = 10.37 min. [Lit.²⁰ mp 219–220 °C, $[\alpha]_D^{25} = +6.1$]. IR (KBr, cm⁻¹): v_{max} 3424, 2958, 2638, 2178, 1630, 1580, 1496. ¹H NMR (D₂O, 300 MHz) δ (ppm): 3.15–2.95 (m, 2H), 2.72–2.63 (m, 1H), 1.52–1.39 (m, 2H), 1.29–1.21 (m, 1H), 0.74–0.71 (dd, 6H, *J*₁ = 5.9, *J*₂ = 3.7 Hz). ¹³C NMR (D₂O, 75.5 MHz) δ (ppm): 177.7, 41.4, 40.5, 25.4, 21.9, 21.7. MS (20 eV): *m/z* 146 (M⁺+H), 128, 102, 89, 73, 55, 43. HR-ESI-TOF: Calcd for C₇H₁₅O₂N [M+H]⁺: 146.11756; found: 146.11797.

4.8.4. 2(R)-([Amino]methyl)-4-methylpentanoic acid, (R)-10b

The general procedure was followed and the crude product was purified by flash chromatography using isopropyl alcohol–methanol–ammonium hydroxide (5:2:1) as eluent to give the desired product in 92% yield, mp 224–226 °C. $[\alpha]_D^{25} = +8$ (*c* 1.0, H₂O) for 99% ee, Chirobiotic T, MeOH–H₂O (80:20), 1 mL/min, 205 nm, *t* = 7.55 min. IR (KBr, cm⁻¹): v_{max} 2958, 2638, 2178, 1630, 1582, 1496. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**10b**. MS (20 eV): m/z 146 (M⁺+H), 128, 102, 89, 73, 55, 43. HR-ESI-TOF: Calcd for $C_7H_{15}O_2N$ [M+H]⁺: 146.11756; found: 146.11780.

4.8.5. 2(*S*)-([Amino]methyl)-3-{1-[(*tert*-butoxy)carbonyl]-1*H*-indol-3-yl}propionic acid, (*S*)-10c

The general procedure was followed and the crude product was purified by silica flash chromatography using isopropyl alcoholmethanol-ammonium hydroxide (6:1:1) as eluent to give the desired product in 65% yield, mp 168–170 °C. [α]_D²⁵ = +7.4 (*c* 0.95, MeOH) for 82% ee, Chirobiotic T, MeOH-H₂O (80:20), 1 mL/min, 230 nm, *t* = 12.23 min. IR (KBr, cm⁻¹): *v*_{max} 3428, 3108, 2972, 2932, 2360, 1728, 1580, 1452, 1256. ¹H NMR (D₂O, 400 MHz) δ (ppm): 7.55–7.21 (m, 2H), 7.16 (s, 1H), 6.92–6.87 (m, 2H), 3.04– 2.92 (m, 3H), 2.80–2.61 (m, 2H), 1.26 (s, 9H), 1.09 (s, 1H). ¹³C NMR (D₂O, 100.5 MHz) δ (ppm): 175.9, 150.6, 84.8, 42.9, 40.1, 27.6, 24.9. MS (20 eV): *m/z* 318 (M⁺), 233, 201, 189, 156, 130, 117, 57, 41. HR-ESI-TOF: Calcd for C₁₇H₂₂O₄N₂ [M+H]⁺: 319.16523; found: 319.16597.

4.8.6. 2(*R*)-([Amino]methyl)-3-{1-[(*tert*-butoxy)carbonyl]-1*H*-indol-3-yl}propionic acid, (*R*)-10c

The general procedure was followed and the crude product was purified by flash chromatography using isopropyl alcohol–methanol–ammonium hydroxide (6:1:1) as eluent to give the desired product in 81% yield, mp 160–162 °C. $[\alpha]_{25}^{D} = -5.9$ (*c* 1.19, MeOH) for 86% ee, Chirobiotic T, MeOH–H₂O (80:20), 1 mL/min, 230 nm, t = 9.10 min. IR (KBr, cm⁻¹): v_{max} 3422, 2974, 2932, 2362, 1790, 1576, 1456, 1370, 1260. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**10c.** MS (20 eV): *m/z* 318 (M⁺), 233, 201, 189, 156, 130, 117, 57, 41. HR-ESI-TOF: Calcd for C₁₇H₂₂O₄N₂ [M+H]⁺: 319.16523; found: 319.16597.

4.9. General procedure for the preparation of derivatives 11a-c

The corresponding amino acid (7.17 mmol, 1 equiv) was suspended in 0.15 M aqueous Na₂CO₃ (14.33 mmol, 2 equiv). The suspension was treated with a solution of Fmoc-OSu (8.6 mmol, 1.2 equiv) in acetone (95 mL) to obtain a homogeneous solution. The solution was stirred for 20 h at room temperature. The solvent was partially evaporated in vacuo and diluted with H₂O (20 mL) and Et₂O (100 mL). The aqueous phase was acidified to pH 2 with 1 M HCl and extracted with EtOAc (2×100 mL). The organic phases were combined, dried over anhydrous Na₂SO₄, and concentrated in vacuo.

4.9.1. 2(S)-[{[(9H-Fluoren-9-ylmethoxy)carbonyl]amino}methyl]-3-methylbutanoic acid, (S)-11a

The general procedure was followed and the crude product was purified by flash chromatography using methanol–dichloromethane (5:95) as eluent to give the desired product in 40% yield, mp 121–123 °C. $[\alpha]_D^{25} = +22$ (*c* 1.0, MeOH) for 99% ee, Chiralcel OD, *i*-PrOH–hexane–TFA; 10:90:0.1, 1 mL/min, 254 nm, *t* = 69.77 min; [Lit.²¹ mp 112 °C, $[\alpha]_D^{25} = +11.5$ (*c* 0.97, CHCl₃)]. IR (KBr, cm⁻¹): ν_{max} 3342, 2964, 1700, 1542, 1448, 1268, 1140. ¹H NMR (CD₃OD, 400 MHz) δ (ppm): 0.95 (d, 3H, *J* = 6.8), 0.99 (d, 3H, *J* = 7), 1.88 (ddd, 1H, $J_1 \approx J_2 = 13.5$, $J_3 = 6.8$ Hz), 2.41 (m, 1H), 3.3 (m, 2H), 4.17 (m, 1H), 4.29 (m, 2H), 4.93 (s, 2H), 7.34 (m, 4H), 7.61 (d, 2H, *J* = 7.4), 7.75 (d, 2H, *J* = 7.4). ¹³C NMR (CD₃OD, 100.5 MHz) δ (ppm): 19.0, 19.4, 28.5, 40.4, 40.5, 47.1, 66.5, 119.6, 124.98, 126.8, 127.4, 141.2, 144.0, 157.4, 176.3.

4.9.2. 2(*R*)-[{(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}methyl]-3-methylbutanoic acid, (*R*)-11a

The general procedure was followed and the crude product was purified by flash chromatography using methanol–dichloromethane (5:95) as eluent to give the desired product in 41% yield, mp 121–123 °C. $[\alpha]_D^{25} = -23$ (*c* 1.0, MeOH) for 95% ee, Chiralcel OD, *i*-PrOH–hexane–TFA; 10:90:0.1, 1 mL/min, 254 nm, *t* = 108.97 min. [Lit.²² $[\alpha]_D^{25} = -12.1$ (*c* 0.97, CHCl₃)]. IR (KBr, cm⁻¹): v_{max} 3342, 2964, 1700, 1540, 1266. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**11a**.

4.9.3. 2(*S*)-[{[(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}methyl]-4-methylpentanoic acid, (*S*)-11b

The general procedure was followed and the crude product was purified by flash chromatography using methanol–dichloromethane (5:95) as eluent to give the desired product in 41% yield, mp 138–140 °C. $[\alpha]_D^{25} = +11.6$ (*c* 0.6, MeOH) for 96% ee, Chiralcel OD, *i*-PrOH–hexane–TFA; 10:90:0.1, 1 mL/min, 254 nm, *t* = 69.75 min. [Lit.²¹ mp 134 °C, $[\alpha]_D^{25} = +10.8$ (*c* 0.6, CHCl₃)]. IR (KBr, cm⁻¹): ν_{max} 3066, 2956, 1550, 1448, 1370, 1164, 1010. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 10.6 (br s, 1H), 7.77–7.31 (m, 8H), 5.3 (br s, 1H), 4.51–4.22 (m, 3H), 3.47–3.12 (m, 2H), 2.76–2.65 (m, 1H), 1.74–1.22 (m, 3H), 0.95 (d, 6H, *J* = 4.4 Hz). ¹³C NMR (CDCl₃, 100.5 MHz) δ (ppm): 180.8, 156.6, 144.0, 127.8, 127.2, 125.2, 120.1, 66.9, 47.3, 43.8, 42.3, 38.6, 25.9, 22.6.

4.9.4. 2(*R*)-[{[(9*H*-Fluoren-9-ylmethoxy)carbonyl]amino}methyl]-4-methylpentanoic acid, (*R*)-11b

The general procedure was followed and the crude product was purified by flash chromatography using methanol–dichloromethane (5:95) to give the desired product in 46% yield, mp 134–135 °C. $[\alpha]_D^{25} = -8.3$ (*c* 0.6, MeOH) for 99% ee, Chiralcel OD, *i*-PrOH–hexane–TFA; 10:90:0.1, 1 mL/min, 254 nm, *t* = 129.72 min. IR (KBr, cm⁻¹): v_{max} 3066, 2956, 1448, 1368, 1164, 1082. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**11b**.

4.9.5. (*S*)-3-({[(Fluoren-9-yl)methoxy]carbonyl}amino)-2-({1-([*tert*-butoxy]carbonyl)-1*H*-indol-3-yl}methyl)propionic acid, (*S*)-11c

The general procedure was followed and the crude product was purified by flash chromatography using methanol–dichloromethane (5:95) as eluent to give the desired product in 65% yield, mp 128–130 °C. $[\alpha]_D^{25} = +12.5$ (*c* 1, CHCl₃) for 90% ee, Chiralcel OD-H, *i*-PrOH–hexane–TFA; 20:80:0.1, 1 mL/min, 210 nm, *t* = 24.8 min. [Lit.^{6c} mp 125–128 °C. $[\alpha]_D^{25} = -5.0$ (*c* 1.04, CHCl₃) for the enantiomer]. IR (KBr, cm⁻¹): v_{max} 3340, 3062, 2978, 2928, 2360, 1550, 1728, 1606, 1452, 1372, 1256, 1158, 1090, 856. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 8.07 (d, 1H, *J* = 7.9 Hz), 7.71 (d, 2H, *J* = 7.3 Hz), 7.59–7.43 (m, 4H), 7.34–7.14 (m, 6H), 4.9 (s, 2H), 4.28 (d, 2H, *J* = 6.8 Hz), 4.12 (t, 1H, *J* = 6.7 Hz), 3.47–3.25 (m, 2H), 3.05–2.74 (m, 3H), 1.61 (s, 9H). ¹³C NMR (CDCl₃, 75.5 MHz) δ (ppm): 157.9, 150.1, 144.2, 141.5, 135.8, 130.8, 127.7, 127.1, 125.2, 124.4, 123.5, 122.6, 119.9, 119.1, 118.5, 115.1, 83.6, 66.8, 47.4, 42.7, 27.4, 26.9, 25.2.

4.9.6. (*R*)-3-({[(Fluoren-9-yl)methoxy]carbonyl}amino)-2-({1-([*tert*-butoxy]carbonyl)-1*H*-indol-3-yl}methyl)propionic acid, (*R*)-11c

The general procedure was followed and the crude product was purified by flash chromatography using methanol–dichloromethane (5:95) as eluent to give the desired product in 58% yield, mp 128–130 °C. $[\alpha]_D^{25} = -18.5$ (*c* 1, CHCl₃) for 88% ee, Chiralcel OD-H, *i*-PrOH–hexane–TFA; 20:80:0.1, 1 mL/min, 210 nm, *t* = 40.38 min. IR (KBr, cm⁻¹): v_{max} 3410, 2974, 2934, 2360, 1724, 1574, 1450, 1374, 1258, 1158, 1090, 856. The ¹H and ¹³C NMR data were identical to those recorded in enantiomeric (*S*)-**11c**.

Acknowledgments

The authors are grateful to Rodrigo González-Olvera and Jorge Vargas-Caporali for technical assistance. The authors are also in-

debted to Conacyt, México, for financial support via grant 60366-Q.

References

- (a) Juaristi, E.; Quintana, D.; Escalante, J. Aldrichim. Acta 1994, 27, 3–11; (b) Cole, D. C. Tetrahedron 1994, 50, 9517–9582; (c) Cardillo, G.; Tomasini, C. Chem. Soc. Rev. 1996, 117–128; (d)Enantioselective Synthesis of β-Amino Acids; Juaristi, E., Ed.; Wiley-VCH: New York, 1997; (e) Juaristi, E.; López-Ruiz, H. Curr. Med. Chem. 1999, 6, 983–1004; (f) Abele, S.; Seebach, D. Eur. J. Org. Chem. 2000, 1–15; (g) Fülöp, F. Chem. Rev. 2001, 101, 2181–2204; (h) Liu, M.; Sibi, M. P. Tetrahedron 2002, 58, 7991–8035; (i) Ma, J.-A. Angew. Chem., Int. Ed. 2003, 42, 4290–4299; (j) Córdova, A. Acc. Chem. Res. 2004, 37, 102–112; (k)Enantioselective Synthesis of β-Amino Acids; Juaristi, E., Soloshonok, V. A., Eds., 2nd ed.; Wiley-VCH: Hoboken, NJ, 2005; (l) Dondoni, A.; Massi, A. Acc. Chem. Res. 2006, 39, 451–463; (m)Synthesis of Non-Natural Amino Acids; Ager, D. J., Ed., 2nd ed.Handbook of Chiral Chemicals; DSM Pharma Chemicals: Raleigh, NC, 2006.
- 2. Gellman, S. H. Acc. Chem. Res. 1998, 31, 173-180.
- (a) Seebach, D.; Matthews, J. L. Chem. Commun. 1997, 2015–2022; (b) Seebach,
 D.; Beck, A. K.; Bierbaum, D. J. Chem. Biodiversity 2004, 1, 1111–1239; (c) Seebach, D.; Gardiner, J. Acc. Chem. Res. ASAP.
- (a) Appella, D. H.; Christianson, L. A.; Karle, I. L.; Powell, D. R.; Gellman, S. H. J. Am. Chem. Soc. **1996**, *118*, 13071–13072; (b) Huck, B. R.; Fisk, J. D.; Guzei, I. A.; Carlson, H. A.; Gellman, S. H. J. Am. Chem. Soc. **2003**, *125*, 9035–9037; (c) Horne, W. S.; Price, J. L.; Keck, J. L.; Gellman, S. H. J. Am. Chem. Soc. **2007**, *129*, 4178– 4180.
- For recent examples see: Lukaszuk, A.; Demaegdt, H.; Szemenyei, E.; Tóth, G.; Tymecka, D.; Misicka, A.; Karoyan, Ph.; Vanderheyden, P.; Vauquelin, G.; Tourwé, D. J. Med. Chem. 2008, 51, 2291-2296; Petersson, E. J.; Schepartz, A. J. Am. Chem. Soc. 2008, 130, 821–823; Schmitt, M. A.; Weisblum, B.; Gellman, S. H. J. Am. Chem. Soc. 2007, 129, 417–428; Sadowsky, J. D.; Fairlie, W. D.; Hadley, E. B.; Lee, H-S.; Umezawa, N.; Nikolovska-Coleska, Z.; Wang, S.; Huang, D. C. S.; Tomita, Y.; Gellman, S. H. J. Am. Chem. Soc. 2007, 129, 139–154; Kritzer, J. A.; Stephens, O. M.; Guarracino, D. A.; Reznik, S. K.; Schepartz, A. Bioorg. Med. Chem. 2005, 13, 11–16; Stephens, O. M.; Kim, S.; Welch, B. D.; Hodsdon, M. E.; Kay, M. S.; Schepartz, A. J. Am. Chem. Soc. 2005, 127, 13126–13127.
- (a) Hintermann, T.; Seebach, D. Synlett **1997**, 437–438; (b) Seebach, D.; Gademann, K.; Schreiber, J. V.; Matthews, J. L.; Hintermann, T.; Jaun, B.; Oberer, L.; Hommel, U.; Widmer, H. Helv. Chim. Acta **1997**, 80, 2033–2038; (c) Micuch, P.; Seebach, D. Helv. Chim. Acta **2002**, 85, 1567–1577.
- See, for example: (a) Steer, D. L.; Lew, R. A.; Perlmutter, P.; Smith, A. I.; Aguilar, M.-I. *Curr. Med. Chem.* **2002**, 9, 811–822; See also: (b) Zubrzak, P.; Williams, H.; Coast, G. M.; Isaac, R. E.; Reyes-Rangel, G.; Juaristi, E.; Zabrockiy, J.; Nachman, R. J. *Biopolymers, Peptide Science* **2007**, 88, 76–82.
- See, for example: Ellmerer-Müller, E. P.; Brössner, D.; Maslouh, N.; Tako, A. Helv. Chim. Acta 1998, 81, 59–65. and references cited therein.
- For examples of the synthesis of β²-amino acids bearing aliphatic side chains see: Juaristi, E.; Quintana, D.; Lamatsch, B.; Seebach, D. J. Org. Chem. 1991, 56, 2553–2557; Juaristi, E.; Quintana, D.; Balderas, M.; García-Pérez, E. Tetrahedron: Asymmetry 1996, 7, 2233–2246; Seebach, D.; Boog, A.; Schweizer, W. B. Eur. J. Org. Chem. 1999, 335–360; Ponsinet, R.; Chassaing, G.; Vaissemann, J.; Lavielle, S. Eur. J. Org. Chem. 2000, 83–90; Gutiérrez-García, V. M.; Reyes-Rangel, G.; López-Ruiz, H.; Juaristi, E. Tetrahedron 2001, 57, 6487–6496; Nagula, G.; Huber, V. J.; Lum, C.; Goodman, B. A. Org. Lett. 2000, 2, 3527–3529; Bedow, J. E.; Davies, S. G.; Smith, A. D.; Russel, A. J. J. Chem. Soc., Chem. Commun. 2004, 2778–2779; Rimkus, A.; Sewald, N. Org. Lett. 2003, 5, 79–80; Eilitz, U.; Lessmann, F.; Seidelmann, O.; Wendisch, V. Tetrahedron: Asymmetry 2003, 14, 189–191; Dursma, A.; Minard, A. J.; Feringa, B. L. J. Am. Chem. Soc. 2003, 125, 3700–3701; Bower, J. F.; Williams, J. M. J. Synlett 1996, 685–686; Bower, J. F.; Junnah, R.; Williams, J. M. J. Chem. Soc., Chem. 1420; Davies, H. M. L.; Venkataramani, C. Angew. Chem., Int. Ed. 2002, 41, 2197–2199; Sibi, M. P.; Patil, K. Angew. Chem., Int. Ed. 2004, 43, 1235–1238.
- See, for example: Gessier, F.; Schaffer, L.; Kimmerlin, T.; Flögel, O.; Seebach, D. Helv. Chim. Acta 2005, 88, 2235–2249.
- (a) Anaya de Parrodi, C.; Juaristi, E.; Quintero, L.; Clara-Sosa, A. Tetrahedron: Asymmetry **1997**, 8, 1075–1082; For reviews on the use of (R) and (S)-αphenylethylamine in the preparation of enantiopure compounds, see: (b) Juaristi, E.; Escalante, J.; León-Romo, J. L.; Reyes, A. Tetrahedron: Asymmetry **1998**, 9, 715–740; (c) Juaristi, E.; León-Romo, J. L.; Reyes, A.; Escalante, J. Tetrahedron: Asymmetry **1999**, *10*, 2441–2495.
- Anaya de Parrodi, C.; Clara-Sosa, A.; Quintero, L.; Pérez, L.; Marañón, V.; Toscano, R. A.; Aviña, J. A.; Rojas-Lima, S.; Juaristi, E. *Tetrahedron: Asymmetry* 2001, 12, 69–79.
- Clara-Sosa, A.; Pérez, L.; Sánchez, M.; Melgar-Fernández, R.; Juaristi, E.; Quintero, L.; Anaya de Parrodi, C. *Tetrahedron* 2004, 60, 12147–12152.
- Reyes-Rangel, G.; Marañón, V.; Avila-Ortiz, C. G.; Anaya de Parrodi, C.; Quintero, L.; Juaristi, E. Tetrahedron 2006, 62, 8404–8409.
- 15. Gage, J. R.; Evans, D. A. Org. Synth. Collective Vol. **1993**, 8, 339–343. 528–531.
- Evans, D. A.; Britton, T. C.; Ellman, J. A.; Dorow, R. L. J. Am. Chem. Soc. 1990, 112, 4011–4030.
- (a) Evans, D. A.; Bartroli, J.; Shih, T. L. J. Am. Chem. Soc. **1981**, 103, 2127–2129;
 (b) Evans, D. A.; Takacs, J. M.; McGee, L. R.; Ennis, M.; Mathre, D. J. B. Pure Appl. Chem. **1981**, 53, 1109–1127; (c) Evans, D. A. Aldrichim. Acta **1982**, 15, 23–32; (d) Evans, D. A.; Nelson, J. V.; Taber, T. R. Top. Stereochem. **1982**, 13, 1–115.

18. The X-ray structure was solved and refined using SHEIX-97, Sheldrick, G. M. Programs for Crystal Structure Analysis; University of Göttingen: Germany, 1997; within whick program version 1.64.05.28, Farrugia, L. J. *J. Appl. Crystallogr.* **1999**, 32, 837–838. Crystal data for (4R,5R)-3-[2(S)-([[(benzyloxy)carbony]]-amino]methyl)-3-methyl-butanoyl]hexahydrobenzoxazo-lidin-2-one, (*R,R,S*)-**9a**: C₂₁H₂₈N₂O₅, *M*_w = 388.45, monoclinic, *P* 21, *a* = 12. 020 Å, *b* = 6.001 Å, *c* = 13.838 Å, α = 90.0°, β = 96.69°, γ = 90.0°, *V* = 2132.47 Å³, crystal size: 0.12 × 0.15 × 0.20 mm³, *R*₁ = 0.0490 (*wR*₂ = 0.1005). Atomic coordinates for the structure reported in this paper have been deposited with the Cambridge

Crystallographic Data Centre. The coordinates can be obtained, on request from the Director, Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 IEZ, UK (fax: +44 1223 336 036. e-mail: deposit@ccdc.cam. ac.uk); Deposition number: CCDC 704760.

- 19. Juaristi, E.; Martínez-Richa, A.; García-Rivera, A.; Sánchez-Cruz, J. S. J. Org. Chem. **1983**, 48, 2603–2606.
- 20. Kim, H.; Jin, Y. Synlett 1998, 1189-1192.
- 21. Guichard, G.; Abele, S.; Seebach, D. Helv. Chim. Acta 1998, 81, 187-206.
- 22. Gellman, S. H.; Kim, B. M.; Park, J. S.; Lee, H. S. J. Org. Chem. 2003, 68, 1575–1578.